

All-trans retinoic acid therapy induces asymmetric division of glioma stem cells from the U87MG cell line

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Received August 7, 2018; Accepted May 17, 2019

DOI: 10.3892/ol.2019.10691

Abstract. The poor therapeutic effect of the current treatments for malignant glioma may be attributed to glioma stem cells (GSCs), which have been demonstrated to divide symmetrically. All-trans retinoic acid (ATRA)-induced differentiation is considered to target GSCs and has been reported to have the capability of eradicating cancer stem cells in specific malignancies. The aim of the present study was to investigate the effects of ATRA on the division mode of GSCs isolated from the U87MG glioblastoma cell line of unknown origin. The expressions of the GSC markers CD133 and nestin were detected using immunocytochemistry to identify GSCs. In addition, the differentiation potency of these GSCs was observed by detecting the expression of glial fibrillary acidic protein, β -tubulin III and galactosylceramidase using immunofluorescent staining. The Numb protein distribution was analyzed in two daughter cells following a GSC division. The results of the present study demonstrated that Numb protein is symmetrically segregated into two daughter cells during GSC division. Furthermore, the present study demonstrated that treatment with ATRA increased the asymmetric cell division of GSCs. In conclusion, these results suggest a therapeutic effect from ATRA-induced asymmetric division of GSCs from the U87MG cell line.

Introduction

Malignant gliomas, which are classified by the World Health Organization as grade III-IV tumors, are extremely lethal primary brain tumors with an incidence of 3-5/100,000 each year between 2000 and 2004 (1). At present, the maximal therapy for malignant gliomas comprises surgical resection followed by radiotherapy and concomitant adjuvant temozolomide (1). However, patients with malignant glioma exhibit a

dismal prognosis (2), with a 5-year survival rate of <3% (3). This poor prognosis is due to therapeutic resistance and tumor recurrence following the combined treatment approach (4). Previously, numerous studies have identified that the resistance of malignant gliomas to current therapy may reside in a relatively small population of glioma stem cells (GSCs), which are hypothesized to substantially influence tumor initiation and serve a critical role in tumor propagation (5-9). The majority of GSCs have been revealed to divide through symmetric cell division and not through asymmetric cell division (10). Recent reports have determined that the frequency of asymmetric division of cancer stem cells is negatively correlated with their proliferative capacity (11,12). A previous study in mouse models have further demonstrated that decreased asymmetry in normal stem cells is associated with neoplastic transformation, while therapies that increase asymmetric division result in decreased numbers of resistant GSCs (13).

All-trans retinoic acid (ATRA)-based differentiation treatment has been observed to target the GSC population and induce therapy-sensitizing and antitumor effects (14,15). Although it has been demonstrated that ATRA-treatment alone is able to induce antitumor effects (5), to the best of our knowledge, it remains unknown whether and how the GSC division mode is affected by treatment with ATRA. Therefore, the present study used a single-cell-based method to observe the differentiation-inducing effects of ATRA on the cell division features of GSCs from the U87MG cell line.

Materials and methods

Culture of U87MG cells. The U87MG cell line (HTB-14; glioblastoma of unknown origin) was purchased from the American Type Culture Collection (Manassas, VA, USA). The cells were maintained in Dulbecco's modified Eagle's medium (DMEM; Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (FBS; Invitrogen; Thermo Fisher Scientific, Inc.) at 37°C in 5% CO₂. The identity of the U87MG cell line was confirmed through comparison with the short tandem repeat profile (Amelogenin, CSF1PO, D13S317, D7S820, D5S818, D16S539, vWA, TH01 and TPOX). This U87MG cells were used in subsequent experiments up to the fifth passage.

Culture of U87MG GSCs. GSCs were isolated as described previously (16). U87MG cells were seeded onto plates at a

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Key words: all-trans retinoic acid, asymmetric division, glioma stem cells, cell line U87

density of 2×10^4 cells/well. The serum-free culture medium (termed tumor stem cell medium) consisted of DMEM/F12 (Gibco; Thermo Fisher Scientific, Inc.), B27 (1x; Gibco; Thermo Fisher Scientific, Inc.), recombinant human epidermal growth factor (20 ng/ml; Sigma-Aldrich, Merck, KGaA, Darmstadt, Germany) and basic fibroblast growth factor (20 ng/ml; Upstate Biotechnology, Inc., Lake Placid, NY, USA) without vincristine. Cultures were incubated in 5% CO₂ at 37°C and half of the medium was replaced with DMEM every 2 days. When primary tumor spheres with diameters of 70–100 μ m were visible under a light microscope, all of the culture medium was discarded and the wells were filled with 1 ml fresh serum-free tumor stem cell medium.

The primary tumor spheres were dissociated enzymatically using Accutase (Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol, into single cells and seeded into a 96-well plate ($2\text{--}3 \times 10^3$ cells/well) with tumor stem cell medium (0.1 ml/well). The culture medium was changed every 3 days. The clonogenic efficacy was examined by calculating the percentage of the wells that contained spheres with >50 cells on day 14. Five fields of vision under a light microscope (x20 magnification) were randomly selected to count the number of spheres. Adherent non-sphere-forming cells and parental U87MG cells were used as controls for the capacity to form tumor cell clones. The morphology of the secondary spheres was examined microscopically. Five fields were randomly selected to examine the morphology of the secondary spheres under a light microscope (x100 magnification).

Immunostaining. GSCs were plated onto poly-L-lysine (Sigma-Aldrich; Merck, KGaA)-coated glass coverslips for 40 h at 37°C in DMEM containing 10% FBS. The cells were fixed with 4% paraformaldehyde (PFA) for 30 min at 4°C and then incubated with primary antibodies against nestin (mouse monoclonal IgG1; 1:200; cat. no. 60091; Stemcell Technologies, Inc., Vancouver, BC, Canada), β -tubulin III (mouse monoclonal IgG1; 1:1,000; cat. no. 60052; Stemcell Technologies, Inc.), glial fibrillary acidic protein (GFAP, rabbit polyclonal; 1:400; cat. no. Z0334; Dako; Agilent Technologies, Inc., Santa Clara, CA, USA), galactosylceramidase (GALC, rabbit polyclonal; 1:20; cat. no. AB142; EMD Millipore, Billerica, MA, USA) or Numb (rabbit polyclonal; 1:100; cat. no. ab155415; Abcam, Cambridge, UK) overnight at 4°C. Appropriate secondary antibodies (Alexa Fluor® 594 donkey anti-mouse; 1:1,000; cat. no. A-21207; Alexa Fluor® 488 donkey anti-rabbit; 1:500; cat. no. A-21202; Alexa Fluor® 488 donkey anti-mouse; 1:400; cat. no. A-21206; and Alexa Fluor® 594 donkey anti-rabbit; 1:500; cat. no. A-21203; Molecular Probes; Thermo Fisher Scientific, Inc.) were then selected for section incubation at room temperature for 2 h. The cell nuclei were counterstained with 4',6-diamidino-2-phenylindole (DAPI; Sigma-Aldrich; Merck KGaA) for 5 min at room temperature for fluorescence microscopy (x200 magnification).

BrdU incorporation. Cells were cultured in DMEM/F12 medium containing 20 ng/ml EGF (Invitrogen; Thermo Fisher Scientific, Inc.). Once pairs were observed, 10 μ g/ml bromodeoxyuridine (BrdU; Invitrogen; Thermo Fisher Scientific, Inc.) was added to the wells for 8–10 h. The washed cells were fixed in 4% PFA at 4°C for 30 min, stained for Numb (rabbit

polyclonal; cat. no. ab155415; Abcam) overnight at 4°C, and then stained with anti-BrdU antibody (cat. no. ab6326; 1:250; Abcam). The cell nuclei were counterstained with DAPI for 5 min at room temperature for fluorescence microscopy (x200 magnification).

Flow cytometric analysis. For flow cytometric analysis, expanded GSCs were collected and evaluated on an FC500 flow cytometer (Beckman Coulter, Inc., Brea, CA, USA). The GSCs were stained with PE-conjugated CD133 clone AC133 antibody (cat. no. 130-113-670; Miltenyi Biotec, GmbH, Bergisch Gladbach, Germany) at a dilution of 1:50 in PBS with 0.5% BSA containing 20% FcR Blocking Reagent (Miltenyi Biotec, GmbH) at 4°C for 10 min. The control was the non-stained cells in the same buffer. Data were analyzed using FlowJo software version 7.6.1 (Tree Star, Inc., Ashland, OR, USA).

Mitotic image analysis. The fluorescence immunostaining intensity was evaluated for each daughter cell to determine the degree of asymmetric distribution of detected molecules between the two daughter cells. The condensed DNA was revealed via staining with DAPI and the morphology of the phase-contrasted image was used to identify the dividing daughter cells. Using Image-Pro Plus v. 6.2 (Media Cybernetics, Inc., Rockville, MD, USA) with the polygon tool, the integrated fluorescent signals of each defined daughter cell in a dividing pair were determined for the stained markers and DNA.

Differentiation assay. U87MG GSC spheres and adherent non-sphere-forming cells were plated on a sterile 24-well glass slide coated with poly-L-lysine (Sigma-Aldrich, Merck KGaA) in DMEM/F12 neurosphere culture medium. These cells were fed with FBS-supplemented culture medium, and coverslips were stained for GFAP, GALC and β -tubulin III as described above on day 2. The nuclei were visualized using DAPI. Quantification of the positive cells stained with each antibody was evaluated as a percentage of the total number of cells counted under five random fluorescence microscope fields (x200 magnification).

ATRA-treatment. The cells in the culture medium were fed with 10% FBS (vehicle group) or FBS combined with ATRA (Sigma-Aldrich; dissolved in DMSO) at 10 nmol/l (ATRA group) (5).

Pair assay. Cells were subjected to a pair assay as described previously (17). In brief, cells were plated at a density of 2,000–3,000 cells/well for 40 h and the cell pairs were observed using time-lapse microscopy, fixed with 4% PFA at 4°C for 30 min, and subjected to immunostaining with Numb primary and secondary antibodies as described above. Nuclei were stained with DAPI, and a total of 200 cell pairs were analyzed using a fluorescence microscope (x200 magnification; Fig. 1).

Statistical analysis. Since paired daughter cells should have equal DAPI, the percentage deviation of the DAPI distribution was calculated in order to quantify the asymmetry, according to the protocol outlined in a previous study (12). For

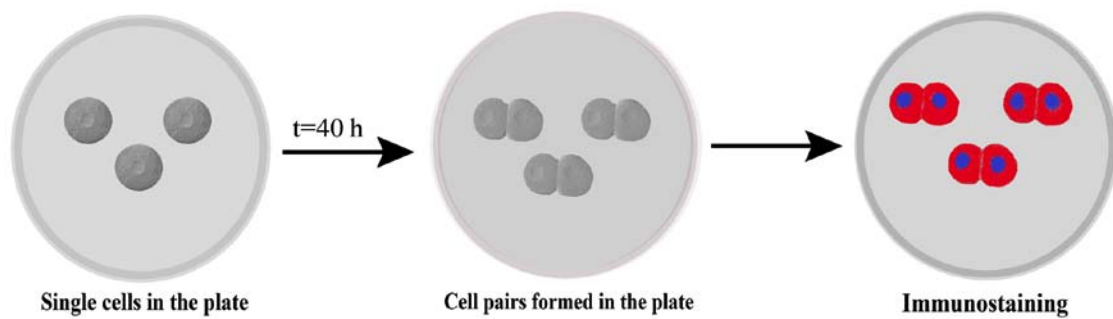


Figure 1. Schematic representation of pair assay. Single cells plated under proliferative conditions enter mitosis and form cell pairs, which were fixed and immunostained. Cell pairs in which proteins partitioned to one daughter cell were considered asymmetrical.

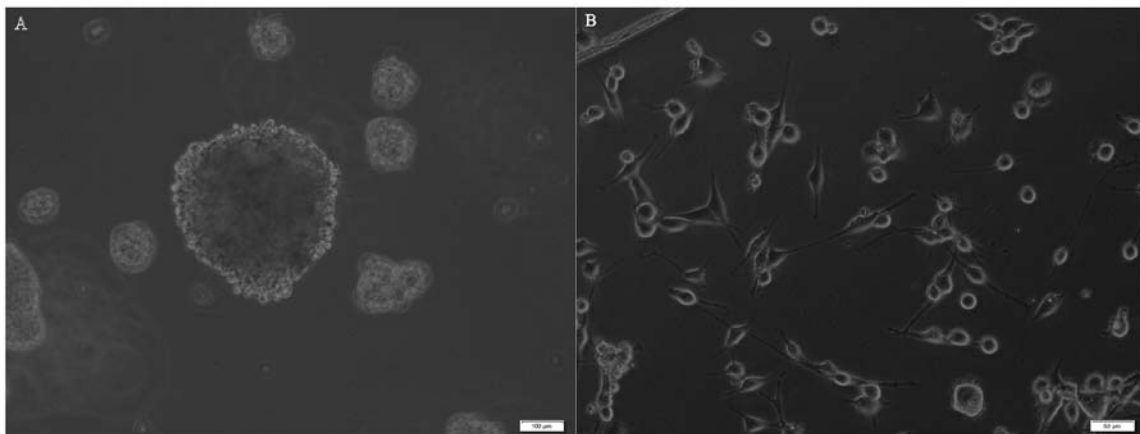


Figure 2. Morphology of tumor spheres. (A) Primary tumor spheres formed in the medium without vincristine. Scale bar, 100 μm . (B) Adherent non-sphere-forming cells. Scale bar, 50 μm .

a minimum of 100 cells, the mean distribution of DAPI was calculated to be 6.848% and the standard deviation was 5.501. Then, the asymmetry cutoff was set to be a difference of >21% between paired daughter cells, based on the >99% confidence interval for DAPI. Based on the 21% cutoff value, each cell division was defined as either symmetrical or asymmetrical.

All experiments were repeated in triplicate, and the data are presented as the mean \pm standard deviation. The data were processed using SPSS 22.0 statistical software (IBM Corp., Armonk, NY, USA). Unpaired Student's t-test was used to determine the significance between groups. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Detection of tumor sphere forming cells with self-renewal capability. U87MG cells were seeded on plates at a density of 2×10^4 /well with progressive increases in the serum-free neural stem cell culture medium. After 5-6 days, 30-50 small spheres per well (each containing 10-20 cells) were observed. At 2 weeks, these spheres had increased their diameters by 10 to 30-fold (Fig. 2A). Numerous adherent non-sphere-forming cells were observed in the bottoms of the wells (Fig. 2B), and remained viable during an 8-week period; however, they did not generate new spheres. Cells in tumor spheres could be passaged for many generations in serum-free neural stem cell culture medium.

Expression of neural stem cell markers. CD133 and nestin are cell surface markers for neural stem cells and are the widely accepted markers for brain GSCs (11). Flow cytometric analysis revealed that these cells had no presence of CD133 on the surface (Fig. 3A and B). The retinoblastoma cell line was used as a positive control (Fig. 3C). The data demonstrated that nestin was positive in the cells from the tumor secondary spheres (Fig. 3D-I).

Multipotency in the differentiation of secondary spheres. An important feature of GSCs are their differentiation capability. Following culture in medium with 10% FBS for 2 weeks, the cells differentiated from tumor secondary spheres were stained positive for GFAP (99.65 ± 7.42 vs. $97.37 \pm 6.06\%$; $P > 0.05$), β -tubulin III (71.25 ± 3.45 vs. $25.33 \pm 3.18\%$; $P < 0.05$) and GALC (19.23 ± 1.41 vs. $10.87 \pm 2.01\%$; $P < 0.05$) compared with the control cells (Fig. 4). Therefore, secondary spheres derived from the single parental cells of primary tumor spheres were able to differentiate into three neural cell lineages.

Asymmetric distribution of Numb is associated with asymmetric GSC divisions. The present study utilized time-lapse lineage tracing to monitor GSC cell divisions at a single-cell resolution (Fig. 5). Cell divisions were recorded using time-lapse microscopy, and cells were subsequently fixed and stained for Numb. Following a 40-h culture, the present study identified the cells that had divided once and generated a cell

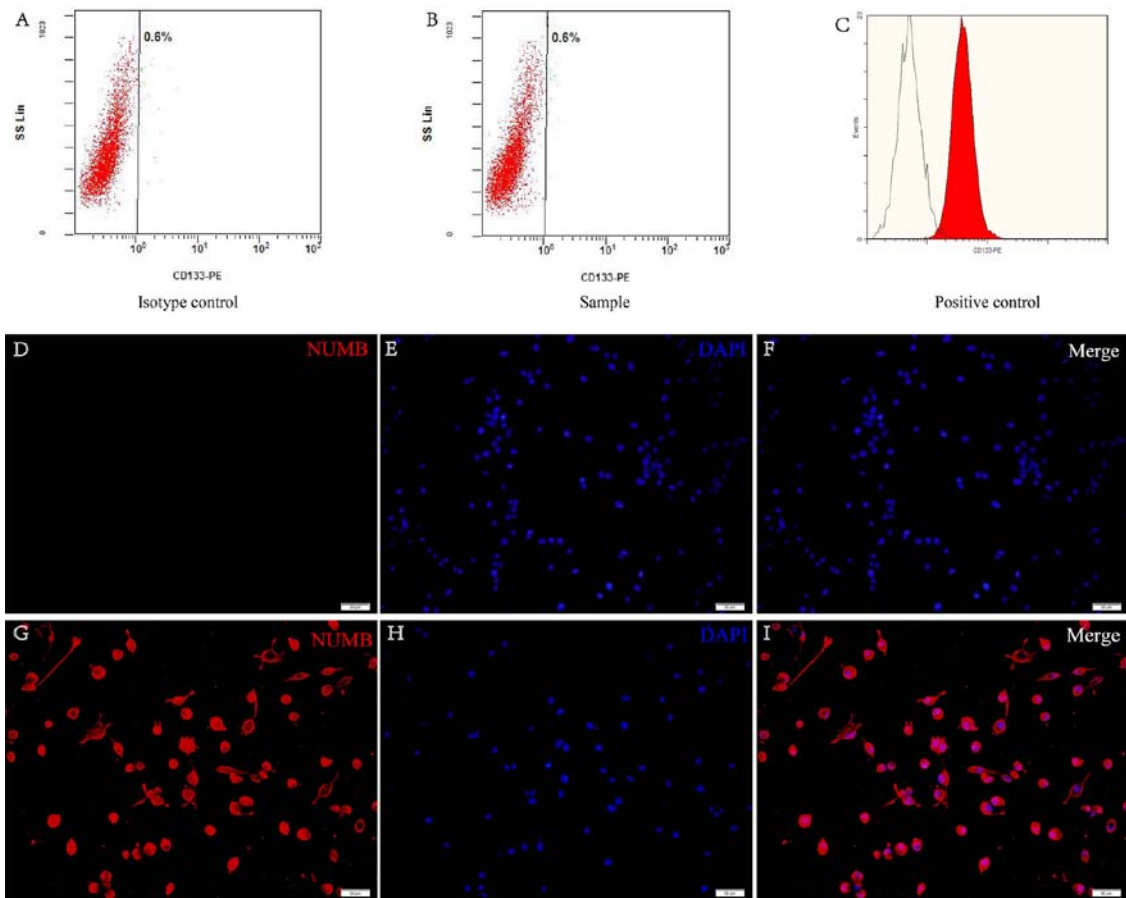


Figure 3. Expression of neural stem cell markers. Characterization of the flow-sorted CD133-negative populations. Flow cytometric analysis demonstrated the (A) isotype control and the (B) CD133-negative cells of the samples that were sorted by FACS. (C) The retinoblastoma cell line was used as a positive control. (D-I) The expression of nestin on neurospheres under fluorescence microscopy. (D-F) Negative expression of nestin in adherent cells. (G-H) Positive expression of nestin in glioma stem cells. Nestin (red) and DAPI (blue). Scale bars, 50 μ m. FACS, fluorescence activated cell sorting.

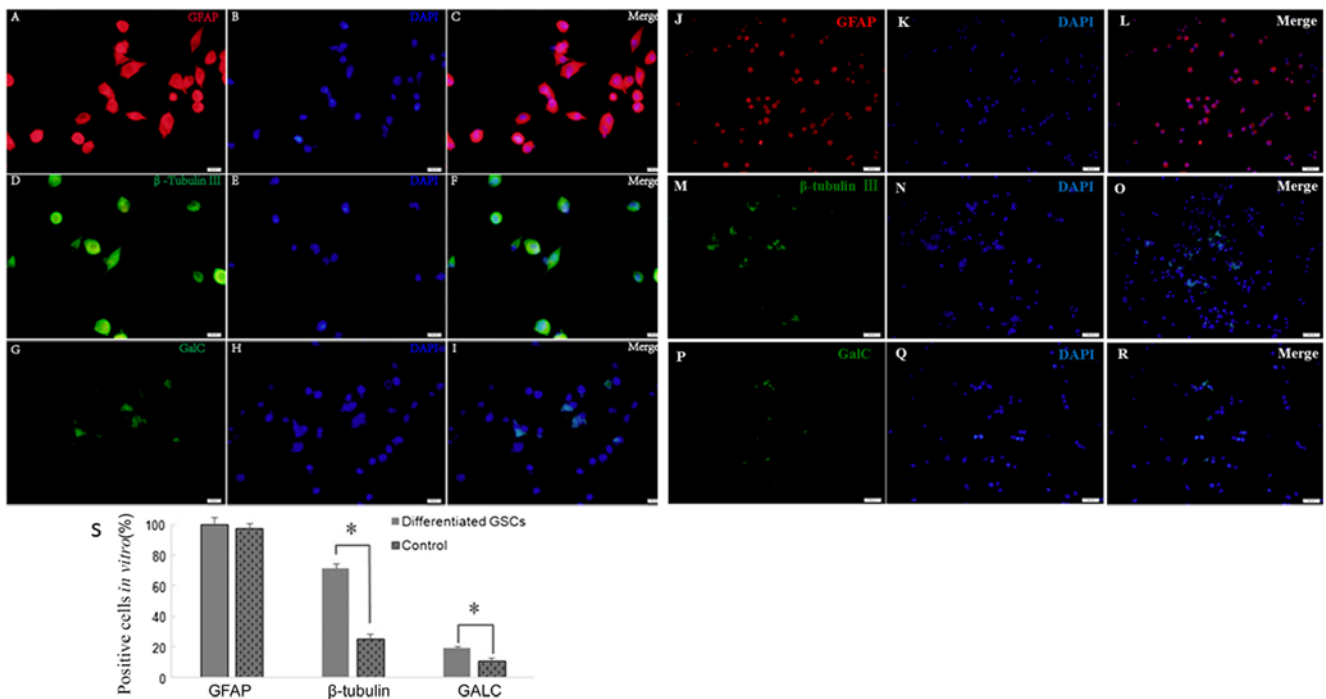


Figure 4. Multipotency of neurosphere cell differentiation observed under a laser confocal scanning microscope. (A-I) Cells were stained for (A-C) GFAP (red), (D-F) β -tubulin III (green) and (G-I) GALC (green) after GSC differentiation. DAPI (blue) indicates nuclei. Scale bars, 20 μ m. (J-L) GSCs were stained for (J-L) GFAP (red), (M-O) β -tubulin III (green) and (P-R) GALC as control. (S) Quantification of cells positively stained by each antibody. * $P < 0.05$. Scale bars, 50 μ m. GFAP, glial fibrillary acidic protein; GALC, galactosylceramidase.

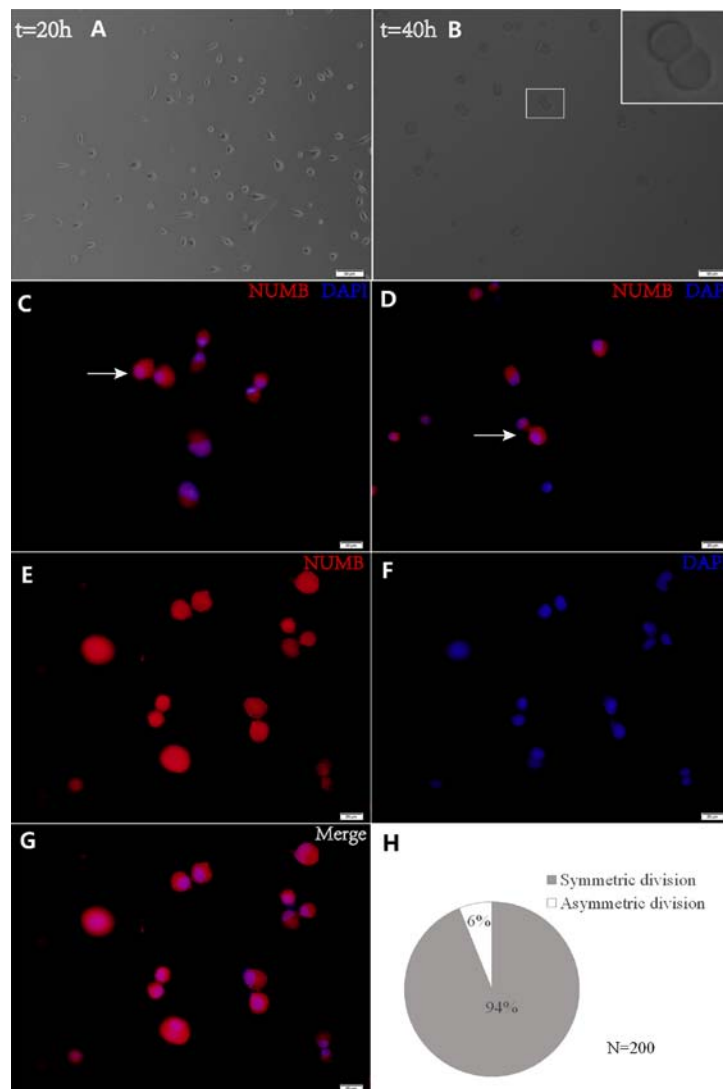


Figure 5. Time-lapse lineage tracing detected cell division. Cell divisions were recorded using time-lapse video microscopy for (A) 20 h and (B) 40 h. Scale bars, 50 μ m. The white box in (B) indicates a pair of daughter cells from a GSC, which is presented at a higher magnification (x800) in the inset. The representative cell pairs with Numb (C) symmetry and (D) asymmetry immunostained for Numb and DAPI (white arrow). (E-G) GSCs are primarily maintained by symmetric cell divisions. Scale bar, 20 μ m. (H) Percentage of Numb distribution features (asymmetric or symmetric) observed from 200 pairs of cells when the GSCs had divided once. GSC, glioma stem cell.

pair (Fig. 5B). Through this analysis, the Numb distribution modes were detected in the daughter cells (Fig. 5C and D). Sister cells with symmetric Numb usually appeared similar. In order to quantify the asymmetry, asymmetrical division was defined as having occurred when the percent deviation of the stained marker between daughter cells was $>21\%$. Using this criterion, it was discovered that the majority of pairs (93.91 ± 1.59 vs. $6.09 \pm 1.57\%$) exhibited symmetry in the Numb distribution (Fig. 5H). These data demonstrate that GSCs are primarily maintained by symmetric, as opposed to asymmetric, cell divisions (Fig. 5E-G). Cells were then treated with BrdU for 8 h and the BrdU distribution was observed in relation to Numb expression by immunostaining in pair assays. The data revealed that BrdU incorporation in the two daughter cells was associated with Numb asymmetry (Fig. 6).

ATRA increased the asymmetric cell division of GSCs. To investigate whether symmetric divisions are affected when ATRA is administered, the present study observed the division

mode when ATRA was added to the culture. Neurospheres were grown in DMEM containing 10% FBS and 10 nmol/l ATRA to induce differentiation. Cell divisions were recorded for 40 h using time-lapse video microscopy. Pairs of daughter cells from single progenitor cells were identified, fixed and stained for Numb. There was a decrease in symmetrical stem cell expansion division incidence to $18.30 \pm 1.43\%$ of the events in the ATRA-treated cells (Fig. 7). This suggests that the cancer stem cells were impaired in their ability to undergo a symmetric division.

Discussion

The results of the present study demonstrated that the GSCs isolated from the U87MG cell line did not express CD133. Using single-cell-based analytical methods and quantitative immunofluorescence, the present study revealed that Numb was widely expressed in GSCs and symmetrically segregated into two daughter cells during the division of GSCs. The

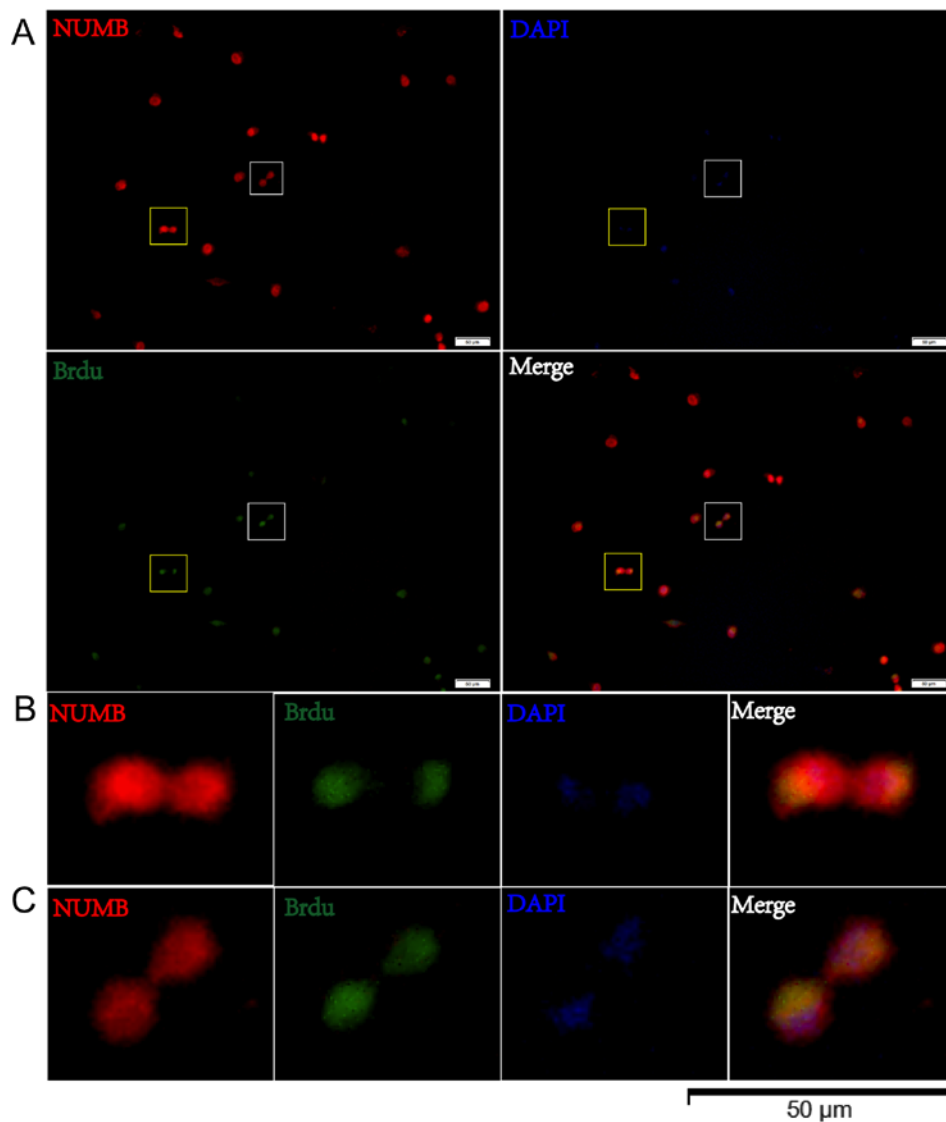


Figure 6. BrdU distribution in paired cells. (A) BrdU incorporation in two daughter cells is associated with Numb asymmetry. The white box in (A) indicates a pair of daughter cells from a GSC with asymmetric BrdU incorporation, which is presented at a higher magnification in (B). The yellow box in (A) indicates a pair of daughter cells from a GSC with symmetric BrdU incorporation, which is presented at a higher magnification in (C). GSC, glioma stem cell.

present study, to the best of our knowledge, is the first to demonstrate, that the Numb symmetrical distribution proportion was decreased following ATRA-based differentiation treatment.

GSCs have been identified in numerous primary culture GBM cells and malignant glioma cell lines (5,18-21). A previous study has confirmed that GSCs are the source of new tumor cells following the administration of temozolomide (22). The universal GSC marker remains a controversial topic due to high interpatient as well as intratumoral variability (8). Various surface markers, including CD133, nestin, Musashi-1, Nanog and platelet-derived growth factor receptor, have been reported to successfully identify GSC populations *in vitro* and *in vivo* (8,14). Additional stem cell marker detection would further support our conclusions. The present study analyzed CD133 expression using flow cytometry and identified that CD133 was negative in glioma cell spheres cells cultured from the U87MG cell line. This result differs from that of previous research, which reported that the majority of U87MG cells

in the spheres were positive for CD133 (20). Further results using immunofluorescence revealed that the CD133-negative cell populations expressed nestin. In addition, the cell populations of the cultured tumor spheres were able to differentiate into cells positive for GFAP, β -tubulin III and GALC, which are representative markers of neuronal, astroglial and oligodendroglial cells (23). These results suggested successful induction of GSCs from the U87MG cell line. However, the lack of an exact evaluation of stemness/differentiation marker expression levels is a limitation of the present study. Although CD133 has been defined as a marker of glioma stem cells, an increasing amount of evidence has demonstrated that the use of CD133 as a unique glioma stem cell marker is insufficient to tag all GSCs. For example, fresh human glioma and gliomasphere cultures express CD133 at low and sometimes barely detectable levels (21). Secondly, CD133-positive and CD133-negative GSCs from cell lines and GBM tumors exhibited cancer stem cell properties (20,24). Thirdly, neither the expression of stemness genes nor the long-term self-renewal

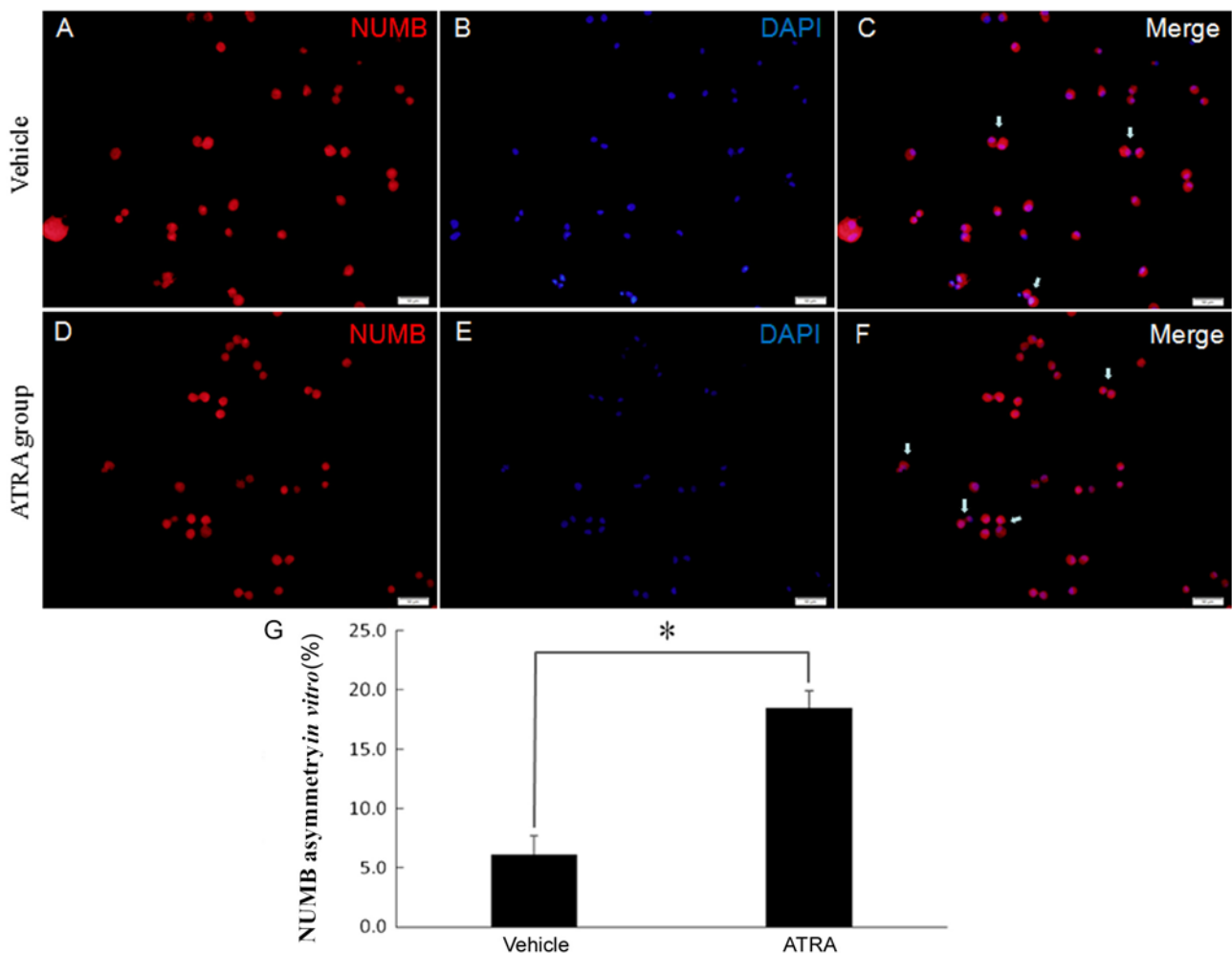


Figure 7. ATRA induces asymmetric cell divisions of GSCs. (A-F) Numb (red) distribution was used to represent GSC division mode in (A-C) vehicle group and (D-F) ATRA group. Cells were counterstained with DAPI (blue) to mark nuclei. White arrows indicate asymmetric pairs. Scale bars, 50 μ m. (G) Quantification of the asymmetric cell pairs demonstrated that treatment with ATRA increased the fraction of cell pairs with an asymmetric Numb distribution. * $P < 0.05$. GSC, glioma stem cell; ATRA, all-trans retinoic acid.

capacities of CD133-positive and CD133-negative cells were significantly different (25). Finally, CD133 negative cells were tumorigenic when implanted into rat brains (26). A previous study demonstrated that the levels of surface CD133 fluctuate during the cell cycle in GSCs (27), indicating that CD133 expression is likely a marker of certain stages of GSC division, rather than a constitutive marker of GSCs.

Lathia *et al* (10) examined a variety of molecules in GSCs and observed that only Numb and CD133 could be asymmetrically segregated. Since the results of the present study demonstrated that CD133 expression was negative in GSCs cultured from the U87MG glioblastoma of unknown origin cell line, the present study used Numb to analyze the GSC division mode. The data revealed that Numb protein was expressed in 99% of GSCs from the U87MG cell line. Using single-cell-based observations, the current study demonstrated that the Numb distribution was predominantly symmetric in the two daughter cells (94%) during GSC division. BrdU incorporation indicates the proliferative ability of cells that were actively replicating their DNA. The results of the present study demonstrated that the BrdU distribution in the two daughter cells was associated with Numb asymmetry. A limitation of the present study is that the exact level of BrdU in paired cells was not measured.

In paraffin-embedded glioblastoma specimens, a previous study indicated that 85% of cells exhibited a symmetric pattern of Numb immunoreactivity (28). Numb is a so-called fate-determining molecule that promotes the differentiation of neural stem cells through antagonizing the notch and hedgehog signaling pathways (29,30). The function of Numb is critical for the occurrence of asymmetric cell division, and different expressions of Numb may indicate cell fate divergence (31). Previous studies have suggested that symmetric determinants exert pivotal functions in tumor initiation, as defects in either the function of fate determinants and regulators of asymmetric division, or the loss of asymmetric division may lead to tumor development (13,32). Although previous data demonstrated that the overexpression of Numb did not induce either differentiation of U87MG cells or alter their morphology, nor that the cell population doubling time was significantly affected (33), until now, numerous findings have demonstrated that Numb is associated with the GSC markers SRY (sex determining region Y)-box 2 and paired box protein Pax-6, as well as GSC survival, proliferation, aggressiveness and therapeutic resistance (12,28,34).

The present results demonstrated a decrease in symmetrical cell division incidence to 82% of the events in ATRA-treated

GSCs from the U87MG cell line. ATRA is a metabolite of vitamin A that is able to induce complete remission in the majority of acute promyelocytic leukemia cases when administered in combination with light chemotherapy and/or arsenic trioxide (35). To date, the efficacy of ATRA to treat solid tumors remains poor (35). A previous study demonstrated that ATRA could reduce the proliferation and invasiveness of U87MG stem-like glioma cells in a dose-dependent manner and decrease tube formation and vascular endothelial growth factor secretion (36). When ATRA was administered in combination with mTOR and PI3K inhibition, there was a synergistic effect resulting in the minimum amount of cellular migration in GSCs (15). It has been revealed that ATRA-based differentiation targeted GSCs and induced GSCs to differentiate, which was mediated by the activation of ERK1/2 (37). The results of the present study demonstrated that differentiation treatment with ATRA induced asymmetric division in GSCs from the U87MG cell line. The underlying mechanism may be associated with the distribution of Numb (12,28,31). In a future study, we aim to investigate how ATRA affects the distributions of p53, E-cadherin and Notch in the paired GSCs in order to understand this underlying mechanism. Asymmetric division is a feature more closely associated with cancer stem cells from early-stage and well-differentiated tumors, while late stage tumors suppress asymmetric division and increase symmetric division (38). Therefore, the increase of asymmetric division following treatment with ATRA may be associated with the suppression of tumorigenesis in GSCs. However, from the present evidence it cannot be concluded that ATRA can be used as a glioblastoma treatment agent.

Although ATRA induces morphological differentiation of GSCs *in vitro*, one of the primary limitations in the present study is that it remains controversial as to whether the cell line U87MG is representative of highly malignant glioma. Therefore, further experiments on primary cell lines are required. Secondly, ATRA-metabolizing enzymes (39) and loss of retinoic acid receptors (40) may render cells insensitive to ATRA effects. Additional studies are necessary in order to investigate how ATRA influences the GSC division mode *in vivo*. In conclusion, the results of the present study demonstrated that ATRA-induced asymmetric division of GSCs from the U87MG glioblastoma cell line of unknown origin.

Acknowledgements

Not applicable.

Funding

This study was supported by the Shaanxi Province Scientific and Technology Development Program (grant no. 2012SF2-03-05) and the Xi'an Jiaotong University Free Exploration Program (grant no. xjj2013059).

Availability of data and materials

The datasets used or analyzed during the current study are available from the corresponding author upon reasonable request.

Authors' contributions

RW designed the study and performed the majority of experiments. CL performed the GSC cultures and immunostaining. RW was a major contributors to the writing of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of the Second Hospital of Xi'an Jiaotong University (Xi'an, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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