

Celecoxib enhances sensitivity to chemotherapy drugs of T-cell lymphoma

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Abstract. Celecoxib is a newly-identified nonsteroidal anti-inflammatory drug, which has been used to treat fever in clinical practice. Celecoxib has been demonstrated to suppress the viability of various human tumor cells. However, the effect of celecoxib on response of T-cell lymphoma to chemotherapy agents remains unclear. The aim of the present study was to investigate the effect of celecoxib on chemosensitivity of human T-cell lymphoma, and to address the underlying mechanism of action. The cytotoxicity of CDDP, epirubicin and VCR on Jurkat and Hut-78 cells treated with celecoxib was assessed by MTT assay, and the half-maximal inhibitory concentration (IC₅₀) value was calculated by Origin 75 software. The effect of celecoxib on apoptosis and intracellular concentration of Rhodamine-123 in Jurkat and Hut-78 cells was analyzed by flow cytometry. The expression of transcription factor p65 (p65), B-cell lymphoma 2 (Bcl-2), Bcl-2-associated X protein (Bax), multidrug resistance 1 (MDR1) and multidrug resistance-associated protein 1 (MRP1) at mRNA and protein levels were detected by reverse transcription quantitative polymerase chain reaction and western blotting, respectively. Proliferation suppression rates and apoptosis levels were significantly increased in Jurkat and Hut-78 cells combined with celecoxib compared with those without celecoxib, when treated with CDDP, epirubicin and VCR. The IC₅₀ values of the chemotherapy agents were lower in Jurkat and Hut-78 cells treated with celecoxib compared with those that were not.

The apoptosis level, expression of Bax and the intracellular concentration of Rhodamine-123 were increased, whereas the expression of p65, Bcl-2, MDR1 and MRP1 were decreased, in celecoxib-treated Jurkat and Hut-78 cells compared with those without celecoxib treatment. These results indicated that celecoxib may enhance the sensitivity of T-cell lymphoma to chemotherapy drugs by inhibiting the expression of multidrug resistance (MDR)-associated proteins via downregulating the activity of the nuclear factor- κ B signaling pathway, suggesting that celecoxib may improve the curative effect of chemotherapy drugs in T-cell lymphoma.

Introduction

T-cell lymphoma is the one of the most common immune subtypes of hematopoietic malignancy originating from lymphoid tissues, accounting for ~15% of all non-Hodgkin's lymphoma in China between 2000 and 2014 (1). At present, chemotherapy agents-based strategies remain the first choice for treatment of T-cell lymphoma, however poor prognosis is inevitable due to the occurrence of multidrug resistance (MDR) (2). The typical MDR phenotype is characterized by the overexpression of the MDR-associated proteins in the cytomembrane that serve as efflux pumps, to exclude the intracellular antitumor agents (3,4). As the prognosis of patients with T-cell lymphoma is greatly impaired by MDR, novel strategies to alleviate drug resistance and improve survival rates are urgently required.

Celecoxib is a newly-identified nonsteroidal anti-inflammatory drug, which is used to treat fever in clinical practice (4). Previously, celecoxib was demonstrated to induce apoptosis in multiple malignant tumor cells, but to have less toxicity and side effects on normal tissue cells (3). Celecoxib was identified to enhance the chemosensitivity and reduce the incidence of acquired MDR in human gastric, colon and breast carcinomas (4). Considering the function of celecoxib in reversing MDR in digestive and gynecologic tumors, we hypothesize that celecoxib may increase the chemosensitivity of T-cell lymphoma.

To the best of our knowledge, the present study revealed, for the first time, that celecoxib enhanced the inhibition effect of conventional chemotherapy drugs on T-cell lymphoma cell lines and significantly increased the percentages of apoptotic

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Abbreviations: CDDP, cis-diamminedichloroplatinum; VCR, vinblastine; IC₅₀, half-maximal inhibitory concentration; MDR, multidrug resistance

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cells. The half maximal inhibitory concentration (IC_{50}) of the representative chemotherapy agents cis-diamminedichloroplatinum (CDDP), epirubicin and vinblastine (VCR) was significantly decreased in T-cell lymphoma cells treated with celecoxib compared with those that were not. Additionally, the expression levels of MDR-associated proteins P-glycoprotein (P-gp), multidrug resistance-associated protein 1 (MRP1), transcription factor p65 (p65) and B-cell lymphoma 2 (Bcl-2) were decreased, whereas the expression level of Bcl-2-associated X protein (Bax) was increased in celecoxib-treated T-cell lymphoma cell lines compared with those that were not treated with celecoxib. By investigating the effect of celecoxib on T-cell lymphoma cells, it was identified that celecoxib may reduce drug resistance in these cells by inactivating the nuclear factor (NF)- κ B pathway. These data indicate that the combination of celecoxib and chemotherapy drugs may be an effective treatment strategy to improve the curative effect of chemotherapy drugs in T-cell lymphoma.

Materials and methods

Cell lines and reagents. The human T-cell lymphoma Jurkat and HuT-78 cell lines were supplied by the Research Center of the Fourth Hospital of Hebei Medical University (Shijiazhuang, China) and cultured in RPMI-1640 complete medium (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) containing 10% fetal calf serum (FCS; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) at 37°C in a humidified incubator with 5% CO_2 . The Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide (PI) double stain kit was supplied by BD Pharmingen; BD Biosciences (San Jose, CA, USA). Rabbit anti-human antibodies against p65 (cat. no. ,76311), P-gp (cat. no. ,168337) and MRP1 (cat. no. ,84320) were obtained from Abcam (Cambridge, MA, USA). Rabbit anti-human antibodies against Bcl-2 (cat. no. ,12789-1-AP) and Bax (cat. no. ,50599-2-Ig) were purchased from ProteinTech Group, Inc. (Chicago, IL, USA). Rabbit anti-human antibodies against β -actin (cat. no. ,AP0060) were obtained from Bioworld Technology, Inc. (St. Louis Park, MN, USA). The horseradish peroxidase (HRP)-conjugated goat anti-rabbit secondary antibody (cat no. 29527) was purchased from Rockland Immunochemicals Inc. (Limerick, PA, USA). TRIzol[®] reagent was obtained from Life Technologies (Thermo Fisher Scientific, Inc.). MTT, Rhodamine-123 and Ficoll separating medium (density: 1.077 g/ml) were purchased from Sigma-Aldrich; Merck KGaA. Go Taq[®] qPCR Master Mix was purchased from Promega Corporation (Madison, WI, USA). The RevertAid[™] First Strand cDNA Synthesis kit was purchased from MBI Fermentas; Thermo Fisher Scientific, Inc. CDDP was purchased from Qilu Pharmaceutical Co., Ltd. (cat no., EA4A7039A; Jinan, China). Epirubicin was purchased from Hisun Pharmaceutical Co., Ltd. (cat no., 17035211; Taizhou, China). VCR was purchased from Shenzhen Main Luck Pharmaceuticals Inc., (cat. no. ,1608V3; Shenzhen, China). Celecoxib was purchased from Pfizer, Inc. (New York, NY, USA; cat. no. ,S52338).

MTT assay. The effect of celecoxib on cell viability of Jurkat and Hut-78 cells was evaluated by the MTT assay. The Jurkat and Hut-78 cells were plated at a density of 1×10^5 cells/well in

200 μ l RPMI-1640 medium supplemented with 10% FCS and treated with celecoxib (20, 40 and 80 μ mol/l) in 96-well plates (Gibco; Thermo Fisher Scientific, Inc.). As a control, Jurkat and Hut-78 cells were not treated with celecoxib. Following incubation for 24, 48 and 72 h at 37°C in a humidified incubator, 10 μ l MTT solution [5 mg/ml in phosphate-buffered saline (PBS)] was added to the cells, which were then incubated again for 4 h at 37°C. Subsequently, 100 μ l DMSO was added to dissolve the formazan crystals. The absorbance at 492 nm was measured using a Titertek Multiskan[™] microplate reader (Flow Laboratories, North Ryde, Australia) to assess the effect of celecoxib on cell viability.

Lymphocyte cell viability assay. The peripheral blood was extracted from 3 cases of healthy volunteers without carcinomas, cardiovascular and endocrine system diseases, who underwent routine physical examination at the Fourth Hospital of Hebei Medical University (Shijiazhuang, China) between April 2016 and May 2016. The median age of the volunteers was 39 years (range, 32-44 years). T lymphocytes was separated from peripheral blood of healthy volunteers (2 females and 1 male) by density gradient centrifugation (240 x g for 20 min) at room temperature, using Ficoll separating medium (density: 1.077 g/ml). MTT assay was used to determine the cytotoxic effect of celecoxib on cell viability of T lymphocytes, as aforementioned. As a control, T lymphocytes were not treated with celecoxib. This research was approved by the ethic committee of the Fourth Hospital of Hebei Medical University and written informed consent was obtained from all participants.

Flow cytometry. The effect of celecoxib on the apoptosis level of Jurkat and Hut-78 cells was assessed by flow cytometry analysis. Jurkat and Hut-78 cells (10^6 /ml) treated with celecoxib at different doses (0, 20, 40 and 80 μ mol/l) were incubated with 5 μ l Annexin V-FITC and PI double stain at room temperature for 15 min. Following staining, cells were resuspended in ice-cold PBS and were analyzed using a flow cytometer (FACSCalibur[™]; BD Biosciences). Data were analyzed using CellQuest Pro version 5.1 software (BD Biosciences).

Western blotting analysis. Jurkat and Hut-78 cells treated with celecoxib at different doses (0, 20, 40 and 80 μ mol/l) were lysed with 500 μ l lysis buffer (1% Triton X-100, 150 mM NaCl, 10 mM Tris-HCl at pH 7.4, 1 mM EDTA, 1 mM EGTA at pH 8.0, 0.2 mM Na_3VO_4 , 0.2 mM phenylmethyl sulphonyl fluoride and 0.5% NP-40). The protein concentration in the lysates was determined by BCA assay. The lysates were then subjected to western blotting analysis to determine the amount of P-gp, MRP1, Bcl-2, Bax, p65 and β -actin protein in cells. Briefly, the total proteins (40 μ g per well) were subjected to 10% SDS-PAGE and were electro-transferred onto a polyvinylidene difluoride membrane. The membranes were blocked in PBS containing 5% FCS for 2 h at room temperature. Membranes were incubated with rabbit anti-human primary antibodies at different dilutions, including monoclonal antibodies to p65 (1:2,000), Bcl-2 (1:1,000), Bax (1:1,000), P-gp (1:1,000), MRP1 (1:1,000) and β -actin (1:5,000) for 8 h at 4°C. The PVDF membranes were visualized in the Odyssey infrared imaging system, after incubation with HRP-conjugated goat anti-rabbit

secondary antibodies (1:5,000) for 2 h at room temperature. The expression levels of these target proteins were calculated as the ratio of the intensity of the specified protein to that of β -actin, using Odyssey v3.0 software (LI-COR Biosciences, Lincoln, NE, USA).

RNA preparation and reverse transcription quantitative polymerase chain reaction (RT-qPCR) analysis. An RT-qPCR assay was used to evaluate the effect of celecoxib on expression of P65, Bcl-2, Bax, MDR1 and MRP1 mRNA in Jurkat and Hut-78 cells. Total RNA was extracted from Jurkat and Hut-78 cells treated with celecoxib at different doses (0, 20, 40 and 80 μ mol/l) using TRIzol reagent and the first strand cDNAs were generated from 1 μ g total RNA using RevertAid™ First Strand cDNA Synthesis kit and incubation at 42°C for 60 min. The resultant cDNAs were amplified by RT-qPCR using GoTaq® qPCR Master. The specific primers sequences (Shanghai Generay Biotech, Co., Ltd., Shanghai, China) and annealing temperature for the target genes are summarized in Table I. The following thermocycling conditions were used for the PCR: 95°C for 10 min; 40 cycles of 95°C for 15 sec, 60°C for 15 sec and 60°C for 1 min. The relative level of expression of each target gene was assessed by the $2^{-\Delta\Delta C_q}$ method (5).

Rhodamine-123 efflux assay. The rhodamine-123 efflux assay was performed to evaluate the efflux pump effect of P-gp and MRP1 in the Jurkat and Hut-78 cells treated with celecoxib at different doses (0, 20, 40 and 80 μ mol/l). The celecoxib-treated cells were co-cultured with Rhodamine-123 (5 mg/ml) for 1 h at room temperature and analyzed using a FACScan flow cytometer (FACSCalibur™, BD Biosciences) to analyze the intensity of fluorescence. Data were analyzed by CellQuest Pro version 5.1 software (BD Biosciences).

Statistical analysis. The data are reported as the mean \pm standard deviation. One-way analysis of variance was performed to determine the significance between groups. Turkey's method was used for multiple comparisons. $P < 0.05$ were considered to indicate a statistically significant difference. Experiments were repeated in triplicate. All statistical analyses were conducted using the SPSS software version 13.0 (SPSS, Inc., Chicago, IL, USA).

Results

Celecoxib may inhibit growth and enhance the sensitivity of T lymphocytes to chemotherapy drugs. Firstly, MTT assays were used to determine the inhibitory effect of celecoxib on the viability of T-cell lymphoma cells. As demonstrated in Fig. 1A, celecoxib significantly suppressed cell viability in Jurkat and Hut-78 cells in a time- and dose-dependent manner compared with the control group ($P < 0.05$), indicating that celecoxib may markedly inhibit the proliferation of T lymphoma cells. Whether celecoxib had an additive effect on the response to chemotherapy agents in T-cell lymphoma was additionally explored. As demonstrated in Fig. 1B, the viability of Jurkat and Hut-78 cells over 24 h was dose-dependently suppressed by CDDP, epirubicin and VCR, with significantly increased inhibition observed in Jurkat and Hut-78 cells treated with chemotherapeutic drugs combined with celecoxib compared

Table I. Primer sequences and annealing temperature for the reverse transcription-quantitative polymerase chain reaction.

Gene	Primer sequence	Annealing temperature, °C
β -actin	F: 5'-GTTGTGATGGGTTC TGA-3'	60
	R: 5'-GAGCAATAGCGTCT GTG-3'	
MRP1	F: 5'-CGCTGAGTTCCTGCGT ACC-3'	60
	R: 5'-TCTGCGGTGCTGTTG TGG-3'	
MDR1	F: 5'-CAGAGGGGATGGTCAG TGTT-3'	60
	R: 5'-CGTGGTGGCAAACAATA CAG-3'	
P65	F: 5'-GGCGAGAGGAGCACAG ATAC-3'	60
	R: 5'-ATCTTGAGCTCGGCAG TGTT-3'	
Bcl-2	F: 5'-GTGAACTGGGGGAGGA TTGT-3'	60
	R: 5'-GGAGAAATCAAACAG AGGCC-3'	
Bax	F: 5'-GGCCGGGTTGTCTGCCCT TTT-3'	60
	R: 5'-CCGCTCCCGGAGGAAG TCCA-3'	

F, forward; R, reverse; MRP1, multidrug resistance-associated protein 1; MDR1, multidrug resistance protein 1; P65, tumor protein 65; Bcl-2, B-cell lymphoma 2; Bax, Bcl-2-associated X protein.

with cells treated with chemotherapy drugs alone ($P < 0.05$). Additionally, the IC_{50} values of chemotherapy agents were significantly reduced in Jurkat and Hut-78 cells treated with celecoxib in a dose-dependent manner compared with those that were not treated with celecoxib ($P < 0.05$) (Fig. 1C). These results suggested that celecoxib may enhance the sensitivity of T-cell lymphoma cells to chemotherapy drugs. In contrast, the viability of normal T cells was not reduced following treatment with celecoxib at the different doses ($P > 0.05$) (Fig. 1D).

To additionally evaluate the facilitating effect of celecoxib on the chemosensitivity of T-cell lymphoma, flow cytometry analysis was performed to determine whether celecoxib induced apoptosis in Jurkat and Hut-78 cells. As demonstrated in Fig. 2, the apoptosis rate of Jurkat and Hut-78 cells treated with celecoxib was significantly increased compared with the cells in the control group ($P < 0.01$), indicating that celecoxib-induced proliferation inhibition of lymphoma cells may be due to a pro-apoptotic effect. Furthermore, when Jurkat and Hut-78 cells were treated with chemotherapy agents combined with celecoxib, the percentage of apoptotic cells

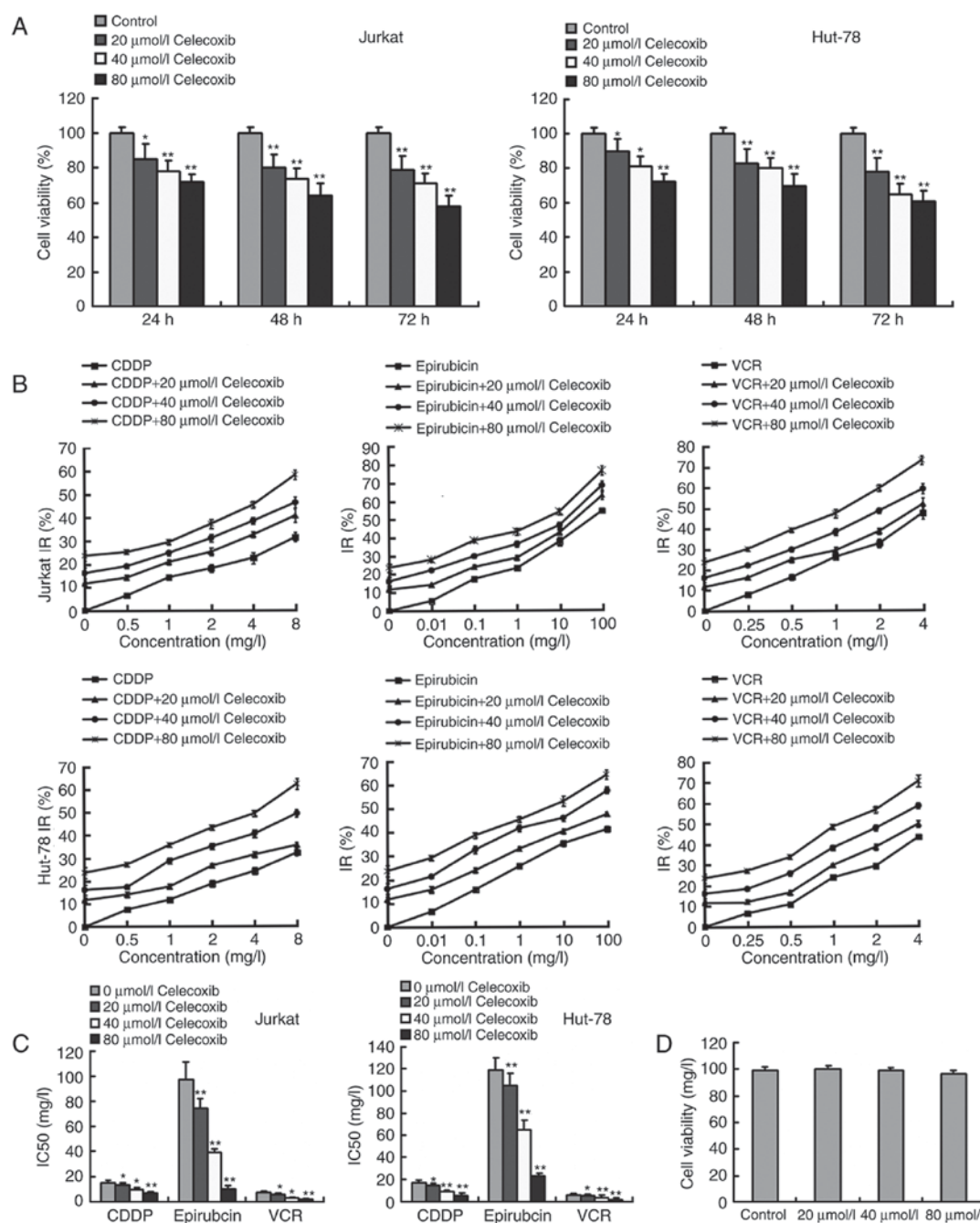


Figure 1. Ability of celecoxib to inhibit proliferation and enhance the chemosensitivity of Jurkat and Hut-78 cells to anticancer drugs. (A) The inhibition effect of celecoxib on Jurkat and Hut-78 cells for 24, 48 and 72 h was determined. (B) The chemotherapy drugs CDDP, epirubicin and VCR dose-dependently inhibited T-cell lymphoma cells proliferation over 24 h. Cells treated with celecoxib were more sensitive to chemotherapy drugs. (C) IC₅₀ values of CDDP, epirubicin and VCR were decreased in Jurkat and Hut-78 cells treated with celecoxib ($P < 0.05$). (D) Effect of celecoxib on T cells was determined. The viability of normal T cells (control) was not reduced following treatment with celecoxib compared with those without celecoxib treatment. Cell viability and IC₅₀ values are presented as the mean \pm standard deviation from three independent experiments. * $P < 0.05$ and ** $P < 0.01$ vs. the control group. CDDP, cis-diamminedichloroplatinum; VCR, vinblastine; IC₅₀, half maximal inhibitory concentration.

was also significantly increased compared with cells treated with chemotherapy agents alone. These data revealed that celecoxib may enhance cytotoxicity of chemotherapy drugs by the pro-apoptotic effect, additionally suggesting that celecoxib is a potential chemosensitizer for T-cell lymphoma treatment.

Celecoxib inhibits the expression of the MDR-associated proteins via the NF- κ B signaling pathway. To address the underlying mechanism that may be involved in the

celecoxib-mediated pro-apoptotic and chemosensitizing effect, the changes of MDR-associated proteins expression in T-cell lymphoma cells was analyzed. As P-gp, encoded by MDR1, and MRP1 are closely associated with the development of MDR, while Bcl-2 and Bax are important factors involved in the induction of apoptosis, their expression was evaluated in Jurkat and Hut-78 cells treated with celecoxib. The results indicated that the expression levels of Bcl-2, MDR1 and MRP1 mRNA and proteins were significantly downregulated,

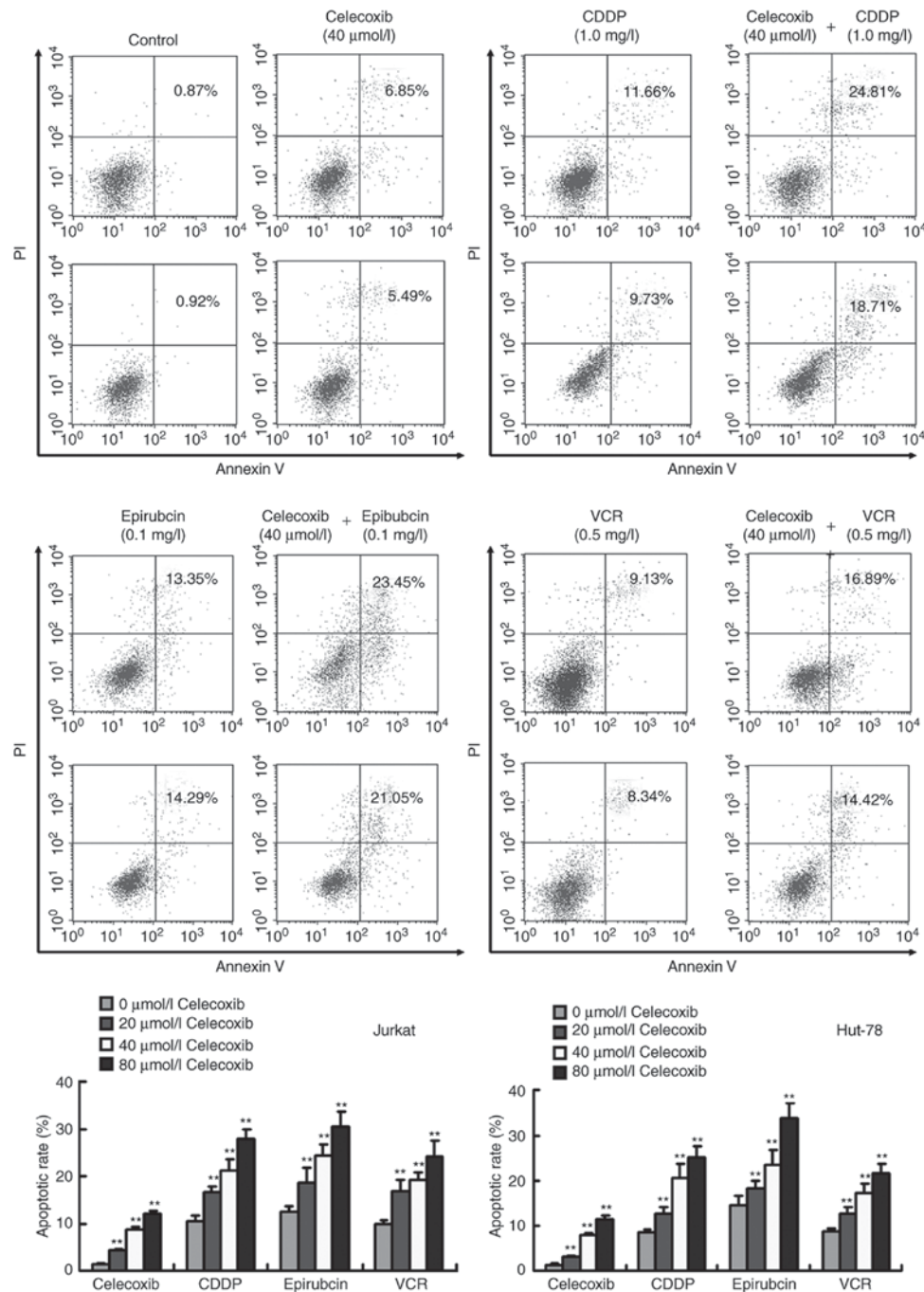


Figure 2. Induction of apoptosis by various chemotherapy drugs in T-cell lymphoma cells treated with or without celecoxib. The effect of celecoxib on the apoptotic rate of Jurkat and Hut-78 cells was detected by flow cytometry analysis. Celecoxib treatment significantly increased the percentage of Annexin V-positive cells compared with that in the control group, when the cells were not treated with chemotherapy agent. Additionally, when Jurkat and Hut-78 cells were treated with CDDP (0.5 mg/l), epirubicin (0.25 mg/l) or VCR (0.01 mg/l), the percentage of Annexin V-positive cells was also significantly higher in cells treated with celecoxib compared with those without celecoxib ($P < 0.01$). Similar results were obtained in three independent experiments. ** $P < 0.01$ vs. the control group. CDDP, cis-diamminedichloroplatinum; VCR, vinblastine; PI, propidium iodide.

whereas the expression of Bax was upregulated in tumor cells treated with celecoxib compared with those that were not treated with celecoxib ($P < 0.05$; Fig. 3A and B).

Furthermore, the effect of celecoxib on the expression of p65, which is involved in regulating cell apoptosis and inducing MDR, was assessed. As indicated in Fig. 3A and B, the expression of p65 was decreased significantly in celecoxib-treated Jurkat and Hut-78 cells. It is well-known that the NF- κ B P65 subunit participates in activating various downstream genes

associated with the regulation of apoptosis and induction of MDR (6-8). Several studies have revealed that Bcl-2 may suppress the activity of multiple pro-apoptotic proteins by binding with P65 (3,6). Additionally, previous studies also confirmed that the majority of the MDR-associated proteins, including P-gp and MRP1, are positively regulated by the NF- κ B pathway, and that blockade of the NF- κ B pathway may suppress their expression (6,7). Therefore, these results suggested that the celecoxib-mediated pro-apoptotic and

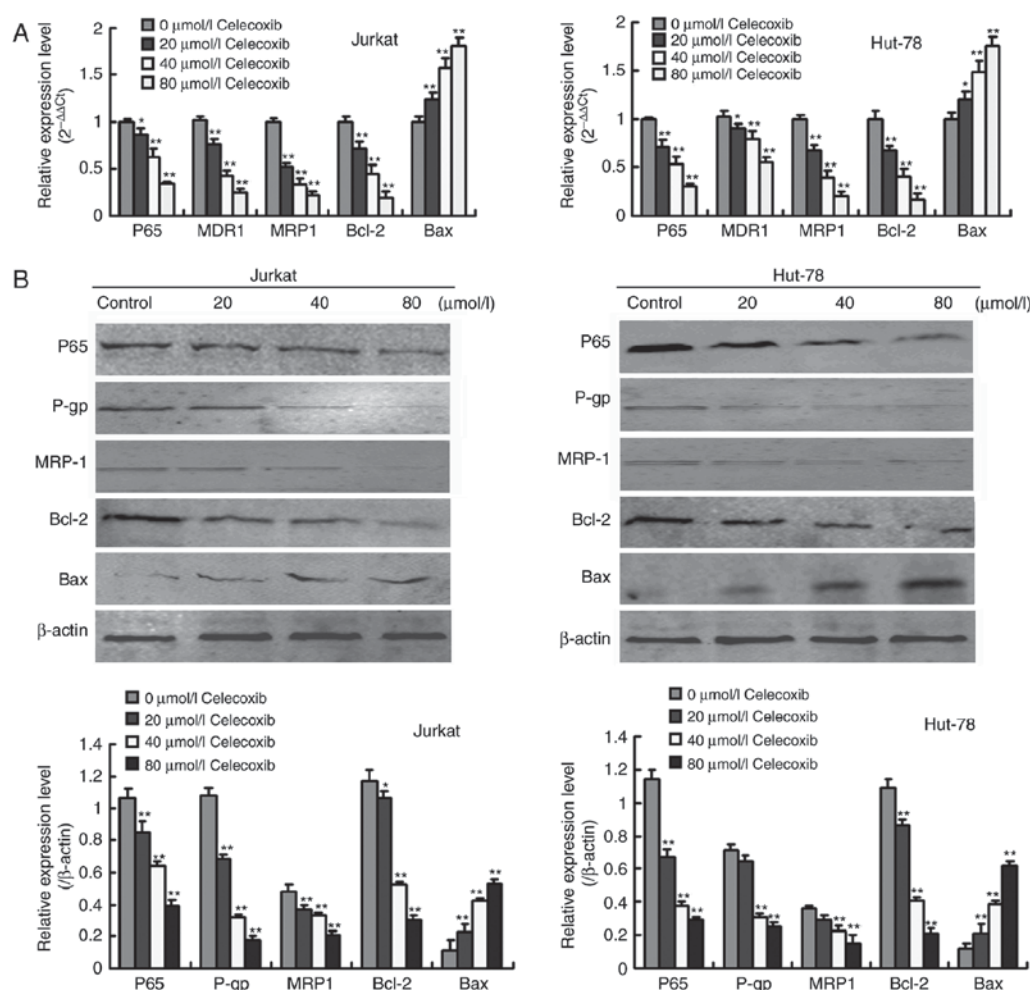


Figure 3. Effect of celecoxib on the expression of proteins associated with chemotherapy sensitivity in Jurkat and Hut-78 cells. (A) Reverse transcription quantitative polymerase chain reaction assays of the expression levels of P65, MDR1, MRP1, Bcl-2 and Bax mRNAs in Jurkat and Hut-78 cells. (B) Western blotting assays of the expression levels of P65, P-gp, encoded by MDR1, MRP1, Bcl-2 and Bax protein expression in Jurkat and Hut-78 cells. β -actin was used as an internal loading control. The experiments presented are representative of three independent experiments. * $P < 0.05$ and ** $P < 0.01$ vs. the control group. P65, tumor protein 65; P-gp, P-glycoprotein; MDR1, multidrug resistance protein 1; MRP1, multidrug resistance-associated protein 1; Bcl-2, B-cell lymphoma 2, Bax, Bcl-2-associated X protein.

chemosensitizing effect on T lymphocytes are associated with decreased expression of MDR-associated proteins Bcl-2, Bax, P-gp and MRP1 via inhibition of the NF- κ B pathway.

Celecoxib inhibits the efflux of intracellular Rhodamine-123 in Jurkat and Hut-78 cells. The effect of celecoxib on the exclusion of intracellular Rhodamine-123 by pump proteins was evaluated by a Rhodamine-123 efflux assay. The results of flow cytometry analysis indicated that the intracellular concentration of Rhodamine-123 was significantly increased in Jurkat and Hut-78 cells treated with celecoxib compared with those that were not treated with celecoxib ($P < 0.01$, Fig. 4), indicating that celecoxib may impair the efflux effect of the P-gp and MRP1 in Jurkat and Hut-78 cells, inhibiting the exclusion and promoting the intracellular accumulation of chemotherapy agents.

Discussion

At present, T-cell lymphoma has one of the highest mortality rates of all non-Hodgkin lymphoma; the average mortality

rate of the patients with T-cell lymphoma remains $>70\%$ (1). Whilst the traditional chemotherapy strategies have markedly improved in the previous 3 decades, MDR remains a great challenge in lymphoma treatment. It was suggested that the application of anticancer agents may induce the overexpression of various MDR-associated proteins and blockage of apoptosis in tumor cells (7). However, the exact mechanism that induces MDR has not been clarified at present. Strategies of avoiding MDR and improving sensitivity to chemotherapy remain the most critical problems in treatment of T-cell lymphoma.

Whilst celecoxib has been identified as an effective anti-tumor agent, characterized by inhibiting proliferation in a variety of tumor cells (8), little is known about its effect on chemosensitivity in T-cell lymphoma. Previous studies have revealed that celecoxib may prevent chemotherapy drug resistance in digestive and gynecologic carcinomas (9-12), but whether it serves a similar role in T-cell lymphoma has not been determined. The present study demonstrated that celecoxib sensitized Jurkat and Hut-78 cells to various chemotherapy drugs by triggering apoptosis and inhibiting the expression of MDR-associated proteins.

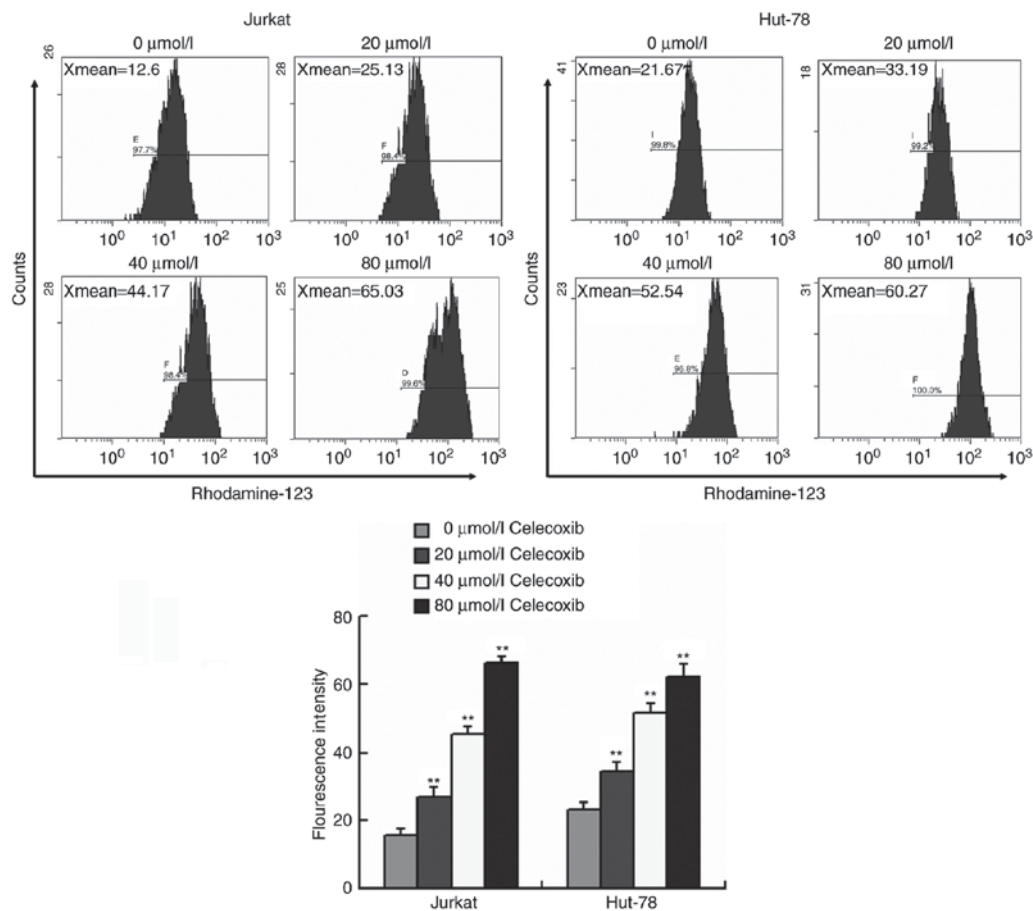


Figure 4. Effect of celecoxib on the intracellular accumulation of Rhodamine-123 in Jurkat and Hut-78 cells. Jurkat and Hut-78 cells were incubated with 5 mg/ml Rhodamine-123 for 1 h, and intracellular fluorescence was measured to assess the pump function of MDR-associated proteins. The fluorescence intensity indicated the intracellular concentration of Rhodamine-123, as demonstrated by the histogram. Data presented are the mean \pm standard deviation from three independent experiments. ** $P < 0.01$ vs. the control group.

Firstly, the inhibitory effect of celecoxib on T-cell lymphoma cells was investigated using an MTT assay. The results demonstrated that celecoxib significantly suppressed the viability of Jurkat and Hut-78 cells and facilitated the sensitivity of cells to conventional chemotherapy drugs, including CDDP, epirubicin and VCR. The IC_{50} value of the chemotherapy drugs was significantly decreased in celecoxib-treated cells compared with those that were not treated with celecoxib. Additionally, the results of the flow cytometry analysis demonstrated that celecoxib may induce apoptosis and enhanced cytotoxicity of the chemotherapy drugs in Jurkat and Hut-78 cells. These data demonstrated that the chemotherapy agents combined with celecoxib improved the efficacy of chemotherapy, suggesting that celecoxib may be a potential chemosensitizer in the clinical treatment of T-cell lymphoma. The present study may provide novel insight into the therapeutic effects of celecoxib, and assist in overcoming chemotherapy resistance in T-cell lymphoma.

To additionally explore the pro-apoptotic and chemosensitizing effect of celecoxib on T-cell lymphoma cells, the role of celecoxib in regulating the expression of MDR-associated proteins was investigated. It has been demonstrated previously that the Bcl2 family members protect against the activation of multiple signaling pathways leading to apoptosis of tumor cells, whereas the activation of Bax family members

will induce cell apoptosis, indicating that Bcl2/Bax axis played a crucial part in regulating the common apoptosis pathway (8-10). In the present study, the celecoxib-treated lymphoma cells exhibited significantly decreased expression of Bcl-2 and increased expression of Bax compared with those that were not treated with celecoxib, suggesting that effect on the Bcl-2/Bax protein ratio may be an important contributing factor to celecoxib-induced apoptosis. Additionally, it has been identified that the abnormal expression of P-gp and MRP1 on the surface of the cell membrane may impair tumor chemosensitivity and lead to MDR by removing chemotherapy drugs from tumor cells (13-16). Overexpression of P-gp and MRP1 in tumor tissues has been associated with poor response to clinical chemotherapy in T-cell lymphoma (15,17). The results of the present study also demonstrated that the expression of P-gp and MRP1 was significantly downregulated in Jurkat and Hut-78 cells treated with celecoxib compared with those that were not treated with celecoxib, indicating that celecoxib may inhibit the efflux of intracellular anticancer agents. In addition, the intracellular concentration of Rhodamine-123 in Jurkat and Hut-78 cells treated with celecoxib was significantly increased, indicating that celecoxib weakened the efflux function of P-gp and MRP1.

To elucidate the signaling pathway affected by celecoxib, the effect of celecoxib on the activity of the NF- κ B pathway

was evaluated. P65, the primary functioning subunit of activated NF- κ B, is an important indicator for predicting MDR in tumors, as it promotes cell viability, causes apoptosis resistance and enhances expression of various MDR-associated proteins including P-gp and MRP1 (7). Furthermore, previous studies have also confirmed that the expression of Bcl-2 may be directly upregulated by activation of the NF- κ B signaling pathway, thereby inhibiting Bax expression and leading to an anti-apoptotic effect in tumor cells (8-11). In the present study, the results suggested that the expression of P65, Bcl2, P-gp and MRP1 was significantly decreased, while the expression of Bax was increased in celecoxib-treated Jurkat and Hut-78 cells compared with those that were not treated with celecoxib. Therefore, P65 may be a key factor affected by celecoxib in regulating the expression of the apoptosis- and MDR-associated proteins.

To conclude, the present study identified that celecoxib may induce apoptosis and improve the sensitivity of T-cell lymphoma cells to chemotherapy by decreasing the expression of P-gp, MRP1 and Bcl-2/Bax ratio via the NF- κ B pathway, and by increasing the intracellular accumulation of anticancer drugs. Celecoxib may therefore be a potential candidate for improving the curative effect of chemotherapy drugs in T-cell lymphomas.

Acknowledgements

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