

# Identification of the association between HMMR expression and progression of hepatocellular carcinoma via construction of a co-expression network

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**Abstract.** The aim of the present study was to identify key genes involved in the progression of hepatocellular carcinoma (HCC). According to the theory of the multi-step process of hepatocarcinogenesis and weighted gene co-expression network analysis, hub genes associated with the progression of HCC were identified using the gene expression profiles of patients with normal to chronic hepatitis/cirrhosis and dysplastic nodules to HCC. An independent dataset was used to verify the association between hub gene and clinical phenotype. The diagnostic and prognostic value of hub genes regarding HCC were evaluated. Gene set enrichment analysis (GSEA) was performed to explore the function of hub genes. A co-expression gene module positively associated with HCC progression was identified. Combined with a protein-protein interaction (PPI) network, a total of 10 common hub genes common to both the module of interest and the PPI network were selected as hub genes. Hyaluronan mediated motility receptor (HMMR) was selected as the candidate gene and was significantly upregulated in HCC at the mRNA and protein expression levels. HMMR is a promising diagnostic biomarker for HCC, and is also associated with its progression. The expression of HMMR was positively correlated with HCC tumor grade, pathological stage, tumor stage and Ishak score. The expression of HMMR was an independent prognostic factor compared with clinicopathological

features. Patients with high expression levels of HMMR exhibited a less favorable prognosis. GSEA identified 6 representative gene sets that were associated with cancer. Overall, HMMR may serve an important role in HCC and may have potential as a biomarker of HCC diagnosis and progression.

## Introduction

Liver cancer, of which 75-85% of cases consist of hepatocellular carcinoma (HCC), was the sixth most commonly diagnosed cancer and the fourth leading cause of cancer-associated death worldwide in 2018 (1). HCC is one of the few types of cancer which has had a continued increase in incidence over the last decade (2). The risk factors for HCC include chronic infection with hepatitis C or hepatitis B virus, high alcohol intake, aflatoxin B1, obesity, smoking and type 2 diabetes (3-5). Hepatocarcinogenesis is considered to be a multistep process evolving from normal to chronic hepatitis/cirrhosis and dysplastic nodules to HCC (6,7). Curative treatment options are limited to surgical resection of the tumor or liver transplantation; however, >70% of patients with HCC will encounter recurrence within 5 years after surgery (8-10). The specific mechanisms underlying the progression from healthy liver to chronic hepatitis/cirrhosis and dysplastic nodules to HCC are still elusive. The investigation of these mechanisms may help identify potential therapeutic targets to prevent the development and recurrence of HCC and biomarkers of these processes may help clinicians monitor this disease progression.

Previously, with the development of high-throughput and microarray technology, gene expression profiles have been used to identify genes associated with the progression of HCC (11-13). However, the majority of these studies focused on the screening of differentially expressed genes without considering the correlations between genes, despite the fact that genes with similar expression patterns may be functionally related (14). Weighted gene co-expression network analysis (WGCNA) can be used to analyze the associations between gene sets and indicators of tumor progression, including tumor stages and grades (15,16). In the present

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study, the various stages of progression from healthy liver to chronic hepatitis/cirrhosis and dysplastic nodules to HCC were treated as phenotypes. It was hypothesized that the expression patterns of certain genes would be closely associated with these phenotypes.

According to the theory of the multistep process of hepatocarcinogenesis (6,7) and WGCNA, the present study aimed to identify network-centric genes associated with HCC progression by constructing a co-expression network using gene expression profiles from normal to chronic hepatitis/cirrhosis and dysplastic nodules to HCC. Hyaluronan mediated motility receptor (HMMR) was identified to exhibit a strong correlation with the progression of HCC and may represent a promising marker for the prevention and treatment of HCC.

## Materials and methods

**Data collection.** The microarray dataset (GSE89377) of HCC was retrieved from the Gene Expression Omnibus (GEO) database ([ncbi.nlm.nih.gov/geo/](http://ncbi.nlm.nih.gov/geo/)) using the R (<http://www.R-project.org/>) package GEOquery (17) in RStudio (<http://www.rstudio.com/>). The GSE89377 dataset was collected using Illumina HumanHT-12 version 3.0 expression beadchip (Illumina, Inc.) and was used to construct co-expression networks and identify hub genes in the present study. There were 13 healthy liver tissue samples and 94 tissues covering 9 stages of HCC progression in the GSE89377 dataset, including low grade chronic hepatitis (n=8), high grade chronic hepatitis (n=12), cirrhosis (n=12), low grade dysplastic nodules (n=11), high grade dysplastic nodules (n=11), early HCC (n=5), HCC with grade 1 (n=9), HCC with grade 2 (n=12) and HCC with grade 3 (n=14). The gene expression profiles had been normalized using quantile normalization with GenPlex version 3.0 by Jung Woo Eun from The Catholic University of Korea. An independent dataset including RNA-sequencing data and clinical information was obtained from The Cancer Genome Atlas (TCGA) database ([cancer.gov/tcga](http://cancer.gov/tcga)) to further verify the association of hub genes and clinical phenotypes. The gene expression profiles of GSE87630 (18) based on the GPL6947 dataset included 64 HCC and 30 non-tumor profiles used to validate the aberrant expression of the hub genes. The gene expression profiles of GSE87630 were processed using the lumi package (19) in R. As these data are publicly available and open-access, ethical approval was not necessary for the present study.

**Weighted gene co-expression network construction.** Probes were filtered by variance as recommended (15), and the 4,881 probes with the highest variance were selected from 48,803 probes. The 'WGCNA' package (15) was used to construct a co-expression network for the 4,881 probes according to the protocols of WGCNA and R software. First, Pearson's correlation matrices were performed for all pair-wise genes. Subsequently, an adjacency matrix was constructed using a power adjacency function [ $\alpha_{mn} = \text{Power}(S_{mn}) = |S_{mn}|^\beta$ ;  $\alpha_{mn}$ , adjacency between two genes;  $S_{mn}$ , Pearson's correlations between two genes].  $\beta$  is a soft-thresholding parameter that emphasizes strong correlations between genes and penalizes weak correlations. In the present study, the power of  $\beta=9$  (scale-free

$R^2=0.85$ ) was chosen in accordance with the scale-free topology criterion (Fig. 1A). Next, the adjacency was transformed into a topological overlap matrix that measured the network connectivity of a gene, defined as the sum of its adjacency with all other genes for network generation (20). Finally, the 'cutreeStaticColor' function was applied to classify similar expression genes into gene modules (minModuleSize=30; mergeCutHeight=0.25).

**Identification of clinically significant modules and functional enrichment analysis.** Gene significance (GS) was defined as the  $\log_{10}$  transformation of the P-value in the linear regression between gene expression and HCC progress. Module significance (MS) was the mean GS for all the genes in a module. In general, the module with the absolute MS ranked first or second (ranked by MS) amongst all modules was considered as the module correlating with HCC progression. In the present study, the module exhibiting the strongest positive correlation with HCC progression was selected for further analysis and termed the primary module. Module eigengenes (MEs) were considered as the major components in the principal component analysis for each gene module and the expression patterns of all genes could be summarized into a single characteristic expression profile within a given module. The correlation between MEs and clinical traits was calculated to identify the relevant module. As there is more potential of oncogene as a marker or therapeutic target (21), the focus was on modules that are positively associated with HCC progression. In addition, functional Gene Ontology (GO) enrichment analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were performed using the 'clusterProfiler' R package (22) in order to uncover the biologic function of genes in the primary module. P-value [adjusted by false discovery rate (FDR)] < 0.01 was set as the cutoff criteria.

**Identification of hub genes.** In the present study, hub genes in the primary module were defined by module connectivity, measured by the absolute value of the Pearson's correlation ( $\text{cor.geneModuleMembership} \geq 0.8$ ) and clinical trait relationship, measured by absolute value of the Pearson's correlation ( $\text{cor.geneTraitSignificance} \geq 0.7$ ) (23). Furthermore, a protein-protein interaction (PPI) network was constructed by uploading all genes in the primary module to the Search Tool for the Retrieval of Interacting Genes (STRING) database (24). Overall, 50 genes with the highest connectivity degree were defined as hub genes in the PPI network. The connectivity degree for each gene in the PPI network was calculated using the 'cytoHubba' (25) plugin in Cytoscape version 3.6.1 software (26). The hub genes common to both co-expression network and PPI networks were selected as 'real' hub genes and included for further analyses.

**Hub gene validation and survival analysis.** The Human Protein Atlas (27) was used to validate the expression of the hub genes at the protein level. The Human Protein Atlas is a publicly available database, all data and images are available for free download and non-commercial use. For validation of the correlation of hub genes and HCC progression, 371 HCC

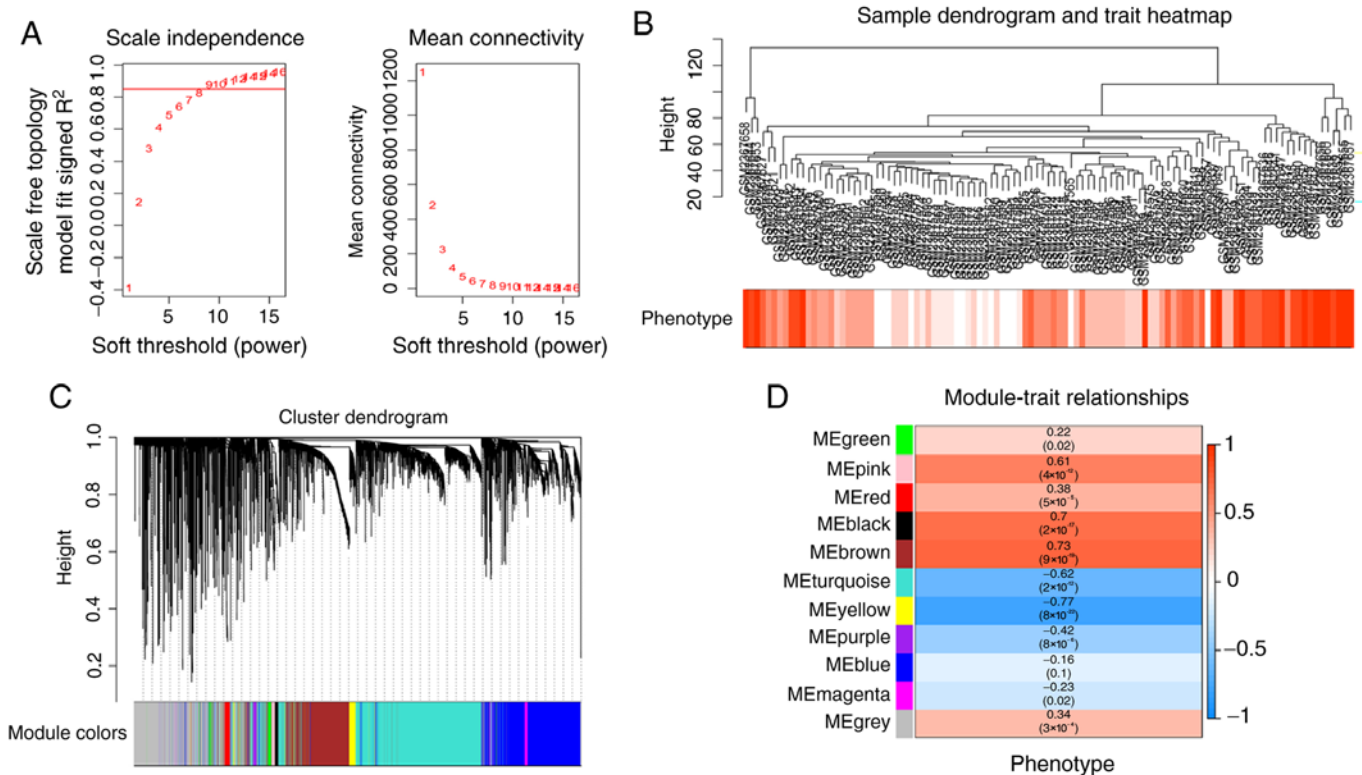


Figure 1. Weighted correlation network analysis in GSE89377. (A) Analysis of the scale-free fit index for various soft-thresholding powers ( $\beta$ ). (B) Clustering dendrogram of 107 samples and the clinical traits. Color intensity is proportional to higher grade (red) based on the theory of the multistep process hepatocarcinogenesis. (C) Dendrogram of genes clustered based on a dissimilarity measure (1-Topological Overlap Matrix). (D) Heatmap of the correlation between module eigengenes and clinical phenotypes in the GSE89377 dataset.

samples from TCGA database were analyzed to calculate the Pearson's correlation coefficient between hub gene expression and certain clinicopathological features. To evaluate the impact of the hub genes on the prognosis of patients with HCC, overall and disease-free survival rate were analyzed using Gene Expression Profiling Interactive Analysis tools (GEPIA) (28). The predictive value for prognosis between hub genes and routine clinicopathological factors were compared using univariate and multivariate Cox regression analyses in an HCC dataset from TCGA (TCGA-LIHC). These clinicopathological factors comprised alpha feto protein (AFP) (29), vascular invasion (30), Ishak score (32), and tumor pathological staging (33).  $P < 0.05$  were set as the cut-off criteria for significance.

**Gene set enrichment analysis (GSEA).** 371 HCC samples from RNA-sequencing data (displayed as read counts) were divided into two groups (high vs. low) according to the expression level of the candidate gene and the median expression value was selected as the cut-off point. The RNA-sequencing data was normalized using the limma package (34) in R. To determine the potential function of candidate gene, GSEA (35) was performed between the 2 groups. Hallmark gene sets summarize and represent specific well-defined biological states or processes and display coherent expression. These gene sets were generated by a computational methodology based on identifying overlaps between gene sets in other MSigDB collections (36) and retaining genes that display coordinate expression. Thus, the Hallmark gene sets (37) were selected

as the reference gene sets. FDR  $< 0.05$  was set as the cut-off criteria.

**Statistical analysis.** The expression levels of the hub genes were analyzed using unpaired Student's t-tests in the comparison of two groups. ANOVA and Dunnett's post-hoc test were used for multiple comparisons using the multcomp package (CRAN.R-project.org/package=multcomp) in R. Univariate/multivariate Cox proportional hazards analyses and Kaplan-Meier survival analysis with log-rank method were used to compare survival between the two groups of patients.  $P < 0.05$  was considered to indicate a statistically significant difference.

## Results

**Weighted co-expression network construction and primary module identification.** Overall, 13 healthy liver samples and 94 samples from different stages (low grade chronic hepatitis, high grade chronic hepatitis, cirrhosis, low grade dysplastic nodules, high grade dysplastic nodules, early HCC, HCC with grade 1, HCC with grade 2 and HCC with grade 3) of HCC progression were included in co-expression analysis (Fig. 1B). In the present study, the power of  $\beta = 9$  (scale free  $R^2 = 0.85$ ) was selected as the soft-threshold to ensure a scale-free network (Fig. 1A) and 11 modules were identified (Fig. 1C). The highest association between module and phenotype was revealed to be between the yellow module and clinical phenotype ( $r = -0.77$ ;  $P = 8 \times 10^{-22}$ ; Fig. 1D); however, the brown module and clinical

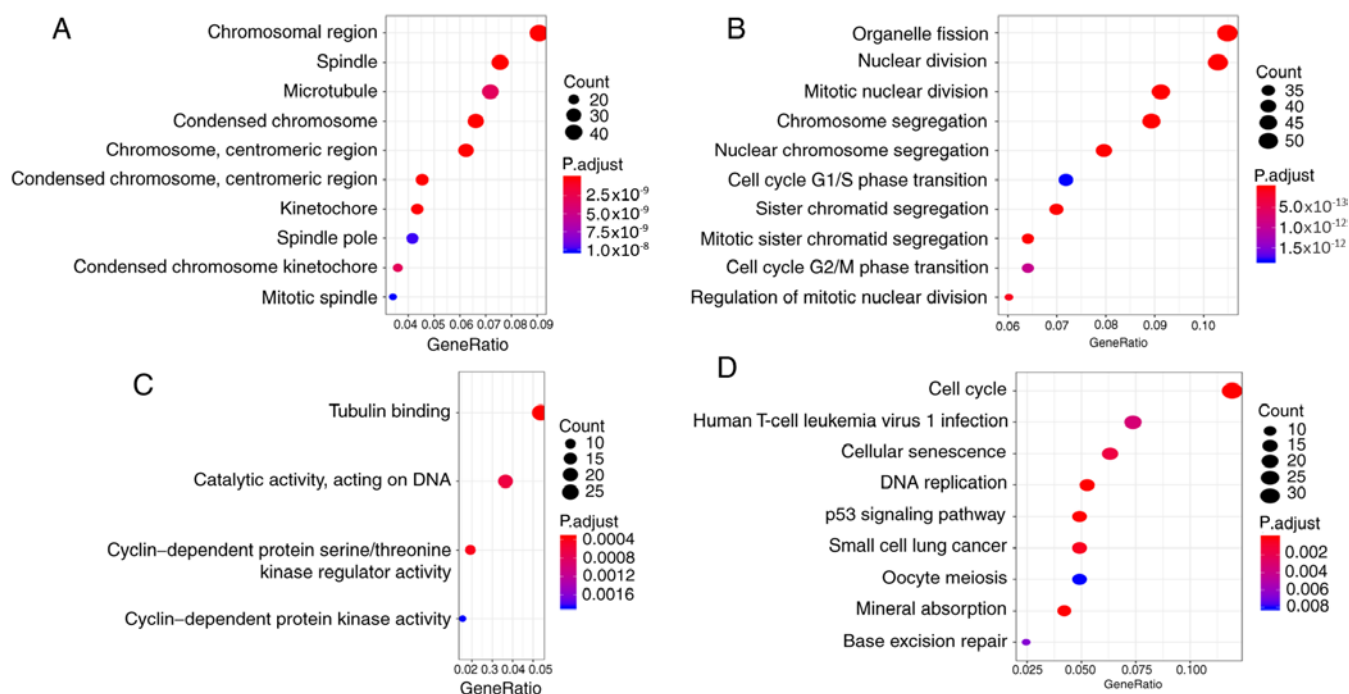


Figure 2. Significantly enriched Gene Ontology annotations and enriched KEGG and Genomes pathways of genes in the brown module. (A) Cellular component. (B) Biological process. (C) Molecular function. (D) KEGG pathway analysis. KEGG, Kyoto Encyclopedia of Genes.

phenotype exhibited the highest positive correlation ( $r=0.73$ ;  $P=9 \times 10^{-19}$ ; Fig. 1D). This indicated that the brown module genes may be oncogenes and the yellow module genes may be tumor suppressor genes. As there is more potential of oncogene as a marker or therapeutic target (21), the brown module was selected and included in subsequent analyses.

To explore the biological relevance of the brown module, GO and KEGG enrichment analyses were performed for 671 genes using the 'clusterProfiler' package. GO enrichment analyses contained three parts: Cellular component (CC; Fig. 2A); biological process (BP; Fig. 2B); and molecular function (MF; Fig. 2C). The brown module genes were involved in mitotic-related CCs and BPs, such as 'microtubule', 'nuclear division' and 'cell cycle G2/M phase transition'. While the brown module genes were involved in kinase-related MFs, such as 'cyclin-dependent protein kinase activity'. The results of enrichment analyses indicated that the brown module genes were involved in various cancer-associated pathways (Fig. 2D), such as 'cell cycle' and 'p53 signaling pathway'. The brown module genes associated with HCC are also involved in some viral infection-related pathways, such as human T-cell leukemia virus 1 infection.

**Identification of hub genes.** In the present study, 20 genes with high connectivity ( $\text{cor.geneModuleMembership}$ ;  $\geq 0.8$ ) and high clinical trait relationship ( $\text{cor.geneTraitSignificance}$ ;  $\geq 0.7$ ) in the brown module were selected as hub genes in WGCNA. A PPI network (Table SI; Fig. 3) was constructed using Cytoscape according to the STRING database and 50 genes with the highest connectivity degree were defined as hub genes in the PPI network. Then, a total of 10 genes (TOP2A, CDC20, CCNB2, PRC1, UBE2C, PTTG1, KIF20A, HMMR,

NUSAP1 and RACGAP1) common to both the co-expression network and the PPI network were selected as 'real' hub genes (Table I). The aberrant expression data of the 10 hub genes were validated in an independent data set (Fig. 4). HMMR was selected as the candidate gene for further analysis due to the few existing reports about its role in HCC.

**Hub gene validation and survival analysis.** The expression of HMMR at the mRNA and protein levels were both significantly higher in HCC tissue compared with healthy liver tissue (Fig. 5A-C). In the GSE89377 dataset, HMMR exhibited diagnostic efficiency for HCC with an area under curve (AUC)=0.949, sensitivity=0.875 and specificity=0.910 (Fig. 5D). Based the results of WGCNA, the expression of HMMR was positively correlated with the progression of HCC ( $\text{cor.geneTraitSignificance}$   $r=0.706$ ;  $P=2.00 \times 10^{-17}$ ). This correlation was validated in the HCC dataset from TCGA ( $r=0.290$ ;  $P=3.57 \times 10^{-6}$ ; Fig. 5E). The expression of HMMR was positively correlated with HCC pathological stage ( $r=0.062$ ;  $P=0.008$ ), tumor (T) stage ( $r=0.069$ ;  $P=4.19 \times 10^{-4}$ ) and Ishak score (Pearson correlation coefficient=0.178;  $P=0.004$ ; Fig. 5E). Using GEPIA tools, it was revealed that patients with higher expression levels of HMMR exhibited significantly shorter overall survival (Fig. 5F, left) and disease-free survival rate (Fig. 5F, right). Furthermore, the expression of HMMR is an independent prognostic factor compared with routine clinicopathological features, not only in overall survival but also disease-free survival rate, in the HCC dataset from TCGA (Tables II and III) (multivariate Cox regression analysis  $P<0.05$ ).

**GSEA.** To analyze the function of HMMR in HCC, GSEA was conducted to compare HMMR with hallmark gene sets. 371 HCC samples were divided into two groups (high vs.



Table I. Common hub genes in the brown module from weighted gene co-expression network analysis in GSE89377 and PPI network.

Probe	Gene	Weighted gene co-expression network analysis		PPI network
		cor.geneModuleMembership	cor.geneTraitSignificance	Connectivity degree
ILMN_1686097	TOP2A	0.956	0.748	142
ILMN_1663390	CDC20	0.961	0.724	121
ILMN_1801939	CCNB2	0.962	0.712	111
ILMN_1728934	PRC1	0.962	0.725	102
ILMN_1714730	UBE2C	0.954	0.710	97
ILMN_1753196	PTTG1	0.946	0.706	92
ILMN_1695658	KIF20A	0.938	0.715	92
ILMN_1781942	HMMR	0.916	0.706	97
ILMN_1726720	NUSAP1	0.933	0.709	86
ILMN_2077550	RACGAP1	0.959	0.717	83

PPI, Protein-Protein Interaction.

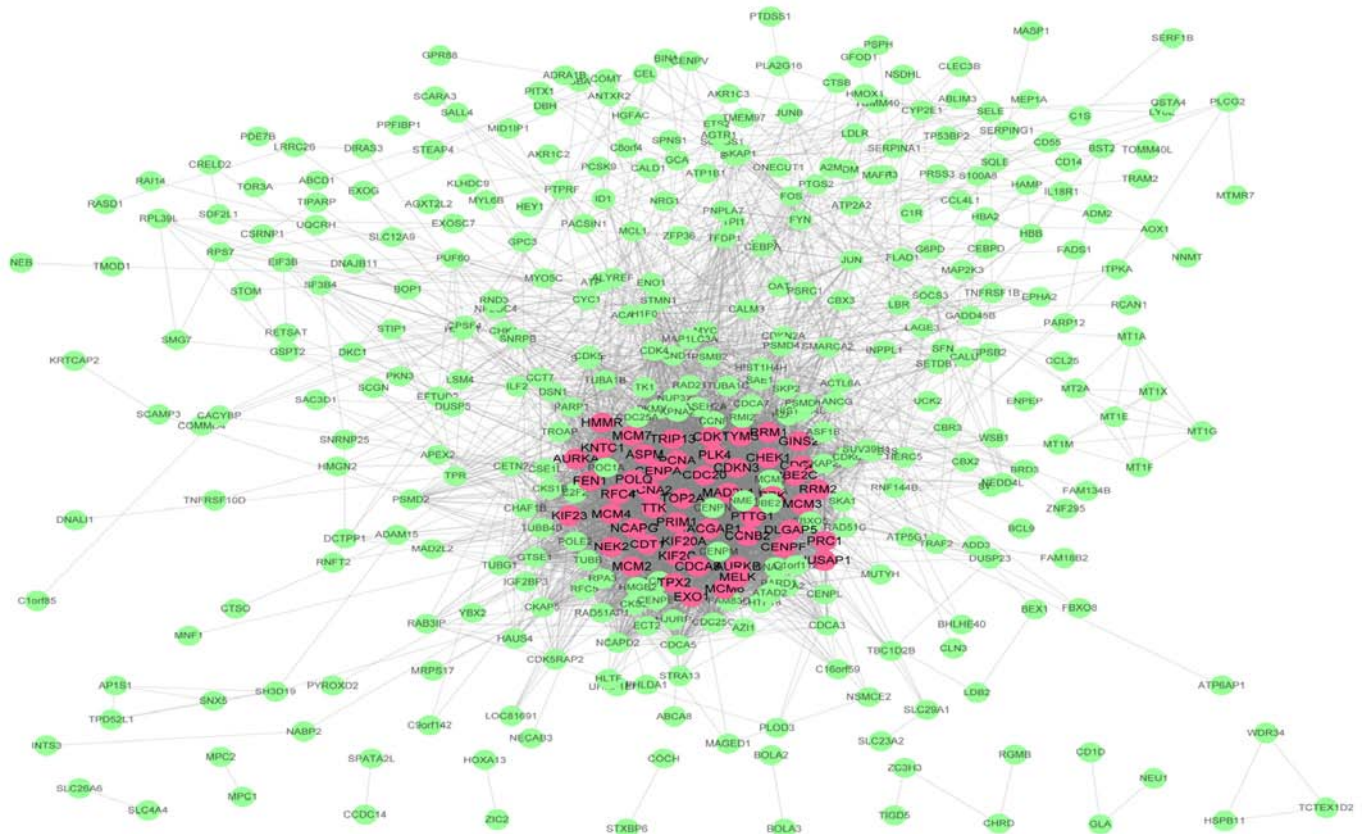


Figure 3. Protein-protein interaction networks of genes in the brown module. Red nodes were 50 genes with the highest connectivity degree and grey edges represent the interaction between proteins.

low) according to the HMMR median expression level (2.62). Under the cut-off criteria of FDR <0.05, a total of 8 functional gene sets were enriched. Overall, 6 representative gene sets were significantly associated with cancer, including 'mitotic spindle', 'G<sub>2</sub>/M checkpoint', 'MYC targets v1', 'E2F targets', 'DNA repair' and 'mTORC1 signaling' (Fig. 6).

## Discussion

In previous years, the concept of multi-step human hepatocarcinogenesis has been well documented (6,7,38). Chronic liver inflammation can result in repeated cell injury, death and regeneration cycles, resulting in subsequent epigenetic

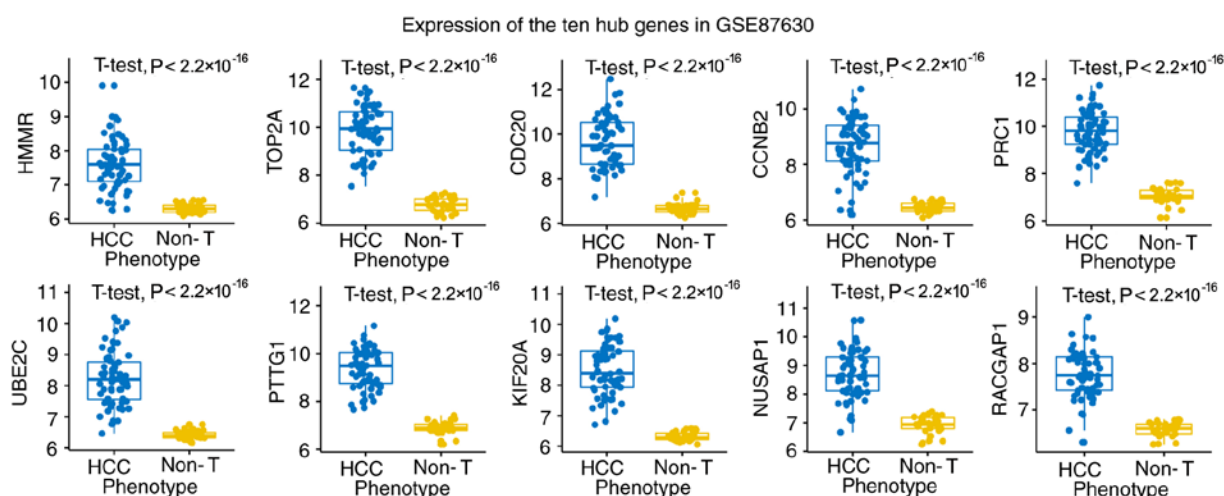


Figure 4. Aberrant expression of hub genes validated in HCC and non-tumor phenotypes of the GSE87630 dataset. HCC, hepatocellular carcinoma. Non-T, non-tumor.

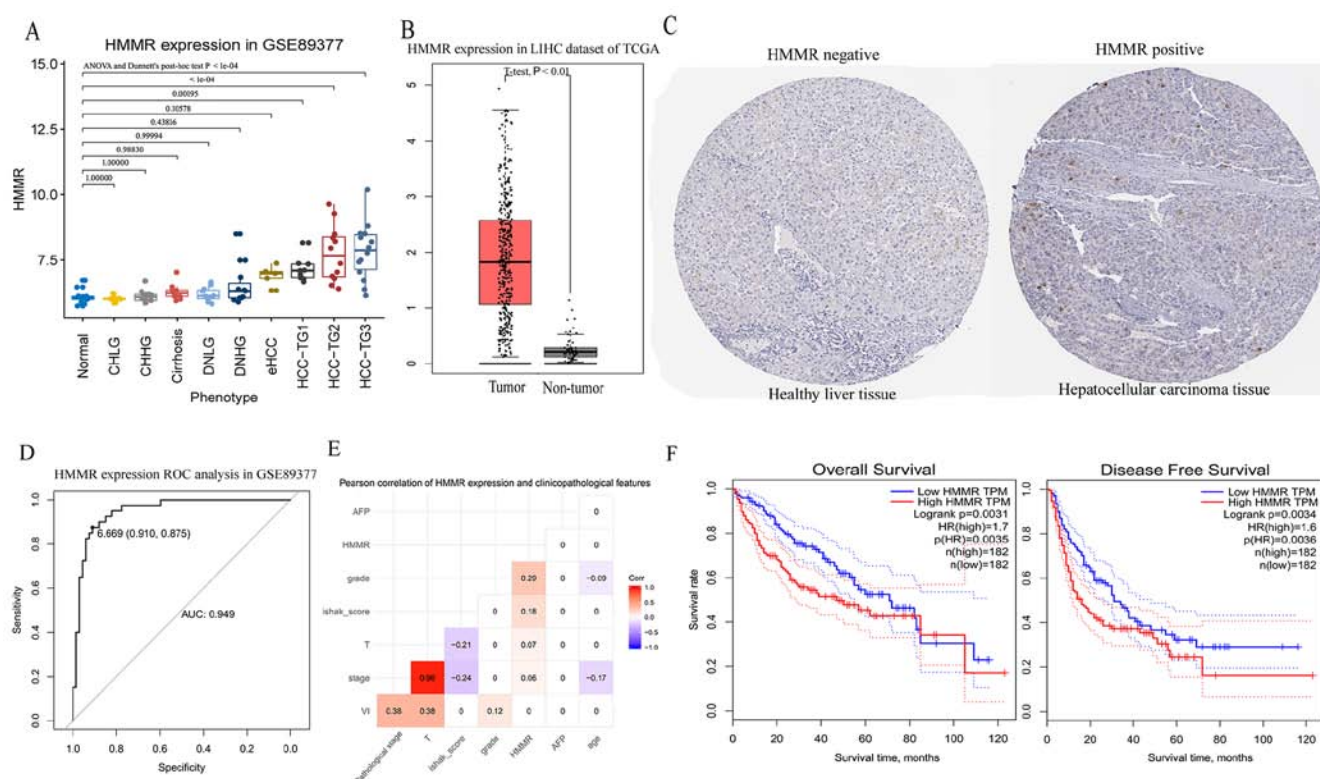


Figure 5. Validation of aberrant expression of HMMR at transcription and protein levels and the prognostic value of HMMR in HCC. (A) Expression of HMMR at different pathological stages from normal to chronic hepatitis/cirrhosis and dysplastic nodules to HCC in the GSE89377 dataset. (B) Expression of HMMR between tumor tissues and non-tumor tissues in hepatocellular carcinoma dataset from TCGA. (C) HMMR protein was upregulated in hepatocellular carcinoma (images.proteinatlas.org/2433/6865\_B\_8\_6.jpg) compared with healthy liver tissue (images.proteinatlas.org/2433/6892\_A\_8\_4.jpg) (antibody CAB002433) using data from the Human Protein Atlas database. The healthy liver tissue was from a female (patient ID: 1846) and the hepatocellular carcinoma tissue was from a male (patient ID: 2325). (D) ROC curves of the expression of HMMR for hepatocellular carcinoma diagnosis in the GSE89377 dataset. (E) Pearson correlation between HMMR expression and routine clinicopathological features. This shows the correlation coefficient when  $P < 0.01$ . (F) Kaplan-Meier curves obtained using the median value of HMMR expression to separate patients into high- and low-expression groups in Gene Expression Profiling Interactive Analysis. HCC, hepatocellular carcinoma; HMMR, Hyaluronan mediated motility receptor; LIHC, liver hepatocellular carcinoma; TCGA, The Cancer Genome Atlas; ROC, receiver operating characteristic; HR, hazard ratio; T, tumor; CHLG, chronic hepatitis with low grade; CHHG, chronic hepatitis with high grade; DNLG, dysplastic nodules with low grade; DNHG, dysplastic nodules with high grade; eHCC, early hepatocellular carcinoma; HCC-TG1, hepatocellular carcinoma with grade 1; HCC-TG2, hepatocellular carcinoma with grade 2; HCC-TG3, hepatocellular carcinoma with grade 3.

and genetic alterations of hepatocytes (39). Phenotypically abnormal precursor hepatic lesions, including cirrhotic nodules, low-grade dysplastic nodules and high-grade

dysplastic nodules dedifferentiate and gradually evolve to HCC (40). This process exists on a biologic continuum and may occur simultaneously at various rates throughout

Table II. Univariate and multivariate COX regression analyses for overall survival in hepatocellular carcinoma dataset of The Cancer Genome Atlas.

Factors	Univariate analysis			Multivariate analysis		
	P-value	HR	HR, 95% CI	P-value	HR	HR, 95% CI
Sex (male vs. female)	0.262	1.225	0.859-1.745	-	-	-
Age, years (>65 vs. ≤65)	0.186	1.265	0.893-1.791	-	-	-
AFP, ng/ml (>20 vs. ≤20) (29)	0.026 <sup>a</sup>	1.641	1.061-2.540	0.563	1.150	0.716-1.846
Vascular invasion (positive vs. negative) (30)	0.155	1.351	0.892-2.047	-	-	-
Child-pugh score (B/C vs. A) (31)	0.184	1.614	0.796-3.270	-	-	-
Ishak score (5-6 vs. 0-4) (32)	0.497	0.829	0.483-1.424	-	-	-
Tumor Grade (G3/4 vs. G1/2)	0.542	1.119	0.780-1.604	-	-	-
Pathological T stage (T3/4 vs. T1/2) (33)	<0.001 <sup>a</sup>	2.537	1.783-3.609	0.889	1.153	0.155-8.607
Pathological stage (III/IV vs. I/II) (33)	<0.001 <sup>a</sup>	2.446	1.687-3.545	0.634	1.620	0.222-11.808
HMMR expression level (high vs. low)	<0.001 <sup>a</sup>	2.136	1.498-3.044	0.007 <sup>a</sup>	1.917	1.192-3.085

<sup>a</sup>P<0.05. T, tumor; HR, hazard ratio; CI, confidence interval; HMMR, hyaluronan mediated motility receptor; AFP, alpha-fetoprotein.

Table III. Univariate and multivariate Cox regression analyses for disease-free survival in HCC dataset of TCGA.

Clinicopathological factors	Univariate analysis			Multivariate analysis		
	P-value	HR	HR (95% CI)	P-value	HR	HR (95% CI)
Sex (male vs. female)	0.919	1.019	0.714-1.454			
Age, years (>65 vs. ≤65)	0.081	1.043	0.739-1.472			
AFP, ng/ml (>20 vs. ≤20) (29)	0.496	1.149	0.771-1.712			
Vascular invasion (positive vs. negative) (30)	0.029 <sup>a</sup>	1.540	1.045-2.268	0.397	1.198	0.788-1.821
Child-pugh score (B/C vs. A) (31)	0.246	1.542	0.742-3.204			
Ishak score (5-6 vs. 0-4) (32)	0.623	1.116	0.721-1.727			
Tumor Grade (G3/4 vs. G1/2)	0.829	1.039	0.733-1.474			
Pathological T stage (T3/4 vs. T1/2) (33)	<0.001 <sup>b</sup>	2.940	2.071-4.173	0.518	0.512	0.067-3.902
Pathological stage (III/IV vs. I/II) (34)	<0.001 <sup>b</sup>	2.885	2.009-4.142	0.126	4.769	0.644-35.330
HMMR expression level (high vs. low)	0.002 <sup>b</sup>	1.697	1.213-2.373	0.038 <sup>a</sup>	1.527	1.024-2.277

<sup>a</sup>P<0.05, <sup>b</sup>P<0.01. HCC, hepatocellular carcinoma; TCGA, The Cancer Genome Atlas; HR, hazard ratio; CI, confidence interval; T, tumor; G, grade; AFP, alpha-fetoprotein; HMMR, Hyaluronan mediated motility receptor expression.

the liver; however, the specific molecular mechanisms underlying this process are yet to be elucidated. In the present study, several modules associated with this process were identified using WGCNA. In particular, genes in the brown module exhibited a strong positive correlation with this process, indicating that gene expression in the module gradually increase as the process progresses. Functional enrichment analysis revealed that genes in the brown module significantly influenced cell cycle-associated biological processes, for example 'cell cycle G<sub>2</sub>/M phase transition', 'cell cycle G<sub>1</sub>/S phase transition' and 'mitotic nuclear division', and cancer-related pathways, including 'p53 signaling pathway' and 'cell cycle'.

Markers which accurately reflect the process from normal to chronic hepatitis/cirrhosis and dysplastic nodules

to HCC are lacking in clinical practice and novel candidate molecules are needed. In the present study, according to the theory of the multistep process of hepatocarcinogenesis and WGCNA, a total of 10 hub genes common to the primary module and PPI network were selected as hub genes, including TOP2A, CDC20, CCNB2, PRC1, UBE2C, PTTG1, KIF20A, HMMR, NUSAP1 and RACGAP1. Previous studies had reported almost all ten genes to be associated with the progression of HCC (41-49). HMMR was chosen as the candidate gene, since few studies have identified its role in HCC (50). HMMR (also known as CD168/IHABP/RHAMM) (51) is highly expressed in various solid tumors and it is described as a cancer-associated antigen, which is involved in both tumorigenesis and progression/metastasis (52-56). HMMR was identified as a



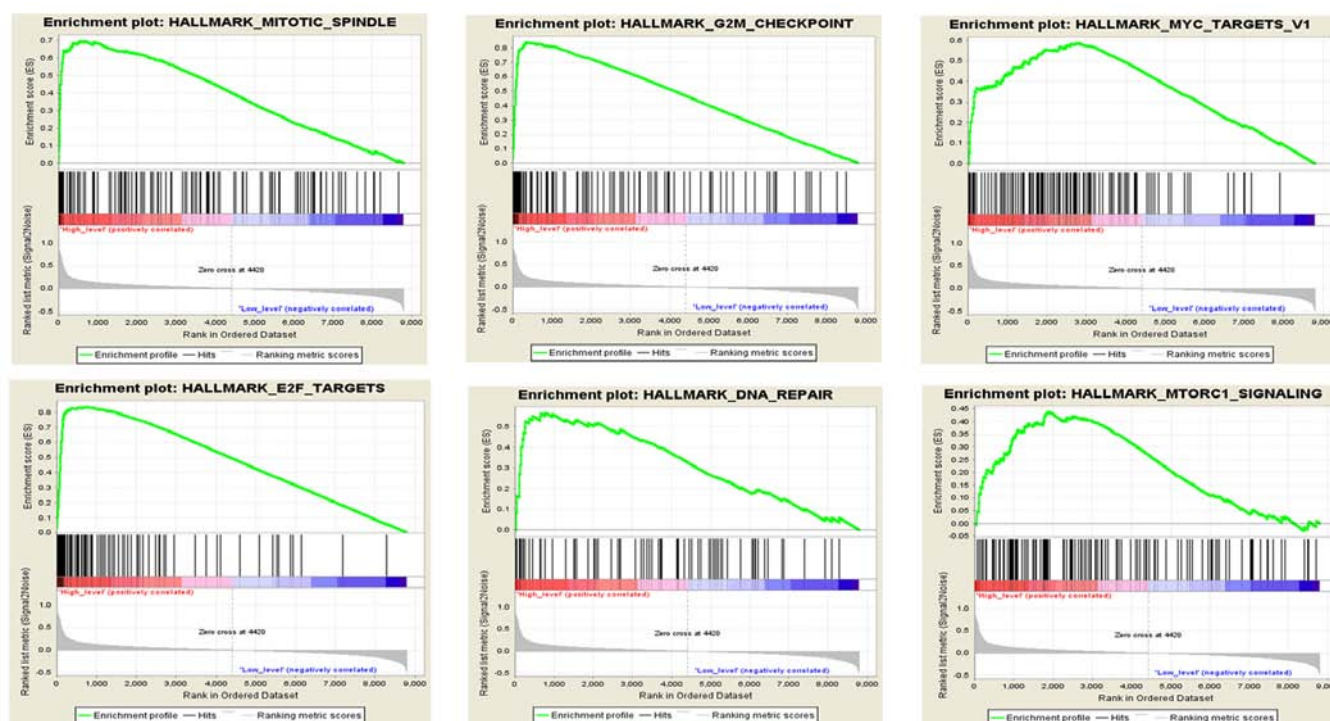


Figure 6. Gene set enrichment analysis. Overall, 6 representative gene sets were significantly enriched in hepatocellular carcinoma samples with high Hyaluronan mediated motility receptor expression.

breast cancer susceptibility gene (57) and was once considered an ideal target antigen for immunotherapy of acute myeloid leukemia (58); however, the association between HMMR and HCC has been rarely reported.

In the present study, HMMR was significantly upregulated in HCC tissue compared with healthy liver tissue at both the mRNA and protein expression levels. HMMR is a promising diagnostic biomarker for HCC (AUC=0.949; sensitivity=0.875; specificity=0.910). In addition, the progression of HCC was associated with the upregulation of HMMR. Notably, the expression of HMMR was positively correlated with HCC tumor grade, pathological stage, T stage and Ishak score. Patients with HCC with higher expression levels of HMMR exhibited significantly shorter overall survival and disease-free survival times. Moreover, the expression of HMMR remained an independent prognostic factor compared with routine clinicopathological features. The current results indicated that HMMR may serve as a biomarker of HCC progression. Thus, patients with HCC and high expression levels of HMMR have a higher risk for recurrence and should be followed up more frequently than the routine schedule.

In order to reveal the function of HMMR in HCC, GSEA was performed. Overall, 6 representative gene sets, including 'mitotic spindle', 'G<sub>2</sub>/M checkpoint', 'MYC targets v1', 'E2F targets', 'DNA repair' and 'mTORC1 signaling', were significantly associated with cancer and enriched in samples with high expression levels of HMMR. This indicates that HMMR may interact with these genes or pathways to promote the progression of HCC. The present findings may improve our understanding of the role of HMMR in HCC and inform future research.

Notably, there were certain limitations to the present study. Firstly, the expression of HMMR was quantified and the values may vary on different platforms. The establishment of a standard is required before being applied to clinical practice. Secondly, as the present study only performed a bioinformatic analysis, including GSEA analysis to help identify the function of HMMR in HCC, it is not clear whether HMMR expression is causal or merely a biomarker of HCC progression. Whether HMMR can be used as a therapeutic target for HCC requires further molecular experimental verification.

In conclusion, the present study revealed that patients with HCC with high expression of HMMR exhibit a less favorable prognosis, suggesting that HMMR may serve an important role in HCC and has potential as a biomarker of HCC diagnosis and progression.

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## Availability of data and materials

The dataset of GSE89377 and GSE87630 are available from the Gene Expression Omnibus (<https://www.ncbi.nlm.nih.gov/geo>). The dataset of TCGA-LIHC are available from The Cancer Genome Atlas ([cancer.gov/tcga](https://cancer.gov/tcga)).



## Authors' contributions

YL designed the study and reviewed the manuscript. DL and XB analyzed the data and wrote the manuscript. ZG and QZ assisted with analyzing the data and writing the manuscript. All authors read and approved the final manuscript.

## Ethics approval and consent to participate

Not applicable.

## Patient consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

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