# High expression of fibronectin 1 indicates poor prognosis in gastric cancer

YANG SUN<sup>1,2</sup>, CHUNLIN ZHAO<sup>1</sup>, YANWEI YE<sup>1</sup>, ZHEN WANG<sup>1</sup>, YUANHANG HE<sup>1</sup>, YULIN LI<sup>1</sup> and HAOXUN MAO<sup>1</sup>

<sup>1</sup>Department of Gastrointestinal Surgery, The First Affiliated Hospital of Zhengzhou University, Zhengzhou, Henan 450052; <sup>2</sup>Department of Breast and Thyroid Surgery, Nanyang Central Hospital, Nanyang, Henan 473000, P.R. China

Received December 13, 2018; Accepted August 1, 2019

DOI: 10.3892/ol.2019.11088

Abstract. Fibronectin 1 (FNI) is involved in the occurrence and development of various tumors and is upregulated in multiple cancer types. FNI has been demonstrated to promote cell proliferation and migration in gastric cancer cell lines. However, the relationship between the expression of FN1 and clinicopathological factors and prognosis is not clear in gastric cancer (GC). The aim of the present study was to investigate the association between FN1 expression and clinicopathology and prognosis of gastric cancer. In this study, 17 publicly available GC cohorts (n=2,376) with gene expression data from the Gene Expression Omnibus (GEO), The Cancer Genome Atlas (TCGA) and Oncomine databases were tested. In addition, FN1 protein expression was validated by immunohistochemistry in a separate cohort (n=190). The meta-analysis results demonstrated an increase in FNI expression at the protein and mRNA level in GC tissues, and the FNI gene was highly expressed at the mRNA level in the advanced T stage (T2 + T3 + T4) group compared with that in the early T stage (T1) group. In addition, the expression of epithelial FNI at the protein level was positively correlated with tumor size. FN1 expression at the protein and mRNA level was a predictor of poor prognosis following radical resection of GC. In conclusion, the expression of FN1 in GC tissues is upregulated compared with adjacent normal tissues, and it is a potential biomarker of poor prognosis in patients with GC.

# Introduction

Gastric cancer is the fifth most commonly diagnosed cancer (5.7% of total cases) and the third leading cause of cancer mortality (8.2% of total cancer mortality) worldwide (1).

E-mail: doctorzhaochunlin@126.com

Gastric cancer is also the third leading cause of cancer-related mortality in China (2). Although surgery combined with radiotherapy, chemotherapy and targeted therapy prolongs survival, the 5-year overall survival rate of patients with advanced gastric cancer remains poor. The 5-year overall survival rates of patients with pathological T stage 2, 3 and 4 disease were 68.3, 33.0 and 24.0% respectively (3,4). Therefore, new biomarkers of gastric cancer to determine prognosis are necessary.

Fibronectin 1 (FNI) mediates the interaction between cells and the extracellular matrix and serves an important role in cell adhesion, migration, growth and differentiation (5). FNI is a ligand for numerous members of the integrin receptor family (6). FN1 is involved in the occurrence and development of various tumors. FN1 activates the PI3K/Akt pathway by binding to its integrin receptor  $\alpha 5\beta 1$  in breast cancer (7). In addition, FNI has been demonstrated to promote cell proliferation and migration in esophageal squamous cell carcinoma, oral squamous cell carcinoma (OSCC), nasopharyngeal carcinoma, colorectal, ovarian, renal and thyroid cancer (8-14). However, little is known about the expression of FN1 in gastric cancer. FN1 is upregulated in GC tissues compared with normal gastric tissues (15). FNI knockdown inhibits cell migration and invasion in vitro, and FOXF1 adjacent non-coding developmental regulatory RNA and microRNA-200c promote the proliferation, migration and invasion of GC cells by negatively targeting FN1 (15-17). Overall, FN1 is a potential biomarker candidate for GC prognosis, but the relationship between FNI expression and clinical factors and prognosis has not been reported, and thus it is necessary to verify and clarify the role of FN1 in GC.

The aim of the present study was to investigate *FN1* gene expression in GC and its association with clinicopathological factors and prognosis by examining 17 publicly available GC cohorts. Furthermore, *FN1* protein expression was validated by immunohistochemistry in a separate cohort. The results demonstrated that *FN1* may serve as a new prognostic marker for GC.

#### Materials and methods

*Data collection.* Microarray data were downloaded from the following datasets in the Gene Expression Omnibus (GEO;

*Correspondence to:* Professor Chunlin Zhao, Department of Gastrointestinal Surgery, The First Affiliated Hospital of Zhengzhou University, 1 East Jianshe Road, Erqi, Zhengzhou, Henan 450052, P.R. China

*Key words:* fibronectin 1, gastric cancer, prognosis, The Cancer Genome Atlas cohort, Gene Expression Omnibus cohort

http://www.ncbi.nlm.nih.gov/geo/), The Cancer Genome Atlas (https://xenabrowser.net/datapages/?cohort=TCGA) and Oncomine (https://www.oncomine.org/resource/login.html): GSE13861, GSE13911, GSE14208, GSE15456, GSE15459, GSE19826, GSE26253, GSE26899, GSE26901, GSE29272, GSE34942, GSE35809, GSE54129, GSE66229, GSE79973, Chen Gastric and TCGA STAD. Several of these datasets have been previously published (18-32). The 17 datasets comprised 2,376 cancer tissues and 294 adjacent normal tissues. Datasets with no clinical data (GSE13861, GSE13911, GSE19826, GSE54129, GSE79973 and Chen Gastric), GSE29272 and TCGA STAD were used to analyze the differences between tumor and adjacent tissues. The remaining datasets were used to analyze the relationship between FN1 expression and clinicopathological factors. Clinical information for the cohorts with respective clinical data included in this study is presented in Table I.

Validation dataset. Immunohistochemistry (IHC) was used for validation. Gastric cancer tissues and adjacent normal gastric tissues were obtained during surgery from 190 randomly selected patients between June 2011 and June 2012 at the First Affiliated Hospital of Zhengzhou University (Zhengzhou, China). The study was approved by the ethics committee of the First Affiliated Hospital of Zhengzhou University. Written informed consent was obtained from all patients. The tissues were fixed with formalin and embedded in paraffin for subsequent experiments. All patients were followed up for  $\geq$ 5 years, and 102 succumbed to any cause during the follow-up period.

IHC. Formalin-fixed paraffin-embedded tissue samples from the IHC cohort were sliced into 4-um sections. A mouse monoclonal antibody against FN1 (cat. no. 66042-1-Ig; ProteinTech Group, Inc.) was used at a 1:600 dilution at pH 9.0. The immunohistochemical staining of the specimens was performed as previously described (16). The results of FN1 expression were separately scored in epithelial cancer cells and intertumoral stroma. The scoring method described by Sung et al (33) was used. For epithelial FN1 (E-FN1) expression, staining intensity and the proportion of stained tumor cells were considered. Staining intensity was classified as follows: 1, weak; 2, moderate; and 3, strong. Positive cells were quantified as a percentage of the total number of tumor cells and assigned to one of the following categories: 0, <5%; 1, 5-24%; 2, 25-49%; 3, 50-74%; and 4, ≥75%. The percentage of positive tumor cells and staining intensity were multiplied to generate an immunoreactivity score (IS) for each case. IS values ranged from 0 to 12; IS $\geq$ 3 was considered positive, whereas IS<3 was considered negative. Stromal FN1 (S-FN1) expression was graded into three categories: No or weak staining, no staining or a low number of FNI-positive strands; moderate staining, fine FNI-positive strands; and strong staining, coarse FNI-positive strands (34).

Statistical analysis. When >1 *FN1* probe was present in a group, the probe with the highest variance was selected for statistical analysis (35). All *FN1* gene expression data normalization and probe summarization were performed by Robust Multichip Analysis and transformed by log2. SPSS 22.0 (IBM Corp.) and RevMan 5.3 (Cochrane Community) were used to perform all statistical analyses.

Independent sample t-tests were used in SPSS for continuous data analysis and Pearson's  $\chi^2$  tests were used for categorical data analysis. The gene expression value was equal to three,  $\geq 1/3$  were defined as high expression and the <1/3 as low expression. Overall survival (OS) rate was analyzed using Kaplan-Meier plots and the log-rank test or Gehan-Breslow-Wilcoxon test. When the two survival functions were parallel, the log-rank test was used, whereas the Gehan-Breslow-Wilcoxon test was used if the data crossed over. A Cox regression model was used to assess the hazard ratio (HR) and perform multivariate analysis. All tests were two-sided, and P<0.05 was considered to indicate a statistically significant difference.

Meta-analyses were performed using RevMan 5.3. First, the heterogeneity between the results of each study was analyzed by the  $\chi^2$  test. The threshold was set to  $\alpha$ =0.100, and the extent of heterogeneity was assessed by combining I<sup>2</sup>. If P>0.10 and I<sup>2</sup>≤50%, the homogeneity between the results was considered high, and the fixed effect model was used; if P≤0.10 or I<sup>2</sup>>50%, the random effects model was used.

# Results

*Patient cohorts*. Data from 17 independent GC cohorts were downloaded from the Gene Expression Omnibus (GEO), The Cancer Genome Atlas (TCGA) and Oncomine, including 2,670 samples, which comprised 2,376 cancer tissues and 294 adjacent normal tissues. Eight of the 17 cohorts included tumor and normal samples. The IHC cohort comprised 190 GC samples and 20 adjacent tissue samples. The clinicopathological characteristics of the patients are presented in Table I.

*FN1 expression in gastric cancer.* A total of eight independent cohorts that included expression data from cancer and normal samples were analyzed; the results revealed upregulated *FN1* mRNA levels in tumor tissues compared with normal tissues (Fig. 1A). Meta-analysis of all the cohorts revealed a significant combined mean difference of 1.99 (P<0.001; Fig. 2A). These results indicated that *FN1* expression was significantly higher in GC tissues compared with that in adjacent normal tissues.

Association between FN1 expression and clinicopathological factors. Compared with that in the early T stage (T1) group, the expression of FN1 was significantly increased in the advanced T stage (T2+T3+T4) group (P=0.002; Fig. 1B) in one cohort, which was further confirmed by meta-analysis in all examined cohorts (P<0.001; Fig. 2B). The expression of FN1 was not associated with differentiation in any cohort (Figs. 1C and 2C). Only two cohorts exhibited increased FN1 expression in patients with high clinical Tumor-Node-Metastasis (TNM) stage (36) (III + IV) compared with that in patients with low clinical TNM stage (I + II) (Fig. 1D). No significant differences between patients with high and low TNM stage were observed in the meta-analysis of all cohorts (Fig. 2D).

High FN1 expression level indicates poor clinical outcomes. Kaplan-Meier survival analysis was performed using clinical data. OS analysis demonstrated that high FN1 expression was associated with unfavorable prognosis compared with low FN1

and a component of an and a particular and an an	iorogram cinut											
Characteristic	IHC cohort, n (%)	, GSE13861, n (%)	IHC cohort, GSE13861, GSE15456, GSE15459, $n (\%) = n (\%) = n (\%) = n (\%) = n (\%)$	GSE15459, n (%)	GSE26253, n (%)	GSE26899, n (%)	GSE26901, n (%)	GSE29272, n (%)	GSE34942, n (%)	GSE35809, n (%)	GSE66229, n (%)	TCGA, n (%)
Sex												
Total	190 (100)	65 (100)	30 (100)	192 (100)	432 (100)	92 (100)	109 (100)	134 (100)	56 (100)	70 (100)	300(100)	415(100)
Male	144 (75.8)	46 (70.8)	17 (56.7)	125 (65.1)	280 (64.8)	73 (79.3)	69 (63.3)	103 (76.9)	36 (64.3)	48 (68.6)	199 (66.3)	268 (64.6)
Female	46 (24.2)	19 (29.2)	13 (43.3)	67 (34.9)	152 (35.2)	19 (20.7)	40 (36.7)	31 (23.1)	20 (35.7)	22 (31.4)	101 (33.7)	147 (35.4)
Median age,	59 (25, 85)	63 (32, 83)	73 (53, 83)	66 (23,92)	53 (23, 74)	59 (36, 83)	58 (28, 74)	59 (23, 73)	69 (43, 84)	67 (32, 85)	64 (24, 86)	67 (30, 90)
years (min, max)												
T stage												
1	15 (7.9)	2 (3.1)	ı	8 (4.2)	I	I	I	ı	I	I	2 (0.7)	22 (5.3)
2	32 (16.8)	23 (35.4)	ı	45 (23.4)	I	I	I	I	I	I	186 (62)	88 (21.2)
3	24 (12.6)	34 (52.3)	I	107 (55.7)	ı	I	I	I	I	I	91 (30.3)	181 (43.6)
4	119 (62.6)	1(1.5)	I	1(0.5)	I	I	I	I	I	I	21 (7)	115 (27.7)
Unknown	(0) (0)	5 (7.7)	I	31 (16.1)	ı	ı	ı	ı	ı	I	0 (0)	9 (2.2)
TNM stage												
I	29 (15.3)	12 (18.5)	6 (20)	31 (16.1)	68 (15.7)	11 (12.0)	38 (29.5)	5 (3.7)	11 (19.6)	13 (18.6)	31 (10.3)	57 (13.7)
II	60 (31.6)	2 (3.1)	4 (13.3)	29 (15.1)	167 (38.7)	18 (19.6)	40 (31)	5 (3.7)	11 (19.6)	16 (22.9)	97 (32.3)	123 (29.6)
III	91 (47.9)	35 (53.8)	15 (50)	72 (37.5)	130 (30.1)	27 (29.3)	36 (27.9)	115 (85.8)	19 (33.9)	33 (47.1)	95 (31.7)	169 (40.7)
IV	10 (5.3)	16 (24.6)	5 (16.7)	60 (31.3)	67 (15.5)	36 (39.1)	15 (11.6)	9 (6.7)	13 (23.2)	7 (10)	77 (25.7)	41 (9.9)
Unknown	(0) (0)	(0) (0)	0 (0)	(0) (0)	(0) (0)	(0) (0)	0 (0)	(0) (0)	2 (3.6)	1 (1.4)	(0) (0)	25 (6.0)
Tumor grade												
Low	48 (24)	20 (30.8)	2 (6.7)	6 (3.1)	I	I	I	1(0.7)	I	2 (2.9)	I	12 (2.9)
Intermediate	66 (33)	15 (23.1)	13 (43.3)	53 (27.6)	ı	I	ı	48 (35.8)	I	22 (31.4)	I	148 (35.7)
High	76 (38)	6 (9.2)	15 (50)	86 (44.8)	ı	I	I	85 (63.4)	I	26 (37.1)	I	246 (59.3)
Undifferentiated	10 (5)	24 (36.9)	0 (0)	47 (24.5)	I	ı	I	(0) (0)	ı	20 (28.6)	I	9 (2.2)
Follow-up endpoint (death)												
Occurred	101 (50.5)	ı	I	95 (49.5)	I	ı	ı	31 (23.1)	27 (48.2)	I	159 (53)	144 (34.7)
Not occurred	99 (49.5)	I	I	97 (50.5)	I	I	I	95 (70.9)	29 (51.8)	I	141 (47)	214 (51.6)
No data	0 (0)	I	ı	(0) (0)	I	I	I	8 (6)	(0) (0)	I	0 (0)	57 (13.7)
data not available; TNM, Tumor-Node-Metastasis.	Tumor-N	ode-Metastasis										

Table I. Clinicopathological characteristics of patients in different datasets.

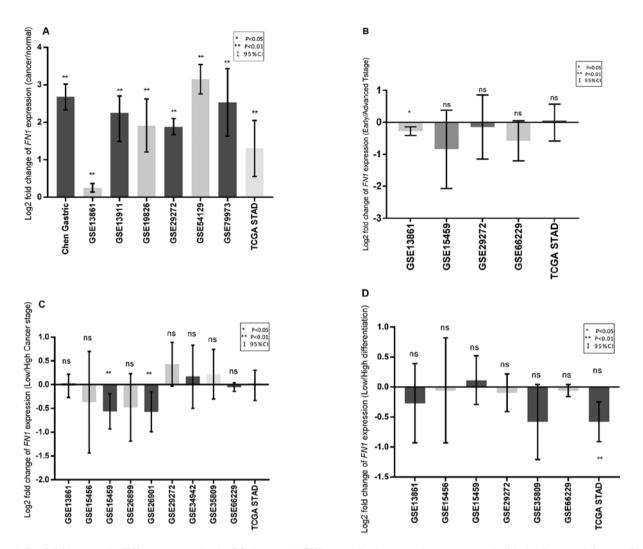


Figure 1. Log2 fold change in *FN1* gene expression in GC tissues. (A) *FN1* expression in tumor tissues compared with that in normal tissues. (B) *FN1* expression in the advanced T stage (T2 + T3 + T4) group compared with that in the early T stage (T1) group. (C) *FN1* expression in the low differentiation group (high tumor grade) compared with that in the high differentiation group (intermediate and low tumor grade). (D) *FN1* expression in the high clinical TNM stage (III+IV) group compared with that in the low clinical TNM stage (I+II) group. Error bars represent 95% confidence interval. \*P<0.05 and \*\*P<0.01 vs. normal tissue. *FN1*, fibronectin 1; TNM, Tumor-Node-Metastasis; ns, non-significant.

expression in four of the six cohorts containing prognostic information (Fig. 3). A meta-analysis of all cohorts validated this result, as it exhibited a significant combined *FN1* hazard ratio (HR) of 1.67 (P<0.001; Fig. 2E). This indicated that the expression of *FN1* is a potential indicator of clinical outcome in patients with GC.

*FN1 immunohistochemistry. FN1* is expressed in cancer cells and the intratumoral matrix in GC (Fig. 4). In the IHC cohort, normal epithelial cells exhibited no *FN1* expression. E-*FN1* expression was positive in 85 of the 190 cases (44.7%). S-FN expression was graded as no/weak in 11 (5.8%), moderate in 71 (37.4%) and strong in 108 (56.8%) cases (Table II). No association was identified between E-*FN1* and S-*FN1* expression (P=0.112; Table III). E-*FN1* expression in GC exhibited a significant association with tumor size (P=0.037), whereas S-*FN1* expression was associated with sex (P=0.027) (Table II).

E-*FN1*-positive patients with GC in the IHC cohort exhibited worse OS compared with E-*FN1*-negative patients (P=0.009; Fig. 5A). S-*FN1* expression exhibited no significant effect on OS (P=0.075, Fig. 5B). In addition, in patients with

high clinical TNM stage (III + IV), E-*FN1* positivity was strongly associated with OS; however, in patients with low clinical TNM stage (I + II), no difference was observed in overall survival between patients with low and high E-*FN1* expression (Fig. 5C and D). E-*FN1* was also confirmed as an independent predictor of overall survival in GC by multivariate analysis (HR, 2.115; 95% CI, 1.343-3.333; P=0.001; Table IV).

# Discussion

In this study, FNI gene expression was analyzed in 17 independent GC cohorts. The results demonstrated an increase in FNI expression in GC compared with normal tissues and a possible increase in the advanced T stage (T2+T3+T4) group compared with that in the early T stage (T1) group in one cohort; however, no association between FNI expression levels and differentiation or clinical TNM stage was identified. In addition, upregulation of the FNI gene may be a predictor of poor prognosis following radical gastrectomy for GC. In summary, the results of the present study support FNI as a biomarker of poor prognosis in GC.

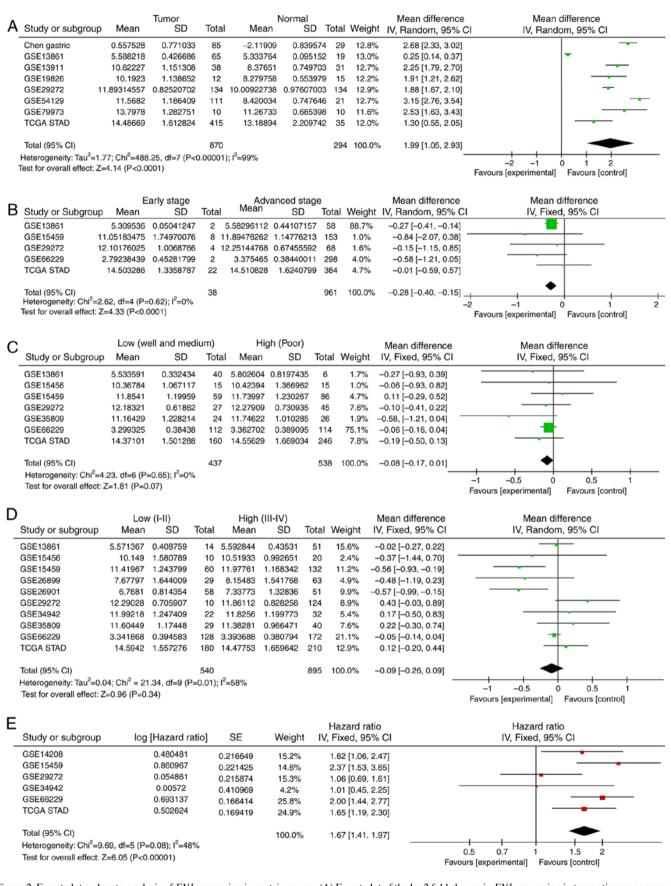


Figure 2. Forest plot and meta-analysis of FNI expression in gastric cancer. (A) Forest plot of the log2 fold change in FNI expression in tumor tissues compared with that in normal tissues. (B) Forest plot of the log2 fold change in FNI expression in the advanced T stage group (T2 + T3 + T4) compared with that in the early T stage group (T1). (C) Forest plot of the log2 fold change in FNI expression in the low differentiation group (high tumor grade) compared with that in the high differentiation group (intermediate and low tumor grade). (D) Forest plot of the log2 fold change in FNI expression in the low 2 fold change in FNI expression in the low 2 fold change in FNI expression in the high differentiation group (intermediate and low tumor grade). (D) Forest plot of the log2 fold change in FNI expression in the high clinical TNM stage (III+IV) group compared with that in the low clinical TNM stage (I+II) group. (E) Forest plot of the comparison of overall survival in patients with gastric cancer with high and low FNI expression (The gene expression value was equal to three, the first two-thirds were defined as high expression and the last one-third as low expression). FNI, fibronectin 1; TNM, Tumor-Node-Metastasis; CI, confidence interval.

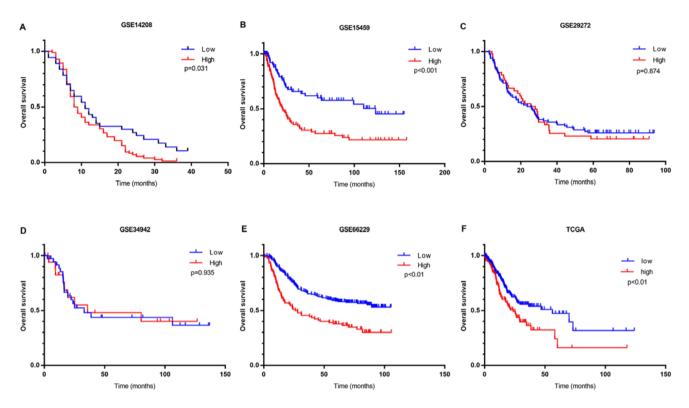


Figure 3. Kaplan-Meier survival curves of the high- and low-*FN1* expression groups in six cohorts. Survival curves for the following datasets: (A) GSE14208; (B) GSE15459; (C) GSE29272; (D) GSE34942; (E) GSE66229 and (F) TCGA. *FN1*, fibronectin 1; TCGA, The Cancer Genome Atlas.

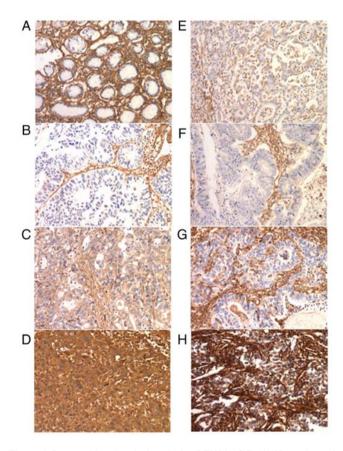


Figure 4. Immunohistochemical analysis of *FN1* in GC. (A) Normal gastric mucosa exhibited negative staining for *FN1*. (B-D) E-*FN1* expression in GS tissues. (B) Expression score, 0; (C) expression score, 6; and (D) expression score, 12. (E-H) S-*FN1* expression in in GS tissues. (E) No expression; (F) low expression; (G) medium expression; and (H) high expression. Magnification, x200. GC, gastric cancer; *FN1*, fibronectin 1; S-*FN1*, stromal *FN1*; E-*FN1*, epithelial *FN1*.

*FN1*, which is an extracellular matrix glycoprotein, is involved in cell proliferation, embryogenesis, wound healing, host defense, epithelial-mesenchymal transition (EMT) and metastasis, as well as oncogenic transformation (5). FN1 is involved in the occurrence and development of various tumors and is upregulated in multiple cancer types, such as esophageal squamous cell carcinoma, colorectal cancer, OSCC, and thyroid cancer (8-10,14). For instance, FN1 is upregulated in OSCC with lymph node metastasis (LNM); FN1 increases the expression of vascular endothelial growth factor C, lymphangiogenesis and LNM through FAK activation and promotes EMT in SAS human OSCC cells (37). FN1 is a key mediator of glioma progression, as its inhibition delays tumor progression and immunosuppression through a mechanism that involves the maintenance of integrin  $\beta 1$ FN receptors (38). In GC, FN1 is highly expressed in tumor tissues compared with that in non-tumor tissues, and knockdown of FN1 represses GC cell proliferation, adhesion and metastasis in vitro (15). The present study aimed to analyze the relationship between FN1 expression in GC and clinicopathological factors and prognoses.

The results of the present study demonstrated that the *FN1* gene was upregulated in gastric cancer tissues compared with that in normal tissues in eight cohorts, and these data were confirmed by meta-analysis of combinations of all datasets. This result was consistent with the results of Xu *et al* (15) and Zhang *et al* (16), who used immunohistochemical methods to analyze tumor and normal tissue specimens from 40 and 52 patients with gastric cancer, respectively. In summary, previous studies have reported that the expression of the *FN1* gene was increased in GC tissues compared with that in normal gastric tissues, but the studies were all small-scale. The present

		Expres E-FN			Expre	ssion of S-FI	N1 (%)	
Characteristic	No. of patients (n=190)	Negative (n=85)	Positive (n=105)	P-value	No/weak (n=11)	Moderate (n=71)	Strong (n=108)	P-value
Sex				0.380				0.027ª
Female	46	18 (39.14)	28 (60.9)		0 (0.0)	13 (28.3)	33 (71.7)	
Male	144	67 (46.5)	77 (53.5)		11 (7.6)	58 (40.3)	75 (52.1)	
Age (years)				0.508				0.361
<60	100	47 (47.0)	53 (53.0)		4 (4.0)	41 (41.0)	55 (55.0)	
≥60	90	38 (42.2)	52 (57.8)		7 (7.8)	30 (33.3)	53 (58.9)	
Tumor diameter (cm)				0.037ª				0.639
<5	114	58 (50.9)	56 (49.1)		8 (7.0)	41 (36.0)	65 (57.0)	
≥5	76	27 (35.5)	49 (64.5)		3 (3.9)	30 (39.5)	43 (56.6)	
T stage				0.742				0.962
T1 + T2	47	22 (46.8)	25 (53.2)		3 (6.4)	18 (38.3)	26 (55.3)	
T3 + T4	143	63 (44.1)	80 (55.9)		8 (5.6)	53 (37.1)	82 (57.3)	
N stage				0.080				0.616
N0 + N1	112	56 (50.0)	56 (50.0)		8 (7.1)	42 (37.5)	62 (55.4)	
N2 + N3	78	29 (37.2)	49 (62.8)		3 (3.8)	29 (37.2)	46 (59.0)	
TNM stage				0.352				0.510
I + II	89	43 (48.3)	46 (51.7)		7 (7.9)	32 (36.0)	50 (56.2)	
III + IV	101	42 (41.6)	59 (58.4)		4 (4.0)	39 (38.6)	58 (57.4)	

Table II. Patient characteristics based on th	e immunohistochemistr	v results of <i>FN1</i> ex	pression in gastric cancer.
rable in rationt enalacteristics based on th	e minimulionistoenemisti	, 1000100 01 1 1 1 0	pression in gasare cancer.

<sup>a</sup>P<0.05, *FN1*, fibronectin 1; S-*FN1*, stromal *FN1*; E-*FN1*, epithelial *FN1*; TNM, Tumor-Node-Metastasis.

Table III. Association between epithelial and stromal expression of FN1 in gastric cancer.

	Expression	of E-FN1		
Expression of S-FN1	Negative (%)	Positive (%)	Total (%)	P-value
No/Weak	7 (63.6)	4 (36.4)	11 (5.8)	
Moderate	34 (47.9)	37 (52.1)	71 (37.4)	
Strong	44 (40.7)	64 (59.3)	108 (56.8)	
Total (%)	85 (100.0)	105 (100.0)	190 (100.0)	0.112

study used multiple cohorts to provide substantial validation of increased *FN1* expression in GC.

To the best of our knowledge, the association between FNI expression and clinicopathological features or patient prognosis, have not been reported previously. In the present study, compared with that in the early T stage group, the expression of FNI was significantly increased in the advanced T stage group, which was further confirmed by meta-analysis in all the examined groups. OS analysis revealed that high FNI expression was associated with unfavorable prognosis in four of the six cohorts containing prognostic information. A meta-analysis of all cohorts further validated this finding. These results indicated that

the expression of FN1 may be a potential indicator of clinical outcomes in patients with GC.

*FN1* is expressed in cancer cells and the intratumoral matrix in GC. Hanamura *et al* (39) reported that the expression of S-*FN1* mRNA was positively correlated with deep invasion and LNM of colon cancer. Bae et al (34) reported that E-*FN1*-positive patients exhibited lower OS and disease-free survival compared with *FN1*-negative breast cancer patients. E-*FN1* was an independent predictor for survival in breast cancer in multivariate analysis, but the expression of S-*FN1* had no significant effect on patient survival (34). In the present study, E-*FN1*-positive patients with GC exhibited worse OS compared with E-*FN1*-negative patients, whereas S-*FN1* 

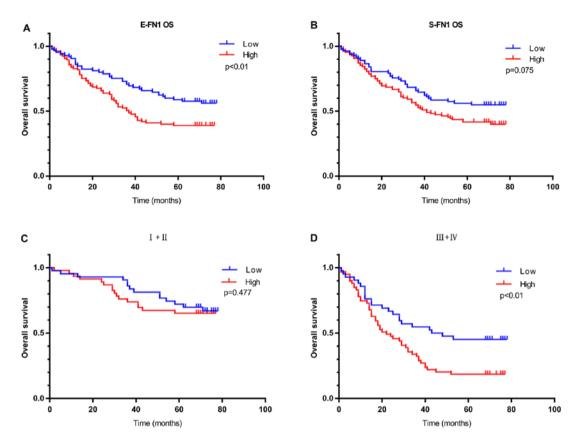


Figure 5. The prognostic significance of *FN1* in patients with gastric cancer based on immunohistochemistry. (A) Kaplan-Meier analysis of OS based on E-*FN1* expression in 190 patients. (B) Kaplan-Meier analysis of OS based on S-*FN1* expression in 190 patients. (C and D) Kaplan-Meier analysis of OS based on S-*FN1* expression in patients with (C) TNM stage I+II (D) and III+IV gastric cancer. *FN1*, fibronectin 1; S-*FN1*, stromal *FN1*; E-*FN1*, epithelial *FN1*; TNM, Tumor-Node-Metastasis; OS, overall survival.

Table IV. Multivariate analysis of overall survival in190 patients with gastric cancer.

		95.0% CI for HR		
Characteristic	HR	Lower	Upper	P-value
E-FN1 expression				0.001 <sup>b</sup>
Negative vs. positive	2.115	1.343	3.333	
Tumor size				0.083
<5 cm vs. ≥5 cm	1.442	0.954	2.181	
Tumor grade				0.024ª
Low + intermediate	1.286	1.034	1.601	
vs. high				
Depth of invasion				$0.028^{a}$
T1+T2 vs. T3+T4	2.352	1.097	5.043	
TNM stage				$0.005^{b}$
I + II vs. III + IV	2.124	1.258	3.585	

<sup>a</sup>P<0.05, <sup>b</sup>P<0.01, HR, hazard ratio; E-*FN1*, epithelial fibronectin 1; TNM, Tumor-Node-Metastasis.

expression had no significant effect on OS. In addition, in patients with high clinical TNM stage (III + IV), E-*FN1* positivity was strongly associated with OS. *FN1* was also confirmed

as an independent predictor of overall survival in patients with GC by multivariate analysis.

Xu *et al* (15) and Zhang *et al* (16) demonstrated no *FN1* expression in the stroma of gastric cancer. In the IHC cohort of the present study, *FN1* was expressed in tumor cells and stromal cells, but not in normal epithelial cells. No association was observed between E-*FN1* and S-*FN1*. E-*FN1* expression in GC was significantly associated with tumor size. Soikkeli *et al* (40) reported that *FN1* is required for tumor and stromal cell growth. It may be speculated in large tumors, the central region is likely to be necrotic, and the expression of *FN1* may promote the migration of tumor cells and reduce necrosis.

In the present study, increased expression of the FN1 gene at the protein and mRNA level in GC tissues was observed; FN1 was highly expressed at the mRNA level in the advanced T stage group compared with that in the early T stage group, and the expression of FN1 at the protein level was positively associated with tumor size. In addition, FN1 expression at the protein and mRNA level was a predictor of poor prognosis following radical resection of GC. In conclusion, the expression of FN1 in GC tissues may be upregulated, and FN1 may be a biomarker of poor prognosis in patients with GC.

#### Acknowledgements

Not applicable.

# Funding

Not applicable.

#### Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author upon reasonable request.

## **Authors' contributions**

YS wrote the manuscript and performed the majority of the experiments. CZ and YS participated in the study design, data acquisition and revision of the manuscript. YL and YH performed immunohistochemistry scoring, followed up the patients and collected clinical information. YY, HM and ZW analyzed the data. All authors read and approved the final manuscript.

## Ethics approval and consent to participate

Written informed consent was obtained from all patients, and the study was approved by the Biomedical Ethics Committee of The First Affiliated Hospital of Zhengzhou University.

#### Patient consent for publication

Patients provided their consent for publication.

#### **Competing interests**

The authors declare that they have no competing interests.

#### References

- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 68: 394-424, 2018.
- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ and He J: Cancer statistics in China, 2015. CA Cancer J Clin 66: 115-132, 2016.
- 3. Katai H, Ishikawa T, Akazawa K, Isobe Y, Miyashiro I, Oda I, Tsujitani S, Ono H, Tanabe S, Fukagawa T, *et al*: Five-year survival analysis of surgically resected gastric cancer cases in Japan: A retrospective analysis of more than 100,000 patients from the nationwide registry of the Japanese gastric cancer association (2001-2007). Gastric Cancer 21: 144-154, 2018.
- Peng PL, Zhou XÝ, Yi GD, Chen PF, Wang F and Dong WG: Identification of a novel gene pairs signature in the prognosis of gastric cancer. Cancer Med 7: 344-350, 2018.
- 5. Pankov R and Yamada KM: Fibronectin at a glance. J Cell Sci 115: 3861-3863, 2002.
- 6. Plow EF, Haas TA, Zhang L, Loftus J and Smith JW: Ligand binding to integrins. J Biol Chem 275: 21785-21788, 2000.
- Korah R, Boots M and Wieder R: Integrin alpha5beta1 promotes survival of growth-arrested breast cancer cells: An in vitro paradigm for breast cancer dormancy in bone marrow. Cancer Res 64: 4514-4522, 2004.
- Xiao J, Yang W, Xu B, Zhu H, Zou J, Su C, Rong J, Wang T and Chen Z: Expression of fibronectin in esophageal squamous cell carcinoma and its role in migration. BMC Cancer 18: 976, 2018.
  Cai X, Liu C, Zhang TN, Zhu YW, Dong X and Xue P: Down-regulation of *FN1* inhibits colorectal carcinogenesis
- 9. Cal X, Liu C, Zhang TN, Zhu YW, Dong X and Xue P: Down-regulation of *FN1* inhibits colorectal carcinogenesis by suppressing proliferation, migration, and invasion. J Cell Biochem 119: 4717-4728, 2018.

- Nakagawa Y, Nakayama H, Nagata M, Yoshida R, Kawahara K, Hirosue A, Tanaka T, Yuno A, Matsuoka Y, Kojima T, *et al*: Overexpression of fibronectin confers cell adhesion-mediated drug resistance (CAM-DR) against 5-FU in oral squamous cell carcinoma cells. Int J Oncol 44: 1376-1384, 2014.
- Wang J, Deng L, Huang J, Cai R, Zhu X, Liu F, Wang Q, Zhang J and Zheng Y: High expression of fibronectin 1 suppresses apoptosis through the NF-κB pathway and is associated with migration in nasopharyngeal carcinoma. Am J Transl Res 9: 4502-4511, 2017.
- 12. Lou X, Han X, Jin C, Tian W, Yu W, Ding D, Cheng L, Huang B, Jiang H and Lin B: SOX2 targets fibronectin 1 to promote cell migration and invasion in ovarian cancer: New molecular leads for therapeutic intervention. OMICS 17: 510-518, 2013.
- Waalkes S, Atschekzei F, Kramer MW, Hennenlotter J, Vetter G, Becker JU, Stenzl A, Merseburger AS, Schrader AJ, Kuczyk MA and Serth J: Fibronectin 1 mRNA expression correlates with advanced disease in renal cancer. BMC Cancer 10: 503, 2010.
- 14. Sponziello M, Rosignolo F, Celano M, Maggisano V, Pecce V, De Rose RF, Lombardo GE, Durante C, Filetti S, Damante G, *et al*: Fibronectin-1 expression is increased in aggressive thyroid cancer and favors the migration and invasion of cancer cells. Mol Cell Endocrinol 431: 123-132, 2016.
- 15. Xu TP, Huang MD, Xia R, Liu XX, Sun M, Yin L, Chen WM, Han L, Zhang EB, Kong R, *et al*: Decreased expression of the long non-coding RNA FENDRR is associated with poor prognosis in gastric cancer and FENDRR regulates gastric cancer cell metastasis by affecting fibronectin1 expression. J Hematol Oncol 7: 63, 2014.
- Zhang H, Sun Z, Li Y, Fan D and Jiang H: MicroRNA-200c binding to *FN1* suppresses the proliferation, migration and invasion of gastric cancer cells. Biomed Pharmacother 88: 285-292, 2017.
- Wang W, Chin-Sheng H, Kuo LJ, Wei PL, Lien YC, Lin FY, Liu HH, Ho YS, Wu CH and Chang YJ: NNK enhances cell migration through alpha7-nicotinic acetylcholine receptor accompanied by increased of fibronectin expression in gastric cancer. Ann Surg Oncol 19 (Suppl): S580-S588, 2012.
  Cho JY, Lim JY, Cheong JH, Park YY, Yoon SL, Kim SM,
- Cho JY, Lim JY, Cheong JH, Park YY, Yoon SL, Kim SM, Kim SB, Kim H, Hong SW, Park YN, *et al*: Gene expression signature-based prognostic risk score in gastric cancer. Clin Cancer Res 17: 1850-1857, 2011.
- D'Errico M, de Rinaldis E, Blasi MF, Viti V, Falchetti M, Calcagnile A, Sera F, Saieva C, Ottini L, Palli D, *et al*: Genome-wide expression profile of sporadic gastric cancers with microsatellite instability. Eur J Cancer 45: 461-469, 2009.
- 20. Kim HK, Choi IJ, Kim CG, Kim HS, Oshima A, Michalowski A and Green JE: A gene expression signature of acquired chemoresistance to cisplatin and fluorouracil combination chemotherapy in gastric cancer patients. PLoS One 6: e16694, 2011.
- Ooi CH, Ivanova T, Wu J, Lee M, Tan IB, Tao J, Ward L, Koo JH, Gopalakrishnan V, Zhu Y, *et al*: Oncogenic pathway combinations predict clinical prognosis in gastric cancer. PLoS Genet 5: e1000676, 2009.
- 22. Chia NY, Deng N, Das K, Huang D, Hu L, Zhu Y, Lim KH, Lee MH, Wu J, Sam XX, et al: Regulatory crosstalk between lineage-survival oncogenes KLF5, GATA4 and GATA6 cooperatively promotes gastric cancer development. Gut 64: 707-719, 2015.
- Lei Z, Tan IB, Das K, Deng N, Zouridis H, Pattison S, Chua C, Feng Z, Guan YK, Ooi CH, *et al*: Identification of molecular subtypes of gastric cancer with different responses to PI3-kinase inhibitors and 5-fluorouracil. Gastroenterology 145: 554-565, 2013.
- 24. Muratani M, Deng N, Ooi WF, Lin SJ, Xing M, Xu C, Qamra A, Tay ST, Malik S, Wu J, et al: Nanoscale chromatin profiling of gastric adenocarcinoma reveals cancer-associated cryptic promoters and somatically acquired regulatory elements. Nat Commun 5: 4361, 2014.
- 25. Cristescu R, Lee J, Nebozhyn M, Kim KM, Ting JC, Wong SS, Liu J, Yue YG, Wang J, Yu K, *et al*: Molecular analysis of gastric cancer identifies subtypes associated with distinct clinical outcomes. Nat Med 21: 449-456, 2015.
- 26. He J, Jin Y, Chen Y, Yao HB, Xia YJ, Ma YY, Wang W and Shao QS: Downregulation of ALDOB is associated with poor prognosis of patients with gastric cancer. Onco Targets Ther 9: 6099-6109, 2016.
- 27. Tao J, Deng NT, Ramnarayanan K, Huang B, Oh HK, Leong SH, Lim SS, Tan IB, Ooi CH, Wu J, et al: CD44-SLC1A2 gene fusions in gastric cancer. Sci Transl Med 3: 77ra30, 2011.

- 28. Lee J, Sohn I, Do IG, Kim KM, Park SH, Park JO, Park YS, Lim HY, Sohn TS, Bae JM, *et al*: Nanostring-based multigene assay to predict recurrence for gastric cancer patients after surgery. PLoS One 9: e90133, 2014.
- 29. Li WQ, Hu N, Burton VH, Yang HH, Su H, Conway CM, Wang L, Wang C, Ding T, Xu Y, *et al*: PLCE1 mRNA and protein expression and survival of patients with esophageal squamous cell carcinoma and gastric adenocarcinoma. Cancer Epidemiol Biomarkers Prev 23: 1579-1588, 2014.
- Wang Q, Wen YG, Li DP, Xia J, Zhou CZ, Yan DW, Tang HM and Peng ZH: Upregulated INHBA expression is associated with poor survival in gastric cancer. Med Oncol 29: 77-83, 2012.
  Wang G, Hu N, Yang HH, Wang L, Su H, Wang C, Clifford R,
- 31. Wang G, Hu N, Yang HH, Wang L, Su H, Wang C, Clifford R, Dawsey EM, Li JM, Ding T, *et al*: Comparison of global gene expression of gastric cardia and noncardia cancers from a high-risk population in china. PLoS One 8: e63826, 2013.
- 32. Wu Y, Grabsch H, Ivanova T, Tan IB, Murray J, Ooi CH, Wright AI, West NP, Hutchins GG, Wu J, *et al*: Comprehensive genomic meta-analysis identifies intra-tumoural stroma as a predictor of survival in patients with gastric cancer. Gut 62: 1100-1111, 2013.
- 33. Sung CO, Park CK and Kim SH: Classification of epithelial-mesenchymal transition phenotypes in esophageal squamous cell carcinoma is strongly associated with patient prognosis. Mod Pathol 24: 1060-1068, 2011.
- 34. Bae YK, Kim A, Kim MK, Choi JE, Kang SH and Lee SJ: Fibronectin expression in carcinoma cells correlates with tumor aggressiveness and poor clinical outcome in patients with invasive breast cancer. Hum Pathol 44: 2028-2037, 2013.

- Sandsmark E, Andersen MK, Bofin AM, Bertilsson H, Drablos F, Bathen TF, Rye MB and Tessem MB: SFRP4 gene expression is increased in aggressive prostate cancer. Sci Rep 7: 14276, 2017.
- 36. Washington K: 7th edition of the AJCC cancer staging manual: Stomach. Ann Surg Oncol 17: 3077-3079, 2010.
- 37. Morita Y, Hata K, Nakanishi M, Omata T, Morita N, Yura Y, Nishimura R and Yoneda T: Cellular fibronectin 1 promotes VEGF-C expression, lymphangiogenesis and lymph node metastasis associated with human oral squamous cell carcinoma. Clin Exp Metastasis 32: 739-753, 2015.
- 38. Sengupta S, Nandi S, Hindi ES, Wainwright DA, Han Y and Lesniak MS: Short hairpin RNA-mediated fibronectin knockdown delays tumor growth in a mouse glioma model. Neoplasia 12: 837-847, 2010.
- Hanamura N, Yoshida T, Matsumoto E, Kawarada Y and Sakakura T: Expression of fibronectin and tenascin-C mRNA by myofibroblasts, vascular cells and epithelial cells in human colon adenomas and carcinomas. Int J Cancer 73: 10-15, 1997.
  Soikkeli J, Podlasz P, Yin M, Nummela P, Jahkola T, Virolainen S,
- 40. Soikkeli J, Podlasz P, Yin M, Nummela P, Jahkola T, Virolainen S, Krogerus L, Heikkila P, von Smitten K, Saksela O and Hölttä E: Metastatic outgrowth encompasses COL-I, *FN1*, and POSTN up-regulation and assembly to fibrillar networks regulating cell adhesion, migration, and growth. Am J Pathol 177: 387-403, 2010.
  - This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.