

Mutations in KRAS codon 12 predict poor survival in Chinese patients with metastatic colorectal cancer

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Abstract. *KRAS* mutations serve a function in tumorigenesis of colorectal cancer (CRC) and guide the use of targeted drugs. However, the prognostic value of *KRAS* mutations and their subtypes remain controversial. The present study aimed to investigate the correlations between *KRAS* mutations and clinicopathological characteristics, and their prognostic significance in Chinese patients with metastatic CRC (mCRC). A total of 135 patients with mCRC were analyzed for *KRAS* mutations. Mutations in codon 12 and 13 were identified in 45 (33.3%) patients. Only 3 patients harbored a mutation of V600E. Compared with male patients, *KRAS* codon 12 mutations were more common in female patients ($P<0.05$). *KRAS* codon 13 mutations tended to arise in right-sided compared with left-sided colon cancer ($P<0.05$). Survival analysis was performed in 101 patients receiving primary tumor resection. Compared with *KRAS* codon 12 wild-type, codon 12 mutations were markedly correlated with a poorer survival (log-rank $P=0.002$). No prognostic significance was revealed in codon 13 mutations. In univariate analysis, mortality risk was significantly increased by subtypes of G12D and G12V [hazard ratio (HR) = 2.313, 95% confidence interval (CI) = 1.069-5.004, $P=0.03$; HR = 2.621, 95% CI = 1.057-6.497, $P=0.04$, respectively]. The results of the present study suggested that codon 12 mutations, in particular G12D and G12V, predicted a negative prognosis in Chinese patients with mCRC. These findings require further confirmation via prospective studies with larger samples.

Introduction

Colorectal cancer (CRC) is one of the leading causes of cancer-associated mortalities in developed countries (1). In the past two decades, the morbidity and mortality of CRC have risen rapidly in Chinese population (2). A systemic therapy involving neoadjuvant therapy, surgical resection and postoperative therapy provides an opportunity for long-term survival. However, the majority of patients with metastatic CRC (mCRC) experience a poor survival rate (3,4). A growing body of research has demonstrated the positive effect of molecularly targeted treatment on the survival rate of patients with mCRC, especially with the use of monoclonal antibodies against epidermal growth factor receptor (EGFR) (5,6).

The tumorigenesis of CRC is a multistep process through the accumulation of genetic alterations. *KRAS* mutations are considered to be an early event in tumorigenesis (7). Proteins expressed by *KRAS* and *BRAF* genes are involved in the Ras/Raf/mitogen-activated protein kinase-extracellular-signal-regulated kinase kinase (MEK)/extracellular-signal-regulated kinase signaling pathway, which is a downstream pathway of EGFR. Mutations in *KRAS* and *BRAF* genes lead to the persistent activation of this pathway and accelerate the proliferation of tumor cells (8). It has been widely accepted that *KRAS* mutations predict the poor efficacy of anti-EGFR therapy in patients with mCRC (9). However, whether *KRAS* mutations are correlated with decreased survival in patients with mCRC remains controversial. Previous studies have indicated that *KRAS* mutations present statistically significant reductions in overall survival (OS) and disease-free survival (DFS) (10-12). Nevertheless, evidence of the association between *KRAS* mutations and poor OS in patients with mCRC is obtained primarily in Western countries and certain Asian countries; few data about prognostic significance of *KRAS* mutations in mCRC are available in Chinese patients (10-12).

Approximately 90% of *KRAS* mutations are located in codon 12 and 13 (13). Certain studies have demonstrated the diversity of biological characteristics in CRC with distinct *KRAS* mutational sites (14,15). An *in vitro* study revealed that tumor cells with codon 12 mutations possess an increased ability for cell transformation compared with those with codon

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13 mutations (16). However, in other studies, codon 13 mutations are considered to be more relevant to the poorer outcome than codon 12 mutations (17,18). Compared with codon 12 mutations, codon 13 mutation (G13D) exhibits increased efficacy of cetuximab treatment (15).

The results from previous studies did not reach a consensus and very few studies were performed to analyze *KRAS* mutation subtypes in Chinese patients with mCRC. Thus, the present study aimed to identify the frequency of *KRAS* and *BRAF* gene mutations in Chinese patients with mCRC, and investigate the prognostic value of distinct codon-specific *KRAS* mutations and their associations with clinicopathological characteristics.

Materials and methods

Study population. Based on the database of Sir Run Run Shaw Hospital (Hangzhou, China), a total of 580 patients were searched with histologically confirmed CRC and imaging confirmed metastasis between January 2010 and June 2016, among which 135 patients were tested for mutations in *KRAS* gene and 128 in *BRAF* gene. Characteristics of sex, age, body mass index (BMI), location of primary tumor, metastatic sites and the time to metastasis were collected. There were 135 patients, including 85 males and 50 females with the age range, 28-82. Survival analysis was performed in 101 patients between January 2010 and September 2015 who received curative resection for primary tumor. Adjuvant therapy included 5-fluorouracil folinic acid in combination with irinotecan (FOLFIRI) or oxaliplatin (FOLFOX); capecitabine in combination with oxaliplatin (XELOX) or capecitabine alone; with or without targeted drug (cetuximab or bevacizumab). A total of 30 patients received neoadjuvant therapy prior to surgery. All the therapies were based on the corresponding National Comprehensive Cancer Network guideline. The present study was approved by the Institutional Ethics Committee of Sir Run Run Shaw Hospital and informed consent was obtained from each participant.

DNA preparation and quantitative polymerase chain reaction. Tissue samples were fixed in 10% formalin at ambient temperature for 6 h. DNA was extracted from formalin-fixed paraffin-embedded samples of primary lesions or biopsy specimens. Genomic DNA was extracted with QIAamp® DNA FFPE tissue kit (Qiagen GmbH, Hilden, Germany) according to the manufacturer's instructions. AmoyDx® *KRAS* Mutation Detection kit and *BRAF* V600 Mutations Detection kit (both from Amoy Diagnostics Co., Ltd., Xiamen, China) were used to detect *KRAS* and *BRAF* status of each DNA sample according to the manufacturer's instructions. The quantitative polymerase chain reaction (qPCR) experiment was performed on Cobas z480 (Roche Molecular Diagnostics, Pleasanton, CA, USA) under the following three stages: one cycle at 95°C for 5 min, 15 cycles at 95°C for 25 sec and 64°C for 20 sec and 72°C for 20 sec, 26 cycles at 93°C for 25 sec and 60°C for 35 sec and 72°C for 20 sec. A result was considered mutation-positive if the Cq value was <30 with a classic S-curve (19). The aforementioned tests were performed in the Molecular Diagnostics Laboratory of Sir Run Run Shaw Hospital.

Statistical analysis. Data was analyzed with χ^2 test or Fisher's exact test to compare proportions. The Student's t-test was used

Table I. Frequency of *KRAS* and *BRAF* mutations in patients with metastatic colorectal cancer.

Amino acid	Case (total)	(%)
<i>KRAS</i>	45 (135)	33.3
G12A	2	
G12D	17	
G12V	12	
G12C	2	
G12S	1	
G13D	11	
<i>BRAF</i>		
V600E	3 (128)	2.3

to compare two groups of continuous data. Data are presented as mean values. The Kaplan-Meier method was performed for survival analysis and log rank test was used to compare the survival distributions. Furthermore, Cox's proportional hazards regression model was chosen to identify the impact of factors on OS. Hazard ratio (HR) was calculated with 95% confidence interval (CI). $P < 0.05$ was considered to indicate a statistically significant difference. All statistical analysis was performed with SPSS statistical software (version 21.0; IBM Corp., Armonk, NY, USA). Survival curves were plotted in Graph Pad Prism (version 6.0; GraphPad Software, Inc., La Jolla, CA, USA).

Results

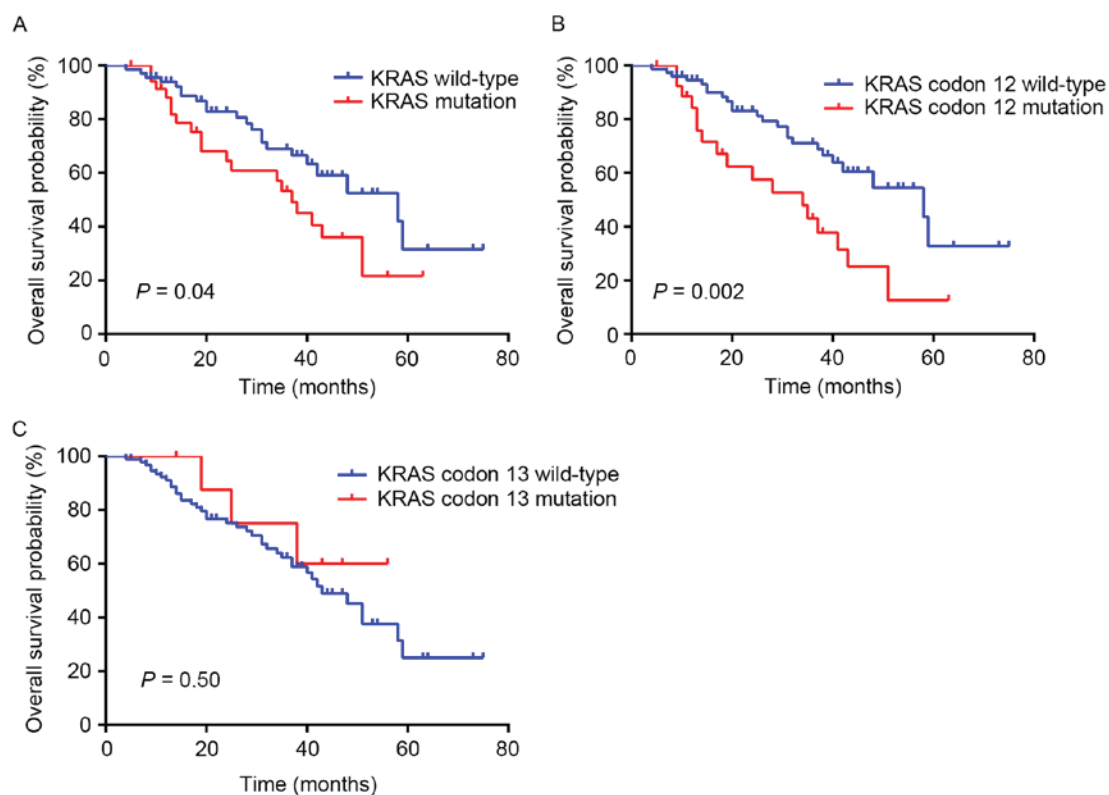
***KRAS* mutation characteristics.** Among the 135 patients with *KRAS* gene detection data, mutations in exon 2 were identified in 45 (33.3%) patients, of which 34 (68.9%) exhibited a single mutation in codon 12, and 11 (31.1%) exhibited a single mutation in codon 13. Codon 12 represented 5 mutational types, while codon 13 represented only 1 (G13D). The most frequently observed mutation was 35G>A (G12D), followed by 35G>T (G12V), 37.8 and 26.7%, respectively. The mutational types G12A, G12C and G12S comprised a small part of the mutations in these 2 codons. Of the 135 patients, *BRAF* codon 600 status was tested in 128 patients. Only 3 patients exhibited a mutation of V600E (3/128, 2.3%) and none of them possessed *KRAS* mutations simultaneously. Mutations are summarized in Table I.

Association between *KRAS* gene mutations and clinicopathological features. The association between clinicopathological features and *KRAS* codon 12, 13, and a 12/13 mutation status are presented in Table II. *KRAS* gene mutations and *KRAS* codon 12 mutations were significantly more common in female patients, compared with male patients ($P < 0.05$). Compared with left-sided colon, right-sided colon experienced a significantly increased number of *KRAS* codon 13 mutations ($P < 0.05$). No other significant associations were identified.

Prognostic value of *KRAS* codon 12 mutation. Among the 135 patients, 101 patients that received curative or palliative resection for CRC were included for survival analysis. In a

Table II. Clinicopathological features according to *KRAS* codon status in 135 patients with metastatic colorectal cancer.

Parameter	No.	KRAS codon 12			KRAS codon 13			KRAS codon 12 and 13		
		Wt	Mutation (n, %)	P-value	Wt	Mutation (n, %)	P-value	Wt	Mutation (n, %)	P-value
Sex				0.03 ^{a,b}			>0.09 ^c			0.04 ^{a,b}
Male	85	69	16 (18.8)		78	7 (8.2)		62	23 (27.1)	
Female	50	32	18 (36.0)		46	4 (8.0)		28	22 (44.0)	
Age (years) ^d	58.2	57.7	59.8	0.34 ^e	58.7	53.3	0.79 ^e	58.3	58.1	0.65 ^e
Body-mass index ^d	22.9	23.1	22.5	0.30 ^e	23.0	22.3	0.82 ^e	23.2	22.4	0.26 ^e
Location				0.44 ^b			0.04 ^c			0.51 ^b
Right-sided	43	34	9 (20.9)		36	7 (16.3)		27	16 (37.2)	
Left-sided	92	67	25 (27.2)		88	4 (4.3)		63	29 (31.5)	
Metastasis				0.81 ^b			0.74 ^b			1.00 ^b
Hepatic	42	32	10 (23.8)		38	4 (9.5)		28	14 (33.3)	
Extrahepatic	93	69	24 (25.8)		86	7 (7.5)		62	31 (33.3)	
Synchronous metastasis				0.71 ^b			0.12 ^b			0.58 ^b
Yes	100	74	26 (26.0)		94	6 (6.0)		68	32 (32.0)	
No	35	27	8 (22.9)		30	5 (14.3)		22	13 (37.1)	

^aP<0.05; ^b χ^2 test; ^cFisher's exact test; ^ddata presented as the mean; ^eStudent's t-test. Wt, wild-type.Figure 1. Kaplan-Meier curves of overall survival according to (A) *KRAS* mutation status, (B) *KRAS* codon 12 mutation status and (C) *KRAS* codon 13 mutation status in 101 patients.

Kaplan-Meier survival analysis of the *KRAS* gene status in the 101 patients, *KRAS* mutations were highly associated with a

poorer survival (log-rank P=0.04; median survival, 37 months in the *KRAS* mutant vs. 58 months in the *KRAS* wild-type;

Table III. Univariate analysis of overall survival for 101 patients.

Variable	Hazard ratio (95% confidence interval)	P-value
Sex		0.17
Male	Ref.	
Female	1.547 (0.832-2.875)	
Age (years)	1.012 (0.985-1.039)	0.39
Location		0.41
Right-sided	Ref.	
Left-sided	0.758 (0.393-1.462)	
Metastasis		0.64
Hepatic	Ref.	
Extrahepatic	1.194 (0.569-2.507)	
Synchronous metastasis		0.16
Yes	Ref.	
No	0.633 (0.336-1.195)	
Targeted drug		0.85
No	Ref.	
Yes	0.937 (0.479-1.834)	
Neoadjuvant therapy		0.78
No	Ref.	
Yes	1.101 (0.569-2.129)	
<i>KRAS</i> status		0.04 ^a
Wt	Ref.	
Mutant	1.884 (1.026-3.462)	
Codon 12 status		<0.01
Wt	Ref.	
Mutant	2.528 (1.369-4.668)	
Codon 13 status		0.49
Wt	Ref.	
Mutant	0.657 (0.202-2.135)	

^aP<0.05. Ref., reference; wt, wild-type.

Fig. 1A). In particular, patients with *KRAS* codon 12 wild-type experienced a median survival of 58 months, which was significantly increased, compared with patients with *KRAS* codon 12 mutation whose median survival was 34 months (log-rank $P=0.002$; Fig. 1B). The survival analysis indicated no difference between patients with and without *KRAS* codon 13 mutation ($P=0.50$; Fig. 1C).

Analysis of prognostic risk factors. Age, sex, the location of primary tumor and metastasis, the time of metastasis, the use of target drug and neoadjuvant therapy, and the *KRAS* gene status were analyzed with the Cox regression model (Table III). With the exception of *KRAS* codon 12 status, none of these factors exhibited a predictive value for poor prognosis. The patients with *KRAS* codon 12 mutations presented a significant decrease in overall survival (HR=2.528, 95% CI=1.369-4.668, $P=0.003$). In comparison, *KRAS* codon

13 mutants demonstrated no significant effect on survival (HR=0.657, 95% CI=0.202-2.135, $P=0.49$). Management of targeted and neoadjuvant therapy was not associated with risk of mortality ($P=0.85$; $P=0.78$, respectively). Further analysis revealed that rather than c.35G>C (G12A) or c.35G>T (G12C), c.35G>A (G12D) and c.35G>T (G12V) were associated with a significantly decreased OS compared with *KRAS* wild-type (HR=2.313, 95% CI=1.069-5.004, $P=0.03$; HR=2.621, 95% CI=1.057-6.497, $P=0.04$, respectively; Table IV).

Discussion

In the present study, the frequencies of *KRAS* and *BRAF* gene mutations were determined in Chinese patients with mCRC. The association between *KRAS* mutations with clinicopathological features was investigated. *KRAS* codon 12 mutations, especially G12D and G12V were revealed to exhibit predictive value for poor overall survival. To the best of our knowledge, studies concerning *KRAS* mutations have primarily been conducted only in the Western population (10,11). The impact of *KRAS* mutation, especially its different mutational sites, on the survival of Chinese population was uncertain and controversial.

The frequency of *KRAS* mutations (33.3%) and *BRAF* mutations (2.3%) in the present study were in accordance with a previous retrospective observational study that also involved Chinese patients with mCRC (34.8 and 3.4%, respectively) (20). However, the *KRAS* and *BRAF* mutation rate was slightly higher in the Western population (37.3 and 6%) (10). Since the morbidity and mortality of CRC in China were different from that in Western countries, the *KRAS* and *BRAF* mutation rates in different regions required further exploration (21).

The majority of *KRAS* mutations have been revealed to occur in codon 12 and 13 (3). The detection of associations between *KRAS* mutations and clinicopathological features had been performed widely; however, the data was lacking when codon 12 and 13 were considered separately (14,22,23). In the present study, compared with male patients, *KRAS* codon 12 mutations were more common in female patients compared with codon 13 mutations. Notably, while *KRAS* mutations (including codon 12 and 13) were not associated with the location of primary tumor, codon 13 mutation (G13D) itself was likely to occur in the tumors arising from right-sided colon. Recently, it was accepted that patients with right-sided colon cancer experienced an inferior OS rate (24). Additionally, increasing evidence suggests that mutations in *KRAS* codon 13 predict a poor outcome (12,18). In the present study, G13D mutation was associated with right-sided colon cancer; however, exhibited no prognostic significance, which may be explained by the small sample size. As for patterns of recurrence, Margonis *et al* (18) suggested that patients with *KRAS* codon 13 mutations, rather than codon 12 mutations possessed an increased risk of extrahepatic recurrence and lung-specific recurrence. Nevertheless, the results of the present study identified no difference in the patterns of tumor progression (including metastatic sites and time to recurrence) according to codon 12 or 13 mutation status.

The distribution of mutations in codon 12 and 13 varied in distinct studies. G12D in the present study was the most prominent subtype which was congruent with data from Caucasian and other Chinese mCRC (25-27). It had been reported that

Table IV. Analysis of overall survival according to *KRAS* codon 12 mutations by Cox regression analysis.

Nucleotide	Amino acid	Total number	Hazard ratio (95% confidence interval)	P-value
Wild-type		74	Ref.	
c.35G>A	G12D	13	2.313 (1.069-5.004)	0.03 ^a
c.35G>T	G12V	10	2.621 (1.057-6.497)	0.04 ^a
c.35G>C	G12A	2	3.308 (0.432-25.358)	0.25
c.34G>T	G12C	2	3.991 (0.928-17.165)	0.06 ^a

^aP<0.05.

different subtypes of *KRAS* mutations may confer variable tumor biology (28,29). A previous study involving surgically resected lung adenocarcinoma revealed that G12C mutation was associated with poorer outcome compared with other subtypes of mutations (28). Another study in Chinese patients with CRC demonstrated that *KRAS* codon 13 mutations, in particular G13D, were associated with significantly decreased OS rates (12). However, in the present study, *KRAS* codon 13 mutations had no effect on OS, but mutations in *KRAS* codon 12 were predictive for poor prognosis. This result was consistent with the evidence from *in vitro* studies, in which codon 12 mutations may increase aggressiveness due to increased transforming capacity and decreased levels of apoptosis (16,30). Previously, meta-analysis also revealed the predictive value that codon 12 mutations possessed by pooling the data from five randomized trials researching patients with mCRC in Western countries (10). In that meta-analysis, G12C was associated with inferior OS compared with *KRAS* wild-type; however, the results of the present study demonstrated that G12D and G12V were associated with decreased OS times. The prognostic value of codon and amino acid specific *KRAS* mutations has been discussed on several cancer types. For example, Bournet *et al* (31) reported that *KRAS* G12D mutant was an independent prognostic factor for unresectable pancreatic cancer. It had been proposed that in non-small cell lung cancer, the G12D mutation subtype was associated with the activation of the phosphoinositide 3-kinase/AKT serine/threonine kinase (AKT) and MEK signaling whereas mutation G12V or G12C preferred to activate Raf and Ral, and decreased growth factor-dependent AKT activation (32,33). These findings led to the present study in which the association between signaling pathways and the role of *KRAS* mutation subtypes on prognosis of patients with CRC were investigated, particularly in patients with mCRC from different regions.

BRAF V600E mutation has also been widely identified in CRC (34). It is considered to be associated with poor clinical outcome and may have confounding effect with *KRAS* mutations (35,36). *BRAF* V600E mutation was not included in the further analysis due to its extremely low mutation rate (2.3%).

There were several limitations to the present study, for example it was a retrospective study with a small sample size. Microsatellite instability and other family members of *RAS* gene that serve a function in CRC were not included. The therapeutic regimens varied between the patients, which may have resulted in the heterogeneity. However, no significant

difference was observed in the number of patients receiving neoadjuvant therapy and targeted therapy between the groups of codon 12 mutation, and codon 12 wild-type (P>0.05; data not shown). Additionally, the Cox regression model identified that whether receiving these two therapies or not was not a factor affecting the survival.

In conclusion, the results of the present study indicate that codon 12 mutations may predict poor OS in Chinese patients with mCRC, and further investigation demonstrated that G12D and G12V served as an indicator of poor prognosis in this specific population. In future clinical studies of CRC, the significance of gene mutation subtypes should be brought into consideration.

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