

Primary mesenchymal chondrosarcoma of the kidney without HEY1-NCOA2 and IRF2BP2-CDX1 fusion: A case report and review

ATSUSHI YAMAGISHI¹, OSAMU ICHIYANAGI², SEI NAITO¹, HIROMI ITO¹, TAKANOBU KABASAWA³, MITSUNORI YAMAKAWA³ and NORIHIKO TSUCHIYA¹

¹Department of Urology, Yamagata University Faculty of Medicine, Yamagata 990-9585; ²Department of Urology, Yamagata Prefectural Kahoku Hospital, Yachi, Kahoku 999-3511; ³Department of Pathological Diagnostics, Yamagata University Faculty of Medicine, Yamagata 990-9585, Japan

Received June 19, 2019; Accepted October 8, 2019

DOI: 10.3892/ol.2019.11143

Abstract. Mesenchymal chondrosarcoma (MC) of the kidney is rare. To the best of our knowledge, the current report is the first case of a giant extraskeletal MC that arose primarily from the right kidney and mimicked renal cell carcinoma at the locally advanced stage (cT3bN0) with vena cava thrombus and multiple pulmonary arterial tumor emboli. Additionally, the literature on renal EMC is reviewed and the possibilities of oncogenic heterogeneity are discussed. A 64-year-old woman was admitted to Yamagata University Hospital for sudden onset of asymptomatic gross hematuria. CT revealed a 90 mm renal mass without calcification in the right kidney and tumor thrombus extending to the inferior vena cava. Radical nephrectomy with thrombectomy was performed. Lung metastasis was detected 2 months later. The patient received systemic chemotherapy, which was only marginally effective. She died of the malignancy 8 months after surgery. Microscopic examination of the tumor revealed typical histology of MC and a lack of HEY1-NCOA2 and IRF2BP2-CDX1 gene fusions in the tumor tissues. Not all MC patients may exhibit chromosomal alterations in the tumor, suggesting the presence of genetically heterogeneous pathways of MC oncogenesis. Further studies are required to confirm the present findings and reinforce the molecular diagnosis of MC.

Abbreviations: MC, mesenchymal chondrosarcoma; EMC, extraskeletal mesenchymal chondrosarcoma; HEY1, hairy/enhancerof-split related with YRPW motif 1; NCOA2, nuclear receptor coactivator 2; STAU2, staufen double-stranded RNA binding protein 2; ZFHX4, zinc finger homeobox 4; IRF2BP2, interferon regulatory factor 2 binding protein 2; CDX1, caudal type homeobox 1

Key words: mesenchymal chondrosarcoma, *HEY1-NCOA2* fusion, *IRF2BP2-CDX1* fusion, tumor thrombus

Introduction

Mesenchymal chondrosarcoma (MC) is a rare neoplasm and represents only 2 to 9% of all chondrosarcomas. Almost 24% of MCs originate from extraskeletal sites, such as head, neck, the extremities, and trunk. The kidney is an extremely rare origin of extraskeletal MC (EMC) (1). In a recent report from Japan (2), primary origins of MC were bone (58%) and soft tissues (42%). MC develops at various sites in the body, including the head and neck (12%), trunk (62%), and extremities (26%). The authors reported 5- and 10-year overall survival rates of 66 and 56%, respectively (2).

In the absence of specific tumor markers and radiographic findings, the diagnosis of MC largely depends on pathologic examination (3,4). The *HEY1-NCOA2* gene fusion is reported to be a specific chromosomal aberration for MC and may play a critical role in diagnosis due to its high sensitivity (3,5-7). Physiologically, HEY1 is considered to function mainly as a transcriptional repressor and *NCOA2* encodes a transcriptional coactivator protein for intranuclear receptors (7). Recently, an *IRF2BP2-CDX1* fusion gene was implicated as another candidate MC oncogene exclusive to the *HEY1-NCOA2* fusion gene (8).

We describe the first case of primary renal EMC mimicking renal cell carcinoma at the locally advanced stage (cT3bN0) accompanied by vena cava thrombus and multiple pulmonary tumor emboli. The microscopic pathology was typical of MC but the *HEY1-NCOA2* and *IRF2BP2-CDX1* gene fusions were not detected.

Case report

Clinical summary. A 64-year-old woman visited our clinic complaining of sudden onset asymptomatic gross hematuria and was hospitalized for further examination. Flank pain and fever were absent. Routine laboratory tests indicated no remarkable abnormality in hematology and urine cytology was negative. No past history was noted, except for liver cystectomy and right mastectomy.

Computed tomography (CT) of the abdomen revealed a giant mass (90x70x67 mm) in the mid to lower pole of the right

Correspondence to: Dr Atsushi Yamagishi, Department of Urology, Yamagata University Faculty of Medicine, Iida-nishi 2-2-2, Yamagata 990-9585, Japan E-mail: giejie100@yahoo.co.jp

kidney. The tumor was hypovascular on contrast-enhanced CT images compared with adjacent normal renal parenchyma, although tumor vessels surrounding the mass were well-developed. In addition, the tumor was characterized as internally heterogeneous, free of calcification, and protruding into the inferior vena cava (IVC) through the right renal vein to form a tumor thrombus up to the level of the caudate lobe of the liver (Fig. 1). Tumor emboli were disseminated to pulmonary arteries, but no distant metastases were identified. The tumor presented with a high standardized uptake value of 12.00 at the maximum in positron emission tomography (PET)-CT. Magnetic resonance imaging (MRI) with delayed contrast-enhancement revealed the heterogeneous nature of the tumor. On plain MRI, the tumor had high, low, slightly low, and slightly low signal intensities on T2-weighted, T1-weighted, diffusion-weighted, and apparent diffusion coefficient mapping images, respectively. Histological examination using percutaneous needle-biopsied specimens of the tumor revealed that it contained atypical chondrocytes and spindle cells with high nuclear-to-cytoplasmic ratio, suggesting renal chondrosarcoma. Radical nephrectomy with intracaval thrombectomy was performed for the initial treatment for the tumor. We successfully en bloc resected the whole kidney and the attached IVC thrombus using an artificial heart-lung machine and clamping IVC just beneath the beginning of the hepatic veins in cooperation with vascular surgeons.

The results of the pathological investigation of the resected specimens are presented in Fig. 2. They had biphasic morphological components with transition zones between round and spindle-shaped hyperchromic tumor cells and contained cartilaginous islands with good differentiation (Fig. 2A and B). The spindle-shaped cells showed indistinct cytoplasmic borders, inconspicuous nucleoli, and a hemangiopericytoma-like pattern (Fig. 2C). Tumor cells invaded into small lympho-vascular structures, renal veins, and renal hilar fat tissues. No metastasis was microscopically confirmed in any of the 11 local lymph nodes surgically resected in total, including para-aortic/vena cava and renal hilar lymph nodes. Immunohistochemically, the tumor was positively stained with antibodies to vimentin, CD99, and Ki-67 (labelling index: 60-70%), without immunoreaction to epithelial membrane antigen, CD34, chromogranin A, and synaptophysin. Expression of S-100 protein was negative in mesenchymal spindle cells, but was slightly positive only in some chondroid cells (Fig. 2D). Based on these pathological features, the tumor was diagnosed as primary renal EMC.

Two months later, she experienced local recurrence and multiple lung metastases. Systemic chemotherapy was delivered and began with three courses of doxorubicin (30 mg/m^2 on day 1-2, every 3 weeks). This was followed by pazopanib (800 mg/day), but the therapy was discontinued due to drug eruption at day 11. Eribulin mesilate was delivered in two courses (1.4 mg/m^2 on day 1 and 8, every 3 weeks). The dose was reduced to 1.1 mg/m^2 in the second cycle because of neutropenia. Palliative irradiation of 20 Gy was given to de novo metastasis to the right 8th rib for pain control. The patient's condition gradually worsened and she finally died of the malignancy 8 months after surgery.

HEY1-NCOA2 and IRF2BP2-CDX1 gene fusion. Standard reverse transcription-polymerase chain reaction (RT-PCR)

Table I. Primer sequences for PCR.

Primer	5'→3'
HEY1_F1	CGAGGTGGAGAAGGAGAGTG
HEY1_F2	ACCGGATCAATAACAGTTTG
HEY1_R	CCCGAAATCCCAAACTCCGA
NCOA2_F	AGCTTTTCCCAGACACGAGG
NCOA2_R1	TCCTGGCTGAGGTATCAC
NCOA2_R2	AGTTGGGCTTTGCAATGTGA
STAU2_F	ACTCCCCCTTGTTCTCCAGT
STAU2_R	TGCCTGGTTATTGTCCGCTT
ZFHX4_F	CCGCTGATGACTGGACAACT
ZFHX4_R	GGTGTTGGTCTTCACCGCTA
βActin_F	CCTCGCCTTTGCCGATCC
βActin_R	GGATCTTCATGAGGTAGTCAGTC
IRF2BP2_F1	CAAGAGCCGCGGGTCTGGAGA
IRF2BP2_F2	GTCAACAGGCCCAAGACCGTGC
IRF2BP2_R	GTGTGGTCCGGTTGGAATGAGGTG
CDX1_F	CCGCAGTACCCCGACTTCTCCAG
CDX1_R1	GTTCAGTGAGCCCCAGATTGGCAG
CDX1_R2	TGATGTCGTGGGCCATCGGC

were used to detect the HEY1-NCOA2 and IRF2BP2-CDX1 chromosomal fusions, as described elsewhere (9). In brief, RNA samples were extracted from surgically resected and fleshly frozen MC tissues with mirVana[™] miRNA Isolation kit (Life Technologies). Total RNA (2 µg) was reverse-transcribed in a 20 μ l reaction volume using a cDNA Reverse Transcription Kit according to the manufacturer's instructions (Applied Biosystems). RT-PCR was run using 1 μ l cDNA in a 50 μ l PCR reaction using AmpliTaq Gold DNA polymerase (Applied Biosystems). Primers for HEY1-NCOA2 fusion, STAU2, and ZFHX4 are shown in Table I and Fig. 3A (all were purchased from Integrated DNA Technologies, IA, USA). The primers for IRF2BP2-CDX1 were the same as reported previously (Integrated DNA Technologies) (8). The PCR conditions were 95°C for 30 sec, 55-60°C (according to the Tm of each primer provided from the manufacturer) for 30 sec, and 72°C for 45 sec after initial denaturation at 95°C for 2 min. Thirty-five cycles were run. RT-PCR of the housekeeping gene encoding β-actin was run on all samples to check the RNA quality. PCR products were checked using 2% agarose gel electrophoresis.

The overall results of RT-PCR are presented in Fig. 3B. The *HEY1-NCOA2* fusion gene was not observed in this patient. Instead of the fusion gene, intact *HEY1* and *NCOA2* genes were successfully detected. The presence of intact *STAU2* and *ZFHX4* genes, each of which should be deleted from the chromosome at the fusion of *HEY1* and *NCOA2* genes (Fig. 3A), was confirmed by RT-PCR in the patient's samples. Similarly, the *IRF2BP2-CDX1* fusion was not detected in the patient (Fig. 4). Taken together, these findings demonstrate that the *HEY1-NCOA2* and *IRF2BP2-CDX1* gene fusions were absent or were not located at the chromosomal breakpoints that were previously reported.



Figure 1. Abdominal computed tomography scan. (A) Plain computed tomography did not reveal calcification in the tumor. (B) The tumor was marginally enhanced and was heterogeneous. (C) The tumor thrombus extended into the IVC and reached the level of the caudal lobe of the liver. The right lobe of the liver had been previously resected. IVC, inferior vena cava.



Figure 2. Microscopic examination of the renal tumor. (A) Islands of well-differentiated cartilage surrounded by hyperchromic cells are visible in a hematoxylin and eosin stained section (magnification, x40). (B) A transition zone was seen between cellular and cartilage components (arrow; hematoxylin and eosin; magnification, x100). (C) A hemangiopericytoma-like pattern was seen (hematoxylin and eosin; magnification, x200). (D) Immunoreaction to S-100 protein was positive in a cartilaginous island, but negative in the surrounding small tumor cells (magnification, x100). This microphotograph shows the same region in (B).

Discussion

Primary renal EMC is extremely rare (1). To our best knowledge, only 16 patients with renal EMC, including the present case, have been reported in the medical literature (Table II) (1,10-23). According to a review article on renal EMC (1), flank pain and hematuria are two most common clinical symptoms. Renal EMC occurs from children to elderly people with the peak incidence in the third decade, equally in men and women (1). No clinical differences in oncological behaviors are indicated between renal and non-renal origins of EMC (1). Despite a paucity of credible evidence for renal EMC, a favorable prognosis might be expected if the tumor is small, locally confined, and completely resected (1). Mimicking advanced renal cell carcinoma (cT3bN0), our case had tumor dissemination in pulmonary arteries at the initial presentation and early metastatic recurrence occurred

in the bilateral lungs soon after radical nephrectomy with IVC thrombectomy.

MC is often difficult to diagnose because of the lack of specific findings. On clinical imaging, MC typically exhibits low CT attenuation with calcification and heterogeneous appearance, T1-low and T2-high/low intensities of MR signals, and strongly metabolic activity on PET-CT (4,24). These radiological findings are not specific to MC and its diagnosis entirely depends on histology. For this reason, MC is usually diagnosed with MC postoperatively. Renal MC is frequently misdiagnosed as renal cell carcinoma, renal pelvic cancer, or other rare malignant tumors, such as Wilms' tumor, in younger cases. In our case, the kidney tumor was initially suspected to be advanced RCC due to IVC thrombus and pulmonary tumor emboli. Low contrast-enhancement in CT and MRI and lack of remarkable findings of blood tests, including leukocyte and neutrophil counts, and serum levels of lactate dehydrogenase



Figure 3. RT-PCR analysis of the *HEY1-NCOA2* fusion. (A) *HEY1* and *NCOA2* gene structures. The upper panel displays the wild type. *HEY1* and *NCOA2* genes are located on chromosome 8q. The *STAU2* and *ZFHX4* genes are located between them. The lower panel displays previously reported fusion genes. (B) Results of RT-PCR. The fusion gene was not detected. The actin band was clearly evident. Normal *NCOA2*, *STAU2* and *ZFX4F* genes were detected. The results affirmed the absence of the fusion gene because those genes should be deleted if *HEY1* and *NCOA2* genes fuses at a certain breakpoint, and also indicated the success of RT-PCR. RT, reverse transcription; *NCOA2*, nuclear receptor coactivator 2; STAU2, staufen double-stranded RNA binding protein 2; *ZFHX4*, zinc finger homeobox 4; *HEY1*, hairy/enhancer-of-split related with YRPW motif 1.



Figure 4. RT-PCR analysis of the *IRF2BP2-CDX1* fusion. (A) *IRF2BP2* and *CDX1* gene structures. The upper panel displays the wild type. *IRF2BP2* is located on chromosome 1q and *CDX1* is located on chromosome 5q. The lower panel displays previously reported fusion genes. (B) Results of PCR. The *IRF2BP2* and *CDX1* gene fusion was not indicated, despite the presence of the actin band. Normal *IRF2BP2* gene was detected as double bands reflecting its two isoforms. Normal *CDX1* was difficult to detect with RT-PCR possibly due to GC richness in the *CDX1* sequence. RT, reverse transcription; *IRF2BP2*, interferon regulatory factor 2 binding protein 2; *CDX1*, caudal type homeobox 1.

and C-reactive protein seem to be atypical for such advanced RCC. Percutaneous needle biopsy helped us to make an accurate pathological diagnosis and formulate sufficient treatment plans for the kidney tumor. However, not the all cases of MC could be successfully diagnosed with needle-biopsied

specimens. Typically, MC histology shows a biphasic pattern, which consists of sheets of undifferentiated, round to spindle cell component, and islands of hyaline cartilage. The diagnosis in a small portion of biopsied tissues remains uncertain when the chondroid structures scattered within MC are not



No.	Year	Author	Age	Sex	Calcification	Size (cm)	Metastasis ^a	Treatment	Follow	Outcome
1	1981	Pitfield J	61	М	+	12	_	_	2 m	Dead
2	1984	Malhotra CM	27	М	+	9	-	RTx, CTx, Mx	69 m	Alive
3	1991	Karanauskas S	15	М	+	ND	+	ND	ND	ND
4	2001	Gomez-B	52	F	+	8	-	-	1 y	Alive
5	2006	Kaneko T	61	F	+	2.5	-	-	6 y	Alive
6	2008	Dantonello TM	24	F	ND	10	-	NAC	1.3 y	Alive
7	2009	Buse S	23	F	+	7	+	AC, RTx	36 m	Alive
8	2012	Xu H	64	М	-	11	+	-	2 m	Dead
9	2014	Gherman V	67	М	-	30	-	-	9 m	Dead
10	2014	Tyagi R	22	F	-	6.5	+	CTx	ND	Alive
11	2015	Rothberg MB	16	F	+	15.2	+	ND	ND	ND
12	2015	Chen D	17	М	-	15	+ ^b	CTx	10 m	Alive
13	2017	Salehipour M	22	М	+	9	-	ND	ND	ND
14	2017	Pani K	24	М	+	8.5	-	AC	6 m	Alive
15	2018	Valente P	35	М	+	20	-	-	18 m	Alive
16	-	Present case	64	F	-	9	-	CTx, RTx	8 m	Dead

Table II. Case series about MC of the kidney.

^aAt the time of diagnosis. ^bNot proved pathologically. MC, mesenchymal chondrosarcoma; M, male; F, female; NAC, neoadjuvant chemotherapy; AC, adjuvant chemotherapy; CTx, chemotherapy; RTx, radiotherapy; Mx, metastasectomy; ND, no data; m, month; y, year.

Table III. Case series about gene fusion of MC.

	Author	HEY1-NCOA2				IRF2E		
Year		n	Positive	Positive ratio (%)	n	Positive	Positive ratio (%)	Assay
2012	Wang L	15	10	67	-	_	_	FISH, RT-PCR
2012	Nyquist KB	4	3	75	4	1	25	FISH, RT-PCR
2012	Nakayama R	10	8	80	-	-	-	FISH
2013	Fritchie KJ	6	6	100	-	-	-	RT-PCR
2014	Panagopoulos I	1	1	100	-	-	-	RT-PCR
2014	Andersson C	1	1	100	-	-	-	RT-PCR
2014	Moriya K	1	1	100	-	-	-	FISH
2015	Sajjad EA	1	1	100	1	0	0	RT-PCR
2015	Bishop MW	6	6	100	-	-	-	FISH
2016	Cohen JN	2	2	100	-	-	-	RT-PCR
2018	Folpe AL	3	3	100	-	-	-	RT-PCR
2018	Toki S	1	1	100	-	-	-	RT-PCR
Total		51	43	84	5	1	20	

MC, mesenchymal chondrosarcoma; RT, reverse transcription; FISH, fluorescence *in situ* hybridization; *NCOA2*, nuclear receptor coactivator 2; *IRF2BP2*, interferon regulatory factor 2 binding protein 2; *CDX1*, caudal type homeobox 1.

incidentally identified (3). This sampling error would be an intrinsic limitation in the histologic diagnosis of biopsied tissue.

Chromosomal aberration of HEY1-NCOA2 fusion is reportedly specific to MC and can be a highly sensitive diagnostic tool (3,5,6). Table III presents 12 articles on the gene fusion in MC that were published in medical literature (3,5,6,8,25-32). The *HEY1-NCOA2* gene fusion is supposed to be absent in another histologic type of sarcoma (3,5,6), but its diagnostic sensitivity is not perfect (67-100%) despite the characteristic histology of MC (Table III). Interestingly, the *IRF2BP2-CDX1* gene fusion in MC may be exclusive to the *HEY1-NCOA2* fusion, suggesting heterogeneous oncogenesis of MC (8,28). In the present study, we failed to detect the *HEY1-NCOA2* and *IRF2BP2-CDX1* gene fusions in renal EMC. Herein, CDX1 expression appeared to be absent in our case (Fig. 4). Specifically expressed in the intestines, CDX1 is a transcription factor to induce epithelial cell differentiation (33,34). In gastric, colorectal and hepatocellular carcinomas, CDX1 acts as a tumor suppressor and low expression of CDX1 are related clinically to poor prognosis of the malignancies (34). Thus, no expression of CDX1 might potentiate malignant nature in the present renal MC. Our results support the possible existence of another oncogenic mechanism in such fusion-negative cases, reflecting on the genetic heterogeneity in MC tumorigenesis to a typical microscopic phenotype.

It might be better to analyze *HEY1*, *NCOA2*, *STAU2* and *ZFHX4* gene expression using the adjacent or normal kidney tissues together with the renal MC for comparison (Fig. 3B). However, we were unable to do additional experiments due to the paucity of the freshly-frozen tissue samples left behind in store. This is a limitation in interpretation of the present results.

In conclusion, we report the first case of primary renal EMC, which progressed to form tumor thrombus in IVC and pulmonary arteries at diagnosis. On pathological inspection of surgical specimens, the tumor exhibited typical microscopic appearance of MC. However, the presence of the *HEY1-NCOA2* nor *IRF2BP2-CDX1* gene fusions in the tumor was detected by RT-PCR, suggesting the possibility of genetically heterogeneous pathways to MC oncogenesis. Further studies will be needed to confirm the present findings and reinforce the molecular diagnosis of MC.

Acknowledgements

Not applicable.

Funding

No funding was received.

Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

AY contributed to conception and design of the study, acquisition of data, interpretation of data and drafted the initial and revised versions of the manuscript and is the corresponding author. OI did conception and design of the study, interpretation of data and assisted with the writing of several drafts of the manuscript. HI participated in analysis and interpretation of data especially for the technical aspects. TK and MY performed the histological examination of the tumor and revised the pathological section of study. SN contributed to the conception, design and intellectual content such as genomic abnormality and tumorigenesis. NT contributed to conception, design and supervision of the study and made final approval of the version to be published. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was performed in accordance with the principles embodied in the Declaration of Helsinki and approved by the Ethical Committee of Yamagata University Faculty of Medicine (approval no. 28, 2019). Written informed consent for publication was provided by the patient.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- Chen D, Ye ZI, Wu X, Shi B, Zhou L, Sun S, Wei B, Yang S, Mao X and Lai Y: Primary mesenchymal chondrosarcoma with bilateral kidney invasion and calcification in renal pelvis: A case report and review of the literature. Oncol Lett 10: 1075-1078, 2015.
- Tsuda Y, Ogura K, Hakozaki M, Kikuta K, Ae K, Tsuchiya H, Iwata S, Ueda T, Kawano H and Kawai A: Mesenchymal chondrosarcoma: A Japanese Musculoskeletal Oncology Group (JMOG) study on 57 patients J Surg Oncol 115: 760-767, 2017.
- 3. Nakayama R, Miura Y, Ogino J, Susa M, Watanabe I, Horiuchi K, Anazawa U, Toyama Y, Morioka H, Mukai M and Hasegawa T: Detection of HEY1-NCOA2 fusion by fluorescence in-situ hybridization in formalin-fixed paraffin-embedded tissues as a possible diagnostic tool for mesenchymal chondrosarcoma. Pathol Int 62: 823-826, 2012.
- Shakked RJ, Geller DS, Gorlick R and Dorfman HD: Mesenchymal chondrosarcoma: Clinicopathologic study of 20 cases Arch Pathol Lab Med 136: 61-75, 2012.
 Wang L, Motoi T, Khanin R, Olshen A, Mertens F, Bridge J,
- Wang L, Motoi T, Khanin R, Olshen A, Mertens F, Bridge J, Cin PD, Antonescu CR, Singer S, Hameed M, *et al*: Identification of a novel, recurrent HEY1-NCOA2 fusion in mesenchymal chondrosarcoma based on a genome-wide screen of exon-level expression data. Genes Chromosomes Cancer 51: 127-139, 2012.
- Fritchie KJ, Jin L, Ruano A, Oliveira AM and Rubin BP: Are meningeal hemangiopericytoma and mesenchymal chondrosarcoma the same? A study of HEY1-NCOA2 fusion. Am J Clin Pathol 140: 670-674, 2013.
- El Beaino M, Roszik J, Livingston JA, Wang WL, Lazar AJ, Amini B, Subbiah V, Lewis V and Conley AP: Mesenchymal chondrosarcoma: A review with emphasis on its fusion-driven biology. Curr Oncol Rep 20: 37, 2018.
- Nyquist KB, Panagopoulos I, Thorsen J, Haugom L, Gorunova L, Bjerkehagen B, Fosså A, Guriby M, Nome T, Lothe RA, *et al*: Whole-transcriptome sequencing identifies novel IRF2BP2-CDX1 fusion gene brought about by translocation t(1;5)(q42;q32) in mesenchymal chondrosarcoma. PLoS One 7: e49705, 2012.
- Ichiyanagi O, Ito H, Takai S, Naito S, Kato T, Nagaoka A and Yamakawa M: A GRIA2 and PAX8-positive renal solitary fibrous tumor with NAB2-STAT6 gene fusion. Diagn Pathol 10: 155, 2015.
- 10. Pitfield J, Preston BJ and Smith PG: A calcified renal mass: Chondrosarcoma of kidney. Br J Radiol 54: 262, 1981.
- Malhotra CM, Doolittle CH, Rodil JV and Vezeridis MP: Mesenchymal chondrosarcoma of the kidney. Cancer 54: 2495-2499, 1984.
- Karanauskas S, Wells RG and Sty JR: Extraosseus uptake with bone scintigraphy. Renal mesenchymal chondrosarcoma with metastasis. Clin Nucl Med 16: 375-377, 1991.
- Gomez-Brouchet A, Soulie M, Delisle MB and Escourrou G: Mesenchymal chondrosarcoma of the kidney. J Urol 166: 2305, 2001.
- Kaneko T, Suzuki Y, Takata R, Takata K, Sakuma T and Fujioka T: Extraskeletal mesenchymal chondrosarcoma of the kidney. Int J Urol 13: 285-286, 2006.
- 15. Dantonello TM, Int-Veen C, Leuschner I, Schuck A, Furtwaengler R, Claviez A, Schneider DT, Klingebiel T, Bielack SS, Koscielniak E, *et al*: Mesenchymal chondrosarcoma of soft tissues and bone in children, adolescents, and young adults. Cancer 112: 2424-2431, 2008.



- 16. Buse S, Behnisch W, Kulozik A, Autschbach F and Hohenfellner M: Primary chondrosarcoma of the kidney: Case report and review of the literature. Urol Int 83: 116-118, 2009.
- 17 Xu H, Shao M, Sun H and Li S: Primary mesenchymal chondrosarcoma of the kidney with synchronous implant and infiltrating urothelial carcinoma of the ureter. Diagn Pathol 7: 125, 2012.
- 18. Gherman V, Tomuleasa C, Bungardean C, Crisan N, Ona V-D, Feciche B, Irimie A and Coman I: Management of renal extraskeletal mesenchymal chondrosarcoma. BMC Surg 14: 107, 2014.
- 19 Tyagi R, Kakkar N, Vasishta RK and Aggarwal MM: Mesenchymal chondrosarcoma of kidney. Indian J Urol 30: 225-227, 2014
- 20. Rothberg MB, Bhalodi AA, Reda EF, Zelkovic P and Franco I: Primary renal mesenchymal chondrosarcoma: A case report. Urology 85: 676-678, 2015.
- Salehipour M, Hosseinzadeh M, Sisakhti AM, Parvin VA, 21. Sadraei A and Adib A: Renal extra skeletal mesenchymal chondrosarcoma: A case report. Urol Case Rep 12: 23-25, 2017.
- 22. Pani K, Yadav M, Priyaa P and Kumari N: Extraskeletal mesenchymal chondrosarcoma at unusual location involving spleen and kidney with review of literature. Indian J Pathol Microbiol 60: 262.2017.
- 23. Valente P, Macedo-Dias J, Lobato C, Reis M and Pina F: Primary mesenchymal chondrosarcoma of the kidney: A case report and review of literature J Cancer Res Ther 14: 694, 2018.
- 24. Chen Y, Wang X, Guo L, Li Y, Deng S, Liu Y and Guo L: Radiological features and pathology of extraskeletal mesenchymal chondrosarcoma. Clin Imaging 36: 365-370, 2012
- 25. Panagopoulos I, Gorunova L, Bjerkehagen B, Boye K and Heim S: Chromosome aberrations and HEY1-NCOA2 fusion gene in a mesenchymal chondrosarcoma. Oncol Rep 32: 40-44, 2014.
- 26. Andersson C, Osterlundh G, Enlund F, Kindblom LG and Hansson M: Primary spinal intradural mesenchymal chondrosarcoma with detection of fusion gene HEY1-NCOA2: A paediatric case report and review of the literature. Oncol Lett 8: 1608-1612, 2014
- 27. Moriya K, Katayama S, Onuma M, Rikiishi T, Hosaka M, Watanabe M, Hasegawa T, Sasahara Y and Kure S: Mesenchymal chondrosarcoma diagnosed on FISH for HEY1-NCOA2 fusion gene. Pediatr Int 56: e55-e57, 2014.

- 28. Sajjad EA, Sikora K, Paciejewski T, Garbicz F, Paskal W, Szacht M, Grajkowska W and Wlodarski PK: Intraparenchymal mesenchymal chondrosarcoma of the frontal lobe-a case report and molecular detection of specific gene fusions from archival FFPE sample. Clin Neuropathol 34: 288-293, 2015.
- 29. Bishop MŴ, Somerville JM, Bahrami A, Kaste SC, Interiano RB, Wu J, Mao S, Boop FA, Williams RF, Pappo AS and Samant S: Mesenchymal chondrosarcoma in children and young adults: A single institution retrospective review. Sarcoma 2015: 608279, 2015
- 30. Cohen JN, Solomon DA, Horvai AE and Kakar S: Pancreatic involvement by mesenchymal chondrosarcoma harboring the HEY1-NCOA2 gene fusion. Hum Pathol 58: 35-40, 2016.
- 31. Folpe AL, Graham RP, Martinez A, Schembri-Wismayer D, Boland J and Fritchie KJ: Mesenchymal chondrosarcomas showing immunohistochemical evidence of rhabdomyoblastic differentiation: A potential diagnostic pitfall. Hum Pathol 77: 28-34, 2018.
- 32. Toki S, Motoi T, Miyake M, Kobayashi E, Kawai A and Yoshida A: Minute mesenchymal chondrosarcoma within osteochondroma: An unexpected diagnosis confirmed by HEY1-NCOA2 fusion. Hum Pathol 81: 255-260, 2018.
- 33. Zheng H, Yang Y, Wang MC, Yuan SX, Tian T, Han J, Ni JS, Wang J, Xing H and Zhou WP: Low CDX1 expression predicts a poor prognosis for hepatocellular carcinoma patients after hepatectomy. Surg Oncol 25: 171-177, 2016.
- 34. Fagerberg L, Hallström BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, Habuka M, Tahmasebpoor S, Danielsson A, Edlund K, et al: Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. Mol Cell Proteomics 13: 397-406, 2014



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.