

Independent prognostic value of HIF-1 α expression in radiofrequency ablation of lung cancer

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Abstract. Radiofrequency ablation (RFA) is widely used in the treatment of lung cancer. Hypoxia-inducible factor-1 α (HIF-1 α) is a crucial transcription factor regulating oxygen homeostasis that is involved in tumor cell metastasis. The present study investigated the impact of HIF-1 α expression and other factors, such as postoperative blood CD4⁺/CD8⁺ ratio, on the prognosis of patients with lung cancer who had received RFA treatment. A total of 80 patients with lung cancer were recruited between January 2011 and October 2016 at The Shenzhen People's Hospital. Lung cancer was confirmed following pathological or histological examination. All patients underwent RFA treatment. Patients were followed up for 6-66 months. HIF-1 α expression in lung cancer tissues was assessed by immunohistochemistry. Multivariate survival analysis was performed using Cox proportional hazards model. The results demonstrated that HIF-1 α level was low in 36 patients and overexpressed in 44 patients with lung cancer. Kaplan-Meier (KM) curve analysis demonstrated that the overall survival time of patients with high HIF-1 α expression was significantly shorter compared with patients with low HIF-1 α expression ($P < 0.05$). Furthermore, the results from the KM model and log-rank test revealed that age, Union for International Cancer Control stage, primary or metastatic cancer, chemotherapy, postoperative blood CD4⁺/CD8⁺ ratio, Eastern Cooperative Oncology Group performance status and HIF-1 α expression had significant effects on overall survival of patients with lung cancer. The results from Cox analysis demonstrated that high HIF-1 α expression, advanced age, clinical staging and chemotherapy were independent risk factors for the prognosis of lung cancer following RFA treatment, and that high HIF-1 α expression was associated

with the increased risk (5.91-fold) of mortality. In conclusion, the present study demonstrated that HIF-1 α expression was increased in lung cancer tissues and was associated with the prognosis of patients with lung cancer who were treated with RFA. These findings suggest that HIF-1 α expression may be considered as a marker for evaluating the prognosis of these patients.

Introduction

Lung cancer is a common malignancy with high morbidity and mortality rates. According to a survey by Chen *et al* (1), the incidence of lung cancer in China ranks now first among all types of cancer. Lung cancer also presents a high rate of metastases (2,3). Metastasis is a complex multi-step process that comprises numerous genes and several factors, including angiogenesis factors, extracellular metal matrix proteases and adhesion molecules (4,5). The development of lung cancer involves many tumor suppressor genes that are down-regulated, such as p53 (6), Rb (7) and Fhit (8), as well as the abnormal overexpression of oncogenes, such as CDCA7 (9), KIF20A (10) and CCNB2 (11). Non-small cell lung carcinoma (NSCLC), which is a type of lung cancer, is divided into four histological subtypes, including adenocarcinoma, squamous cell carcinoma, large cell carcinoma, and NSCLC not otherwise specified (12). Adenocarcinoma and squamous cell carcinoma represent ~80% of all NSCLC cases (13). The various types of lung cancer possess distinct histological features and display different biological behavior (14), which influence the choice of treatment and the prognosis of patients with lung cancer (15). Determining the molecular profiles of all types of lung cancer in order to develop novel therapies is therefore essential (16). The present study aimed to investigate the effect of hypoxia on all histological types of lung cancer. It has been demonstrated that a hypoxic microenvironment can inhibit tumor apoptosis and promote DNA repair, increasing therefore cancer invasion and metastasis and promoting radio-chemotherapy resistance (17,18).

HIF-1 α is a crucial transcription factor that regulates oxygen homeostasis, serving therefore a pivotal role in tumor hypoxia (19). An increased expression of HIF-1 α has been observed in various types of human cancer, including NSCLC, and can be associated with poor prognosis in some cases (20,21). HIF-1 α level is regulated by hypoxic factors,

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such as limited oxygen concentration, and is associated with tumor differentiation and invasion (22,23). Radiofrequency ablation (RFA) is a minimally-invasive interventional treatment for local tumors that promotes tumor cell apoptosis and necrosis through high temperatures (24). RFA also stops blood supply to the peripheral blood vessels of the tumor in order to reduce metastasis (25). In addition, RFA has demonstrated satisfactory clinical effects in the treatment of patients with primary lung cancer and lung metastases (26). Subsequently, the 5-year survival rate of patients with lung cancer is 25-61% (27). The 3-year survival rate of patients with lung cancer reaches 57% when radiotherapy and chemotherapy are combined (28). Although RFA is effective for the treatment of lung cancer, it also induces several complications that can severely affect the prognosis of patients (29-32). The results from our previous study demonstrated that local recurrences of lung cancer caused by the overgrowth of residual tumor following RFA treatment are driven by HIF-1 α (33). However, whether HIF-1 α could be considered a prognostic factor for patients with lung cancer following RFA treatment remains unknown. The present study analyzed the clinical data and survival time of 80 patients with lung cancer who underwent RFA in order to investigate the effect of HIF-1 α expression on the prognosis of these patients.

Materials and methods

Clinical data collection. A total of 80 patients diagnosed with lung cancer by histopathological analysis and who had received RFA treatment between January 2011 and October 2016 at the Department of Thoracic Surgery of The Shenzhen People's Hospital were included in the present study. The cohort consisted of 66 men and 14 women with an average age of 62.14 \pm 9.41 years (age range, 41-80 years). Of the 80 patients, 61 patients were smokers or ever smokers (76.32%). The inclusion criteria were as follows: i) Patients unable to undergo or unsuitable for surgical treatment due to multiple tumors, recurrences and severe cardiovascular disease as confirmed by pathological or cytological examination (including percutaneous biopsy, lymph node biopsy, fiberoptic bronchoscopy brushing and biopsy, pleural effusion cytology and sputum cytology); ii) patients with complete clinical data; and iii) patients with definite staging. Patients with second primary malignancy and who were lost during follow-up were excluded. During the surgery, tumor tissues and tumor adjacent tissues (located 2 cm way from tumors) were collected. The baseline characteristics of all patients are summarized in Table I. Following RFA treatment tissue samples were stained with hematoxylin and eosin (H&E) and examined for pathological changes by light microscopy. The ablation zone was treated by needle biopsy to evaluate the expression of HIF-1 α by immunohistochemistry (IHC). Peripheral blood (10 ml) was collected at 1 week following RFA and directly sent to the laboratory for the analysis of T lymphocyte subsets (CD4⁺/CD8⁺). Their association with the prognosis of patients with lung cancer was also analyzed.

Main instruments and reagents. HIF-1 α (1:500; cat. no. RM242) rabbit anti-human monoclonal antibody and

SP immunoassay kit were purchased from Boster Biological Technology. The goat anti-rabbit secondary antibody (1:1,000; cat. no. BA1056) was provided by Wuhan Boster Biological Engineering Technology Co., Ltd. Fluorescein isothiocyanate- and antigen-presenting cells-CD4 and CD8 were purchased from BD Biosciences. Inverted Microscope (Olympus Corporation). The tissue embedder (Sakura), slicer (Leica Microsystems GmbH), oscillator (Shanghai Jinghong Experimental Equipment Co., Ltd.), plastic staining frame (Fuzhou Maixin Biotech Co., Ltd.), computer image analysis system (HP), expandable anchor RF electrode needle (Beijing Blade Photoelectric Technology Development Co., Ltd.). A total of 12 microelectrode needles can be released radially with a diameter of 4.0 cm, when fully opened to cover the ablation area as much as possible, the local temperature can reach 90-100°C during treatment and the thermal coagulation necrosis diameter was 5-6 cm), RF-2000 radiofrequency treatment instrument (Beijing Blade Photoelectric Technology Development Co., Ltd.; radiofrequency, 500 kHz; maximum output power, 200 W) and Siemens 64 row SOMATOM Perspective dual source spiral CT (Siemens, AG were used in experiments.

RFA treatment. All the patients undergoing lung RFA were admitted as inpatients at the Department of Thoracic Surgery of The Shenzhen People's Hospital and placed in observation for a minimum of one night. All patients completed at least one post-procedure chest radiograph. A total of 57 patients received lung RFA under general anesthesia, whereas 23 patients were treated under local anesthesia. Briefly, CT-guided radiofrequency needle was inserted into the lung cancer tissue and the microelectrode was opened to connect the radiofrequency current. The average duration of the RFA treatment was 50 min with a mean and median impedance of 439 and 446 units, respectively. Technical success was defined as complete ablation of all target lesions. When the RFA operation was completed, a needle biopsy was performed immediately for the detection of HIF-1 α expression.

Patients' follow-up. The follow-up (6-66 months) was performed by electronic medical record inquiry and telephone calls. Overall survival was calculated from the first treatment with RFA (not from the diagnosis of lung cancer). The association between HIF-1 α expression and patient prognosis was analyzed by unpaired two-tailed Student's t-test. Kaplan-Meier (KM) analysis was used to construct the survival curve. Cox proportional hazards model was applied for multifactor survival analysis.

IHC. The dissected tissue was fixed in 4% polyoxymethylene for two days, decalcified in 10% EDTA at 48°C for 2 weeks and sliced into 5- μ m sections. Sections were subjected to antigen retrieval in sodium citrate buffer (1 M, pH 6.0) at 99°C for 20 min and incubated with primary antibody against HIF-1 α (1:1,000; cat. no. RM242; Boster Biological Technology) overnight at 4°C. The slides were then incubated with a biotin-conjugated goat anti-rabbit secondary antibody (1:1,000; cat. no. BA1003; Wuhan Boster Biological Engineering Technology Co., Ltd.) at room temperature for 45 min. The slides were examined with a Nikon Eclipse Ti light microscope under a x40 objective

Table I. Univariate analysis of the clinical characteristics of patients with lung cancer.

Characteristics	Number	Median survival time (days)	χ^2 value	P-value
Sex			1.035	0.596
Male	66	578		
Female	14	432		
Age (years)			11.769	0.001
<60	30	983		
\geq 60	50	480		
Primary or metastasis			32.387	<0.001
Primary cancer	67	836		
Metastatic cancer	13	356		
UICC stage (67 cases)			58.084	<0.001
I/II	34	1,124		
III/IV	33	467		
Postoperative blood CD4 ⁺ /CD8 ⁺			83.053	<0.001
\leq 1.65	47	457		
>1.65	33	1,145		
Chemotherapy			3.967	0.042
<3 cycles	16	516		
\geq 3 cycles	23	813		
ECOG rating			51.674	<0.001
0	13	1,145		
1	24	983		
2	31	480		
3	12	301		
Smoking			0.326	0.568
Yes	61	568		
No	19	578		
HIF-1 α expression			79.266	<0.001
Low expression	36	1,124		
High expression	44	438		

ECOG, Eastern Cooperative Oncology Group; HIF-1 α , hypoxia-inducible factor-1 α ; UICC, Union for International Cancer Control.

(Nikon Instruments). A positive expression for HIF-1 α was detected when brownish-yellow particles appeared in the nucleus, membrane or cytoplasm. Results of the IHC were analyzed using semi-quantitative scoring method as described by Lin *et al* (34). Briefly, staining was scored as negative (-), mild (+), moderate (++) or strong (+++) for <1, 1-10, 10-50 or >50% of cell nuclear stain, respectively. In the present study, moderate (++) and strong (+++) scores defined high HIF-1 α expression, whereas negative (-) and mild (+) scores defined low HIF-1 α expression.

CD4⁺/CD8⁺ measurement by flow cytometry. FACS lysing solution (BD Biosciences) was used to lyse red blood cells in the peripheral blood, which was followed by two washes twice with PBS. CD4⁺/CD8⁺ was measured using the BD Simultest™ CD4/CD8 kit (BD Biosciences), according to manufacturer's instructions, through incubation with

FITC-labeled CD4 and PE-labeled CD8 antibody for 15-30 min at room temperature. Flow cytometry was used to analyze the residual white blood cells, and the proportions of the lymphocyte subsets were calculated using FlowJo software version 10 (FlowJo LLC).

Statistical analysis. SPSS 19.0 software (IBM Corp.) was used for data analysis. All data are expressed as the means \pm standard error of the mean. Statistical differences were analyzed by unpaired two-tailed Student's t-test for the comparison between two groups. The skewed distribution was described by median \pm interquartile range (M \pm Q). Independent samples were compared by rank sum test. All values described represent the means \pm standard deviation. KM method was used to construct the survival curve of patients with lung cancer. Significance test was based on the log-rank method. Cox ratio risk model was applied for the multivariate

Table II. Comparison of the survival rate of patients with lung cancer after radiofrequency ablation treatment.

Group	Number	1 year (%)	2 years (%)	3 years (%)	4 years (%)	5 years (%)
Primary cancer	67					
Stage I/II	34	100.00	94.12	55.88	23.53	5.88
Stage III/IV	33	72.73	9.09	3.03	0.00	0.00
Metastatic cancer	13	46.15	0.00	0.00	0.00	0.00
Total	80	80.00	43.75	25.00	10.00	2.50

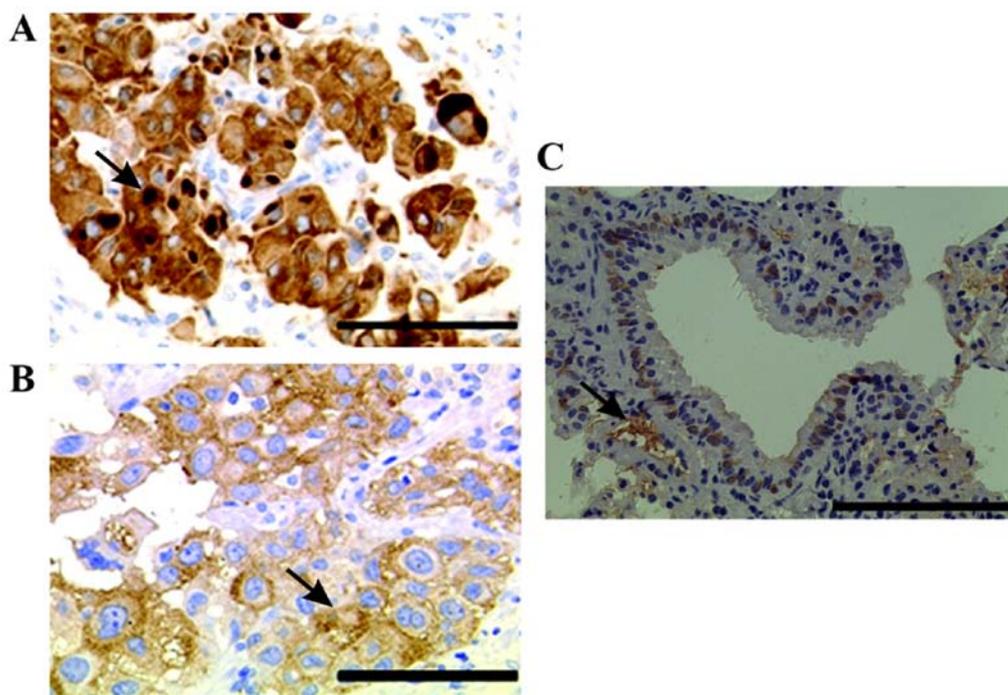


Figure 1. Immunohistochemistry of HIF-1 α in lung cancer and adjacent tissues. (A) Some cancer cells contained intracytoplasmic macroglobular spots strongly positive for HIF-1 α . Magnification, x20. Scale bar, 100 μ m. (B) Some tumor cells presented a perinuclear halo positive for HIF-1 α . Magnification, x40. Scale bar, 50 μ m (C) IHC staining of tumor adjacent tissues. Magnification, x10. Scale bar, 200 μ m Arrows indicate the positive staining of HIF-1 α . HIF-1 α , hypoxia-inducible factor-1 α .

survival analysis. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Patient characteristics. A total of 80 patients with lung cancer were included in the present study (Table I). The median age of the patients was 61.6 ± 9.8 (range, 41-80) years. The cohort consisted of 66 men (82.5%) and 14 women (17.5%). Furthermore, 61 patients were smokers or ever-smokers (76.3%). The Eastern Cooperative Oncology Group (ECOG) performance status was used to assess the patient's physical condition and the score of all patients was < 3 . A total of 13 patients (16.3%) suffered from metastatic pulmonary cancer. According to the results from the survival rate, no patients with metastatic cancer and stage III/IV primary cancer (according to The Eighth Edition of TNM Staging of Lung Cancer) survived for 5 years (35), and only two patients with stage I/II primary cancer survived for 5 years (Table II). The overall 5-year survival rate was only 2.5% (Table II).

HIF-1 α expression in lung cancer tissues. The results demonstrated that HIF-1 α was mainly localized in the nucleus and cytoplasm of lung cancer cells (Fig. 1A). Furthermore, only little HIF-1 α expression was detected in normal lung tissues (Fig. 1C). In addition, certain cancer cells containing intracytoplasmic macroglobular spots were strongly positive for HIF-1 α (Fig. 1B), and some tumor cells exhibited a positive perinuclear aureole. Representative image of HIF-1 α staining is presented in Fig. 1B. Among the 80 cases of lung cancer tissues analyzed, the level of HIF-1 α expression was as follows: 9 cases (11.3%) had negative staining (-), 27 cases (33.8%) had mild staining (+); 13 cases (16.3%) had moderate staining (++), and 31 cases (38.8%) had strong staining (+++). Subsequently, among all patients with lung cancer, 36 (45%) displayed low HIF-1 α expression and 44 (55%) presented high HIF-1 α expression.

Prognostic value of HIF-1 α expression and other factors in patients with lung cancer treated by RFA. The follow-up time of the whole group was 6-66 months. Two patients

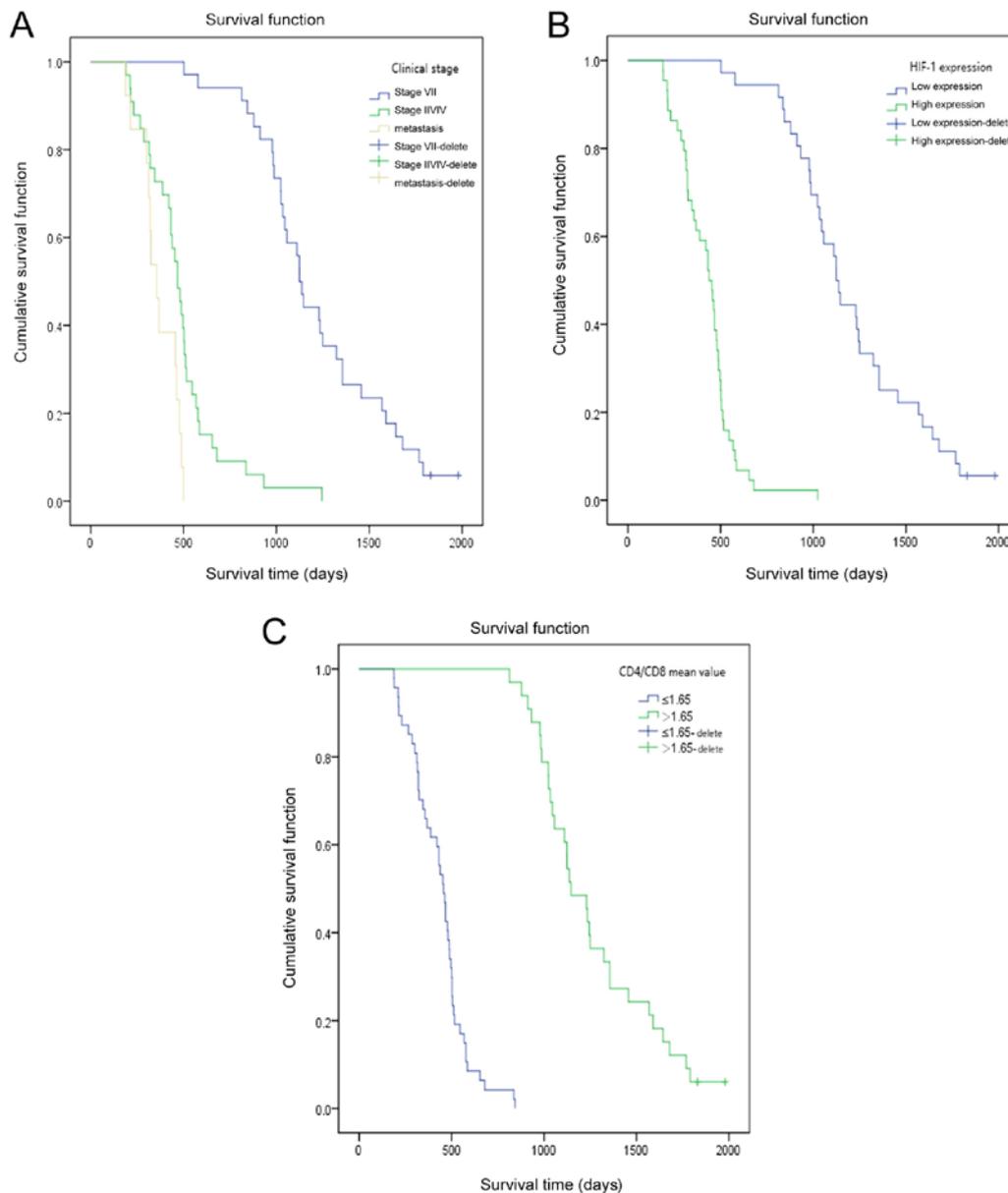


Figure 2. Overall survival of patients with lung cancer at different time points after RFA treatment. (A) Survival curve analysis of patients with lung cancer after RFA treatment according to the clinical stage. (B) Survival curve analysis of patients with lung cancer after RFA treatment according to HIF-1 α expression. (C) Survival curve analysis of patients with lung cancer after RFA treatment according to the postoperative level of CD4⁺/CD8⁺. HIF-1 α , hypoxia-inducible factor-1 α ; RFA, radiofrequency ablation.

survived until the end of the follow-up. The median survival time was 573 days. The KM model and log-rank test were used to analyze the effect of clinical factors on the overall survival (OS). The results demonstrated that factors, including age, Union for International Cancer Control (UICC) stage, primary or metastatic cancer, chemotherapy, postoperative CD4⁺/CD8⁺, ECOG rating and HIF-1 α expression significantly affected the OS (Table I). The average value of CD4⁺/CD8⁺ was 1.65 (range, 0.15-4.58). Furthermore, decreased survival was observed in patients with older age, high UICC stage, lower CD4⁺/CD8⁺ level (expression of CD4 and CD8 by flow cytometry are presented in Fig. S1), high ECOG rating and high HIF-1 α expression (Table I). In addition, a significant difference between the OS of patients with primary cancer stage I/II and of patients with primary cancer stage III/IV was observed ($\chi^2=58.084$; $P<0.001$; Table II). Furthermore, a

clear difference was observed in the survival rate of patients with primary and metastatic carcinoma ($P<0.001$; $\chi^2=32.387$; Table II). The associations between HIF-1 α expression, postoperative blood CD4⁺/CD8⁺ or UICC staging and the OS of patients with lung cancer at different time points following RFA treatment are presented in Fig. 2. The results from KM survival curves demonstrated that patients with high HIF-1 α expression, high UICC stage and low CD4⁺/CD8⁺ had a significantly shorter OS.

Associations between HIF-1 α expression and clinicopathological characteristics in patients with lung cancer treated by RFA. The association between HIF-1 α expression and the clinical characteristics of patients, including sex, age, tumor metastasis, UICC stage, ECOG rating and smoking status was evaluated. As presented in Table III, HIF-1 α expression

Table III. Association between HIF-1 α expression and patients' clinical characteristics.

Characteristics	Number	Low HIF-1 α expression (%; n=36)	High HIF-1 α expression (%; n=44)	χ^2 value	P-value
Sex				1.011	0.315
Male	66	28 (42)	38 (58)		
Female	14	8 (57)	6 (43)		
Age (years)				0.485	0.486
<60	30	15 (50)	15 (50)		
\geq 60	50	21 (42)	29 (58)		
Primary or metastasis				8.729	0.003
Primary cancer	67	35 (52)	32 (48)		
Metastatic cancer	13	1 (8)	12 (92)		
UICC stage (67 cases)				18.6	<0.001
I/II	34	24 (71)	10 (29)		
III/IV	33	6 (18)	27 (82)		
ECOG rating				45.801	<0.001
0	13	12 (92)	1 (8)		
1	24	20 (83)	4 (17)		
2	31	6 (19)	25 (81)		
3	12	2 (17)	10 (83)		
Smoking				0.67	0.413
Yes	61	29	32		
No	19	7	12		

ECOG, Eastern Cooperative Oncology Group; HIF-1 α , hypoxia-inducible factor-1 α ; UICC, Union for International Cancer Control.

Table IV. Cox regression analysis of independent factors in the prognosis of patients with lung cancer.

Variables	β	SE	Wald χ^2	P-value	OR value	95% CI
HIF-1 α expression	1.777	0.720	6.090	0.014	5.910	1.441-24.230
Advanced age	0.836	0.277	9.086	0.003	2.307	1.340-3.397
Clinical stage	-3.548	1.111	10.202	0.001	0.029	0.003-0.254
Chemotherapy or not	-1.488	0.366	16.523	<0.001	0.226	0.110-0.463

CI, confidence interval; HIF-1 α , hypoxia-inducible factor-1 α ; β , estimated regression coefficient; SE, standard error; Wald, Wald statistic.

was not associated with sex, age or smoking status ($P>0.05$). However, HIF-1 α expression was significantly associated with tumor metastasis, UICC stage or ECOG rating ($P<0.05$) in patients with lung cancer treated with RFA.

Determination of independent factor of prognosis in patients with lung cancer by Cox regression analysis. The results from Cox regression analysis demonstrated that the high expression of HIF-1 α , advanced age, clinical stage and chemotherapy could be considered as independent risk factors for the prognosis of patients with lung cancer following RFA treatment (Table IV). Among all cases, the OR (odds ratio) value of high HIF-1 α expression, advanced age, clinical staging and chemotherapy was 5.910-, 2.307-, 0.029- and 0.026-fold, respectively ($P=0.014$ and 95% confidence interval (CI),

1.441-24.230 for high expression of HIF-1 α ; $P=0.003$ and 95% CI, 1.340-3.397 for advanced age; $P=0.001$ and 95% CI, 0.003-0.254 for clinical staging; and $P<0.001$ and 95% CI, 0.110-0.463 for chemotherapy). Furthermore, the OR value of high HIF-1 α expression was higher than that of advanced age, clinical staging and chemotherapy. HIF-1 α expression may therefore represent a more effective independent prognostic factor in patients with lung cancer treated by RFA compared with age, clinical stage and chemotherapy.

Discussion

RFA becomes a widely accepted treatment for primary lung cancer in patients who are not candidates for subsegment resection or lobectomy (36). Compared with radiotherapy

and chemotherapy, RFA focuses exclusively on the tumor area without causing any damage to the normal surrounding tissue (37). In addition, compared with surgical treatment, RFA is a minimally-invasive method (38). In advanced lung cancers, RFA is extremely effective for palliative care (39); however, it induces several complications, including local recurrence, which severely affects the prognosis of patients. Previous studies indicated that RFA causes deep hypoxia in the tissues that are adjacent to the resected area (40). The presence of hypoxia has been reported in several types of solid tumor and is associated with the aggressiveness, proliferation, and angiogenesis potential of the tumor (41). HIF-1 α is induced in hypoxia and serves a crucial role in the pathological process of inflammation, hypoxia, vascular permeability, apoptosis, pulmonary fibrosis and lung cancer (42,43). In addition, HIF-1 α regulates the expression of *PGK-1*, *VEGF* and other target genes to induce blood vessel and tissue proliferation that in turn promotes tumor growth (44,45). Hypoxia inhibits HIF-1 α degradation and enhances HIF-1 α DNA binding activity, leading to high HIF-1 α expression. VEGF expression is therefore elevated, binds to vascular endothelial surface receptors and activates the ischemic transduction pathway (46,47).

It has been reported that HIF-1 α expression is increased in several types of cancer, including lung, prostate, breast and colon carcinomas, which are the main causes of cancer-associated mortality (48). In the present study, 36 and 44 patients with lung cancer exhibited a low and high HIF-1 α expression, respectively. Furthermore, the results from KM curve demonstrated that patients with high HIF-1 α expression exhibited significantly shorter OS compared with patients with low HIF-1 α expression. The results from KM model and log-rank test demonstrated a significant effect of HIF-1 α expression on the OS of patients with lung cancer. The results from Cox regression analysis indicated that high HIF-1 α expression could be considered as an independent risk factor for patients with lung cancer following RFA treatment. Furthermore, HIF-1 α expression may affect tumor angiogenesis, proliferation and metabolism, which may influence tumor cell sensitivity to RFA treatment. It has been reported that HIF-1 α level is increased under hypoxic condition, which results in increased tumor angiogenesis via *VEGF* and a subsequent increase in tumor size, expansion of hypoxic areas, resistance to RFA treatment and poor prognosis of patients with lung cancer (49). Consistently, the results from the present study demonstrated that patients with lower HIF-1 α expression presented elevated survival time compared with patients with higher HIF-1 α expression. In addition, HIF-1 α expression was significantly associated with tumor metastasis, UICC stage and ECOG rating.

Lung cancer cells secrete numerous immunosuppressive factors that can inhibit the anti-tumor immunity (50,51). Furthermore, RFA treatment of lung cancer is effective and can be monitored in real time (52). A previous study demonstrated that RFA therapy for lung cancer can improve the cellular immune function, due to the direct killing effect of antigens and synergistic killing effect of cytokines released by sensitized T cells. This leads to the correction of immune imbalance of Th1/Th2 cells, increase CD4⁺ and CD8⁺ cell levels, eliminate the sources of tumor-derived inhibitors

and restore the anti-tumor immunity (53,54). For example, the decreased proportions of CD4⁺ cells (T helper cells), CD4/CD8 ratios and B cells, and the increased number of CD8⁺/CD28 T lymphocytes and regulatory T (Treg) cells have been demonstrated in patients with lung cancer (55,56). In the present study, the levels of CD4⁺ and CD8⁺ cells were increased following RFA treatment, which suggested that RFA treatment may restore the imbalanced immune function in patients with lung cancer. In addition, results from the KM model and log-rank test demonstrated that some factors, including age, UICC stage, primary or metastatic cancer, chemotherapy, postoperative blood levels of CD4⁺/CD8⁺, ECOG rating and HIF-1 α expression had a significant effect on the OS of patients with lung cancer following RFA treatment. Furthermore, Cox regression analysis revealed that high HIF-1 α expression, advanced age, clinical staging and chemotherapy may be considered as independent risk factors for the prognosis of patients with lung cancer after RFA treatment, which indicated that changes in T lymphocyte subsets after treatment may be associated with patient prognosis. UICC staging may also be considered as a prognostic factor for patients with lung cancer after RFA treatment. In addition, the long-term effect of treatment with RFA combined with chemotherapy for patients with advanced lung cancer was better than that of chemotherapy or RFA treatment alone. However, since the present study only included a limited number of patients, further investigation including a larger sample size is therefore required in order to confirm the prognostic value of HIF-1 α expression in patients with lung cancer. Although the present study determined the effect of HIF-1 α expression on the prognosis of patients with lung cancer after RFA treatment, the underlying mechanisms involved and the correlation between HIF-1 α expression and patients' immunity required further investigation.

In conclusion, the present study analyzed HIF-1 α expression in patients with lung cancer. The results demonstrated that patients treated with RFA and with high HIF-1 α expression had significantly shorter OS compared with patients with low HIF-1 α expression. Furthermore, high HIF-1 α expression was significantly associated with tumor metastasis, UICC stage and ECOG rating. HIF-1 α expression may therefore be considered as an effective independent prognosis factor in patients with lung cancer treated by RFA. The detection of HIF-1 α expression may therefore be crucial for evaluating the prognosis of patients with lung cancer who underwent RFA treatment.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JW and GD designed the experiments. JW, XL and ZR performed the experiments. GD and BP analyzed the data. JW and XL wrote the manuscript. All authors read and approved the final version of the manuscript and agreed to be accountable for all aspects of the research to ensure that the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate

This study was approved by the Clinical Medical Research Ethics Committee of the First Affiliated Hospital of Anhui Medical University (approval number AF/SC-08/02.0). All patients provided informed consent prior to the study.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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