

# Comprehensive analysis of differentially expressed long non-coding RNAs in non-small cell lung cancer

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**Abstract.** Non-small cell lung cancer (NSCLC) is the primary subtype of lung cancer. Long non-coding RNAs (lncRNAs) have been reported to serve prominent roles in cancer progression. However, the expression patterns and potential roles of lncRNAs in NSCLC remain to be elucidated. In the present study, four public datasets were analyzed to identify differentially expressed lncRNAs (DELncs) in NSCLC. A further dataset, GSE19188, was analyzed to validate the findings. A total of 38 upregulated and 31 downregulated lncRNAs were identified in NSCLC, compared with samples from healthy controls. Among these, 12 lncRNAs were associated with the progression of NSCLC, and dysregulated between high grade (stage III and IV) and low grade (stage II) NSCLC samples. Moreover, dysregulation of lncRNA-SIGLEC17P, GGTA1P, A2M-AS1, LINC00938, GVINP1, LINC00667 and TMPO-AS1 was associated with overall survival time in patients with NSCLC. Co-expression analyses, combined with the construction of protein-protein interaction networks, were performed to reveal the potential roles of key lncRNAs in NSCLC. The present study revealed a series of lncRNAs involved in the progression of NSCLC, which may serve as novel biomarkers for the disease.

## Introduction

With the development of high-throughput methods, a number of studies have revealed the roles of non-coding RNAs, including microRNAs (miRNA) (1), long non-coding RNAs (lncRNAs) (2) and small nucleolar RNAs (snoRNAs) (3).

lncRNAs are a subclass of RNA transcripts >200 bps in length (4). Emerging studies have indicated that lncRNAs are abnormally expressed in various types of human cancer, including prostate cancer (5), breast cancer (6), hepatocellular carcinoma (7) and lung cancer (8). These lncRNAs were involved in multiple biological processes, including the cell cycle, invasion, proliferation and drug resistance, by influencing the targets of transcriptional (cis/trans) and post-transcriptional regulation (9). Specific lncRNAs, including GHET1 (10), LINP1 (11), NEAT1 (12) and MIR100HG (13) were reported to be oncogenes. However, various other lncRNAs, including GAS5 (14) and LINC00961 (15), were implicated in cancer suppression. Therefore, exploring the functions of lncRNAs may provide novel insights to identify diagnostic and prognostic markers of cancer.

Lung cancer has become the leading cause of cancer-associated mortality worldwide (16), of which non-small cell lung cancer (NSCLC) is the primary subtype (16). Despite the development of various treatment methods, including radiotherapy, chemotherapy and targeted therapy, the five-year survival rate of NSCLC remains low, at ~15% (17). In previous reports, lncRNAs were identified as novel regulators of lung cancer progression; the transforming growth factor  $\beta$ -induced lncRNA-TBILA promoted NSCLC progression via cis-regulation of human germinal center-associated lymphoma HGAL, and activation of S100A7/JAB1 (18). However, in NSCLC, the potential roles of a great number of lncRNAs remain unknown.

The aim of the present study was to identify novel therapeutic and prognostic biomarkers for NSCLC. Previously published datasets were analyzed to identify differentially expressed lncRNAs (DELnc). To determine the functions of these DELncs, a series of bioinformatics analyses were performed, including Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes analysis.

## Materials and methods

**lncRNA classification pipeline.** A pipeline described by Zhang *et al* (19) was employed to re-annotate the microarray data. Entries labeled as 'NR' or annotated with 'lncRNA', 'processed transcripts', 'non-coding' or 'misc\_RNA' in Ensembl annotations, were retained.

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**Microarray data and pre-processing.** The following datasets were downloaded from the Gene Expression Omnibus (GEO) database (<https://www.ncbi.nlm.nih.gov/geo/>) to identify DElncs: GSE19804, GSE27262, GSE18842 and GSE19188. GSE19804 was reported by Lu *et al* (20) and contained 65 normal samples and 91 female lung cancer samples; GSE27262 included 25 normal and 25 stage I lung adenocarcinoma patients (21); GSE18842 included 45 normal and 46 NSCLC samples (22). A further dataset, GSE19188, which includes 65 normal and 91 lung cancer samples, was used for validation of the findings (23). In addition, The Cancer Genome Atlas Lung Adenocarcinoma dataset (TCGA-LUAD), based on RNA-sequence methodology was also downloaded to analyze the expression patterns of lncRNAs in LUAD samples. In the present study, the lncRNAs whose expression in NSCLC samples were higher than that in normal samples (with a fold change (FC)  $\geq 2$  and  $P < 0.05$ ), were considered to be upregulated. The lncRNAs whose expression in NSCLC samples was lower than that in normal samples (with an  $FC \leq 0.5$  and  $P < 0.05$ ), were considered to be downregulated.

**Functional group analysis.** Bioinformatics analysis was conducted using the Database for Annotation, Visualization and Integrated Discovery system (DAVID version 6.8; <http://david.ncifcrf.gov/>), to identify the relevant biological functions of any high-throughput gene functional analysis (24).  $P < 0.05$  was considered to indicate a statistically significant difference.

**Protein-protein interaction (PPI) network and module analysis.** Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) online software was used to construct PPI networks ([https://string-db.org/cgi/input.pl?sessionId=AUH42ZEWajP&input\\_page\\_show\\_search=on](https://string-db.org/cgi/input.pl?sessionId=AUH42ZEWajP&input_page_show_search=on)). PPIs with a combined score  $> 0.4$  were considered to be significant. Cytoscape software (<https://cytoscape.org/>) was used to visualize the PPI networks.

**Survival analysis.** The Kaplan-Meier plotter (<http://www.kmplot.com/analysis>) is a public dataset including 54,675 genes on survival using 2,437 lung cancer samples, with a mean follow-up period of 49 months. The median expression of lncRNAs was selected as the cut-off point to divide patients with NSCLC into high- and low-expression groups.

**Statistical analysis.** Data are presented as the mean  $\pm$  standard deviation. All statistical analyses were performed using SPSS 17.0 software (SPSS, Inc.). Statistical comparisons between groups were performed using the Mann-Whitney U test.  $P < 0.05$  was considered to indicate a statistically significant difference.

## Results

**Identification of DElncs in NSCLC.** In order to identify key lncRNAs involved in the progression of NSCLC, comprehensive analysis of four public datasets were performed. A total of 638 upregulated and 294 downregulated lncRNAs were identified in the GSE19804 dataset (Fig. 1A); 525 upregulated and 216 downregulated lncRNAs were identified in the GSE27262 dataset (Fig. 1B); 379 upregulated and 508 downregulated

lncRNAs were identified in the GSE18842 dataset (Fig. 1C); and 489 upregulated and 223 downregulated lncRNAs were identified in TCGA dataset (Fig. 1D). Hierarchical clustering revealed systematic variations in the expression of lncRNAs in NSCLC samples.

Integrated analysis of DElncs in NSCLC was also performed. The upregulation of 38 lncRNAs (Fig. 2A), and the downregulation of 31 lncRNAs (Fig. 2B) was observed in lung cancer compared with normal, healthy samples. Of note, a number of lncRNAs identified as prominent regulators in lung cancer progression were upregulated, including TUG1, DLEU2 and DANCR. However, the functions of the majority of lncRNAs in human disease require further investigation.

**Verification of DElncs in NSCLC.** In order to verify the expression pattern of the aforementioned DElncs in NSCLC, the validation dataset GSE19188 was analyzed. Upregulation of 38, and downregulation of 31 lncRNAs in the training datasets were identified as dysregulated in GSE19188. Hierarchical clustering revealed systematic variations in the expression of lncRNAs in the NSCLC, by analysis of GSE19188 (Fig. 2C).

**Identification of lncRNAs associated with the progression of NSCLC.** Associations between the expression of 69 DElncs and the clinicopathological features of NSCLC were determined. The expression levels of lncRNA-GVINP1, RPL32P3, EPB41L4A-AS2, ILF3-AS1, LINC00938, LINC00667, A2M-AS1, GUSBP11, GGTA1P, BCRP3 and SIGLEC17P were suppressed, while lncRNA-TMPO-AS1 expression was upregulated in high grade (stage III and IV) in comparison to low grade (stage II) NSCLC samples (Fig. 3).

**Dysregulation of DElncs is associated with altered survival time of patients with NSCLC.** To evaluate the prognostic value of DElncs in NSCLC, Kaplan-Meier analysis was performed using TCGA LUAD dataset. Patients with NSCLC were divided into low- and high-lncRNA expression groups, with the median expression level selected as the cut-off. Among these lncRNAs, higher expression of lncRNA-SIGLEC17P, GGTA1P, A2M-AS1, ILF3-AS1, LINC00938 and GVINP1, in addition to lower expression of lncRNA-TMPO-AS1, was associated with longer survival time in patients with LUAD (Fig. 4). Furthermore, the Kaplan-Meier-plotter database was used to ascertain the potential prognostic values of the aforementioned lncRNAs. It was revealed that higher expression levels of lncRNA-SIGLEC17P, GGTA1P, A2M-AS1, LINC00667 and GVINP1, and lower expression levels of lncRNA-ILF3-AS1, LINC00938 and TMPO-AS1 were associated with higher survival rates in patients with NSCLC (Fig. 5). Collectively, these results suggested that DElncs may serve as novel biomarkers for the prognosis of patients with NSCLC.

**Construction of key lncRNA-associated PPI networks in NSCLC.** Co-expression analysis was performed for the construction of PPI networks to reveal the potential roles and molecular mechanisms of key lncRNAs in NSCLC. The Pearson's correlation coefficient was calculated for the lncRNA-mRNAs, using their expression levels in TCGA dataset. lncRNA-mRNAs pairs with  $|R| > 0.7$  were considered

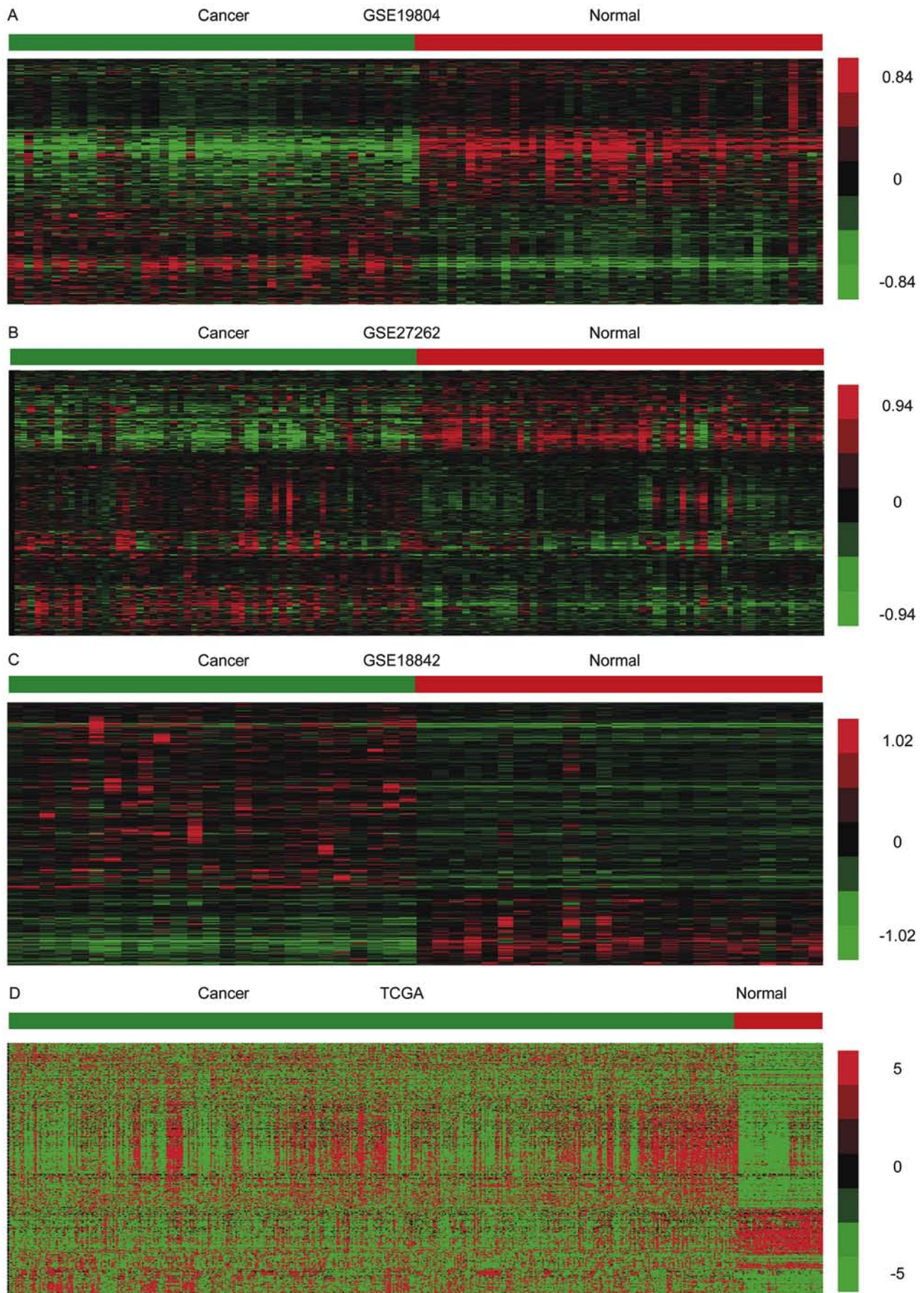


Figure 1. Identification of differentially expressed long non-coding RNAs in NSCLC. Differentially expressed lncRNAs in NSCLC vs. normal tissues, obtained from (A) GSE19804, (B) GSE27262, (C) GSE18842 and (D) TCGA LUAD datasets are represented with heatmaps. Red indicates high relative expression and green indicates low relative expression. NSCLC, non-small cell lung cancer; TCGA LUAD, The Cancer Genome Atlas Lung Adenocarcinoma.



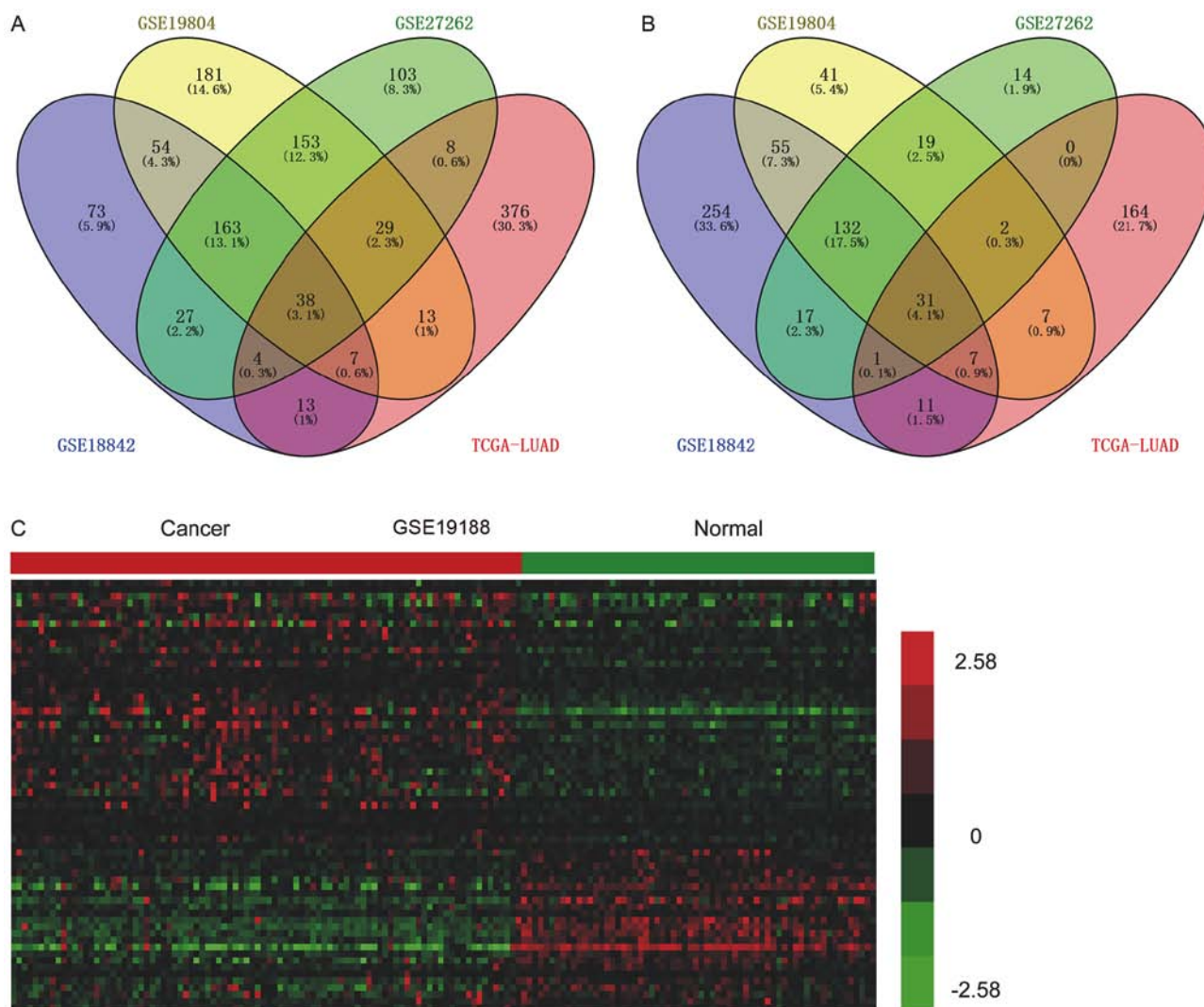


Figure 2. Verification of DElncs associated with NSCLC, using a validation dataset. Four-way Venn diagrams display the overlap between (A) upregulated and (B) downregulated lncRNAs associated with NSCLC, from the analysis of GSE18842, GSE19804, GSE27262 and TCGA LUAD. (C) Verification of the DElncs in NSCLC using GSE19188. Red indicates high relative expression and green indicates low relative expression. DElncs, differentially expressed lncRNAs; NSCLC, non-small cell lung cancer; TCGA LUAD, The Cancer Genome Atlas Lung Adenocarcinoma.

to be reliable. Subsequently, the STRING database was used to investigate PPIs among key lncRNA genes that were co-expressed (combined score >0.4).

The A2M-AS1 mediated PPI network included 67 proteins and 282 edges (Fig. 6A). The GVINP1 mediated PPI network included 28 proteins and 378 edges (Fig. 6B). The LINC00667 mediated PPI network included 23 proteins and 244 edges (Fig. 6C). The GGTA1P mediated PPI network included 47 proteins and 329 edges (Fig. 6D). TMPO-AS1 mediated PPI network included 118 proteins and 6075 edges (Fig. 6E). The ILF3-AS1 mediated PPI network included 13 proteins and 78 edges (Fig. 6F). The SIGLEC17P mediated PPI network included 65 proteins and 560 edges (Fig. 7A). The LINC00938 mediated PPI network included 33 proteins and 176 edges (Fig. 7B).

**Functional analysis of key lncRNAs in NSCLC.** Bioinformatics analysis was performed using the DAVID system. GO analysis revealed that lncRNA-GVINP1 was associated with the regulation of B cell receptor signaling pathway, inflammatory

response, positive regulation of T cell proliferation, immune response and chemotaxis (Fig. 8A); A2M-AS1 was associated with cell adhesion, angiogenesis, positive regulation of GTPase activity, leukocyte migration and vasculogenesis (Fig. 8B); SIGLEC17P was associated with cell adhesion, leukocyte migration, signal transduction, positive regulation of GTPase activity and angiogenesis (Fig. 8C); ILF3-AS1 was involved in regulating cilium morphogenesis, cilium assembly, cilium movement, axoneme assembly and protein homotetramerization (Fig. 8D); TMPO-AS1 was associated with cell division, DNA replication, mitotic nuclear division, sister chromatid cohesion and DNA repair (Fig. 8E); LINC00938 was involved in regulating transcription, cilium morphogenesis, intraciliary retrograde transport, skin development and cilium assembly (Fig. 8F); LINC00667 was involved in regulating transcription, apoptotic process, one-carbon metabolic process, negative regulation of endothelial cell migration and mitotic cytokinesis (Fig. 8G); and GGTA1P was associated with leukocyte migration, inflammatory response, cell adhesion, immune response and signal transduction (Fig. 8H).

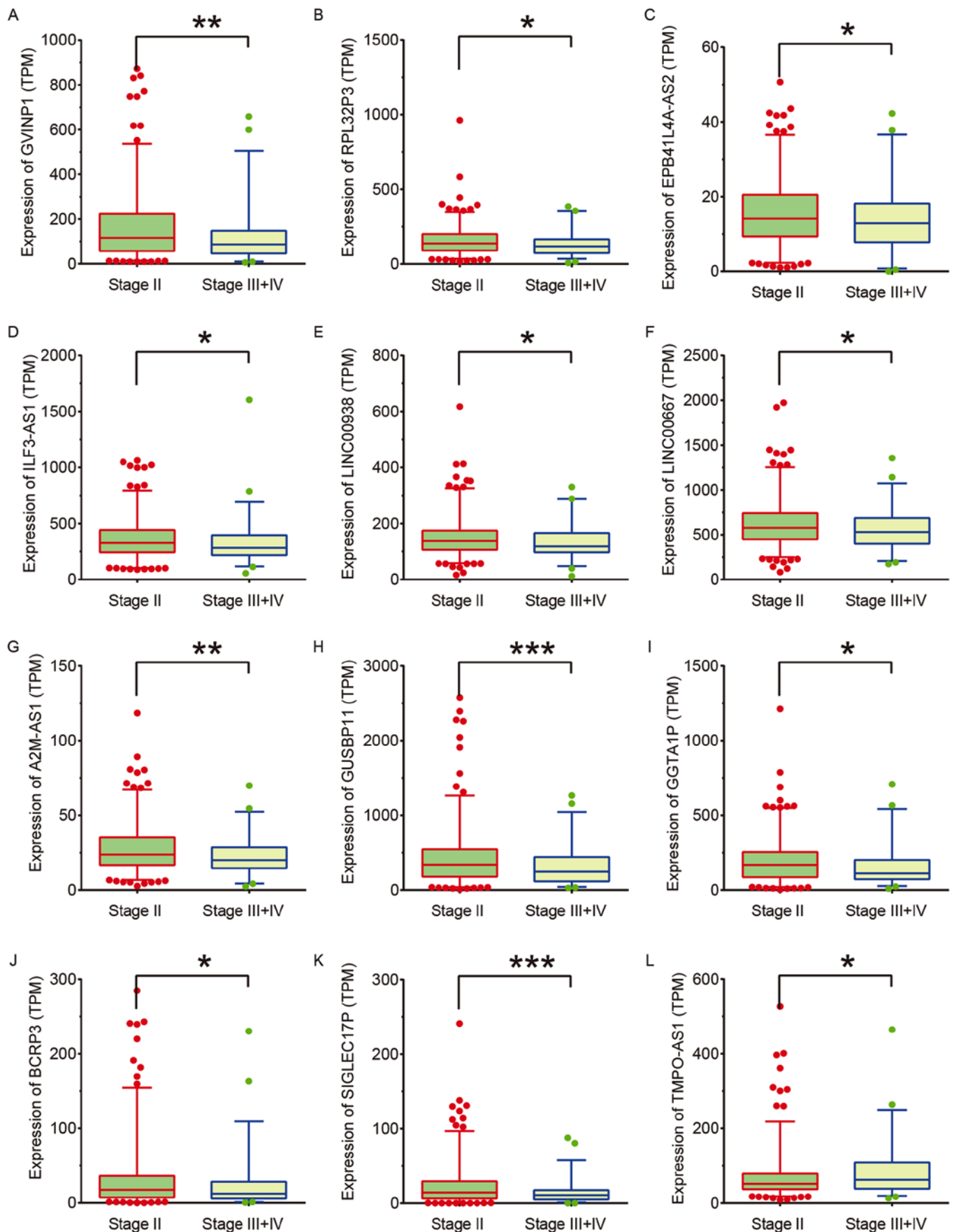


Figure 3. Identification of lncRNAs associated with NSCLC progression. (A) lncRNA-GVINP1, (B) RPL32P3, (C) EPB41L4A-AS2, (D) ILF3-AS1, (E) LINC00938, (F) LINC00667, (G) A2M-AS1, (H) GUSBP11, (I) GGTA1P, (J) BCRP3, and (K) SIGLEC17P expression were suppressed, and (L) TMPO-AS1 expression was upregulated in high grade (stage III and stage IV) compared with low grade (stage II) NSCLC samples. \*P<0.05; \*\*P<0.01; \*\*\*P<0.001. DElncs, differentially expressed lncRNAs; NSCLC, non-small cell lung cancer; TPM, transcripts per kilobase (of exon model per million mapped reads).

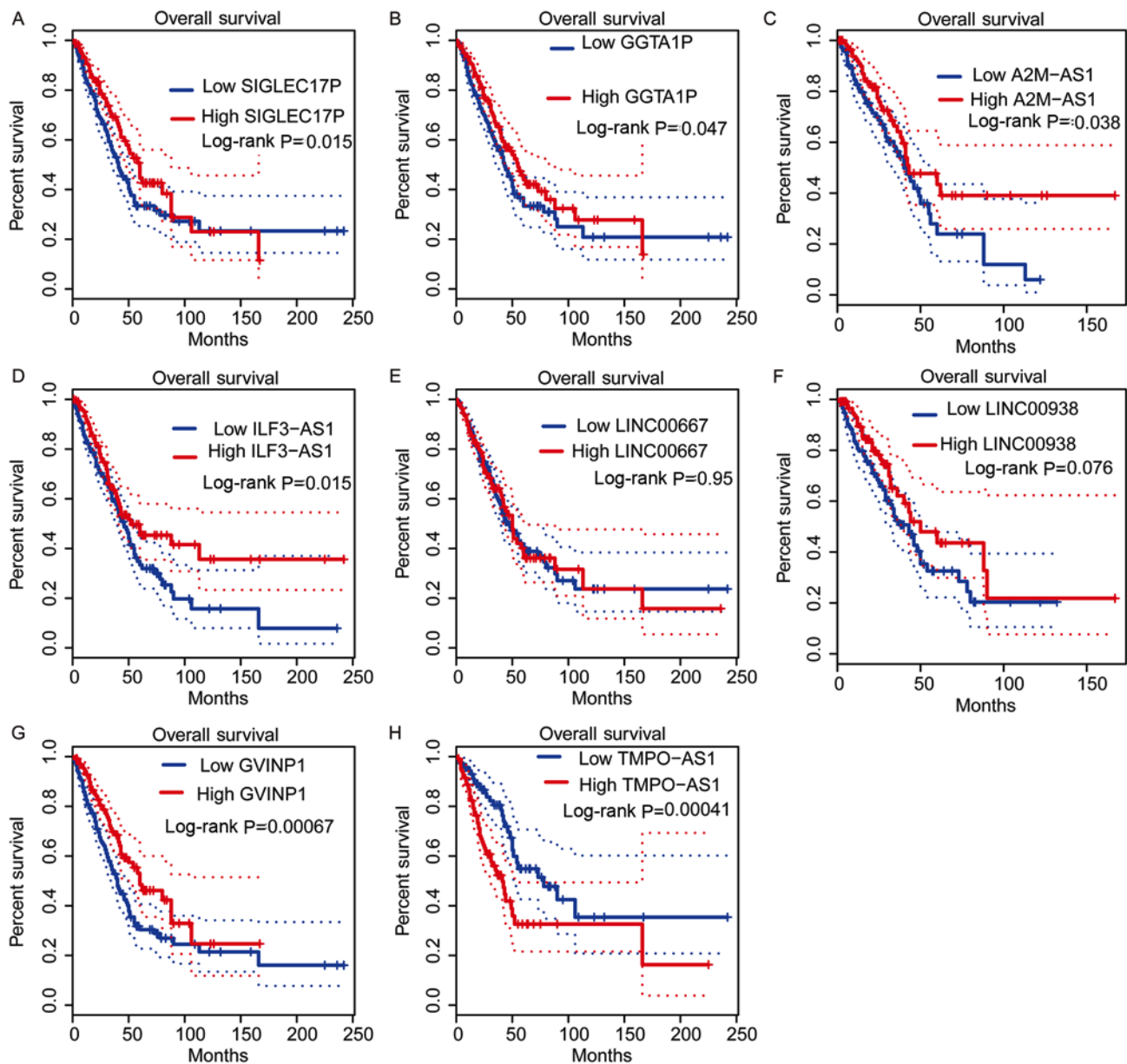


Figure 4. Dysregulation of DElncs is associated with survival time in NSCLC, as determined by analysis of TCGA dataset. TCGA LUAD analysis revealed that higher expression levels of (A) lncRNA-SIGLEC17P, (B) GGTA1P, (C) A2M-AS1, (D) ILF3-AS1 were associated with longer survival times in patients with LUAD. However, the expression of (E) lncRNA-LINC00667 was not significantly associated with prognosis in LUAD patients. Higher expression levels of (F) LINC00938 and (G) GVINP1, and lower expression of (H) TMPO-AS1 were associated with longer survival times in patients with LUAD. Red dotted lines represent the 95% confidence interval of the high group survival curves. Blue dotted lines represent the 95% confidence interval of the low group survival curves. Differentially expressed lncRNAs; NSCLC, non-small cell lung cancer; TCGA LUAD, The Cancer Genome Atlas Lung Adenocarcinoma; lncRNA, long non-coding RNA.

## Discussion

NSCLC is one of the leading causes of cancer-associated mortality worldwide. Previous studies have demonstrated that lncRNAs serve a prominent role in the progression of NSCLC, as either oncogenes or tumor suppressors; lncRNA-MALAT-1 enhanced NSCLC cell motility (25) and lncRNA-CCAT2 promoted the invasion of NSCLC cells (26). However, lncRNA-SIK1-LNC suppressed the proliferative and invasive abilities of NSCLC cells (27). lncRNA-PICART1 suppressed the proliferation of NSCLC cells by inhibiting JAK2/signal transducer and activator of transcription 3 signaling (28). Of note, other reports have demonstrated that the dysregulation

of specific lncRNAs may serve as an indicator of NSCLC. For instance, lncRNA-HOTAIR (29), MALAT1 (25), CCAT2 (26) and H19 (30) were upregulated, and lncRNA-MEG3 (31) and TUG1 (32) were downregulated in lung cancer. However in NSCLC, the expression patterns and molecular functions of the majority of lncRNAs remained to be investigated.

In the present study, using various public datasets, DElncs were identified as diagnostic biomarkers for NSCLC. The upregulation of 38 lncRNAs was observed in NSCLC compared with normal tissues, in addition to the suppression of a further 31 lncRNAs. Among these lncRNAs, HCG11 (33), CASC15 (34), TUG1 (32), DLEU2 (35) and DANCR (36) were dysregulated during cancer progression. lncRNA-HCG11 was

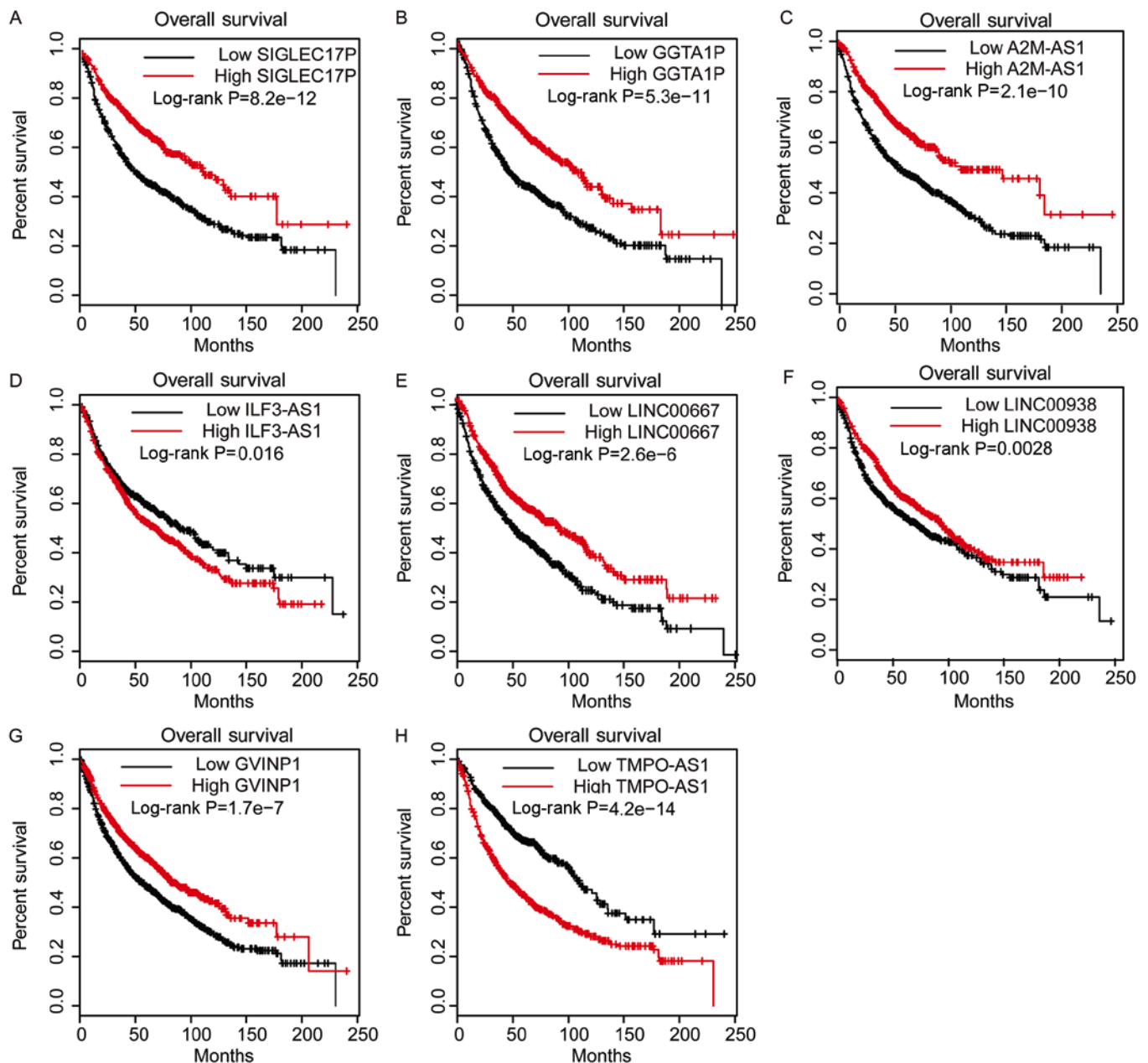


Figure 5. Dysregulation of DElncs is associated with survival time in NSCLC, as determined by analysis of the Kaplan-Meier-plotter database. Higher expression levels of (A) lncRNA-SIGLEC17P, (B) GGTA1P, (C) A2M-AS1, (F) LINC00938, and (G) GVINP1, and lower expression of (D) ILF3-AS1, (E) LINC00667 and (H) TMPO-AS1 were associated with longer survival times in patients with NSCLC. Differentially expressed lncRNAs; NSCLC, non-small cell lung cancer; lncRNA, long non-coding RNA.

reported to be downregulated, and to predict poor prognosis in prostate cancer. Additionally, lncRNA-CASC15 promoted the progression of various types of cancer, including melanoma, colon cancer, gastric cancer and hepatocellular carcinoma. TUG1 is widely reported to be an oncogene, regulating cancer cell proliferation, apoptosis, invasion and glutamine metabolism. In NSCLC, lncRNA-TUG1 was reported to be involved in regulating the growth and chemoresistance of NSCLC cells, in association with LIMK2b, BAX and HOXB7 (37). Of note, the majority of these lncRNAs, including EPB41L4A-AS2, ILF3-AS1, LINC00938, LINC00667 and A2M-AS1, have not been reported in human cancers.

In the present study, the association of these abnormally expressed lncRNAs with patient stage and survival

time was assessed. It was revealed that the expression of lncRNA-GVINP1, RPL32P3, EPB41L4A-AS2, ILF3-AS1, LINC00938, LINC00667, A2M-AS1, GUSBP11, GGTA1P, BCRP3 and SIGLEC17P was suppressed, and that lncRNA-TMPO-AS1 expression was upregulated in high grade (stage III and IV) compared with low grade (stage II) NSCLC samples. Furthermore, Kaplan-Meier analysis illustrated that the overexpression of GVINP1, A2M-AS1, GGTA1P, SIGLEC17P, ILF3-AS1 and LINC00938, and the downregulation of TMPO-AS1 were associated with longer survival times in patients with LUAD. This demonstrated that these lncRNAs may serve as novel biomarkers for NSCLC.

For the most part, the functions of lncRNAs in cancer remain uninvestigated. Co-expression and bioinformatics



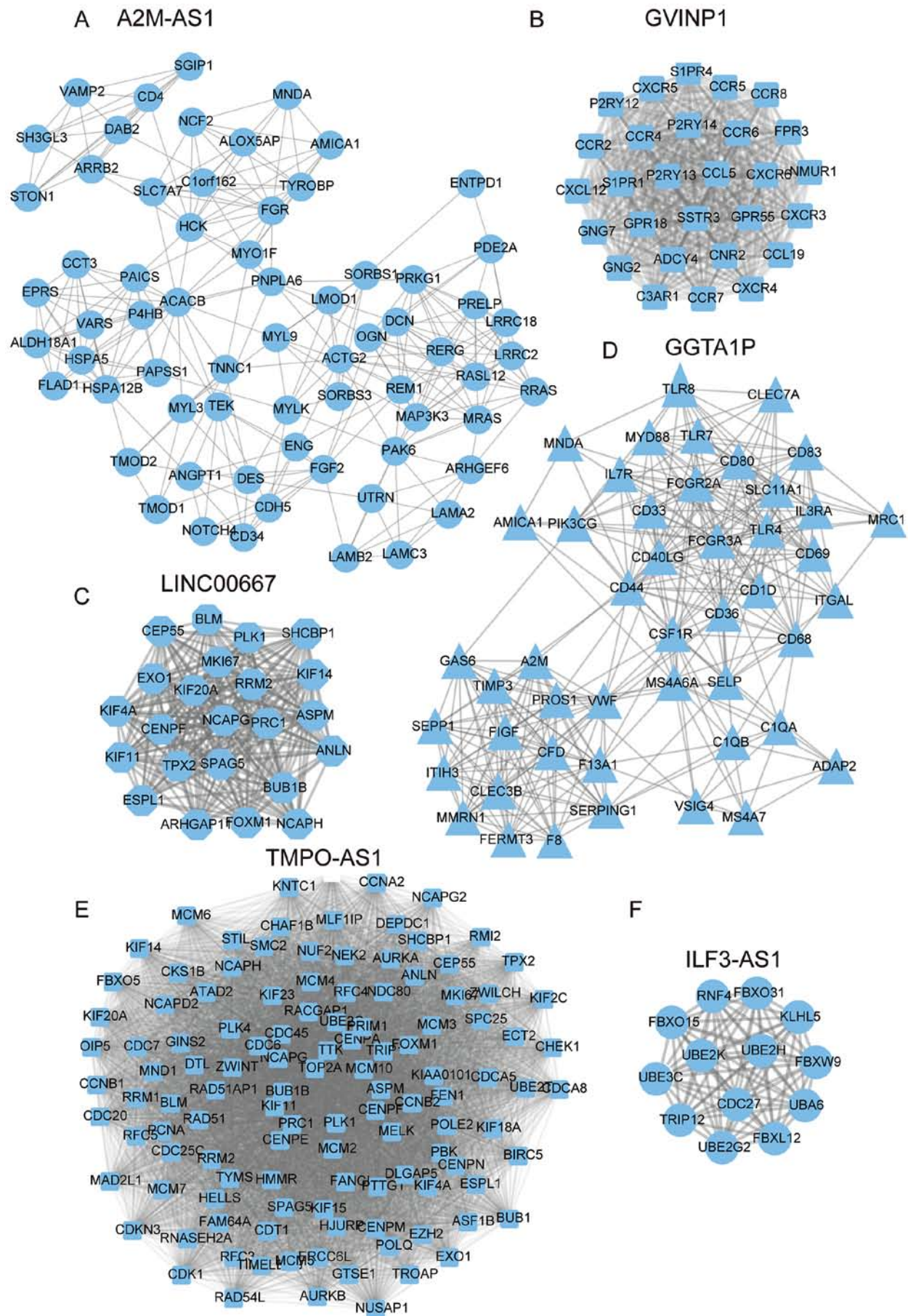


Figure 6. Construction of key lncRNA-associated PPI networks in NSCLC. lncRNA-associated PPI networks for (A) lncRNA-A2M-AS1, (B) GVINP1, (C) LINC00667, (D) GGTA1P, (E) TMPO-AS1 and (F) ILF3-AS1 in NSCLC. lncRNA, long non-coding RNA; PPI, protein-protein interaction; NSCLC, non-small cell lung cancer.



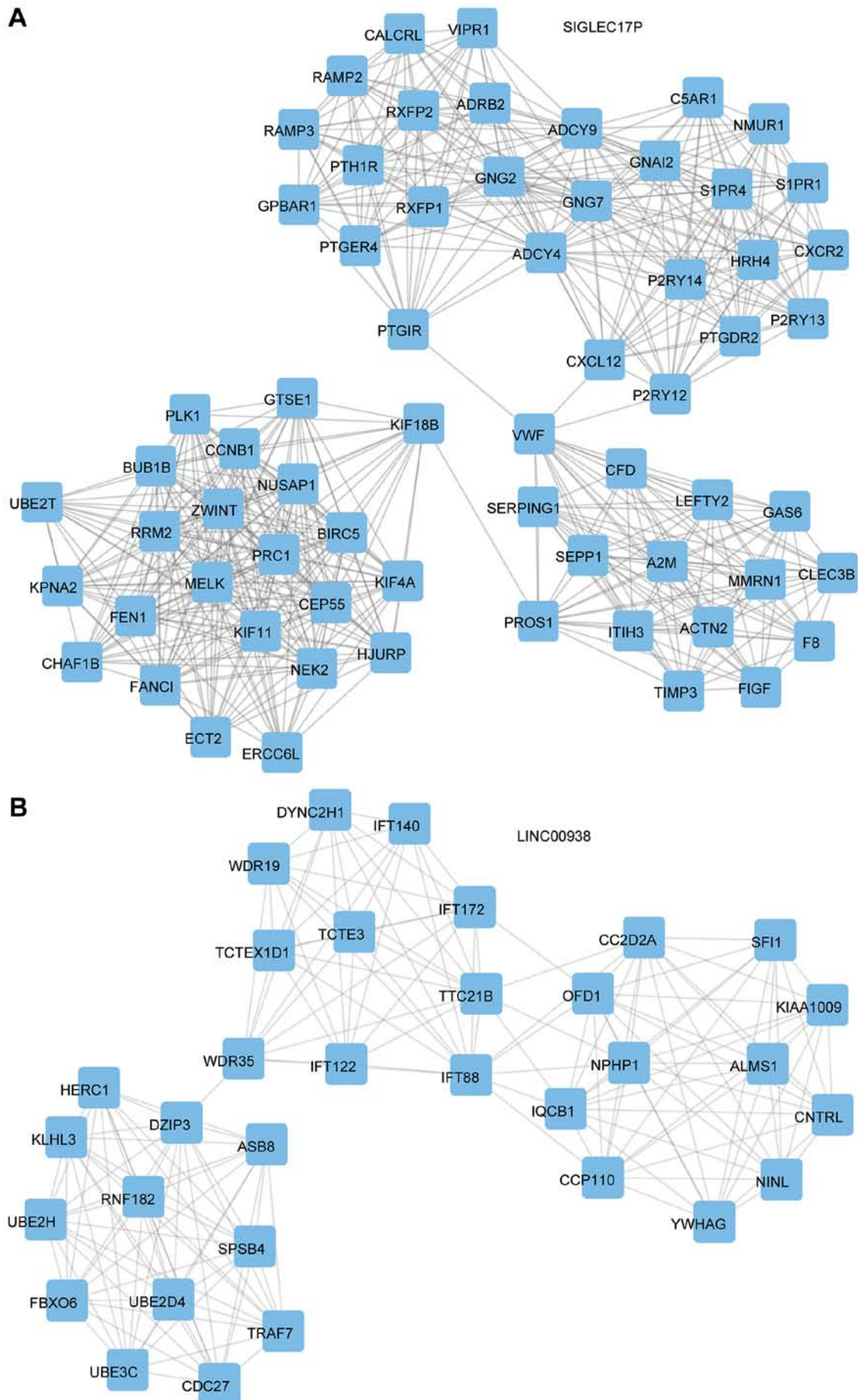


Figure 7. Construction of key IncRNA-associated PPI networks in NSCLC. IncRNA-associated PPI networks for (A) IncRNA-SIGLEC17P and (B) LINC00938 in NSCLC. IncRNA, long non-coding RNA; PPI, protein-protein interaction; NSCLC, non-small cell lung cancer.

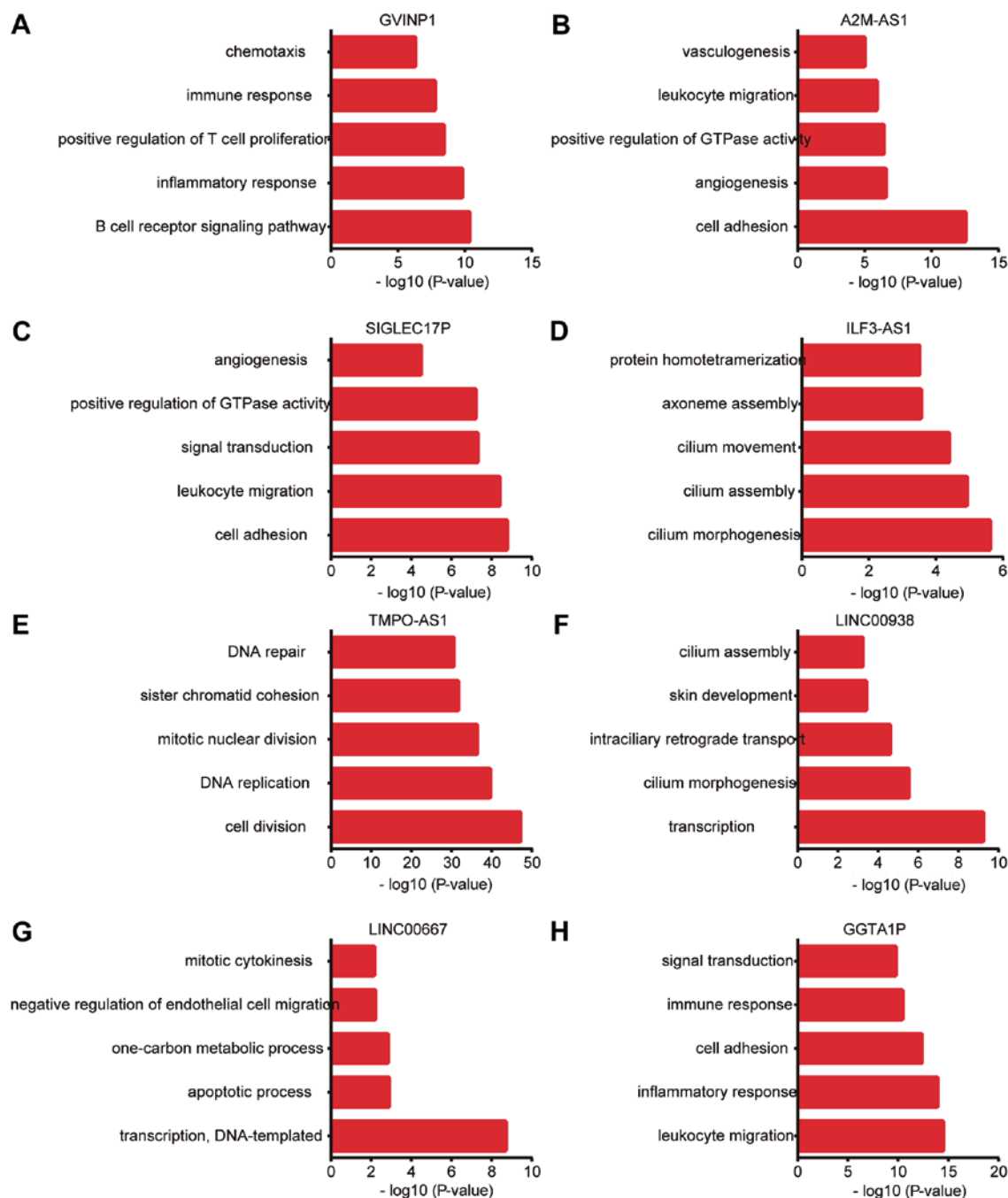


Figure 8. Bioinformatics analysis of the functional roles of key lncRNAs in NSCLC. Bioinformatics analysis revealed the functional roles of (A) lncRNA-GVINP1, (B) A2M-AS1, (C) SIGLEC17P, (D) ILF3-AS1, (E) TMPO-AS1, (F) LINC00938, (G) LINC00667 and (H) GGTA1P in NSCLC. NSCLC, non-small cell lung cancer; lncRNA, long non-coding RNA.

analysis has been widely used to identify the potential functions and molecular mechanisms of novel lncRNAs in human diseases (38,39); using bioinformatics analysis, Jiao *et al* (40) revealed the roles of lncRNAs associated with polycystic ovary syndrome. In the present study, lncRNA-associated PPI modules in NSCLC progression were constructed. GO bioinformatics analysis indicated that lncRNA-LINC00667 was involved in the regulation of transcription and apoptotic process, and that lncRNA-GVINP1 was involved in regulating B cell receptor signaling and the inflammatory response. lncRNA-A2M-AS1 and SIGLEC17P were associated with cell adhesion regulation, and GGTA1P was associated with

leukocyte migration, inflammatory response and immune response. lncRNA-ILF3-AS1 was involved in regulating cilium morphogenesis and cilium assembly, whilst LINC00938 was involved in regulating transcription and cilium morphogenesis, and TMPO-AS1, with cell cycle and DNA repair progression.

The present study identified potential, novel biomarkers for the progression of NSCLC. However, one limitation should be noted; the expression levels of these lncRNAs in NSCLC tumor and serum samples were not detected using clinical samples. Further validation of their expression in NSCLC samples is required, and may strengthen their prognostic value as potential biomarkers for NSCLC.

In conclusion, 38 upregulated, and 31 downregulated NSCLC-associated lncRNAs were identified through comprehensive analysis of public datasets. A total of 12 lncRNAs were associated with NSCLC progression. Furthermore, Kaplan-Meier-plotter analysis illustrated for the first time, that higher expression levels of lncRNA-GVINP1, A2M-AS1, GGTA1P, SIGLEC17P, ILF3-AS1 and LINC00938, and lower expression levels of lncRNA-TMPO-AS1 were associated with longer survival time in patients with NSCLC. Key lncRNA-associated PPI networks were constructed, and bioinformatics analysis performed to provide insights for the identification of novel biomarkers for NSCLC.

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### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

### Authors' contributions

WF and HZ conceived and designed the study. GS acquired the data, and TL analyzed the data. WF and WZ drafted the manuscript and were involved in the conception of the study. LL performed the statistical analysis. All authors read and approved the manuscript and agree to be accountable for all aspects of the research, ensuring that the accuracy and integrity of any part of the work was appropriately investigated and resolved.

### Ethics approval and consent to participate

Not applicable.

### Patient consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

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