

A novel approach to glioma therapy using an oncolytic adenovirus with two specific promoters

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Abstract. Gliomas are the most common type of primary brain tumor in adults, where more than half of the cases are malignant, and the prognosis is poor. The early viral 1A (E1A) protein has been widely recognized to be essential for adenoviral replication and production of progeny virions in human cells, a process that is regulated by human telomerase reverse transcriptase. The p53 gene, as a tumor suppressor, regulates diverse cellular processes, including cell cycle arrest, cell autophagy, senescence and apoptosis. Dysfunction of the p53 pathways is common in malignant gliomas. Exogenous expression of p53 during adenovirus replication in human cancer cells may accelerate cell death and improve the release of early virus progeny. In the present study, a conditionally replicative adenovirus (CRAd) Ad-Tp-E1A-Gp-p53, which expressed functional p53 protein when replicating in cancer cells, was constructed. Next, the level of p53 expression in U251 cells was determined by western blot analysis, and the inhibitory effect of Ad-Tp-E1A-Gp-p53 on U251 cells was detected via an MTT assay. The results indicated that p53 expression was upregulated with an increase in the multiplicity of infection (MOI) of Ad-Tp-E1A-Gp-p53. Additionally, the inhibitory effects of Ad-Tp-E1A-Gp-p53 in different groups were significantly different ($P<0.05$), with the inhibition ratio of the experimental groups being higher, compared with the control group ($P<0.05$). Furthermore, the inhibition ratio increased with increases in the MOI of Ad-Tp-E1A-Gp-p53. Therefore, the expression of functional p53 and that of E1A may increase

the potency of CRAd, and overexpression of p53 through CRAd is a promising approach to more effective treatments in a number of human cancer types.

Introduction

Approximately 250,000 people worldwide are diagnosed with brain cancer annually. Unfortunately, only ~20% of all brain tumors are of primary origin at the time of diagnosis, while ~80% are metastatic (1). Gliomas are the most common primary brain tumor in adults and more than half are malignant (2). The current therapy for gliomas is multimodal, including surgical resection, radiotherapy and chemotherapy. However, due to the invasive properties of the cells, these treatment options are not always effective. Furthermore, the prognosis of patients with glioblastoma is poor and of invariable recurrence, with a median survival time of slightly >1 year (3,4). In the search for alternative treatment modalities, oncolytic viruses have recently received increasingly widespread attention for their potential in novel cancer therapy (5,6). Oncolytic viruses are effective for tumor therapy as they have the capacity to selectively replicate in tumor cells, to spread viral progeny in tumor tissues and to eventually induce tumor cell apoptosis (7). In this respect, conditionally replicative adenoviruses (CRAds) appear to be attractive anticancer agents and are currently being evaluated in clinical trials (8). The replication of CRAds occurs only in tumor cells and not in normal cells, which reflects the safety of this agent.

Conditionally replicative adenoviruses are designed to exert intrinsic anticancer activity through selective replication in cancer cells and to induce lysis in order to destroy these cells (9). Oncolytic viruses have been developed by adding specific promoter elements and therapeutic genes, including the human telomerase reverse transcriptase (hTERT) gene to control gene expression essential for adenoviral replication, and the p53 gene to promote the dissolution of tumor cells (7,10). In addition, the release of viral progeny from infected tumor cells offers a potential to amplify the oncolytic effects of CRAds by lateral spread in a solid tumor (6,9).

Self-replicated adenoviruses have previously been demonstrated to affect the progression of cancer cells *in vitro* by expressing p53. Promoters have been used previously to increase adenoviral replication or p53 expression, but the use of two promoters at same time is rare (6,10). In the present

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Abbreviations: E1A, early viral 1A; CRAd, conditionally replicative adenovirus; hTERT, human telomerase reverse transcriptase; GFAP, glial fibrillary acidic protein; CPE, cytopathic effect; SDS, sodium dodecyl sulfate

Key words: human telomerase reverse transcriptase, glial fibrillary acidic protein, early viral 1A, p53, conditionally replicative adenovirus

study, to enhance the oncolysis of CRAAd on tumor cells, a P74-Tp-Gp53 plasmid, containing the p53 gene, the early region 1A (E1A) gene and two tumor-specific promoters hTERT and glial fibrillary acidic protein (GFAP), was constructed. The plasmid p74-Tp-Gp53 was restructured with the plasmid of an adenovirus skeleton, PPE3, into the recombinant oncolytic adenovirus, Ad-Tp-E1A-Gp-p53, to selectively replicate in tumor cells and to restrain cell growth by expressing the p53 gene. Therefore, this approach may serve as a promising therapeutic agent for the treatment of numerous types of cancer.

Materials and methods

Cell culture. The human glioma U251 and T98G cell lines, and the human embryonic lung MRC-5 cell line as well as the 293 cell line were cultured in Dulbecco's modified Eagle's medium (DMEM) plus GlutaMAX™ (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA), supplemented with 10% heat-inactivated fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc.), 100 U/ml penicillin and 100 g/ml streptomycin at 37°C in 5% CO₂.

Plasmid construction. hTERT and GFAP were designed according to the sequences published in GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>; accession numbers, NM_198253 and M67446, respectively), and were synthesized by Generay Biotech Co., Ltd. (Shanghai, China). Next, restriction sites, *Bgl*III (5'-AGATCT-3') and *Hind*III (5'-AAGCTT-3'), were added upstream and downstream of hTERT. Subsequently, hTERT and GFAP were cloned into the Dual-Luciferase® Reporter vector, pGL3-Basic (empty vector; Promega Corporation, Madison, WI, USA), resulting in the plasmids pGL3-hTERTp and pGL3-GFAP, respectively. Using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), plasmids were transfected into the MRC-5 (11), U251 and T98 G cells for 2 h. A total of 24 h after transfection, the activity of the promoters was detected using the Dual-Luciferase reporter gene detection system (Invitrogen; Thermo Fisher Scientific, Inc.). Cells in the normal control group remained intact and pRL-TK (Promega Corporation) was used as a control reporter vector, and was used in combination with any experimental reporter vector to co-transfect mammalian cells. Renilla promoters (Promega Corporation) were co-transfected as an internal control. Firefly luciferase activity was normalized to renilla luciferase activity for individual analysis.

To enhance the activity of pGL3-GFAP, four optimized HIF-binding site ophBS enhancer sites (5'-TACGTGCAGTACGTGCAGTACGTGCAGTACGTGCAG-3') were cloned (Invitrogen; Thermo Fisher Scientific, Inc.) into *Mlu*I/*Xho*I sites of pGL3-GFAP, resulting in the plasmid pGL3-ENGfAP. pGL3-ENGfAP was then identified by excising with two different double enzyme systems (*Mlu*I/*Xho*I and *Pvu*I/*Pvu*II) and the excision product was detected by 2% agarose gel electrophoresis.

A 1,836 bp fragment was obtained from the double digestion of pGL3-ENGfAP with *Mlu*I/*Hind*III followed by *Eco*RI/*Xba*I and cloned into the *Eco*RI/*Xba*I sites of PCA19 to generate the plasmid PCA19-ENGfAP. Subsequently, two

different enzyme systems (*Eco*RI/*Xba*I and *Pst*I) and 2% agarose gel electrophoresis were used to verify the generation of PCA19-ENGfAP. Next, the early viral1A (E1A) gene was cloned into *Nco*I/*Sal*I sites of pGL3-hTERT, and the plasmid pGL3-hTERT-E1A (P74-Tp) was generated. The p53 gene was excised by double digestion of PENTER15-p53 (Invitrogen; Thermo Fisher Scientific, Inc.) with *Sal*I and *Eco*RI and was cloned into the *Sal*I and *Eco*RI sites in PCA19-ENGfAP, resulting in the plasmid PCA19-Gp53. The plasmid PCA19-Gp53 was excised by digestion with *Eco*RI, *Sal*I and *Pvu*II, and the excised product was detected by 2% agarose gel electrophoresis. Additionally, the 3,765 bp fragment excised from PCA19-Gp53 using *Bgl*II was cloned into the 8,465 bp fragment excised from pGL3-hTERT-E1A (P74-TP) using *Bgl*II, to yield the plasmid p74-Tp-Gp53. A different enzyme system (*Bam*HI and *Hind*III; Invitrogen; Thermo Fisher Scientific, Inc.) was used to identify the p74-Tp-Gp53 plasmid prior to detection of the excised fragments by 2% agarose gel electrophoresis.

Recombination of oncolytic adenovirus. The shuttle plasmid p74-Tp-Gp53 and the adenovirus vector PPE3 (both Invitrogen; Thermo Fisher Scientific, Inc.) were co-transfected into well-cultured 293 cells using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.) to complete the virus recombination. Following 9 days of incubation at 37°C, virus plaques were observable and were purified three times. Subsequently, 293 cells were alternately frozen and thawed to collect the restructured oncolytic adenovirus, Ad-Tp-E1A-Gp-p53. Polymerase chain reaction was then performed to determine the recombination of the Ad-Tp-E1A-Gp-p53 oncolytic adenovirus.

DNA extraction and PCR. Genomic DNA of the oncolytic adenovirus, Ad-Tp-E1A-Gp-p53, was extracted using a Blood Genome DNA Extraction kit (Takara Biotechnology Co., Inc., Dalian, China) according to the manufacturer's protocol and was stored at -80°C. To determine the recombination sites of the oncolytic adenovirus, PCR was performed with repeated three times using the following primers: GT154+GT156 forward, 5'-CCCACCGGTCACAGACGCCAGGAC-3' and reverse, 5'-GTGGCCGGGGCCAGGGCTTCCC-3'; W267+W268 forward, 5'-CCGGACGATATTGAACAATGGTTC-3' and reverse, 5'-GTGAAATATTCTCCATCCAGTGG-3'; and W331+W332 forward, 5'-CGACGCGTCCCTCTAGATACGTGCAGTACGTGCAGTACGTGCAGTACGTGCAGAA-3' and reverse, 5'-CCGCTCGAGTTCCCACACATCAGCCTGGAGAGAT-3' to amplify hTERT, p53 and GFAP, respectively. The PCR reaction was set at an initial denaturation step of 3 min at 95°C followed by 35 cycles of 95°C for 40 sec, 58°C for 40 sec, 72°C for 90 sec and, finally, 72°C for 10 min. The PCR products were subsequently detected by 2% agarose gel electrophoresis.

Detection of viral titer by 50% cell culture infection dose (TCID₅₀). 293 cells (permissive to viral infection) were seeded onto 96-well plates (10⁴ cells/well). After 24 h, eight serial 10-fold dilutions of freeze/thaw lysates from infected cells were seed into a 96-well plate (100 µl/well). After 10 days of incubation at 37°C, the observable cytopathic effect (CPE)

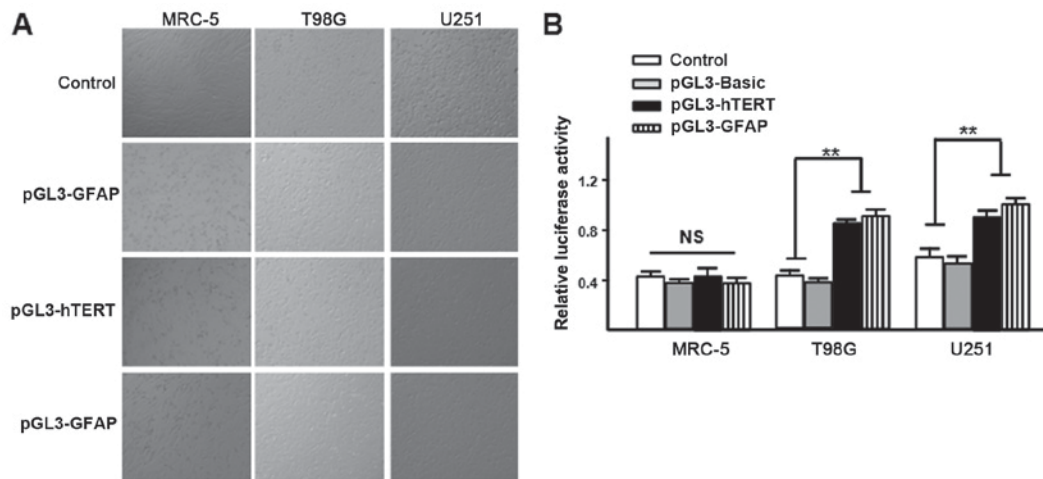


Figure 1. Activity of pGL3-hTERTp and pGL3-GFAP in different cells. (A) The plasmids (pGL3-hTERTp and pGL3-GFAP) were transfected into the cultured MRC-5 (11), U251 and T98 G cells (magnification, x100). (B) The relative luciferase activity of pGL3-hTERTp and pGL3-GFAP was detected using a dual-luciferase reporter assay. Data are presented as the mean \pm standard deviation (n=3). **P<0.01. NS, not significant. hTERT, human telomerase reverse transcriptase; GFAP, glial fibrillary acidic protein.

per dilution was counted, and the ratio between infected and unaffected wells was determined. The viral titer was then calculated using the following equation: $TCID_{50} = 10^{L+d(s-0.5)}$, where L is the log of the lowest dilution (e.g., If 10^{-1} is the minimum dilution degree, L=1), d is the dilution coefficient (e.g., If there is a 10-fold dilution, d=1), and s is the sum of the CPE rate observed at each dilution.

Western blot analysis. U251 cells were infected with Ad-Tp-E1A-Gp-p53 at different multiplicities of infection (MOIs; 0, 1, 10, 100 and 1,000). A bicinchoninic acid assay was used to determine protein concentrations. After 48 h of incubation, infected U251 cells were lysed using lysis buffer [0.125 M TRIS-HCl (pH 6.8), 2% SDS and proteinase inhibitor; Abcam, Cambridge, UK]. Total protein (20 μ g/lane) was then separated on a SDS polyacrylamide gel (12.5%) and blotted onto HyBond N membranes (EMD Millipore, Billerica, MA, USA). Following blocking with a 5% skimmed milk solution in TBS with 0.1% Tween-20 for 2 h at room temperature, adenoviral p53 expression was detected using the following primary antibodies: Anti-p53 [DO-1] (cat. no. ab1101; dilution, 1:1,000; Abcam) and anti-GAPDH [6C5] (cat. no. ab8245; dilution, 1:2,000; Abcam), and incubated for 12 h at 4°C. GAPDH (cat. no. ab8245; dilution, 1:2,000; Abcam) served as a loading control. Membranes were then incubated with a horseradish peroxidase-conjugated goat anti-mouse immunoglobulin G secondary antibody (cat. no. ab6789; dilution, 1:5,000; Abcam) for 2 h at room temperature. The antigen-antibody complexes were visualized using an enhanced chemiluminescence detection plus kit (cat. no. PE0010; EMD Millipore), according to the manufacturer's protocol.

Detection of the growth inhibiting effect of Ad-Tp-E1A-Gp-p53 on U251 cells by MTT. Tumor U251 cells were seeded onto 96-well plates (10^4 cells/well) and were infected with freeze/thaw lysates of infected cells at different MOIs (0, 1, 10, 100 and 1,000) following incubation for 24 h at 37°C (Control, 0 MOI; Group 1, 1 MOI; group 2, 10 MOI; group 3,

100 MOI; group 4, 1,000 MOI). After 2 h, the culture solution was discarded and 5% FBS/DMEM was added. After incubation for 72 h at 37°C, MTT (5 mg/ml) was added to the 96-well plates (20 μ l/well). After 4 h, 200 μ l dimethyl sulfoxide was added to the 96-well plates, and then the 96-well plates were agitated to dissolve the purple formazan. Subsequently, optical density (OD) values were measured at 490 nm. The cell growth inhibition rate was calculated using the following equation: Inhibition rate (%)=(control group OD value-experimental group OD value)/control group OD value.

Statistical analysis. All analyses were performed using SPSS software package (version 19.0; IBM Corp., Armonk, NY, USA). Statistical differences between all groups (experimental groups and the control group) were compared using one-way analysis of variance, followed by the Student-Newman-Keuls post hoc test. Data are expressed as the mean \pm standard deviation of three independent experiments. P<0.05 was considered to indicate a statistically significant difference.

Results

Activity of pGL3-hTERTp and pGL3-GFAP. The plasmids, pGL3-hTERTp and pGL3-GFAP, were transfected into the MRC-5, U251 and T98 G cells with pRL-TK (Invitrogen; Thermo Fisher Scientific, Inc.) at a ratio of 50:1, respectively. After 24 h of incubation, the activity was detected using a dual-luciferase reporter assay. It was observed that the hTERT promoter is active in U251 and T98G glioma cells, but it is not active in human MRC-5 cells, and the same finding was obtained for the GFAP promoter (Fig. 1). These results indicate that there is a good level of specificity of the hTERT and GFAP promoters to glioma cells.

Verification of pGL3-ENGFP, PCA19-ENGFP, PCA19-Gp53, P74-Tp-Gp53 plasmids and the recombinant adenovirus Ad-Tp-E1A-Gp-p53. The results of agarose gel electrophoresis revealed that two predicted fragments were

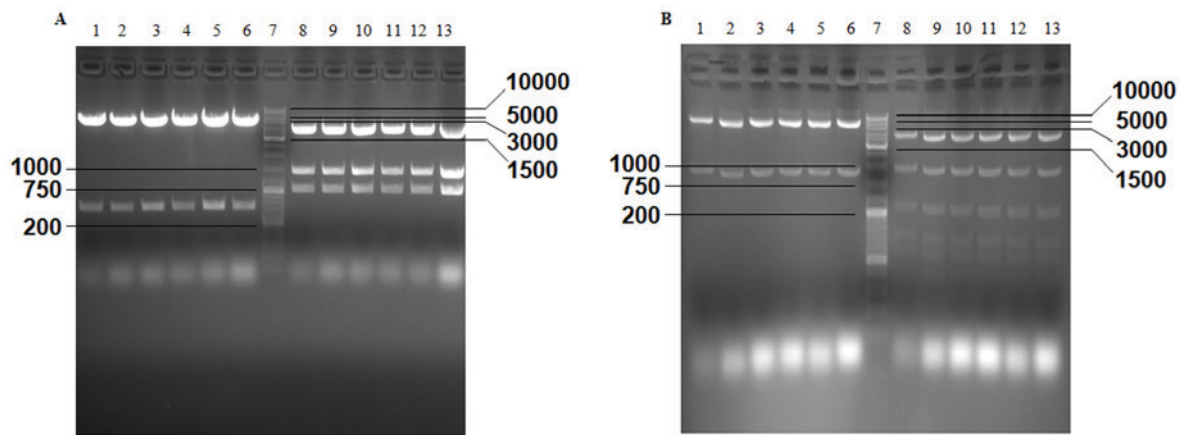


Figure 2. Verification of pGL3-ENGfap and PCA19-ENGfap plasmids by gel electrophoresis, according to the size (bp) of excised fragments. (A) Excised fragments of pGL3-ENGfap were detected by gel electrophoresis. Lanes 1-6, double digestion with *Mlu*I and *Xho*I; lane 7, GeneRuler™ DNA Ladder mix (10,000 bp); and lanes 8-13, double digestion with *Pvu*I and *Pvu*II. (B) Excised fragments of PCA19-ENGfap were detected by gel electrophoresis. Lanes 1-6, double digestion with *Eco*RI and *Xba*I; lane 7, GeneRuler™ DNA Ladder Mix; and lanes 8-13, digestion with *Pst*I. GFAP, glial fibrillary acidic protein.

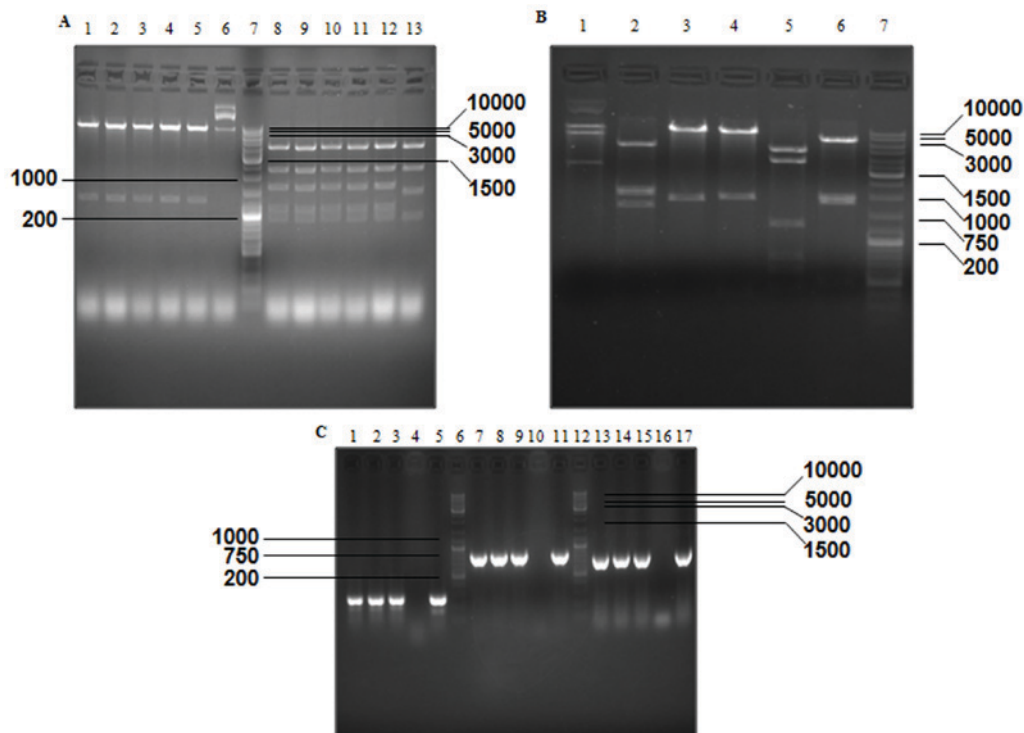


Figure 3. Verification of PCA19-Gp53 and p74-Tp-Gp53 plasmids by gel electrophoresis, according to the size (bp) of excised fragments. (A) Excised fragments of PCA19-Gp53 detected by gel electrophoresis. Lanes 1-6, double digestion with *Eco*RI and *Sal*I; lane 7, GeneRuler™ DNA Ladder mix (10,000 bp); lanes 8-13, digestion with *Pvu*II. (B) Excised fragments of P74-Tp-Gp53 detected by gel electrophoresis. Lanes 1-6, digestion of PCA19-Gp53 by *Bgl*II, *Sac*I and *Xba*I, *Nco*I and *Sal*I, *Bam*HI, *Pst*I and *Nco*I, respectively; and lane 7, GeneRuler™ DNA Ladder Mix (10,000 bp). (C) DNA fragments of Ad-Tp-E1A-Gp-p53 were detected by gel electrophoresis. Lanes 1-3, amplification of Ad-Tp-E1A-Gp-p53 using the GT154+GT156 primer; lanes 4, 10 and 16, negative control; lane 5, amplification of plasmid P74-TP using the GT154+GT156 primer; lanes 6 and 12, GeneRuler™ DNA Ladder Mix; lanes 7-9, amplification of Ad-Tp-E1A-Gp-p53 using the W267+W268 primer; lane 11, amplification of plasmid PENTER-p53 using the W267+W268 primer; lanes 13-15, amplification of Ad-Tp-E1A-Gp-p53 using the W331+W332 primer; and lane 17, amplification of plasmidpGL3-GFAP by using the W331+W332 primer. E1A, early viral 1A; GFAP, glial fibrillary acidic protein.

obtained from pGL3-ENGfap, which was double digested with *Mlu*I and *Xho*I, and three fragments were obtained from the double digestion of pGL3-ENGfap with *Pvu*I and *Pvu*II (Fig. 2A). Following double digestion of PCA19-ENGfap with *Eco*RI and *Xba*I and digestion with *Pst*I, two and seven predicted fragments were obtained (Fig. 2B). *Eco*RI+*Sal*I and *Pvu*II were used to identify PCA19-Gp53, and two and

seven predicted fragments were generated, respectively (Fig. 3A). As a result of the excision with *Bgl*II, *Sac*I+*Xba*I, *Nco*I+*Sal*I, *Bam*HI, *Pst*I and *Nco*I, different numbers of predicted fragments (2, 5, 6, 4, 7 and 5, respectively) were obtained from p74-Tp-Gp53 (Fig. 3B). Additionally, the results demonstrated that partial fragments of TERTp (size, 266 bp), p53 (size, 847 bp) and GFAP (size, 744 bp) were

successfully amplified from Ad-Tp-E1A-Gp-p53 using PCR (Fig. 3C).

Viral titer of p74-Tp-Gp53. Using the aforementioned equation for $TCID_{50}=10^{L+d(s-0.5)}$, a viral titer of 4.15×10^{11} PFU/ml Ad-Tp-E1A-Gp-p53 was calculated.

Expression of functional p53 in cancer cells infected with adenovirus. To investigate whether Ad-Tp-E1A-Gp-p53 affects the expression of p53, western blot analysis was performed and the levels of p53 expression in the Ad-Tp-E1A-Gp-p53-infected U251 cells at different MOIs were compared. As indicated in Fig. 4A, infection with Ad-Tp-E1A-Gp-p53 was able to upregulate p53 expression in U251 cells, and the level of p53 expression gradually increased with an increase in MOI.

Inhibitory effect of Ad-Tp-E1A-Gp-p53 on U251 cell growth. To evaluate the efficacy of viral transmission and the therapeutic potential of Ad-Tp-E1A-Gp-p53 *in vitro*, U251 cells derived from human brain gliomas were infected with Ad-Tp-E1A-Gp-p53 at different MOIs and the inhibition rate was calculated (Fig. 4B). The inhibitory effect of different MOIs of Ad-Tp-E1A-Gp-p53 was significantly different ($P<0.05$), in which the inhibition ratios of the experimental groups 1-4 were significantly higher compared with the control group ($P<0.05$). Additionally, the inhibition ratio of the four experimental groups exhibited an increasing trend as the MOI of Ad-Tp-E1A-Gp-p53 increased.

Discussion

In the present study, pGL3-hTERTp and pGL3-GFAP plasmids were successfully constructed and due to low activity, four optHBS enhancer sites were cloned into pGL3-GFAP. Additionally, a recombinant p74-Tp-Gp53 plasmid was constructed to specifically express E1A and p53 in tumor cells. Furthermore, Ad-Tp-E1A-Gp-p53 was demonstrated to suppress the growth of U251 cells through the expression of functional p53.

With the development of molecular biology and in-depth knowledge of viral gene function, it has become possible to genetically re-engineer viruses, and these may be used to selectively target tumor cells through the use of adenoviruses and herpesviruses (12,13). Conditionally replicative adenoviruses (CRADs) have recently presented as novel agents for cancer therapy and the use of these CRADs has been evaluated in preclinical trials (14). These evaluations have already demonstrated the potential of these adenoviruses in the therapy of malignant brain tumors. For example, the conditionally replicative adenovirus, ONYX-015, has entered into clinical trials for malignant glioma (6,15,16). The release and spread of conditionally replicative adenoviruses progeny depend on the replication efficiency of the adenovirus in cancer cells and their oncolytic capacity to induce death in cancer cells at the late stages of infection (9).

Since the E1A protein has been identified and widely recognized to be essential for adenoviral replication and the production of progeny virions in human cells, a number of telomerase promoter-regulated adenoviral vectors retain E1A genes (17). Additionally, the catalytic component of hTERT is

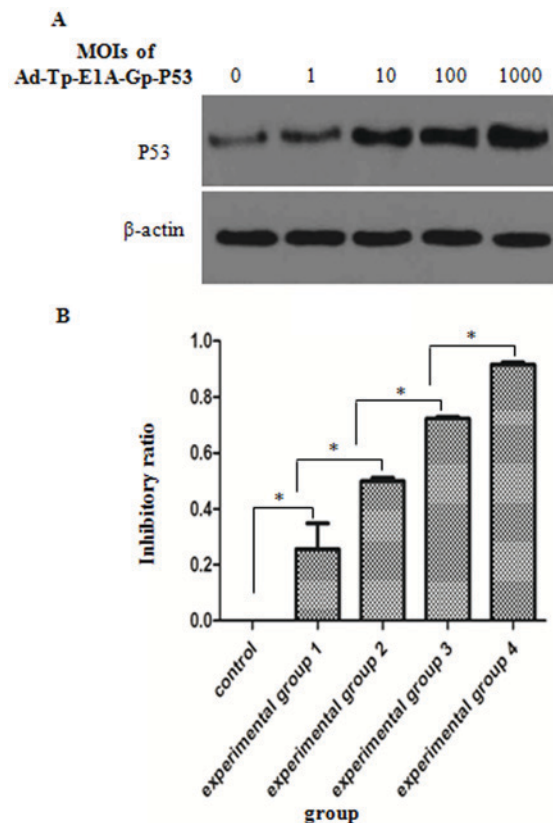


Figure 4. Adenovirus Ad-Tp-E1A-Gp-p53 suppresses the growth of U251 cells. (A) Quantitative analysis of p53 expression via western blot analysis. β -actin gene was used as an internal control. (B) Inhibitory effect of Ad-Tp-E1A-Gp-p53 on the growth of U251 cells as detected by MTT. Data are presented as the mean \pm standard deviation ($n=3$). * $P<0.05$. Group 1, 1 MOI; group 2, 10 MOI; group 3, 100 MOI; group 4, 1,000 MOI. MOI, multiplicity of infection. E1A, early viral 1A.

not expressed in the majority of primary somatic human cells, whereas major cancer cells are able to reactivate telomerase by transcriptional upregulation of hTERT (18). Furthermore, the hTERT gene, as a promoter, controls the gene expression that is essential for adenoviral replication (19). Therefore, hTERT is usually used to induce E1A expression and adenovirus replication in tumor cells. In the present study, hTERT was cloned together with the E1A gene into pGL3-Basic to induce E1A expression in tumor cells. GFAP, which was used in the present study as a promoter, has been previously demonstrated to be able to promote p53 expression and is downregulated in glioblastoma cells (20,21). GFAP and hTERT have been demonstrated to be glioma specific and are not expressed in MRC-5 cells (11), and these findings were confirmed in the present study.

The p53 gene, as a tumor suppressor, regulates diverse cellular processes, including cell cycle arrest, cell autophagy, senescence and apoptosis (22). In normal cells, the interaction between cellular proteins and encoded proteins, including cellular p53 proteins with viral E1A proteins, is necessary to complete the viral life cycle. However, when this interaction is dysregulated, normal cells gain the ability to develop into tumor cells (9,23).

The p53 suppressor protein is one component of the cell apoptosis pathway that is exploited by adenoviruses. Unfortunately, p53 is frequently inactivated by genetic

alterations in ~50% of all types of human cancer. For example, dysfunction of the p53 signaling pathway is common in malignant gliomas, which leads to a non-functional p53 pathway and inhibition of CRAd-induced cell death (6,24).

It was reported in a previous study that p53 expression was higher in high-grade brain glioma compared with low-grade brain glioma ($P < 0.05$), and the expression was not associated with patient sex or age, or the size of the tumor, demonstrating that p53 serves an important role in the occurrence and development of brain glioma (25). Therefore, restoring the function of wild-type p53 in p53-inactivated tumor cells is a potential treatment for certain types of tumor. In recent years, adenovirus-mediated p53 gene therapy has been rapidly developed as a promising antitumor strategy and has been employed in a number of preclinical experiments and clinical studies (22,25,26). In this regard, p53 is a common gene inserted in CRAd to control tumor cell growth and to cause cell apoptosis.

A number of experiments involving the use of p53 in CRAd have been performed. For example, Chen *et al* (26) demonstrated that p53-induced microRNA-107 inhibited brain tumor cell growth, which indicates that it serves as a tumor suppressor and thus, may be used as a target for glioma therapy. Additionally, Yang *et al* (27) suggested a combination of recombinant adenovirus-p53 (rAd-p53) with fractionated stereotactic radiotherapy (fSRT) is an effective and relatively safe method for the treatment of primary hepatocellular carcinoma (HCC) compared with fSRT monotherapy, indicating that rAd-p53 combined with fSRT may be preferred as a local method to treat primary HCC if the patients are unable to undergo surgery or refuse operation.

In the present study, a conditionally replicating adenovirus (Ad-Tp-E1A-Gp-p53) was constructed using the two plasmids pGL3-hTERT-E1A (p74-Tp) and PCA19-Gp53. Next, the expression level of functional p53 in adenovirus-infected U251 cells was detected by western blot analysis, and the results revealed that the expression of p53 was upregulated with an increasing MOI. Additionally, the inhibitory effect of Ad-Tp-E1A-Gp-p53 on U251 cells was detected by MTT and the results indicated that the inhibitory effects of Ad-Tp-E1A-Gp-p53 in different experimental groups (with different MOIs) were significantly different ($P < 0.05$), with the inhibition ratio of the experimental groups being higher compared with the control group ($P < 0.05$). Furthermore, the inhibition ratio increased with increases in MOI, indicating that the expression of functional p53 enhances the inhibitory effect of Ad-Tp-E1A-Gp-p53 on U251 cells.

In conclusion, in the present study, p74-Tp, PCA19-Gp53 and Ad-Tp-E1A-Gp-p53 were constructed. It was also demonstrated that hTERT and GFAP were able to promote the expression of their downstream genes *in vitro*. Furthermore, Ad-Tp-E1A-Gp-p53 was able to suppress the growth of U251 cells and the inhibitory effect increased with increasing MOIs of Ad-Tp-E1A-Gp-p53. Therefore, Ad-Tp-E1A-Gp-p53 may contribute to more effective treatment for different types of human cancer to enhance the potency of CRAd through the expression of functional p53. Although the present study demonstrated that the constructed CRAd represses the growth of glioma cells, there were certain limitations. The human MRC-5 cell line was used rather than normal

glial cells to verify glioma specificity of the constructed virus and thus, future experiments should use normal glial cells as a control to further verify the effectiveness of the constructed virus.

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