

Silencing of miR-19a-3p enhances osteosarcoma cells chemosensitivity by elevating the expression of tumor suppressor PTEN

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Abstract. MicroRNAs (miRNAs/miRs) are small non-coding RNAs, which serve important roles in tumor progression. The present study analyzed the role of miR-19a-3p in the chemosensitivity of osteosarcoma (OS) cells. Overexpression of miR-19a-3p was observed in OS cells and a cisplatin-resistant MG63 cell line was subsequently constructed. It was observed that miR-19a-3p inhibitor transfection suppressed cell proliferation and decreased the expression of Ki67 and PCNA compared with the cisplatin treatment group. miR-19a-3p inhibitor transfection also promoted apoptotic rate, increased the expression of Bcl-2 associated X, apoptosis regulator (Bax) and markedly decreased the expression of Bcl-2 compared with the cisplatin treatment group. These results elucidated that silencing of miR-19a-3p enhanced chemosensitivity of OS cells to Cisplatin, through suppressing cell proliferation and promoting cell apoptosis during treatment with Cisplatin. Bioinformatics study and luciferase reporter assays indicated that PTEN was a target of miR-19a-3p, and western blotting demonstrated that PTEN expression was negatively regulated by miR-19a-3p in OS cells. In addition, overexpression of PTEN decreased cell proliferation, but increased apoptotic rate compared with the cisplatin treatment group. It was observed that inhibition of PTEN by BpV(HOpic) upregulated cell proliferation and downregulated apoptotic rate compared with the Cisplatin-treated miR-19a-3p inhibitor group, indicating that inhibition of PTEN expression

counteracted the effect of the miR-19a-3p inhibitor on the regulation of chemosensitivity in OS cells. Taken together, overexpression of miR-19a-3p was observed in OS cell lines and that downregulation of miR-19a-3p enhanced the chemosensitivity of OS cells to Cisplatin, by elevating the expression of the tumor suppressor, PTEN.

Introduction

Osteosarcoma (OS) is one of the most commonly diagnosed types of malignant bone cancer in adolescents and young adults (1). The long-term disease-free survival of patients with OS remains low (~65%) and disease-free survival of patients exhibiting metastasis have a lower survival rate (2). Due to the improvement of chemotherapy drugs, the 5-year survival rate of OS patients has increased in recent decades (3). However, chemoresistance remains a major problem, and contributes to the low 5-year survival rate (4). Previous studies have indicated that various oncogenes and tumor suppressors are involved in the chemoresistance of OS cells (5,6). However, the existing understanding of the mechanism of chemoresistance in OS is limited and further investigation is urgently required.

MicroRNAs (miRNAs/miRs), a group of small non-coding RNAs that regulate gene expression by binding to the 3'-untranslated region (3'-UTR) of mRNAs, serve important roles in various cellular processes, including proliferation, apoptosis, invasion and differentiation (7,8). miRNAs are dysregulated in various types of cancer, acting as oncogenes or tumor suppressors during tumorigenesis (9). Evidence demonstrated that miRNA serves a regulatory role in the chemosensitivity of various types of cancer cells. miRNA-107 enhanced chemosensitivity to paclitaxel in non-small cell lung cancer by targeting the antiapoptotic factor Bcl-2 (10). miRNA-625 has been demonstrated to increase chemosensitivity in glioma by directly targeting AKT serine/threonine kinase 2 AKT2 (11). The aim of the present study was to elucidate the regulatory role of miR-19a-3p in the chemosensitivity of OS cells. miR-19a-3p was reported to enhance proliferation and insulin secretion, while inhibiting the apoptosis of pancreatic β -cells through the inhibition of suppressor of cytokine signaling 3 (SOCS3) (12). However, the role of miR-19a-3p

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Abbreviations: OS, osteosarcoma; miRNAs, microRNAs; Bax, Bcl-2 associated X, apoptosis regulator; 3'-UTR, 3'-untranslated region; PTEN, phosphatase and tensin homolog; BMSCs, bone marrow derived stroma cells; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; CCK-8, Cell Counting Kit-8

Key words: miR-19a-3p, osteosarcoma, chemosensitivity, PTEN

in the regulation of chemosensitivity is unclear and requires further examination.

Down-regulation of phosphatase and tensin homolog (PTEN), a tumor suppressor gene, was reported in various human primary tumors (13). Previous studies have also demonstrated that PTEN was associated with chemosensitivity of cancer cells. The study of Shen *et al* (14) indicated that downregulation of miRNA-147 expression increased the chemosensitivity of gastric cancer cells to 5-fluorouracil by directly targeting PTEN. Using bioinformatics analysis, it was revealed in the present study that PTEN was a putative target gene of miR-19a-3p. Accordingly, the hypothesis was that miR-19a-3p may be involved in the regulation of chemosensitivity through targeting PTEN in OS cells.

In the present study, overexpression of miR-19a-3p was identified in OS cells and that silencing of miR-19a-3p enhanced the chemosensitivity of OS cells by elevating the expression of PTEN. These results will assist in the understanding of the underlying mechanism of involvement of miR-19a-3p in regulating chemosensitivity of OS cells.

Materials and methods

Cell culture and induction of cisplatin resistant cells. Bone marrow-derived stroma cells (BMSCs) were purchased from BeNa Culture Collection (Beijing, China) and the OS cell lines, MNNG/HOS, U-2 OS, MG63, Saos-2, were purchased from the American Type Culture Collection (Manassas, VA, USA). Cells were cultured in DMEM (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.) at 37°C in 5% CO₂. Cisplatin was purchased from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany) and dissolved in PBS. For the induction of cisplatin-resistant cells, MG63 cells were treated by gradually increasing doses of cisplatin in the cell culture medium throughout the passages for 6~8 months. The cells were maintained in the presence of 5 µM cisplatin in the culture medium for 48 h in every alternate passage. Bpv(HOpic) (Merck KGaA, Darmstadt, Germany), a PTEN inhibitor, was used to treat cells at a concentration of 1 µM.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). The expression level of miR-19a-3p in the cell lines was measured via RT-qPCR. Total RNA was extracted from the cells using the TRIzol (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. cDNA was synthesized using a miScript reverse transcription kit (Qiagen, GmbH, Hilden, Germany) and the PCR reaction was performed using the SYBR Premix Ex Taq™ II kit (Takara Bio, Inc., Otsu, Japan), according to the manufacturer's instructions. The primers used were as following, miR-19a-3p forward, 5'-GGGGGGGTGTGCAAATCT-3', and reverse, 5'-GTGCGTGTCTGGAGTCG-3'; U6, forward, 5'-GCTTCGGCAGCACATATACTAAAAT-3', and reverse, 5'-CGCTTCACGAATTTGCGTGTCTAT-3'. The amplification protocol included an initial denaturation step at 95°C for 10 min, followed by 40 cycles of 95°C for 15 sec and 60°C for 60 sec. The expression levels were calculated using the 2^{-ΔΔC_q} method with U6 used for normalization (15).

Transfection. Cells were seeded into 96-well plates to reach 60% confluence for transfection. miR-19a-3p mimics (cat. no. MIMAT0000073) and inhibitor (cat. no. MIMAT0021837) were purchased from Invitrogen; (Thermo Fisher Scientific, Inc.). According to manufacturer's protocol, transfections were performed using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. For overexpression of PTEN, the full-length PTEN sequence (5'-UUCACAUCCUACCCCUUUGCACU-3') obtained from Invitrogen (Thermo Fisher Scientific, Inc.) was cloned into a pcDNA3.0 vector (Invitrogen; Thermo Fisher Scientific, Inc.). Cells were transfected with pcDNA3.0-PTEN using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. After transfection for 48 h, cells were collected for subsequent experimentation.

Cell Counting Kit-8 (CCK-8) assay. Cells were seeded at 5x10³ per well in 96-well plates and incubated for 0-5 days with and without cisplatin treatment (3 µM). A total of 10 µl of CCK-8 solution (Beyotime Institute of Biotechnology, Shanghai, China) was incubated at 37°C for 2 h. The absorbance was measured at 450 nm using a microplate spectrophotometer (Molecular Devices, LLC, Sunnyvale, CA, USA). Triplicate wells were used in each group.

Western blotting. According to the manufacturer's protocol, proteins were extracted from cells using RIPA lysis buffer (Beyotime Institute of Biotechnology). Homogenized samples were washed with ice-cold PBS and centrifuged at 10,000 x g for 15 min at 4°C. The supernatant was collected and the protein concentration was determined using a BCA protein assay kit (Beijing Solarbio Science & Technology Co., Ltd., Beijing, China). A total of 20 µg protein were loaded per lane and separated by 10% SDS-PAGE and then transferred into PVDF membranes (Merck KGaA). The membrane was blocked with 5% bovine serum albumin for 1 h at room temperature and then incubated with the following primary monoclonal antibodies (at a 1:1,000 dilution): Anti-Ki67 (cat. no. ab92742; Abcam, Cambridge, UK), anti-PCNA (cat. no. 2586; Cell Signaling Technology, Inc., Danvers, MA, USA), anti-Bax (cat. no. ab32503; Abcam), anti-Bcl-2 (cat. no. ab32124; Abcam), anti-PTEN (cat. no. ab79156; Abcam) and GAPDH (cat. no. 5174; Cell Signaling Technology, Inc.) and gently agitated at 4°C overnight. Then the membranes were incubated with horseradish peroxidase-conjugated secondary antibodies (Anti-rabbit IgG, HRP-linked Antibody, cat. no. 7074; Cell Signaling Technology, Inc.; dilution 1:3,000) for 1 h at room temperature. An enhanced chemiluminescence system (Bio-Rad Laboratories, Inc., Hercules, CA, USA) was used for the detection of antibody-bound proteins, according to manufacturer's instructions. Analysis was performed using ImageJ software (version 1.48, National Institutes of Health, Bethesda, MD, USA).

Cell apoptosis assay. An Annexin V-FITC/propidium iodide (PI) apoptosis detection kit (Multisciences, Shanghai, China, <http://liankebio.biomart.cn/>) was used for the cell apoptosis assay, according to manufacturer's protocol. A total of

3×10^5 cells were washed with PBS, resuspended and incubated with 5 μ l Annexin V-FITC and 10 μ l PI. Cell apoptosis was analyzed using a flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA). FACS data was processed using FlowJo (version 7.5.5; Tree Star Inc., Ashland, OR, USA).

MiRNA target prediction. Genes containing the binding sites of the miR-19a-3p 3'-UTR were obtained using TargetScan (http://www.targetscan.org/vert_71/), MiRanda (<http://www.microrna.org/microrna/home.do>), PICTAR (<http://www.pictar.org/>) and miRDB (<http://mirdb.org/miRDB/custom.html>) target prediction algorithms.

Luciferase reporter assay. The fragment of PTEN containing the target sequence 5'-UUUGCAC-3', Invitrogen, Thermo Fisher Scientific, Inc.) of miR-19a-3p was cloned into pGL3 Luciferase Reporter Vectors (Promega Corporation, Madison, WI, USA) to form the reporter vector PTEN-wild-type (PTEN WT), while the PTEN-mutated-type (PTEN MUT) contained the mutated binding site. The cells were co-transfected with PTEN WT or PTEN MUT and miR-19a-3p mimics using Lipofectamine[®] 2000, according to manufacturer's protocol. Following transfection for 48 h, luciferase activity was measured using the Dual-Luciferase[®] Reporter assay system kit (Promega Corporation), according to the manufacturer's instructions. Renilla luciferase activity was used for normalization.

Statistical analysis. All data are presented as the mean \pm standard deviation. Statistical analysis was performed by Student's t-test or one-way analysis of variance with Student-Newman-Keuls post hoc test, using SPSS 19.0 (IBM Corp., Armonk, NY, USA). $P > 0.05$ was considered to indicate a statistically significant difference.

Results

miR-19a-3p is overexpressed in OS cells. To investigate the role of miR-19a-3p in OS, its expression level in OS cells was detected using RT-qPCR. The expression of miR-19a-3p that was revealed in OS cells, MNNG/HOS, U-2 OS, MG63 and Saos-2, was higher than that in normal BMSCs ($P < 0.01$; Fig. 1A). Transfection with miR-19a-3p mimics or inhibitor increased and decreased the expression of miR-19a-3p significantly in U-2 OS and MG63 cells, respectively ($P < 0.01$; Fig. 1B). The expression of miR-19a-3p was highest in MG63 cells, thus, MG63 cells were selected for the following experiments.

Inhibition of miR-19a-3p enhances chemosensitivity of OS cells. A Cisplatin-resistant MG-63 cell line was constructed, as described in the Materials and methods section. Cisplatin treatment did not significantly affect cell proliferation or apoptosis of cisplatin-resistant MG-63 cells. However, transfection with the miR-19a-3p inhibitor downregulated the rate of cell proliferation and decreased the expression of the proliferation-associated proteins, Ki67 and PCNA. Transfection with miR-19a-3p inhibitor NC did not have a significant effect compared with the Cisplatin group ($P > 0.05$; Fig. 2A-C), suggesting that downregulation of miR-19a-3p suppressed the proliferation of Cisplatin-resistant OS cells

under Cisplatin treatment. Transfection with miR-19a-3p inhibitor increased cell apoptotic rate, while miR-19a-3p mimics did not have a significant effect compared with the Cisplatin group ($P > 0.05$; Fig. 2D and E). Transfection with miR-19a-3p inhibitor increased the expression of Bax and decreased the expression of Bcl-2 compared with the Cisplatin group ($P < 0.05$; Fig. 2F and G), suggesting that downregulation of miR-19a-3p expression promoted apoptosis of Cisplatin-resistant OS cells under Cisplatin treatment. The data indicates that miR-19a-3p inhibitor increases the chemosensitivity of OS cells to Cisplatin.

PTEN is a target of miR-19a-3p in OS cells. To explore the chemosensitivity regulatory mechanism of miR-19a-3p in OS cells, the public database, TargetScan was used to search for potential targets of miR-19a-3p. It was revealed that the PTEN mRNA 3'-UTR contained highly conserved sequences, which were targeted by miR-19a-3p (Fig. 3A). Expression of endogenous PTEN protein was detected in OS cells. PTEN expression was revealed to be significantly downregulated in MNNG/HOS, U-2 OS, MG63 and Saos-2 cells compared with BMSCs cells ($P < 0.05$, $P < 0.01$; Fig. 3B and C). Expression of PTEN was downregulated in the miR-19a-3p mimics group and upregulated in the miR-19a-3p inhibitor group, compared with the control group ($P < 0.05$; Fig. 3D and E). To confirm the association between PTEN and miR-19a-3p in OS cells, a luciferase reporter assay was conducted. It was revealed that co-transfection with miR-19a-3p mimics and PTEN WT led to a significant decrease in luciferase activity. However, co-transfection with miR-19a-3p mimics and PTEN MUT did not have a significant effect on luciferase activity ($P > 0.01$; Fig. 3F). In summary, these results indicated that PTEN was a target of miR-19a-3p and that the expression of PTEN was negatively regulated by miR-19a-3p in OS cells.

Overexpressed PTEN enhances the chemosensitivity of OS cells. MG-63 cells were transfected with pcDNA3.0-PTEN to increase the protein expression level of PTEN ($P < 0.05$; Fig. 4A and B). It was demonstrated that overexpressed PTEN decreased cell proliferation and increased apoptotic rate compared with the Cisplatin group ($P < 0.05$; Fig. 4C and D). The results indicate that overexpressed PTEN enhances the chemosensitivity of OS cells.

PTEN inhibitor BpV(HOpic) counteracts the effect of miR-19a-3p inhibitor in the regulation of chemosensitivity. To investigate the interaction between PTEN and miR-19a-3p, the PTEN inhibitor, BpV(HOpic), was used in the present study. It was observed that BpV(HOpic) treatment increased cell proliferation and decreased apoptotic rate compared with the Cisplatin-treated miR-19a-3p inhibitor group ($P < 0.05$; Fig. 5A and B). These results indicate that inhibition of PTEN counteracted the effect of the miR-19a-3p inhibitor on the regulation of chemosensitivity in OS cells.

Discussion

In the treatment of OS, the combination of chemotherapy and surgery has improved the overall 5-year survival rate of patients with OS (16). Cisplatin is one of the most commonly

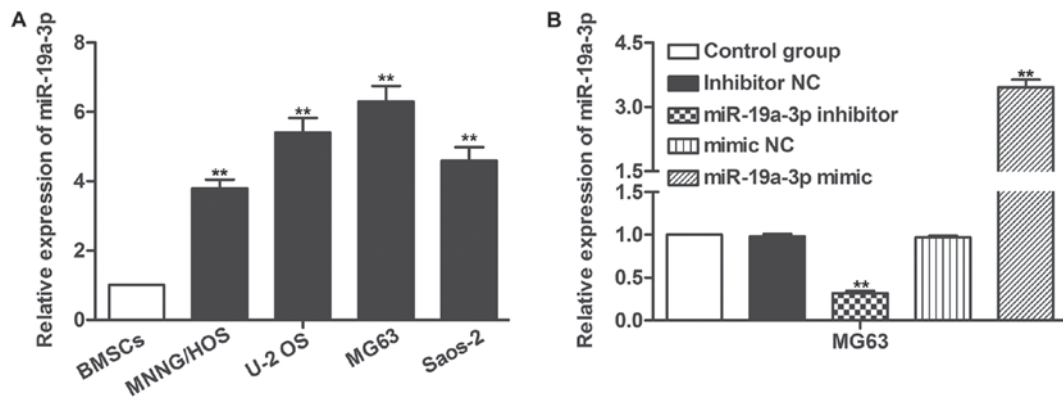


Figure 1. miR-19a-3p is overexpressed in OS cells. (A) The relative expression of miR-19a-3p was compared among BMSCs and OS cell lines by RT-qPCR. ** $P < 0.01$ compared with BMSCs. (B) Relative expression of miR-19a-3p was measured in MG63 cells transfected with miR-19a-3p mimic or inhibitor by RT-qPCR. ** $P < 0.01$ compared with the control group. The error bars represent mean \pm standard deviation of three independent experiments. miR, microRNA; OS, osteosarcoma; BMSCs, bone marrow derived stroma cells; RT-qPCR, reverse transcription-quantitative polymerase chain reaction.

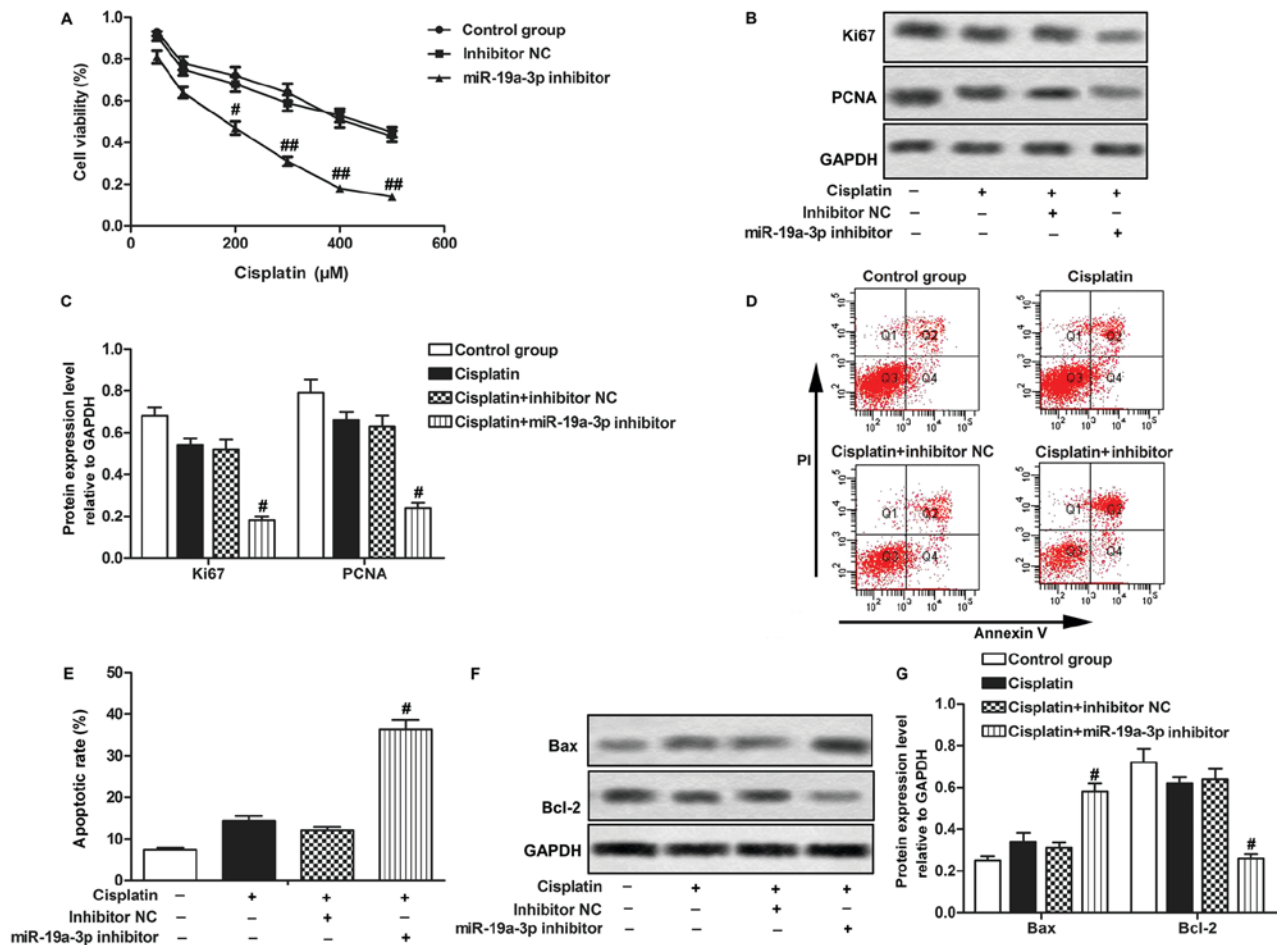


Figure 2. miR-19a-3p inhibitor enhances chemosensitivity of OS cells. A Cisplatin-resistant cell line was constructed, and Cisplatin-resistant cells were transfected with miR-19a-3p mimics or inhibitor. Cisplatin treatment of cisplatin-resistant cells was used as a control. (A) Cell proliferation was measured by CCK-8 assay. (B) Expression of Ki67 and PCNA protein was detected through western blotting. (C) Quantification of the protein expression of Ki67 and PCNA. (D) Apoptosis was detected by flow cytometric analysis. (E) Quantification of cell apoptotic rate. (F) Expression of Bax and Bcl-2 protein was detected through western blotting. (G) Quantification of Bax and Bcl-2 protein expression. The error bars represent means \pm standard deviation of three independent experiments. # $P < 0.05$, ## $P < 0.01$ compared with Cisplatin group. miR, microRNA; OS, osteosarcoma; CCK-8, Cell Counting Kit-8; PCNA, proliferating cell nuclear antigen; Bax, BCL2 associated X, apoptosis regulator.

used platinum-based anticancer drugs for the treatment of OS, interacting with nucleophilic N⁷-sites of purine bases to induce DNA damage that leads to cell death (17). However,

Cisplatin resistance can develop and limit the effectiveness of chemotherapeutics. Therefore, understanding the molecular mechanism of chemoresistance to Cisplatin is essential to

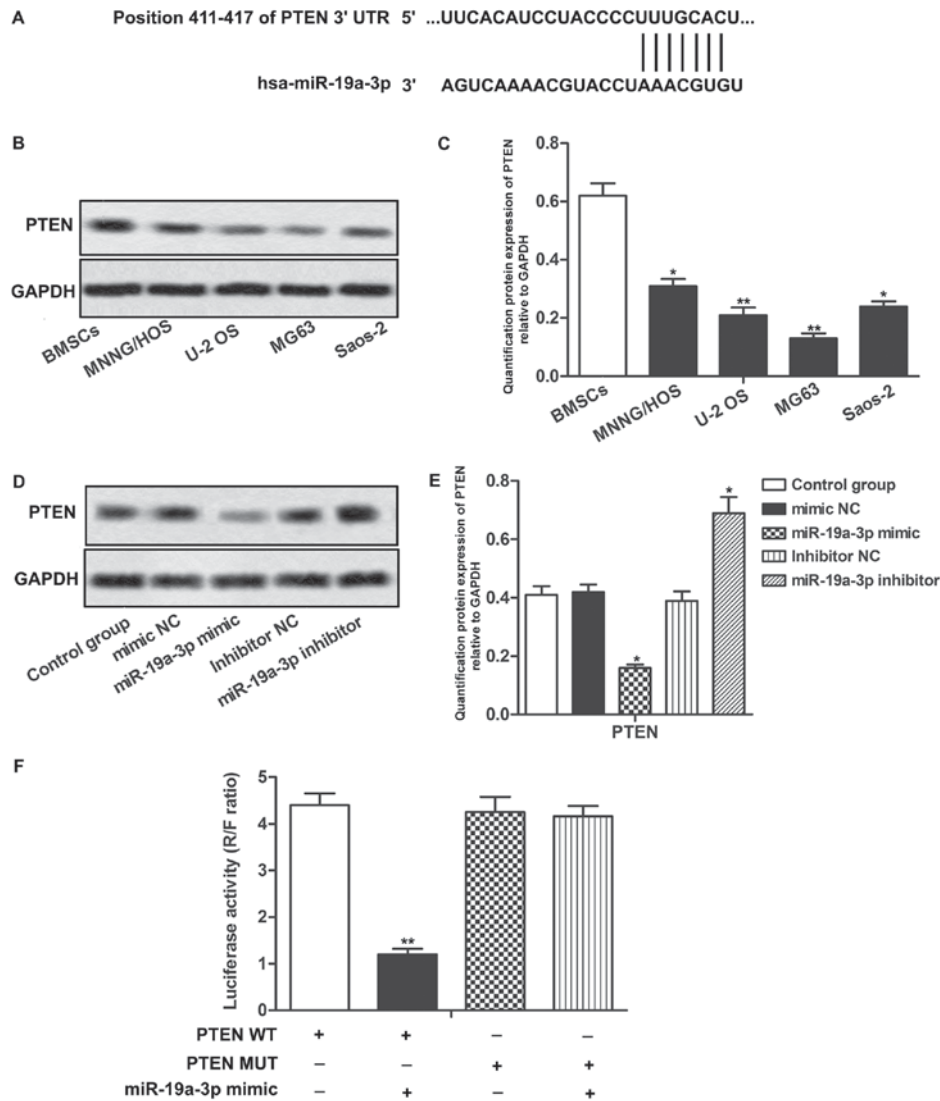


Figure 3. PTEN is a target of miR-19a-3p in OS cells. (A) The putative binding site of miR-19a-3p in 3'-UTR of PTEN. (B) Protein expression of PTEN in OS cell lines and bone marrow-derived stromal cells was measured through western blotting. (C) Quantification of the protein expression of PTEN in MNNG/HOS, U-2 OS, MG63, Saos-2. * $P<0.05$, ** $P<0.01$ compared with BMSCs group. (D) Relative expression of PTEN in MG63 cells transfected with miR-19a-3p mimics or inhibitor was measured through western blotting. (E) Quantification of the protein expression level of PTEN relative to GAPDH. * $P<0.05$, compared with control group. (F) Results of the luciferase reporter assay. ** $P<0.01$ compared with PTEN WT group. PTEN, phosphatase and tensin homolog; miR, microRNA; OS, osteosarcoma; 3'-UTR, 3'-untranslated region; BMSC, bone marrow derived stroma cell; WT, wild type; MUT, mutated; NC, negative control; R, *Renilla*; F, *Firefly*.

develop a more effective treatment for OS. In the present study, it was demonstrated that silencing of miR-19a-3p enhanced chemosensitivity to Cisplatin in OS cells by up-regulating the expression of PTEN. These results can assist in finding a better therapeutic strategy against Cisplatin resistance in OS.

miRNAs, a class of non-coding regulatory RNAs, have been reported to be involved in tumor progression (18,19). As miRNAs act as oncogenes or tumor suppressor genes, they constitute a large gene network for the regulation of various cellular processes during tumorigenesis (9). Evidence suggests that numerous miRNAs are associated with chemoresistance in various types of cancers. For example, miR-491 has been demonstrated to inhibit chemoresistance of OS cells by targeting α B-crystallin (20). miR-608 has been demonstrated to inhibit gemcitabine chemoresistance

by regulating resistance genes in pancreatic cancer cells (21). Previous studies have reported that miR-19a-3p expression was correlated with cell proliferation and autophagy (12,22). In addition, the serum level of miR-19a-3p has been reported to be significantly higher in patients with colorectal cancer compared with healthy patients (23). Similarly, it was demonstrated that the expression of miR-19a-3p was upregulated in OS cells compared with normal BMSCs. In the present study a cisplatin-resistant MG-63 cell line was constructed to investigate the role of miR-19a-3p in the chemosensitivity of OS cells. It was observed that Cisplatin treatment alone did not significantly affect proliferation or apoptosis of cisplatin-resistant MG-63 cells. However, Cisplatin treatment combined with miR-19a-3p inhibitor transfection suppressed cell proliferation and decreased the expression of

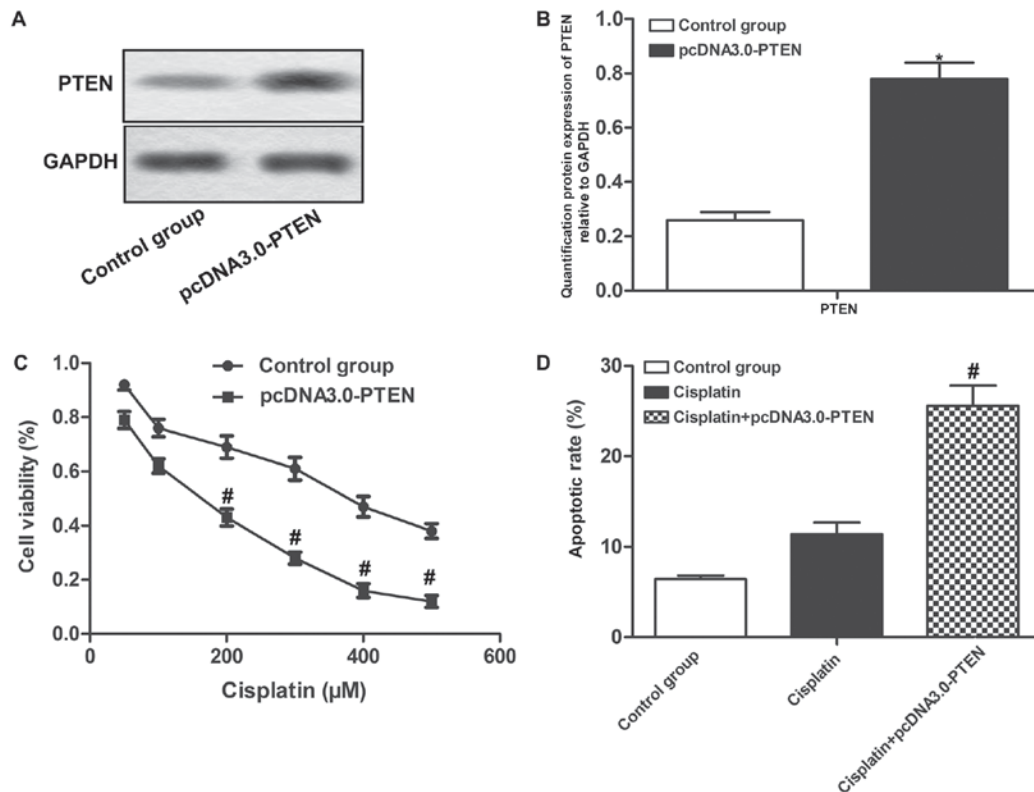


Figure 4. Overexpression of PTEN enhances the chemosensitivity of OS cells. Cisplatin-resistant MG63 cells transfected with pcDNA3.0-PTEN received Cisplatin treatment and Cisplatin-resistant MG63 cells as control. (A) Relative protein expression of PTEN was compared between the control group and the pcDNA3.0-PTEN group through western blotting. (B) Quantification of the protein expression of PTEN. (C) Proliferation was measured by CCK-8 assay. (D) Cell apoptosis was detected by flow cytometric analysis. The error bars represent means \pm standard deviation of three independent experiments. * $P < 0.05$ compared with control group and # $P < 0.05$ compared with Cisplatin group. PTEN, phosphatase and tensin homolog; OS, osteosarcoma; CCK-8, Cell Counting Kit-8.

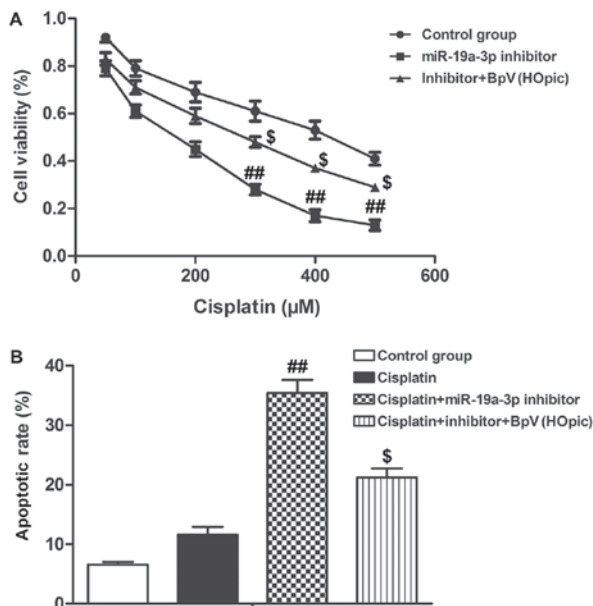


Figure 5. PTEN inhibitor, BpV (HOpic), counteracts the effect of miR-19a-3p inhibitor-transfection on the regulation of chemosensitivity. Cisplatin-resistant MG63 cells transfected with miR-19a-3p inhibitor received Cisplatin treatment with or without BpV(HOpic) treatment. (A) Cell proliferation was measured using CCK-8 assay. (B) Cell apoptosis was detected by flow cytometric analysis. The error bars represent the mean \pm standard deviation of three independent experiments. ## $P < 0.05$ compared with Cisplatin group and \$ $P < 0.05$ compared with Cisplatin + miR-19a-3p inhibitor group. PTEN, phosphatase and tensin homolog; CCK-8, Cell Counting Kit-8.

proliferation-associated proteins, including Ki67 and PCNA, compared with the Cisplatin treatment group. Cisplatin treatment combined with miR-19a-3p inhibitor transfection increased the apoptotic rate, increased the expression of Bax, and decreased the expression of Bcl-2 compared with the Cisplatin treatment group. In summary, these results suggest that the silencing of miR-19a-3p enhances chemosensitivity to Cisplatin in OS cells by suppressing cell proliferation and promoting cell apoptosis in treatment with Cisplatin.

PTEN is a tumor suppressor and its expression is often downregulated in various types of tumors (24,25). Similarly, downregulation of the expression of PTEN in OS cells was observed in the present study. Silencing of PTEN has been reported to induce activation of the PI3K/AKT pathway, which leads to cell survival and apoptosis resistance (26,27). Previous studies have reported that PTEN is regulated by miRNAs in many types of cancers. Liu *et al* (26) indicated that miR-206 inhibited head and neck squamous cell carcinoma (HNSCC) cell progression by targeting HDAC6 via the PTEN/AKT/mTOR pathway. miR-130a increased cell proliferation and tumor growth through repression of PTEN (28). miRNA-21 enhanced chemoresistance to Cisplatin by negatively regulating PTEN in epithelial ovarian cancer (29). The miRNA/PTEN axis may serve an important role in the regulation of tumorigenesis. In the present study, bioinformatics tools demonstrated that the 3'-UTR of the PTEN gene

harbored a putative binding site for miR-19a-3p. Expression of PTEN was significantly downregulated in the miR-19a-3p mimics group and significantly upregulated in the miR-19a-3p inhibitor group, indicating that the expression of PTEN was negatively regulated by miR-19a-3p. To further examine the interaction between PTEN and miR-19a-3p, a luciferase reporter assay was performed and the results suggested that miR-19a-3p directly targeted PTEN in OS cells. In order to examine the effect of PTEN in the chemosensitivity of OS cells, cisplatin-resistant MG-63 cells were transfected with pcDNA3.0-PTEN to up-regulate the expression of PTEN. The present study revealed that Cisplatin treatment combined with pcDNA3.0-PTEN transfection decreased cell proliferation and increased apoptotic rate compared with Cisplatin treatment alone, suggesting that overexpression of PTEN enhanced chemosensitivity of OS cells to Cisplatin. Therefore, silencing of miR-19a-3p may result in upregulation of the expression of PTEN in OS cells. To verify this result, Bpv(HOPic), a PTEN inhibitor, was used to treat miR-19a-3p inhibitor-transfected cells. It was observed that inhibition of PTEN upregulated cell proliferation and down-regulated apoptotic rate compared with the Cisplatin-treated miR-19a-3p inhibitor group, indicating that inhibition of PTEN counteracted the effect of the miR-19a-3p inhibitor on the chemosensitivity of OS cells.

Taken together, the results of the present study indicate a role of miR-19a-3p in the chemosensitivity of OS cells. miR-19a-3p was overexpressed in OS cell lines and downregulation of miR-19a-3p enhanced chemosensitivity to Cisplatin by elevating the expression of tumor suppressor PTEN in OS cells. Therefore, an miR-19a-3p/PTEN axis may be a potential target for the treatment of OS.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

BZ interpreted the main data regarding the cell survival and apoptosis analysis. JNZ was responsible for western blot and statistical analysis. YL was responsible for study design and drafting of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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