

Helicobacter pylori induces epithelial-mesenchymal transition in gastric carcinogenesis via the AKT/GSK3 β signaling pathway

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Abstract. *Helicobacter pylori* (*H. pylori*) is a main risk factor for gastric cancer (GC). Epithelial-mesenchymal transition (EMT) is involved in the development and progression of *H. pylori*-associated GC. However, the exact molecular mechanism of this process remains unclear. The AKT/GSK3 β signaling pathway has been demonstrated to promote EMT in several types of cancer. The present study investigated whether *H. pylori* infection induced EMT, and promoted the development and metastasis of cancer in the normal gastric mucosa, and whether this process was dependent on AKT activation. The expression levels of the EMT-associated proteins, including E-cadherin and N-cadherin, were determined in 165 gastric mucosal samples of different disease stages by immunohistochemical analysis. The expression levels of E-cadherin, N-cadherin, AKT, phosphorylated (p-) AKT (Ser473), GSK3 β and p-GSK3 β (Ser9) were further determined in *H. pylori*-infected Mongolian gerbil gastric tissues and cells co-cultured with *H. pylori* by immunohistochemical analysis and western blotting. The results indicated that the expression levels of the epithelial marker E-cadherin were decreased, whereas the expression levels of the mesenchymal marker N-cadherin were increased during gastric carcinogenesis. Their expression levels were associated with *H. pylori* infection. Furthermore, *H. pylori* infection resulted

in downregulation of E-cadherin expression and upregulation of N-cadherin expression in Mongolian gerbils and GES-1 cells. In addition, an investigation of the associated mechanism of action revealed that p-AKT (Ser473) and p-GSK3 β (Ser9) were activated in GES-1 cells following co-culture with *H. pylori*. Furthermore, following pretreatment of the cells with the AKT inhibitor VIII, the expression levels of E-cadherin, N-cadherin, p-AKT and p-GSK3 β did not show significant differences between GES-1 cells that were co-cultured with or without *H. pylori*. The levels of p-AKT and p-GSK3 β were increased in *H. pylori*-infected Mongolian gerbils. In conclusion, the present study demonstrated that *H. pylori* infection activated AKT and resulted in the phosphorylation and inactivation of GSK3 β , which in turn promoted early stage EMT. These effects were AKT-dependent. This mechanism may serve as a prerequisite for GC development.

Introduction

Although its incidence is declining, gastric cancer (GC) is still a leading cause of cancer-associated mortality and a major socio-economic burden worldwide (1). *Helicobacter pylori* (*H. pylori*) infection is a major risk factor of GC (2). Environmental factors are subordinate to the effect of *H. pylori* infection, which has been listed as a class I carcinogen for GC by the World Health Organization since 1994 (2). *H. pylori* infection causes gastric mucosal inflammation, as well as genetic and epigenetic changes, which lead to genetic instability in host gastric epithelial cells (2). However, the exact molecular mechanism of *H. pylori*-associated GC is not yet clear (3). At present, several studies have demonstrated that *H. pylori* induces the epithelial-mesenchymal transition (EMT) process in gastric epithelial cells, which promotes invasion and metastasis of GC via the TGF- β , NF- κ B and Hippo signaling pathways (4-6).

EMT is a process in which epithelial cells lose their polarity and transform their morphology by developing fiber-like structures (7). Simultaneously, the contacts with the surrounding cells and cell adhesion are reduced and an increased migratory and invasive activity is noted. Both of these processes promote tumor metastasis (7). The molecular

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changes of EMT are mainly characterized by the down-regulation of the expression of epithelial cell markers and the upregulation of the expression of mesenchymal phenotypic markers (8). In particular, the absence of E-cadherin is an important EMT signal and a prerequisite for tumor cell invasion (9). The functional molecules of mesenchymal cells are mainly skeletal proteins derived from mesenchyme or embryos (e.g., N-cadherin) that are normally not expressed or expressed at low levels in epithelial cells, while the expression of these molecules is increased during the EMT process (10).

Multiple signaling pathways are involved in the development of EMT (11). For example, hepatocyte growth factor/scatter factor and early growth response gene 1 synergistically activate the MAPK signaling pathway, which enhances the activity of snail and inhibits the expression of the epithelial marker E-cadherin, promoting EMT (12). The AKT/GSK3 β signaling pathway has attracted considerable attention due to its involvement in EMT (13). AKT, which is also known as protein kinase B, is a serine/threonine protein kinase associated with a number of signaling pathways that promote cell proliferation, survival and inhibition of apoptosis (14). GSK3 β is a serine/threonine protein kinase that acts downstream of AKT (15). Unlike other kinases, GSK3 β is normally dephosphorylated in its active state. The phosphorylation of GSK3 β (Ser9) by phosphorylated (p-)AKT leads to inhibition of its activity (15). Activation of AKT results in GSK3 β phosphorylation, which reduces β -catenin degradation, promotes Snail expression and eventually induces EMT in combination with T cell factor/lymphocyte enhancer factor (13,16).

Previous studies demonstrated that *H. pylori* activated the AKT signaling pathway in human gastric epithelial cells (GES-1), thereby causing disorders in cell proliferation and apoptosis (17,18). However, to the best of our knowledge, the role of the AKT/GSK3 β signaling pathway in *H. pylori*-associated EMT is not yet clear. The aim of the present study was to investigate whether *H. pylori* infection induces EMT and promotes the development and metastasis of cancer in normal gastric mucosa. In addition, the present study investigated whether this process was dependent on AKT activation. The pathogenic mechanism of *H. pylori* was thoroughly examined and the data provided a novel theoretical basis for the development of prevention strategies for *H. pylori* infection-associated diseases.

Materials and methods

Patients and sample collection. Between January 2007 and September 2012, 165 formalin-fixed (10% formalin at room temperature for 24 h) and paraffin-embedded gastric tissue samples from 165 patients who underwent gastroduodenoscopy were obtained from the Digestive Centre of The First Affiliated Hospital of Nanchang University (Nanchang, China). Patients aged between 18-70 years and without previous *H. pylori* treatment history were enrolled in the present study, and patients who underwent gastric surgery were excluded. The clinical characteristics of patients were stratified according to pathological diagnosis and are shown in Table I. The patients had a median age of 55 years (range, 36-69 years) and 86 patients (52%) were male. Among these patients, 19 out of the 39 who were diagnosed with chronic non-atrophic gastritis (CNAG),

19 out of the 40 who had intestinal metaplasia (IM), 19 out of the 39 who had dysplasia (Dys) and 22 out of the 47 who had GC were *H. pylori*-positive. The age and gender distribution of these patients were compared and the data revealed no significant differences ($P>0.05$; Table I). Gastric tissue samples were collected from the gastric antrum and from the individual lesions of each patient. Pathological diagnosis and classification were based on the criteria of the World Health Organization and the updated Sydney system (19,20). A non-atrophy gastritis diagnosis was made when all biopsies indicated a non-atrophy gastritis result (19,20). The rapid urease test and Giemsa staining were used for the detection of *H. pylori* infection, as previously described (21). The present study was approved by the Human Ethics Committee of the First Affiliated Hospital of Nanchang University (Nanchang, China). All patients signed an informed consent form for their participation in the study.

***H. pylori* strain and culture conditions.** The wild-type *H. pylori* CagA⁺ VaCA⁺ strain (ATCC43504; American Type Culture Collection) was used in the present study. Briefly, *H. pylori* was cultured on Campylobacter agar (Oxoid; Thermo Fisher Scientific, Inc.) plates with 5% sheep blood (Gibco; Thermo Fisher Scientific, Inc.) and incubated at 37°C with 5% CO₂ for subsequent *in vitro* experiments. Following 24 h of incubation, *H. pylori* strains were grown in Brucella broth (Gibco; Thermo Fisher Scientific, Inc.) with 10% sheep serum (Gibco; Thermo Fisher Scientific, Inc.) for 16 h under microaerophilic conditions (5% O₂, 10% CO₂ and 85% N₂). *H. pylori* co-culture experiments with GES-1 cells were conducted at an MOI of 50:1.

Animals and infections. Male Mongolian gerbils (specific pathogen-free; age, 5-8 weeks; weight, 30-50 g; obtained from Zhejiang Academy of Medical Sciences (Zhejiang, China) <http://www.zjams.com.cn/>) were utilized in the present study between August 2018 and June 2019. A total of 79 gerbils were maintained in an isolated clean room with regulated temperature (20-22°C), humidity (~55%) and a 12/12-h light/dark cycle, with *ad libitum* access to rodent diet and water as previously described, animal health and behavior were monitored twice a day, and no gerbils were found dead during experiments (22). Following fasting for 12 h, the animals were orogastrically challenged with 0.2 ml sterile Brucella broth, which was used as a control group (n=25) or *H. pylori* strains (n=54) at 10 time points (every 3 days). The Mongolian gerbils were humanely euthanized via CO₂ inhalation (30% of the chamber volume per min) at 9 months after the challenge and gastric tissues were harvested. Euthanasia was confirmed by observation of each gerbil for lack of respiration and faded eye color. The First Affiliated Hospital of Nanchang University Ethics Committee (Nanchang, China) approved all experiments and procedures.

Cell line and reagents. The immortalized human gastric cell line GES-1 (Beijing Institute for Cancer Research, Beijing, China) was grown in DMEM (Gibco; Thermo Fisher Scientific, Inc.) with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.) and 1% penicillin/streptomycin at 37°C with 5% CO₂. The AKT inhibitor VIII was purchased from Merck KGaA.

Table I. Clinical characteristics of patients with different stages of gastric carcinogenesis.

Characteristic	CNAG	IM	Dys	GC	P-value
No.	39	40	39	47	
Age, years (mean \pm SD)	50.80 \pm 14.08	54.32 \pm 12.18	56.68 \pm 10.81	55.06 \pm 14.01	>0.05
Male, n	20	21	20	25	>0.05
Female, n	19	19	19	22	>0.05
<i>H. pylori</i> positive, n	19	19	19	22	>0.05
<i>H. pylori</i> negative, n	20	21	20	25	>0.05

CNAG, chronic non-atrophic gastritis; IM, intestinal metaplasia; Dys, dysplasia; GC, gastric cancer; *H. pylori*, *Helicobacter pylori*. One-way ANOVA test (age) or the χ^2 test (sex, *H. pylori* infection status) were used to determine the differences among groups.

Western blotting. GES-1 cells were co-cultured with *H. pylori* strain 43504, and total protein was extracted in RIPA lysis buffer (Beyotime Institute of Biotechnology). Protein concentration was determined using a BCA assay kit (Thermo Fisher Scientific, Inc.). Protein (20 μ g/lane) from each sample was separated by 10% SDS-PAGE and transferred to PVDF membranes, as previously described (21). Following blocking with 5% non-fat milk at room temperature for 1 h, the blots were incubated with the primary antibodies overnight at 4°C and with HRP-conjugated secondary antibodies (dilution, 1:5,000; cat. no. ZB-2301; Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd.) at 4°C for 4 h. The protein bands were visualized using the Super-Signal West Femto Maximum Sensitivity Substrate Kit (Thermo Fisher Scientific, Inc.) and exposed by the Bio-Rad Image Lab system (Bio-Rad Laboratories, Inc.). The band intensities of the target proteins were normalized to those of β -actin using Image Lab software (version 5.2.1; Bio-Rad Laboratories, Inc.). The primary antibodies and their sources were as follows: E-cadherin (anti-E-cadherin antibody; dilution, 1:1,000; cat. no. ab1416; Abcam), N-cadherin (anti-N-cadherin antibody; dilution, 1:1,000; cat. no. ab18203; Abcam), AKT (anti-AKT antibody; dilution, 1:1,000; cat. no. 4691; Cell Signaling Technology, Inc.), p-AKT (Ser473) (anti-p-AKT antibody; dilution, 1:1,000; cat. no. 9271; Cell Signaling Technology, Inc.), GSK3 β (anti-GSK3 β antibody; dilution, 1:1,000; cat. no. 9315S; Cell Signaling Technology, Inc.), p-GSK3 β (Ser9) (anti-p-GSK3 β antibody; dilution, 1:1,000; cat. no. 9322; Cell Signaling Technology, Inc.) and β -actin (β -actin antibody; dilution, 1:1,000; cat. no. sc-1615-R; Santa Cruz Biotechnology, Inc.).

Immunohistochemical analysis. Biopsy specimens and animal gastric tissues were processed for immunohistochemical analysis as previously described (21). Briefly, specimens were sectioned at 4- μ m thickness and baked at 70°C for 1.5 h. Subsequently, sections were de-paraffinized in xylene and rehydrated with gradient ethanol (100, 95 and 85% ethanol). The sections were repaired with citrate buffer solution (TransGen Biotech Co., Ltd.) at 100°C for 15 min, and then washed in PBS solution (TransGen Biotech Co., Ltd.) three times. Following blocking with 3% H₂O₂ solution at room temperature for 8 min and incubation with 5% goat serum (Beyotime Institute of Biotechnology) at room temperature for 30 min, sections were incubated with the primary antibodies at

4°C overnight and with secondary antibodies (dilution, 1:100; 100 μ l per section; cat. no. PV-6000; Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd.) at 37°C for 30 mins. Sections were stained with DAB solution (Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd.) and evaluated under a fluorescence microscope (Olympus bx53 biomicroscope; Olympus Corporation). The samples were stained with anti-E-cadherin antibody (biopsy and gastric tissues; dilution, 1:300; cat. no. ab1416; Abcam), anti-N-cadherin antibody (biopsy and gastric tissues; dilution, 1:800; cat. no. ab18203; Abcam), anti-AKT antibody (gastric tissues; dilution, 1:500; cat. no. ab8805; Abcam), anti-p-AKT antibody (Ser473) (gastric tissues; dilution, 1:500; cat. no. ab66138; Abcam), anti-GSK3 β antibody (gastric tissues; dilution, 1:400; cat. no. ab98081; Abcam) and anti-p-GSK3 β antibody (Ser9) (gastric tissues; dilution, 1:200; cat. no. ab75814; Abcam). Two pathologists (Department of Pathology, The First Affiliated Hospital of Nanchang University, Nanchang, China) who were blinded to the clinicopathological data scored (0-12 points) and assessed (grade 1-4) the stained sections by evaluating and calculating the intensity (0-3 points) and frequency (0-4 points) of staining. The relative expression levels of the proteins were reported as grades as follows: Grade 1 (0-2 points), grade 2 (3-5 points), grade 3 (6-8 points) and grade 4 (9-12 points) (18). Grading discrepancies were re-reviewed and discussed to obtain a final score.

Statistical analysis. The results are presented as the mean \pm standard deviation of three independent experiments or the percentage compared with the control samples. The χ^2 and Mann-Whitney tests were used to analyze the statistical difference between two groups. One-way ANOVA followed by Tukey's post hoc test or the Kruskal-Wallis test followed by Bonferroni post hoc test were used to determine the differences among multiple groups based on the normality of the data (SPSS software; version 18.0; SPSS, Inc.). P \leq 0.05 was considered to indicate a statistically significant difference.

Results

Expression levels of EMT-associated proteins in gastric carcinogenesis. To evaluate the expression levels of EMT-associated proteins in patients with gastric carcinogenesis, an immunohistochemical analysis of human gastric tissues of different gastric lesions was performed. The expression

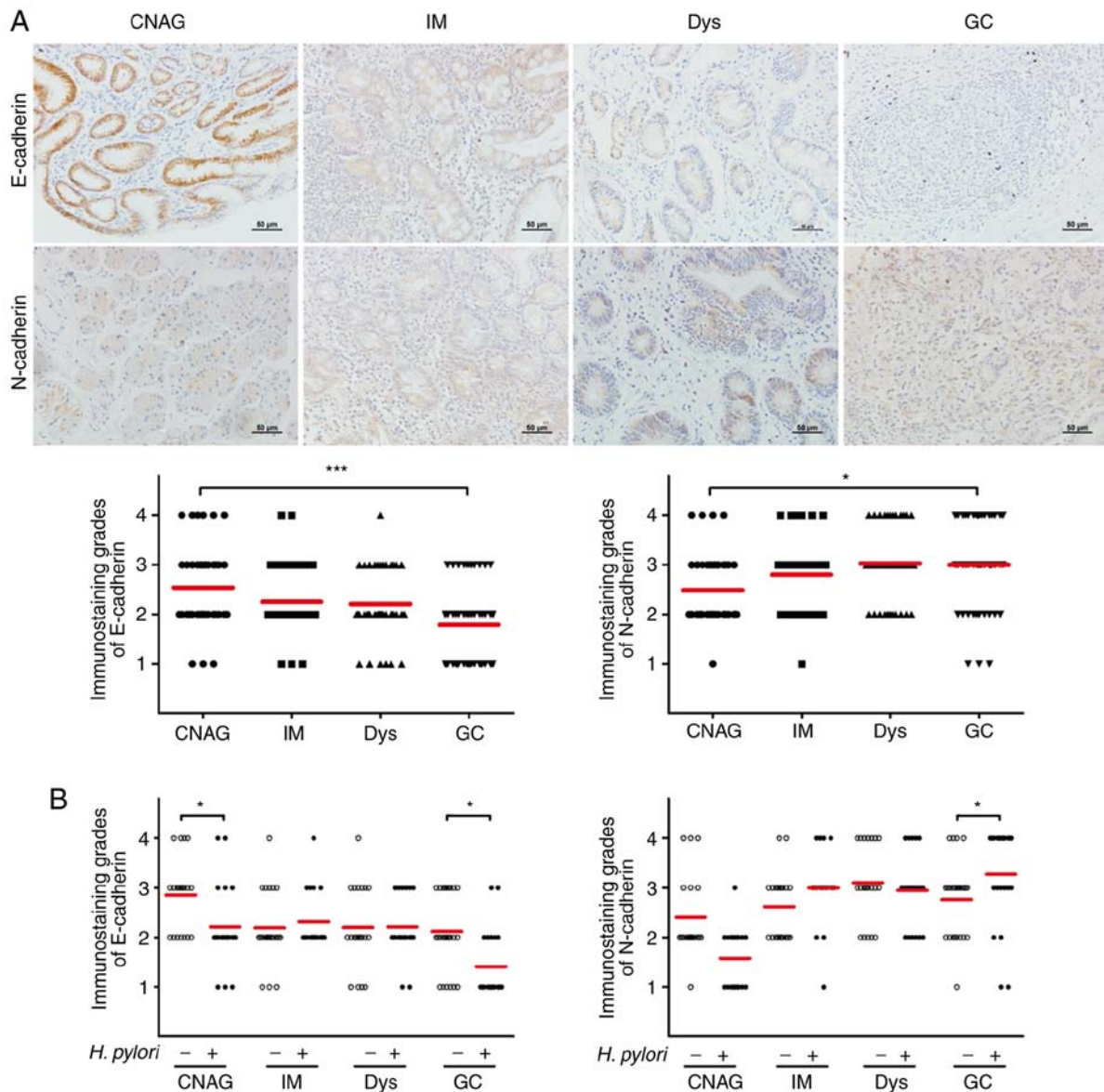


Figure 1. Detection of the expression levels of EMT-associated proteins in gastric carcinogenesis. (A) Expression levels of the EMT-associated markers E-cadherin and N-cadherin in 165 gastric tissue samples at different disease stages (CNAG, IM, Dys and GC) were detected by immunohistochemical analysis. The expression grades were assessed by the staining intensity and frequency. Scale bar, 50 μ m. (B) Expression levels of E-cadherin and N-cadherin were assessed in 165 gastric tissue samples at different disease stages according to *H. pylori* infection status. *H. pylori* infection was assessed by rapid urease tests and Giemsa staining. * $P < 0.05$, *** $P < 0.001$. EMT, epithelial-mesenchymal transition; CNAG, chronic non-atrophic gastritis; IM, intestinal metaplasia; Dys, dysplasia; GC, gastric cancer; *H. pylori*, *Helicobacter pylori*.

levels of the epithelial marker E-cadherin were decreased at the IM stage compared with CNAG stage and continued to decrease in the GC stage. E-cadherin expression was significantly reduced in GC tissues compared with in CNAG tissues ($P < 0.001$). However, the expression levels of the mesenchymal marker N-cadherin were increased at the IM stage compared with CNAG stage and continued to increase in the subsequent stages of gastric carcinogenesis. N-cadherin expression was significantly increased in GC tissues compared with in CNAG tissues ($P < 0.05$; Fig. 1A). These results indicated that EMT may be involved in the initiation of the early stage of gastric carcinogenesis.

H. pylori is a known important risk factor that triggers gastric carcinogenesis (2). The association between *H. pylori* infection and the expression levels of E-cadherin and

N-cadherin was examined in the patients. The immunohistochemical results indicated that the expression levels of E-cadherin were significantly lower following *H. pylori* infection compared with those of non-infected patients ($P < 0.05$) in CNAG and GC stage, whereas the expression levels of N-cadherin were significantly higher following *H. pylori* infection compared with those of non-infected patients ($P < 0.05$; Fig. 1B) in GC stage. There was no significant difference in E-cadherin and N-cadherin expression in IM and Dys stages. Overall, these results suggested that *H. pylori* infection may induce EMT.

H. pylori infection induces EMT in *GES-1* cells and Mongolian gerbils. The initial observation demonstrated an association between *H. pylori* infection and the expression

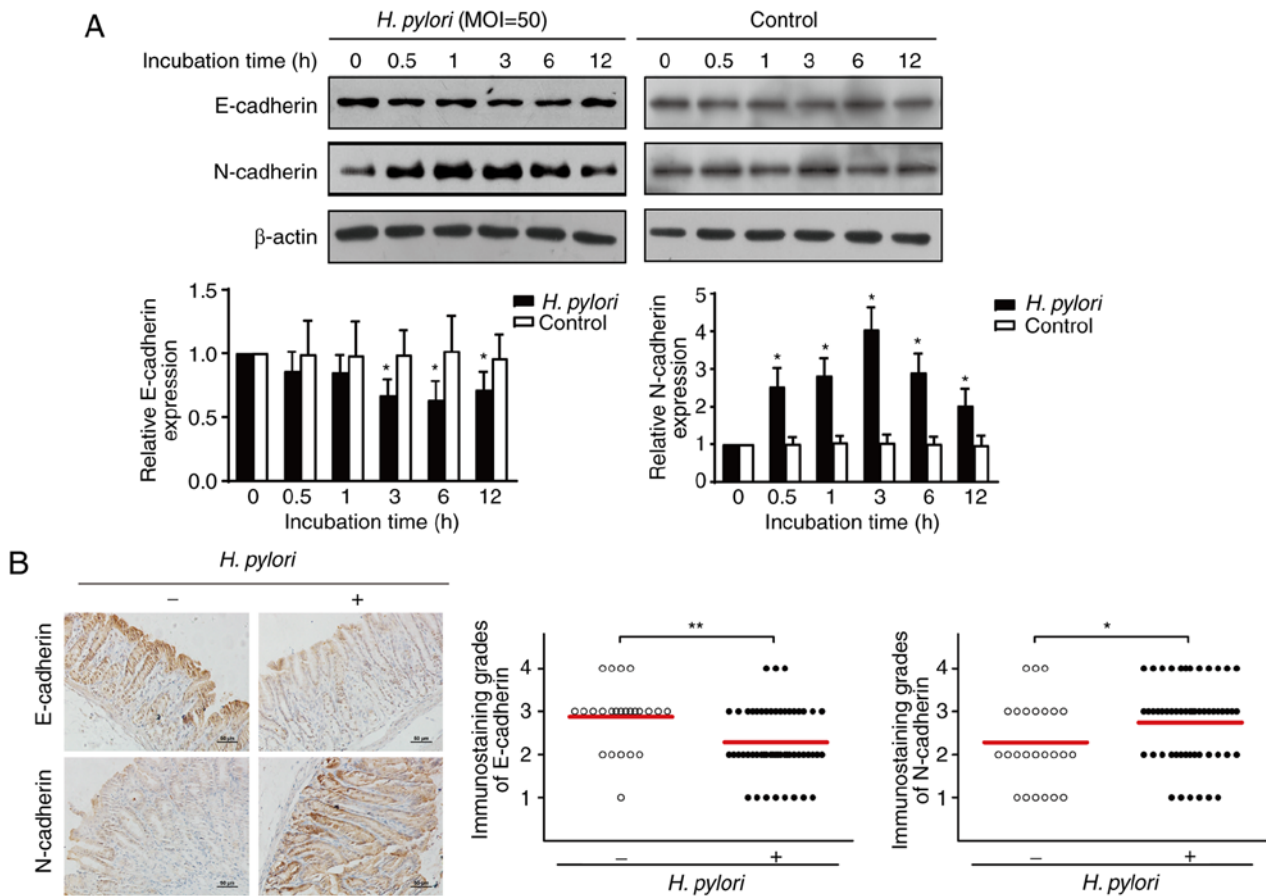


Figure 2. *H. pylori* infection induces EMT in GES-1 cells and Mongolian gerbils. (A) Expression levels of the EMT-associated markers E-cadherin and N-cadherin were detected by western blot analysis in GES-1 cells following 0, 0.5, 1, 3, 6 and 12 h of co-culture with or without *H. pylori*. The statistical analysis of three independent experiments is presented. (B) Expression levels of the EMT-associated markers E-cadherin and N-cadherin were detected by immunohistochemical analysis of the gastric mucosa of Mongolian gerbils with or without *H. pylori* infection. The expression grades were assessed by the intensity and frequency of staining. Scale bar, 50 μ m. * P <0.05, ** P <0.01 vs. 0 h or as indicated. EMT, epithelial-mesenchymal transition; *H. pylori*, *Helicobacter pylori*.

levels of EMT-associated markers in human gastric tissues. Subsequently, the effects of *H. pylori* infection on EMT were investigated in gastric epithelial GES-1 cells and in an animal model. GES-1 cells were co-cultured with *H. pylori* and cellular proteins were extracted at different time points. Protein detection was performed by western blotting. The results demonstrated that *H. pylori* infection significantly decreased the expression levels of E-cadherin following 3 h of co-culture, whereas it increased the expression levels of N-cadherin following 0.5 h of co-culture (P <0.05; Fig. 2A). Subsequently, immunohistochemical analysis was performed in order to examine the expression levels of E-cadherin and N-cadherin in gastric tissues of Mongolian gerbils following *H. pylori* infection and in the corresponding tissues of the control group. *H. pylori* infection significantly increased the expression levels of N-cadherin and decreased the expression levels of E-cadherin (P <0.05; Fig. 2B). In summary, these *in vivo* and *in vitro* data demonstrated that *H. pylori* infection may induce EMT.

H. pylori-induced EMT is regulated via the AKT/GSK3 β signaling pathway. The molecular mechanisms of *H. pylori* infection were investigated with regard to EMT induction. The AKT/GSK3 β signaling pathway has previously been

demonstrated to induce EMT (13). To investigate whether the AKT/GSK3 β signaling pathway was involved in *H. pylori*-induced EMT, the effects of *H. pylori* infection on the AKT/GSK3 β signaling pathway were further investigated in GES-1 cells. Western blot analysis demonstrated that the relative expression ratio of p-AKT (p-AKT/AKT) was significantly increased in GES-1 cells following incubation of the cells with *H. pylori* for 0.5, 1, 3 and 6 h, and no significant difference was observed at 12 h compared with in cells in the group without *H. pylori* incubation (P <0.05; Fig. 3A). The relative expression ratio of p-GSK3 β (p-GSK3 β /GSK3 β) was significantly increased in GES-1 cells that were incubated with *H. pylori* for 3, 6 and 12 h compared with in cells in the group without *H. pylori* incubation (P <0.05; Fig. 3A).

The role of AKT in *H. pylori*-induced EMT was further explored. As shown in Figs. 2 and 3, *H. pylori*-infected GES-1 cells exhibited a prominent increase in the levels of N-cadherin, p-AKT and p-GSK3 β , whereas a decrease was noted in the expression levels of E-cadherin compared with the corresponding expression levels of the control group (P <0.05). The results were similar in GES-1 cells that were pre-treated with DMSO. *H. pylori* infection did not result in a change in the expression levels of E-cadherin, N-cadherin, p-AKT and p-GSK3 β in GES-1 cells that were pre-treated with the AKT

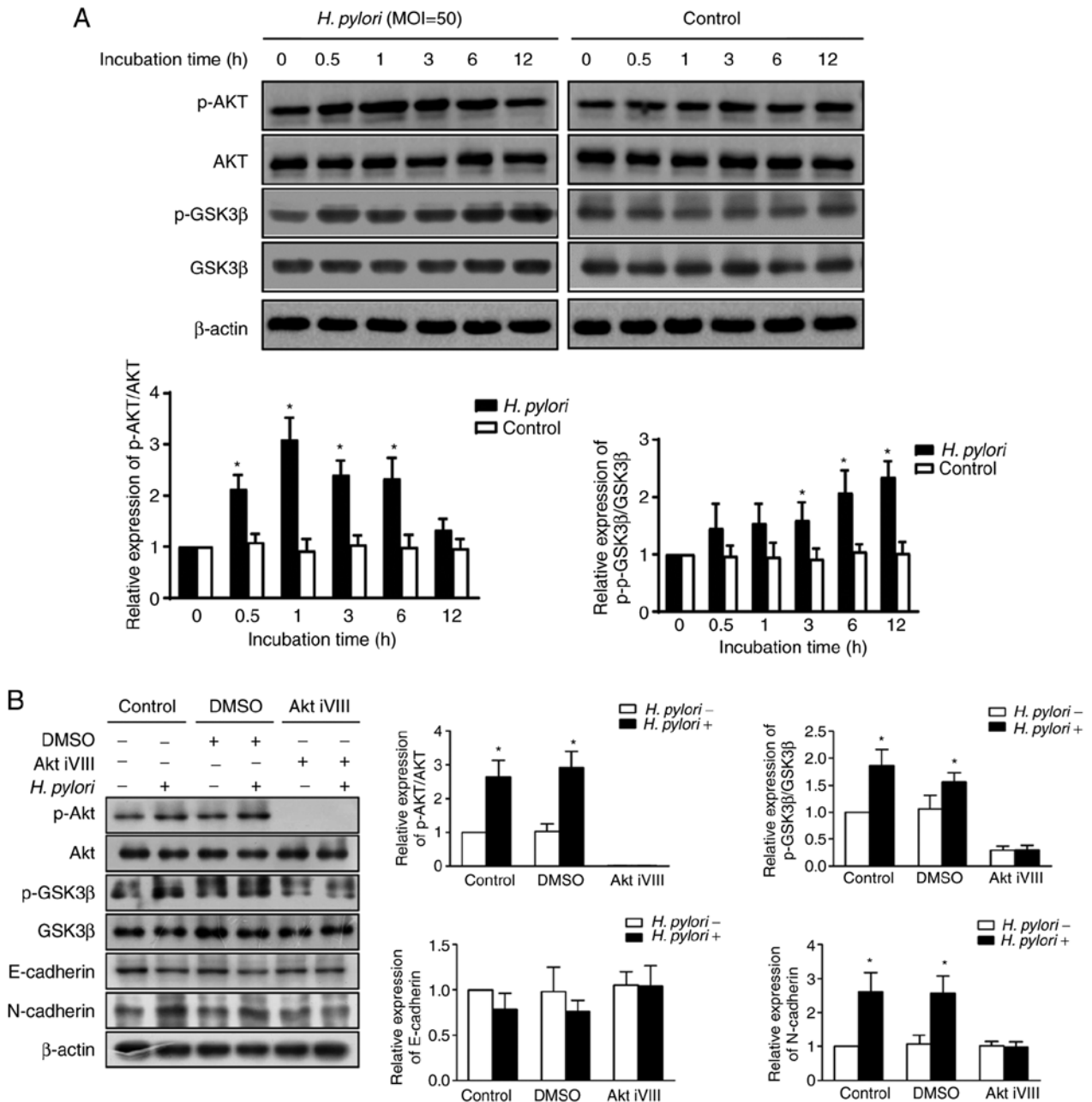


Figure 3. *H. pylori*-induced epithelial-mesenchymal transition is regulated via the AKT-GSK3 β signaling pathway. (A) Expression levels of the AKT-GSK3 β signaling pathway-associated proteins AKT, p-AKT (Ser473), GSK3 β and p-GSK3 β (Ser9) were detected by western blotting in GES-1 cells following 0, 0.5, 1, 3, 6 and 12 h co-culture with or without *H. pylori*. The statistical analysis of three independent experiments is shown. (B) GES-1 cells were pretreated with DMSO, the Akt iVIII or control medium. The expression levels of the AKT-GSK3 β signaling pathway-associated proteins were detected by western blot analysis in GES-1 cells with or without *H. pylori* infection. The statistical analysis of three independent experiments is shown. * $P < 0.05$ vs. 0 h in (A) and *H. pylori* (-) group in (B). Akt iVIII, AKT inhibitor VIII; *H. pylori*, *Helicobacter pylori*; p-, phosphorylated.

inhibitor VIII ($P > 0.05$; Fig. 3B). Therefore, *H. pylori*-induced EMT may be blocked by AKT inhibition, suggesting that *H. pylori*-induced EMT may be regulated by the AKT/GSK3 β signaling pathway.

AKT and GSK3 β are activated in H. pylori-infected Mongolian gerbils. To further validate the role of the AKT/GSK3 β signaling pathway in *H. pylori*-induced EMT, immunohistochemical analysis was performed to assess the expression levels of AKT, p-AKT, GSK3 β and p-GSK3 β in gastric tissues of Mongolian gerbils following *H. pylori* infection and they were compared with those of the control

group. Immunohistochemical assays indicated that although the expression levels of AKT and GSK3 β did not exhibit significant differences between the two groups ($P > 0.05$), *H. pylori* infection significantly increased the expression levels of p-AKT and p-GSK3 β ($P < 0.05$; Fig. 4). Overall, these results further validated the role of the AKT/GSK3 β signaling pathway in *H. pylori*-induced EMT.

Discussion

H. pylori infection is the main risk factor for GC and is responsible for the initiation of gastric carcinogenesis in

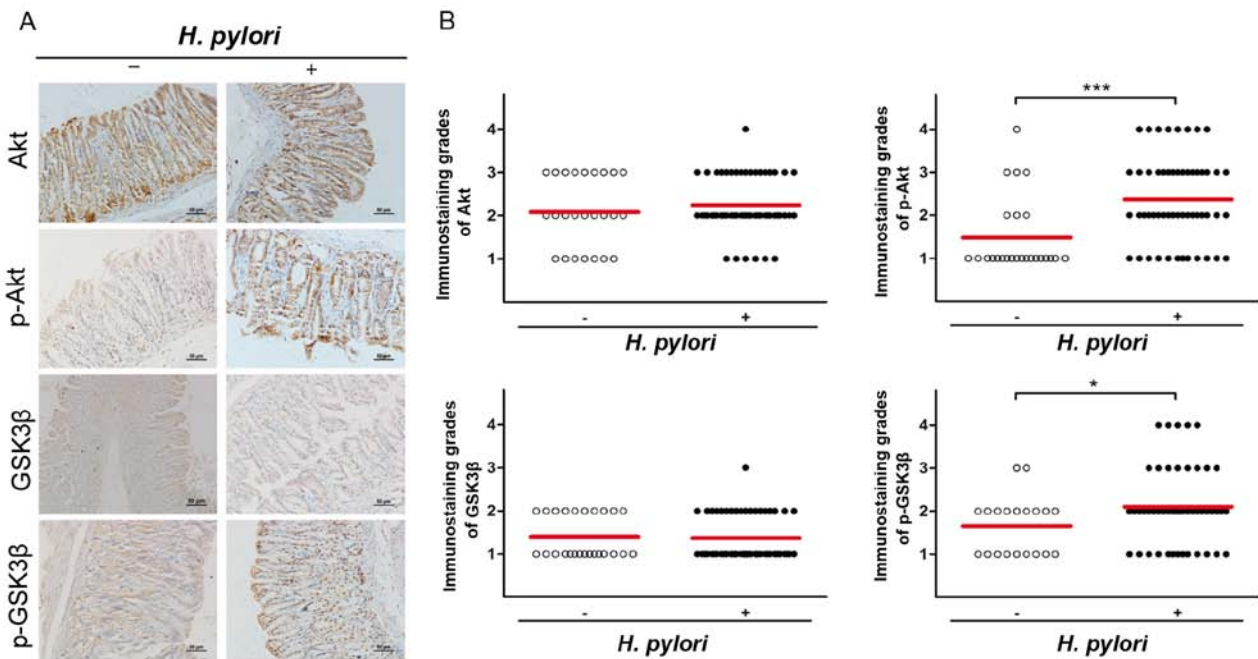


Figure 4. AKT-GSK3 β signaling pathway activation in *H. pylori*-infected Mongolian gerbils. (A) Protein expression levels of the AKT, p-AKT (Ser473), GSK3 β and p-GSK3 β (Ser9) were detected by immunohistochemical analysis in the gastric mucosa of Mongolian gerbils with or without *H. pylori* infection. Scale bar, 50 μ m. (B) Expression grades were assessed by the staining intensity and frequency. *P<0.05, ***P<0.001. *H. pylori*, *Helicobacter pylori*; p-, phosphorylated.

different developmental stages, including CNAG, IM, Dys and eventually GC (23). It has been reported that various oncogenic signaling pathways are involved in the pathogenic effects of *H. pylori* (24).

Sun *et al* (25) assessed the expression levels of E-cadherin by immunohistochemical analysis in 58 GC tissues, 40 adjacent tissues and 42 CNAG tissues. Sun *et al* (25) demonstrated that the expression levels of E-cadherin in specimens of patients with GC were significantly lower than those of patients with CNAG, which is consistent with the present findings. Furthermore, the present study highlighted that the changes in the expression levels of the EMT markers E-cadherin and N-cadherin were initially noted at the IM stage of gastric adenocarcinoma formation, suggesting that EMT may contribute to the initiation of gastric adenocarcinoma. Subsequently, the specimens were divided into two groups according to *H. pylori* infection status and the data demonstrated that at the CNAG and GC stages, the expression levels of E-cadherin in the *H. pylori*-positive group were significantly lower than those in the *H. pylori*-negative group, while at the GC stage, the expression levels of N-cadherin were significantly higher in the *H. pylori*-positive group than in the *H. pylori*-negative group. A recent study by Li *et al* (6) indicated that *H. pylori* infection reduces E-cadherin expression via the YY1 associated protein 1 signaling pathway in patients with CNAG, which is consistent with the results of the present study. These results strongly suggest that *H. pylori* infection initiates the EMT process at an early stage of gastric carcinogenesis, which may subsequently promote growth, metastasis and invasion of GC.

Subsequently, GES-1 cells were infected with *H. pylori in vitro*, which induced EMT, as detected by decreased E-cadherin expression and increased N-cadherin expression

in GES-1 cells in the early stage of *H. pylori* infection. In addition, the effect of *H. pylori* infection on EMT was further demonstrated by experiments in *H. pylori*-infected Mongolian gerbils. *H. pylori* infection induced a decrease in E-cadherin levels and an increase in N-cadherin expression, which was consistent with the *in vitro* results.

Activation of the AKT signaling pathway is involved in the EMT process by directly affecting the expression of specific transcription factors, such as Twist, Snail and Slug, the expression of integrin-linked kinase and extracellular matrix or by interacting with other signaling pathways, such as the TGF- β , NF- κ B, Ras and Wnt/ β -catenin signaling pathways (13). Activation of AKT/GSK3 β can increase the transcriptional activity and intracellular levels of β -catenin, which exerts a positive regulatory effect on the Wnt signaling pathway (26,27). The present results indicated that the levels of p-AKT and p-GSK3 β in GES-1 cells began to increase following 0.5 and 3 h, respectively, of co-culture with *H. pylori* and that this trend of p-AKT persisted until the 9 h time point and the trend for p-GSK3 β persisted until the 12 h time point. Furthermore, GES-1 cells were pre-treated with the AKT inhibitor VIII or DMSO, which was used as a negative control. Following 1 h of co-culture with *H. pylori*, the protein expression levels of p-AKT, p-GSK3 β , N-cadherin and E-cadherin in GES-1 cells that were pre-treated with the AKT inhibitor VIII did not change significantly compared with those noted in the cells that were not infected with *H. pylori*. These results indicated that inhibition of AKT activity blocked the development of EMT, demonstrating that the *H. pylori*-associated EMT process was AKT-dependent.

To further validate the role of the AKT/GSK3 β signaling pathway in *H. pylori*-induced EMT, the levels of AKT, p-AKT, GSK3 β and p-GSK3 β were examined in Mongolian gerbils

with or without *H. pylori* infection. Although no significant differences were noted in the expression levels of AKT and GSK3 β between the two groups, *H. pylori* infection activated phosphorylated AKT and phosphorylated GSK3 β . These results were consistent with the *in vitro* findings.

There were a few limitations of the present study. Although the role of the AKT/GSK3 β signaling pathway in *H. pylori*-induced EMT was thoroughly examined, other signaling pathways involved in the mechanism underlying this process require further exploration. Although the present study focused on the role of the AKT/GSK3 β signaling pathway, whether this signaling pathway interacts with other signaling pathways in *H. pylori*-induced EMT remains to be explored. In addition, there is still a gap between activation of AKT and EMT, and the exact mechanism by which the AKT/GSK3 β signaling pathway regulates EMT is not clear. Future studies should investigate this gap in further.

In conclusion, the present study indicated that *H. pylori* infection activated AKT and resulted in the phosphorylation and inactivation of GSK3 β , which promoted early-stage EMT development. These effects were AKT-dependent and were blocked by application of the AKT inhibitor VIII.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

YO, GL, JH and NLu conceived and designed the study. YO and GL performed the experiments, and NLu analyzed the data. WX and ZY collected human specimens and analyzed immunohistochemical data. CX, CZ and JC provided assistance with analyses of the data and technical issues. YO, NLi and YZ interpreted the data and drafted the manuscript. NLu supervised the study. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The use of human tissues was approved by the Human Ethics Committee of the First Affiliated Hospital of Nanchang

University. All patients signed an informed consent form for their participation in the study protocol. The First Affiliated Hospital of Nanchang University Ethics Committee approved all animal experiment protocols.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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