

A validation study for the utility of serum microRNA as a diagnostic and prognostic marker in patients with osteosarcoma

YOSHIHIRO ARAKI¹, NAOFUMI ASANO^{2,3}, NORIO YAMAMOTO¹,
KATSUHIRO HAYASHI¹, AKIHIKO TAKEUCHI¹, SHINJI MIWA¹, KENTARO IGARASHI¹,
TAKASHI HIGUCHI¹, KENSAKU ABE¹, YUTA TANIGUCHI¹, HIROTAKA YONEZAWA¹,
SEI MORINAGA¹, YOHEI ASANO¹, TAKESHI YOSHIDA⁴, RIKINARI HANAYAMA⁴,
JUNTARO MATSUZAKI⁵, TAKAHIRO OCHIYA⁶, AKIRA KAWAI^{3,7} and HIROYUKI TSUCHIYA¹

¹Department of Orthopedic Surgery, Graduate School of Medical Sciences, Kanazawa University, Kanazawa, Ishikawa 920-8641; ²Department of Orthopedic Surgery, Keio University School of Medicine, Tokyo 160-8582; ³Department of Musculoskeletal Oncology, National Cancer Center Hospital, Tokyo 104-0045; ⁴WPI Nano Life Science Institute (NanoLSI), Kanazawa University, Kanazawa, Ishikawa 920-1192; ⁵Division of Pharmacotherapeutics, Keio University Faculty of Pharmacy, Tokyo 105-8512; ⁶Department of Molecular and Cellular Medicine, Tokyo Medical University, Tokyo 160-0023; ⁷Division of Rare Cancer Research, National Cancer Center Research Institute, Tokyo 104-0045, Japan

Received July 24, 2022; Accepted March 3, 2023

DOI: 10.3892/ol.2023.13808

Abstract. In our previous study, osteosarcoma advanced locally, and metastasis was promoted through the secretion of large number of small extracellular vesicles, followed by suppressing osteoclastogenesis via the upregulation of microRNA (miR)-146a-5p. An additional 12 miRNAs in small extracellular vesicles were also detected $\geq 6x$ as frequently in high-grade malignancy with the capacity to metastasize as in those with a low metastatic potential. However, the utility of these 13 miRNAs for determining the prognosis or diagnosis of osteosarcoma has not been validated in the clinical setting. In the present study, the utility of these miRNAs as prognostic and diagnostic markers was therefore assessed. In total, 30 patients with osteosarcoma were retrospectively reviewed, and the survival rate was compared according to the serum miRNA levels in 27 patients treated with chemotherapy and surgery. In addition, to confirm diagnostic competency for osteosarcoma, the serum miRNA levels were compared with

those in patients with other bone tumors (n=112) and healthy controls (n=275). The patients with osteosarcoma with high serum levels of several miRNAs (miR-146a-5p, miR-1260a, miR-487b-3p, miR-1260b and miR-4758-3p) exhibited an improved survival rate compared with those with low levels. In particular, patients with high serum levels of miR-1260a exhibited a significantly improved overall survival rate, metastasis-free survival rate and disease-free survival rate compared with those with low levels. Thus, serum miR-1260a may potentially be a prognostic marker for patients with osteosarcoma. Moreover, patients with osteosarcoma had higher serum miR-1261 levels than those with benign or intermediate-grade bone tumors and thus may be a potential therapeutic target, in addition to being useful for differentiating whether or not a bone tumor is high-grade. A larger investigation is required to clarify the actual utility of these miRNAs in the clinical setting.

Correspondence to: Professor Norio Yamamoto, Department of Orthopedic Surgery, Graduate School of Medical Sciences, Kanazawa University, 13-1 Takaramachi, Kanazawa, Ishikawa 920-8641, Japan
E-mail: norinori@med.kanazawa-u.ac.jp

Abbreviations: miRNA, microRNA; AJCC, American Joint Committee on Cancer; ROC, receiver operating characteristic; NCCN, National Cancer Center Hospital; NCCN, National Comprehensive Cancer Network

Key words: validation, utility, serum miRNA, osteosarcoma, diagnosis, prognosis, marker

Introduction

MicroRNAs (miRNAs) consist of 20-30 bases and play an important role in fostering communication among cells in various organisms (1). Tumor cells also affect the surrounding normal cells via miRNAs, encouraging changes in the tumor microenvironment to facilitate tumor progression (2-6). These miRNAs have been reported to be transmitted to the target cells via tumor-derived small extracellular vesicles (3-6). Thus, these miRNAs in circulating blood may be useful as prognostic or diagnostic markers of tumors (7-11).

Sarcoma is a rare cancer, and osteosarcoma typically occurs in children, adolescents and young adults (12,13). Thus far, the diagnosis of osteosarcoma has generally depended on the histological findings of a biopsy specimen. The prognosis of these patients has been reported to be associated with the

presence of metastasis at presentation, a delayed initiation of treatment, a poor response to chemotherapy and small amounts of mature osteoclasts in biopsy specimens (14-19). Compared with these factors, however, circulating biomarkers assessed using blood tests are simpler and easier to evaluate, especially in children. Previously, several serum miRNAs associated with the prognosis or diagnosis of osteosarcoma have been reported, including circulating miR-663a, miR-25-3p, miR-487a, miR-493-5p, miR-501-3p and miR-502-5p (20-25).

In our previous study, osteosarcoma advanced locally and metastasis was promoted through the secretion of large number of small extracellular vesicles, followed by the suppression of osteoclastogenesis via the upregulation of miR-146a-5p (26). Several other miRNAs were also identified (miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907) that were included in small extracellular vesicles derived from high-grade osteosarcoma cells with the capacity to metastasize at rates $\geq 6x$ than in those with low metastatic potential (26). However, to the best of our knowledge, the utility of the 13 identified miRNAs for determining the prognosis or diagnosis of high-grade osteosarcoma has not yet been validated in the clinical setting.

Therefore, in the present study, the association of these miRNAs with the prognosis of patients with osteosarcoma and their potential utility in the diagnosis of osteosarcoma and differentiation from other bone tumors was investigated.

Materials and methods

Patient enrollment. A total of 43 patients with osteosarcoma were treated at The National Cancer Center Hospital (NCCH) in Tokyo, Japan, from January 2007 to June 2013. Among these patients, 13 were excluded due to a previous history of chemotherapy or surgical treatment at another institution (n=7), metastatic lesions at presentation (n=2), the histology of dedifferentiated osteosarcoma from low-grade osteosarcoma (n=2) and absence of microarray data (n=2). The remaining 30 patients who had no previous history of treatment for the histology of primary osteosarcoma without metastatic lesions at presentation and whose microarray data were sufficiently obtained from their serum samples, were retrospectively reviewed in the present study. A total of 27 patients underwent standard treatment with the combination of chemotherapy and surgery in accordance with the National Comprehensive Cancer Network (NCCN) guidelines (27) for osteosarcoma during the follow-up period at the NCCH. In total, 3 patients did not undergo either neoadjuvant or adjuvant chemotherapy. Of these 3 patients, 1 patient was assumed to have a low-grade malignant bone tumor before surgery, while another patient did not receive adjuvant chemotherapy due to the patient's refusal to receive further treatment after surgery. The other patient opted to receive surgery and radiotherapy instead of chemotherapy.

Patients with a histological diagnosis of osteochondroma (n=15), enchondroma (n=11) and fibrous dysplasia (n=10) as benign tumors, giant cell tumors of the bone (n=28), synovial chondromatosis (n=5) and osteoblastoma (n=3) as intermediate-grade tumors, Ewing sarcoma (n=26), chordoma (n=8) and chondrosarcoma (grade 2-3) (n=6) as high-grade tumors

[according to the histological grade in the 2020 WHO classification (12)], who were treated at the NCCH from January 2007 to June 2013 and whose microarray data were sufficiently obtained from their serum samples at presentation before any treatment, were also included in the study.

The present study was conducted in accordance with the 1975 Declaration of Helsinki. It was approved by The NCCH Institutional Review Board (Tokyo, Japan; approval nos. 2004-050, 2013-111 and 2015-266). Written informed consent was obtained from each participant and/or their family.

Data collection on patient and control characteristics. The age (mean and range), sex, tumor location (limb or trunk), tumor size (mean and range), staging [using the American Joint Committee on Cancer (AJCC) 8th edition] (28), surgery, chemotherapy, radiation therapy, distant metastasis, local recurrence, follow-up periods (mean and range), oncological outcomes at the final follow-up time and the levels of 13 serum miRNAs (miR-146a-5p, miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907) at the first visit were retrospectively investigated in all patients with osteosarcoma selected for study using medical records. The age (mean and range), sex and the level of the 13 serum miRNAs at the first visit in patients with other types of bone tumors (osteochondroma, enchondroma, fibrous dysplasia, giant cell tumors of the bone, synovial chondromatosis, osteoblastoma, Ewing sarcoma, chordoma and chondrosarcoma) and healthy controls were also reviewed. The 13 serum miRNA levels in all patients in the present study were used in microarray experiments in a previous study conducted at the NCCH, which are publicly available (25). A microarray analysis was not newly conducted in the present study.

miRNA mimics and inhibitor transfection. To determine the effects of miR-1261, 70 pmol of miRNA hairpin-inhibitor transfection control with Dy547 (miR hairpin inhibitor-Ctrl; cat. no. IP-004500-01-05) or hsa-miR-1261 hairpin-inhibitor (cat. no. IH-301388-01-0002) (both from Horizon Discovery Ltd.; PerkinElmer, Inc.) was transfected into 143B cells [highly aggressive, k-ras activated, human osteosarcoma cells (OS)] obtained from the ATCC using Lipofectamine™ RNAiMAX (Thermo Fisher Scientific, Inc.), and 70 pmol of miRNA mimic transfection control with Dy547 (miR mimic-Ctrl; cat. No. CP-004500-01-05) or hsa-miR-1261 mimic (cat. no. C-301388-00-0002) (both from Horizon Discovery Ltd.; PerkinElmer, Inc.) was transfected into HOS cells (non-aggressive, wild-type k-ras, human OS) obtained from the ATCC using Lipofectamine RNAiMAX. These two cell lines were cultured at 37°C in DMEM (Nacal Tesque, Inc.) with 10% heat inactivated FBS (Biowest) and 100 units/ml Penicillin G and 100 µg/ml Streptomycin (FUJIFILM Wako Pure Chemical Corporation) for 1 day before transfection. Both cells were cultured at 37°C in DMEM after transfection for 2 days. To confirm the transfection efficacy of the cells, uptake of the Dy547-labelled control in each cell line was observed using a BZ-9000 phase-contrast and fluorescence microscope (Keyence Corporation) and the positive cells were counted by a BZ-II Analyzer, version 1.42 (Keyence Corporation) (Fig. S1).

Cell viability and cell proliferation assays. 143B and HOS cells were seeded at 3×10^3 cells/200 μ l/96-well (n=3) after the aforementioned miRNA transfection. After incubating for 2 days, cellular viability was evaluated using WST-8 assay reagent (Nacalai Tesque, Inc.) for an hour.

After the culture of cells (5×10^4 cells/500 μ l/4-well glass slide) for 2 days, the cells were fixed using 4% paraformaldehyde 500 μ l/well at room temperature for 20 min. Cells were then blocked using serum-free unconjugated blocking liquid (cat. no. X0909; EnVision Systems; Dako; Agilent Technologies, Inc.) at room temperature for 10 min. For immunocytochemistry, the primary anti-rabbit polyclonal antibody for Ki-67 (1:200; cat. no. NB500-170; Novus Biologicals, LLC) was incubated with the cells at 4°C for 1 day. Anti-mouse IgG conjugated with peroxidase-labeled polymers (cat. no. K4001; EnVision Systems; Dako; Agilent Technologies, Inc.) was incubated with the cells at room temperature for 30 min as the secondary antibody. DAB-Chromogen staining was conducted for 3 min at room temperature (cat. no. GV825; EnVision Systems; Dako; Agilent Technologies, Inc.). The percentage of the Ki-67⁺ cells per total cells was measured by phase-contrast and fluorescence microscopy using a BZ-9000 and BZ-II Analyzer version 1.42. (KEYENCE).

Reverse transcription-quantitative PCR (RT-qPCR). To detect mRNA, total RNA extraction was performed on 143B and HOS cells that had been seeded at 6×10^5 cells/3 ml/60-mm dish after miRNA transfection and cultured at 37°C for 2 days, using a Fast Gene RNA Basic Kit (Nippon Gene Co., Ltd.). The mRNA was reverse transcribed using 100 ng RNA and 10 μ l reaction mix from a ReverTra Ace qPCR RT Kit (Toyobo Life Science) according to the manufacturer's protocol (37°C for 15 min, 50°C for 5 min, 98°C for 5 min and held at 4°C). After diluting the cDNA with 90 μ l dH₂O, 4 μ l was incubated with 16 μ l Fast SYBR Green Master Mix (Thermo Fisher Scientific, Inc.) in a LightCycler96 system (Roche Diagnostics K.K.). The thermocycler conditions were as follows: 95°C for 20 sec, 45 cycles of 95°C for 3 sec (denaturation) and 60°C for 30 sec (combination of annealing and extension). At the end of the qPCR, specificity was confirmed using melting peak analysis. Paired primers are listed in Table SI. mRNA expression of each gene was normalized to GAPDH using $2^{-\Delta\Delta Cq}$ (29).

Statistical analysis. Using receiver operating characteristic (ROC) curves, the optimal threshold values for miR-146a-5p, miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907 in patients with osteosarcoma were obtained when the Youden index was maximal, from which death from disease, distant metastasis and freedom from disease were predicted. The 27 patients with osteosarcoma who underwent standard treatment with a combination of chemotherapy and surgery in accordance with the NCCN guidelines were divided into two groups based on the area under the curve values (low vs. high level) determined by ROC curves analyses. The overall survival rate, distant metastasis-free survival rate and disease-free survival rate were plotted using the Kaplan-Meier method for each group and compared by the log-rank test. P<0.05 was considered to indicate a statistically significant difference. The overall

survival was defined as the period until the death of the patient since the osteosarcoma diagnosis was made. The distant metastasis-free survival was defined as the period until a distant metastasis was detected by an imaging examination. The disease-free survival was defined as the period of no evidence of disease.

For differentiation of osteosarcoma, serum miRNA levels at presentation in 30 patients with osteosarcoma were compared with those in patients with other types of bone tumors (benign, intermediate and high-grade) and healthy controls. Using the level of each miRNA (miR-146a-5p, miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907) in the 10 bone tumor groups and the osteosarcoma group, the Kruskal-Wallis test was performed to differentiate osteosarcoma and, as a post-hoc test, all 11 groups (including the healthy control group) were compared by Steel-Dwass multiple tests. P<0.05 was considered to indicate a statistically significant difference.

Cell viability using the WST-8 assay, the percentage of Ki-67⁺ cells, and the change in miRNA expression levels for miR-1261 target genes between HOS cells transfected with hsa-miR-1261 mimic and HOS cells transfected with miRNA mimic control, or between 143B cells transfected with hsa-miR-1261 hairpin-inhibitor and 143B cells transfected with miRNA hairpin-inhibitor control were compared by unpaired Student's t-test. P<0.05 was considered to indicate statistically significant difference.

All statistical analyses were performed using EZR version 1.61 (Saitama Medical Center, Jichi Medical University), which is a graphical user interface for the R software program (The R Foundation for Statistical Computing). More precisely, it is a modified version of R commander designed to add statistical functions frequently used in biostatistics (30,31).

Results

Patient characteristics. The osteosarcoma study population included 20 male and 10 female patients with a mean age of 24 (range, 10-68) years old. The histological findings were high-grade osteosarcoma in all cases. Tumors were located in the pelvis in 3 patients and in the appendicular skeleton in 27 patients. No distant metastasis was observed at presentation in all cases. The mean tumor size was 9.7 cm in the greatest dimension (range, 6-18 cm). The tumor stage was IIA in 13 patients, IIB in 14 patients and III in 3 patients according to the AJCC 8th staging classification. In total, 27 patients underwent surgical treatment after neoadjuvant chemotherapy, and thereafter received adjuvant chemotherapy. The mean follow-up period was 108 (range, 26-182) months. Local recurrence was observed in 1 patient at 48 months after surgery, and distant metastasis was observed in 11 patients at a mean follow-up period of 23 (range, 10-37) months. In total, 6 patients died of disease at a mean follow-up period of 49 (range, 26-90) months. At the final follow-up, a total of 19 patients were free from disease (Table I).

The characteristics of the patients with other types of bone tumors were as follows: i) 8 male and 7 female patients with a mean age of 27 (range, 6-48) years old with osteochondroma;

Table I. Patient characteristics (n=30).

Patient characteristic	Value
Mean age (range), years	24 (10-68)
Sex, n	
Male	20
Female	10
Mean follow-up period (range), months	108 (26-182)
Tumor location, n	
Pelvis	3
Femur	17
Tibia	10
Mean tumor size (range), cm	9.7 (6-18)
Tumor stage, n	
IIA	13
IIB	14
III	3
Treatment, n	
Surgery	30
Chemotherapy (neoadjuvant + adjuvant)	27
Neoadjuvant	28
Doxorubicin + cisplatin + MTX	19
Doxorubicin + cisplatin + ifosfamide	9
Adjuvant	28
Radiotherapy	2
Clinical outcome, n	
Progression-free	18
Local recurrence	1
Distant metastasis	11
Oncological outcome at final follow-up, n	
DOD	6
AWD	5
Disease-free	19

MTX, methotrexate; DOD, dead of disease; AWD; alive with disease.

ii) 7 male and 4 female patients with a mean age of 42 (range, 11-75) years old with enchondroma; iii) 7 male and 3 female patients with a mean age of 31 (range, 12-68) years old with fibrous dysplasia; iv) 15 male and 13 female patients with a mean age of 35 (range, 10-67) years old with giant cell tumor of the bone; v) 3 male and 2 female patients with a mean age of 52 (range, 36-63) years old with synovial chondromatosis; vi) 1 male and 2 female patients with a mean age of 16 (range, 15-18) years old with osteoblastoma; vii) 19 male and 7 female patients with a mean age of 27 (range, 8-65) years old with Ewing sarcoma; viii) 6 male and 2 female patients with a mean age of 60 (range, 26-92) years old with chordoma; and ix) 3 male and 3 female patients with a mean age of 48 (range, 19-76) years old with chondrosarcoma [Grade 2-3; graded using the WHO Classification of Tumors, Soft Tissue and Bone Tumors, 5th edition (12)]. The healthy controls were 150 male and 125 female individuals with a mean age of 52 (range, 35-80) years old.

Survival rates. The optimal cut-off points and areas under the curve of the miRNAs (miR-146a-5p, miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907) for predicting death from disease, distant metastasis or freedom from disease are shown in Table II.

A comparison of the overall survival according to miRNA levels is shown in Figs. 1A and 2. The patients with high serum levels of miR-146a-5p (P=0.04), miR-1260a (P=0.04) and miR-487b-3p (P=0.04) were significantly associated with an improved overall survival compared with those with low levels. By contrast, the serum levels of miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907 were not associated with the overall survival.

A comparison of the distant metastasis-free survival according to miRNA values is shown in Figs. 1B and 3. The patients with high serum levels of miR-1260a (P=0.04), miR-487b-3p (P=0.04), miR-1260b (P=0.03) and miR-4758-3p (P=0.04) were significantly associated with an improved distant metastasis-free survival compared with those with low levels. By contrast, the serum levels of miR-146a-5p, miR-6720-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907 were not associated with the distant metastasis-free survival.

A comparison of the disease-free survival according to miRNA level is shown in Figs. 1C and 4. The patients with high serum levels of miR-1260a (P=0.04), miR-1260b (P=0.02) and miR-4758-3p (P=0.04) were significantly associated with an improved disease-free survival compared with those with low levels. By contrast, the serum levels of miR-146a-5p, miR-487b-3p, miR-6720-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-1261, miR-7975, miR-4664-5p and miR-3907 were not associated with the disease-free survival.

Differentiation from other types of bone tumors. A comparison of serum miRNAs levels in patients with bone tumors and healthy controls is shown in Figs. 1D and 5. The levels of serum miR-1261 in patients with osteosarcoma were significantly higher than those in the healthy controls (P<0.05) and patients with benign (osteochondroma, P<0.05) or intermediate-grade bone tumor (giant cell tumor of the bone, P<0.05). In addition, the levels of serum miR-1261 tended to be lower for patients with other benign bone tumors (enchondroma and fibrous dysplasia) and other intermediate-grade bone tumors (synovial chondromatosis and osteoblastoma) compared with patients with osteosarcoma, although significant differences were not observed. The serum levels of miR-1261 were not significantly associated with those in patients with other high-grade bone tumors. By contrast, the serum levels of miR-146a-5p, miR-1260a, miR-487b-3p, miR-6720-3p, miR-1260b, miR-4758-3p, miR-4690-3p, miR-4286, miR-6765-3p, miR-7975, miR-4664-5p and miR-3907 in patients with osteosarcoma were not markedly different from those in patients with benign bone tumors, intermediate-grade bone tumors or other high-grade bone tumors, although the levels were significantly higher than those in healthy controls (P<0.05).

Table II. Optimal cut-off values, AUC, sensitivity and specificity of variables to predict death from disease, distant metastasis or freedom from disease.

miRNA	Overall survival rate			Distant metastasis-free survival rate			Disease-free survival rate					
	Cut-off value	AUC	Sensitivity, %	Specificity, %	Cut-off value	AUC	Sensitivity, %	Specificity, %	Cut-off value	AUC	Sensitivity, %	Specificity, %
miR-146a-5p	6.621	0.881	100	67	6.693	0.718	80	59	6.693	0.694	80	59
miR-1260a	6.621	0.881	100	62	6.693	0.753	90	59	6.693	0.741	90	59
miR-487b-3p	6.621	0.889	100	67	6.693	0.741	90	59	6.621	0.682	70	65
miR-6720-3p	6.695	0.833	100	62	6.695	0.629	70	59	6.695	0.747	80	65
miR-1260b	7.735	0.802	100	48	6.693	0.694	60	82	6.693	0.724	60	82
miR-4758-3p	6.621	0.865	100	62	6.693	0.735	90	59	6.693	0.782	90	59
miR-4690-3p	7.849	0.633	100	40	7.346	0.510	78	59	7.346	0.438	70	56
miR-4286	7.378	0.579	100	48	7.378	0.653	90	53	7.378	0.700	90	53
miR-6765-3p	6.285	0.746	67	81	6.693	0.688	80	59	6.693	0.612	70	53
miR-1261	9.984	0.675	83	62	9.984	0.565	60	59	9.984	0.665	70	65
miR-7975	6.621	0.778	100	57	6.693	0.671	90	53	6.693	0.724	90	53
miR-4664-5p	6.019	0.762	67	81	6.621	0.612	70	59	6.019	0.659	50	82
miR-3907	6.621	0.841	100	57	6.693	0.700	90	53	6.693	0.753	90	53

AUC, area under the curve; miR, microRNA; miRNA, microRNA.

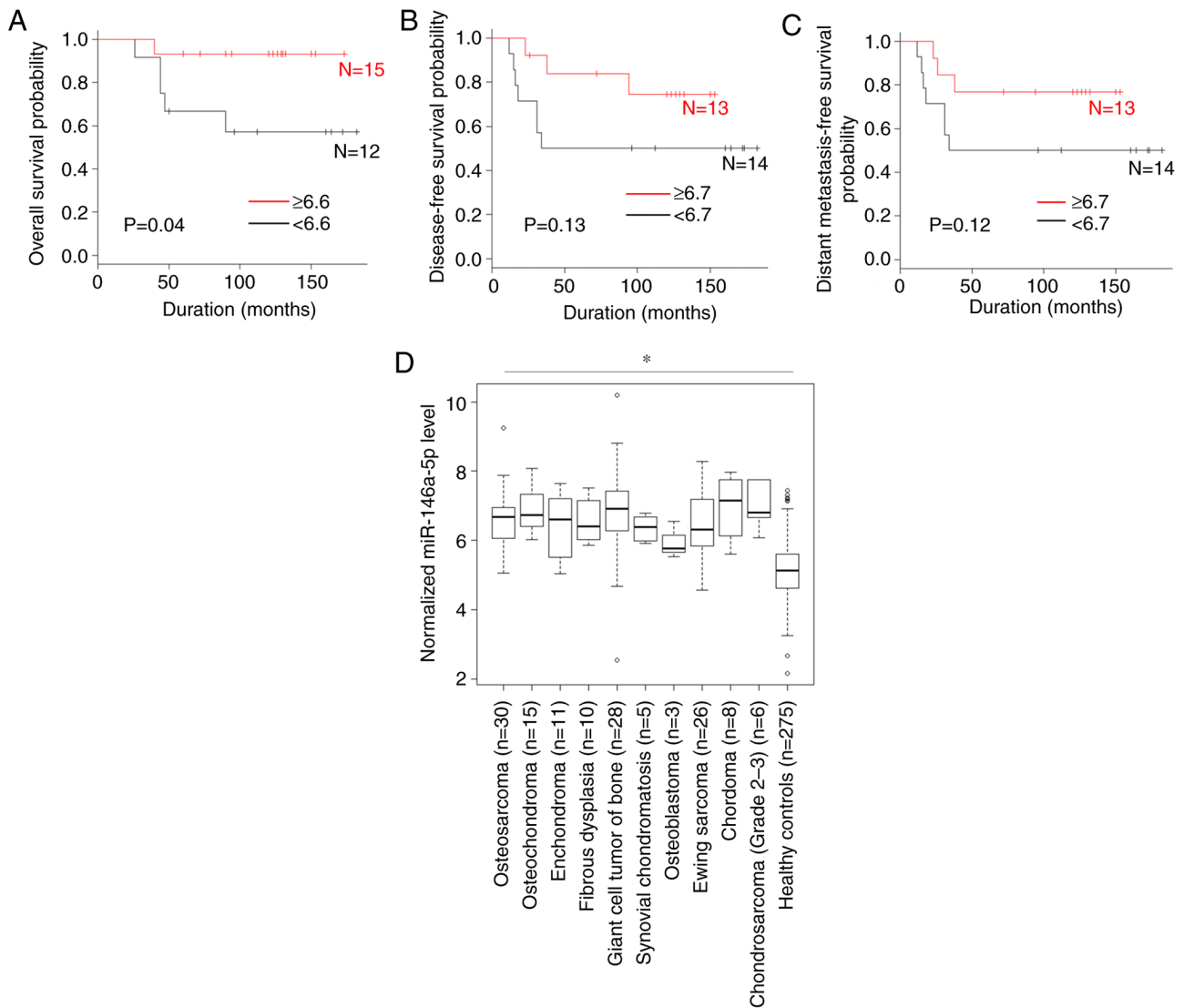


Figure 1. A comparison of the overall rate, distant metastasis-free rate and disease-free survival rate according to the levels of miR-146a-5p, and the serum levels of miR-146a-5p in patients with bone tumors and healthy controls. (A) Overall survival, (B) distant metastasis-free survival and (C) disease-free survival., (D) A comparison of serum levels of miR-146a-5p in patients with bone tumors and healthy controls. Only the level in patients with osteosarcoma was significant compared with the healthy controls. * $P < 0.05$. miR, microRNA.

Effect of miR-1261 on cellular viability and proliferation.

The inhibition of miR-1261 in 143B cells did not suppress cell viability ($P=0.35$; Fig. 6A), and the addition of miR-1261 to HOS cells did not promote cell viability ($P=0.42$; Fig. 6B). The amount of Ki-67⁺ 143B cells appeared to decrease with the inhibition of miR-1261 (Fig. 7A-C), and the amount of Ki-67⁺ HOS cells appeared to increase with the transfection of miR-1261 (Fig. 7D-F), although neither assay showed a statistically significant difference ($P=0.11$ and $P=0.19$, respectively).

Effect of miR-1261 on target genes. In reference to the list of predicted targets for hsa-miR-1261 (32), three genes associated with osteosarcoma were selected for screening, cadherin 6 (CDH6) (33), kinesin family member 26B (KIF26B) (34) and SET domain containing 2, histone lysine methyltransferase (SETD2) (35,36). The inhibition of miR-1261 in 143B cells significantly increased the mRNA levels of CDH6 and SETD2, but there was no significant difference in the mRNA level of KIF26B (Fig. 8A). The addition of miR-1261 mimic to HOS

cells significantly decreased the mRNA levels of KIF26B and SETD2, but there was no significant difference in the mRNA levels of CDH6 (Fig. 8B).

Discussion

The present study is a retrospective observational study to validate the utility of 13 serum miRNA levels as prognostic and diagnostic markers in the clinical setting using pretreatment patient data, as was first noted in our previous basic study (26). The patients with osteosarcoma with high serum levels of several types of miRNA (miR-146a-5p, miR-1260a, miR-487b-3p, miR-1260b and miR-4758-3p) exhibited an improved survival rate compared with those with low levels. In particular, patients with high serum levels of miR-1260a exhibited an improved overall survival rate, metastasis-free survival rate and disease-free survival rate compared with those with low levels. Of further note, patients with osteosarcoma exhibited higher levels of serum miR-1261 than patients with benign or intermediate-grade bone tumors.

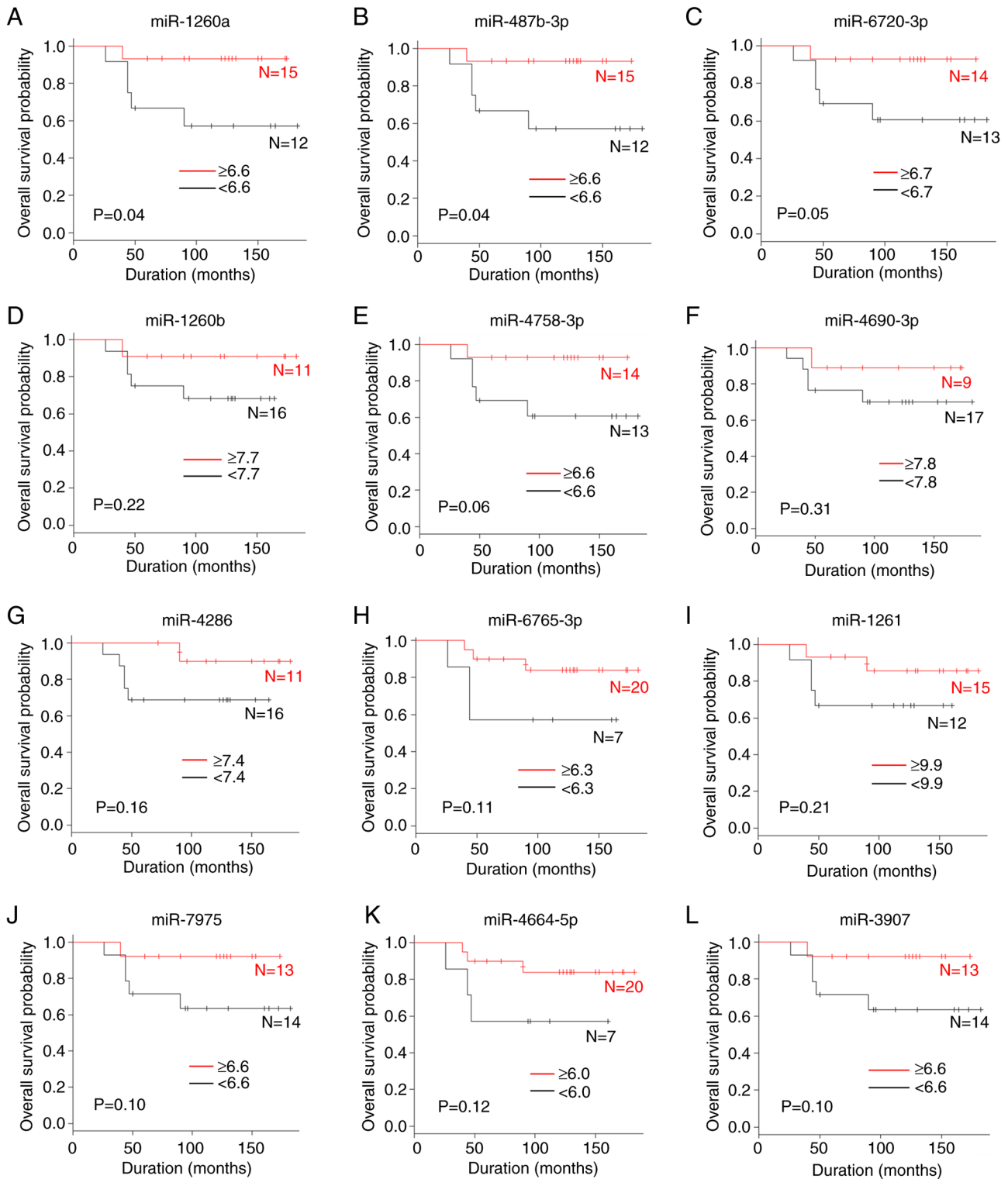


Figure 2. A comparison of the overall survival rate according to miRNA levels. (A) miR-1260a, (B) miR-487b-3p, (C) miR-6720-3p, (D) miR-1260b, (E) miR-4758-3p, (F) miR-4690-3p, (G) miR-4286, (H) miR-6765-3p, (I) miR-1261, (J) miR-7975, (K) miR-4664-5p and (L) miR-3907. miR, microRNA.

In our previous *in vivo* and *in vitro* study, highly aggressive osteosarcoma advanced locally and promoted metastasis via the secretion of large number of small extracellular vesicles (26). 143B cells, k-ras-activated human osteosarcoma cells with the capacity to metastasize, secreted greater amounts of small extracellular vesicles (exosomes) than HOS cells, which are k-ras-wild-type human osteosarcoma

cells with a low metastatic potential. The tumor-derived exosomes contained various types of miRNAs, including miR-146a-5p (37,38). miR-146a-5p suppressed osteoclastogenesis in the tumor bone microenvironment, and patients with osteosarcoma with few mature osteoclasts on histology of a biopsy specimen exhibited a worse prognosis and higher rates of lung metastasis than those with numerous

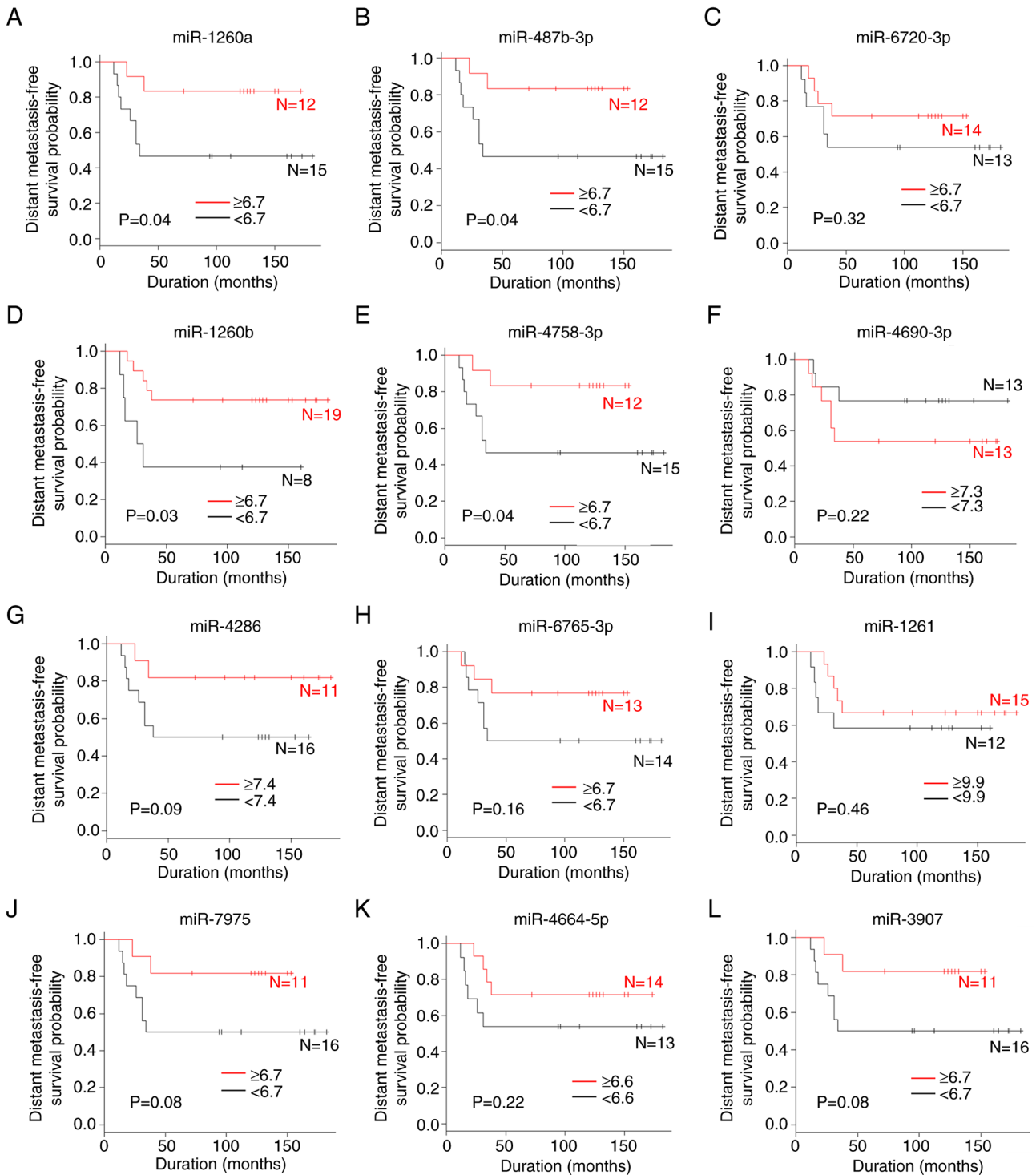


Figure 3. A comparison of the distant metastasis-free survival rate according to miRNA levels. (A) miR-1260a, (B) miR-487b-3p, (C) miR-6720-3p, (D) miR-1260b, (E) miR-4758-3p, (F) miR-4690-3p, (G) miR-4286, (H) miR-6765-3p, (I) miR-1261, (J) miR-7975, (K) miR-4664-5p and (L) miR-3907. miR, microRNA.

成熟破骨细胞 (19,26)。因此，在那些具有高水平 miR-146a-5p 的患者中，预后较差，与那些具有低水平者相比。本研究描述的另外 12 种 miRNAs 也在 k-ras 激活的 143B 细胞中比在 k-ras 野生型 HOS 细胞中更频繁地出现 ≥ 6 倍。类似地，其他 12 种 miRNAs 的高水平也预期反映较差的预后。

然而，本研究意外地展示了关于患者预后的一些相反的结果。例如，具有高水平某些类型的 miRNAs (miR-146a-5p, miR-1260a, miR-487b-3p, miR-1260b 和 miR-4758-3p) 的患者，其生存率与低水平者相比有所改善，这可能是由于分析中患者数量较少、miRNAs 的不同截断值、

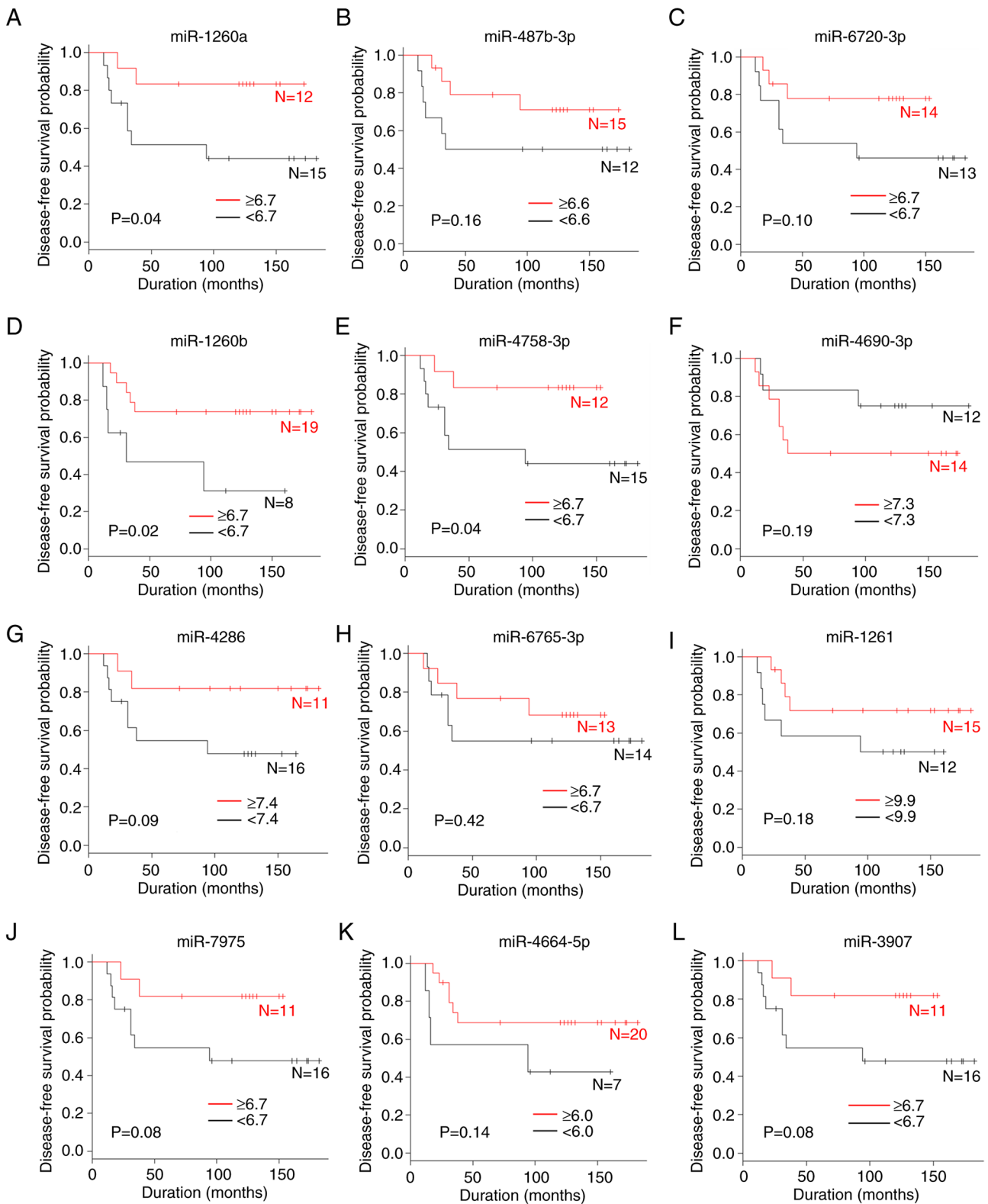


Figure 4. A comparison of the disease-free survival rate according to miRNA levels. (A) miR-1260a, (B) miR-487b-3p, (C) miR-6720-3p, (D) miR-1260b, (E) miR-4758-3p, (F) miR-4690-3p, (G) miR-4286, (H) miR-6765-3p, (I) miR-1261, (J) miR-7975, (K) miR-4664-5p and (L) miR-3907. miR, microRNA.

decomposition of miRNAs in the serum (39) or the inconsistency in miRNA levels in the serum and tumor tissues.

The upregulation of miR-1260a was previously reported to be a biomarker of lung adenocarcinoma (40). However, the serum levels of miR-1260a were shown to be decreased in ovarian and prostate cancer and increased in the postoperative

state for metastatic melanoma (41-43). Furthermore, miR-1260a was also reported to improve osteogenesis and angiogenesis (44), so high serum levels of this miRNA may contribute to an improved survival rate and have potential utility as a prognostic marker for some types of cancer, including osteosarcoma. In a recent study, 143B cells had higher levels of

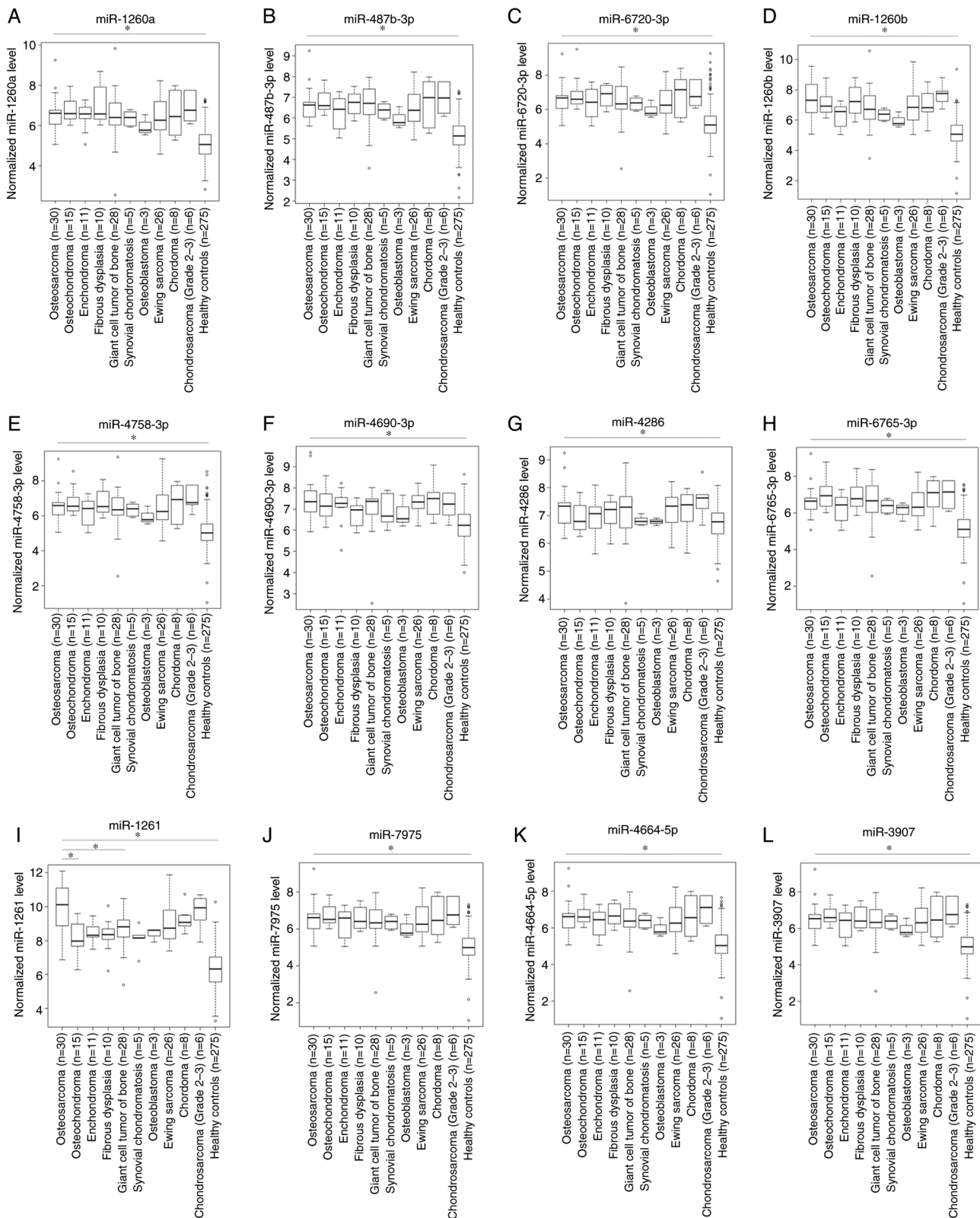


Figure 5. A comparison of serum miRNAs levels in patients with bone tumors and healthy controls. (A) miR-1260a, (B) miR-487b-3p, (C) miR-6720-3p, (D) miR-1260b, (E) miR-4758-3p, (F) miR-4690-3p, (G) miR-4286, (H) miR-6765-3p, (I) miR-1261, (J) miR-7975, (K) miR-4664-5p and (L) miR-3907. * $P < 0.05$. miR, microRNA.

miR-1260a than HOS cells (26), however, high-grade malignant cells may also generate some miRNAs that have a health benefit, not just those that negatively influence health (45-48). Serum miR-1260a may be a potential prognostic marker for

patients with osteosarcoma, although a further investigation is required to validate this.

Numerous miRNAs have been reported to be associated with the diagnosis and prognosis of patients with

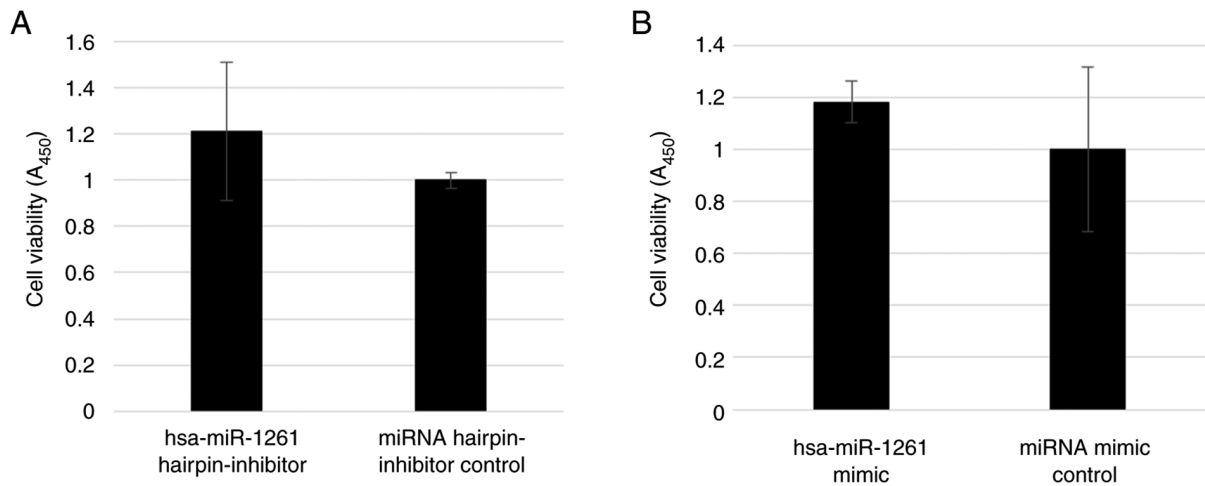


Figure 6. Cell viability. (A) 143B cells transfected with hsa-miR-1261 hairpin-inhibitor and miRNA hairpin-inhibitor control. (B) HOS cells transfected with hsa-miR-1261 mimic and miRNA mimic control. miR, microRNA.

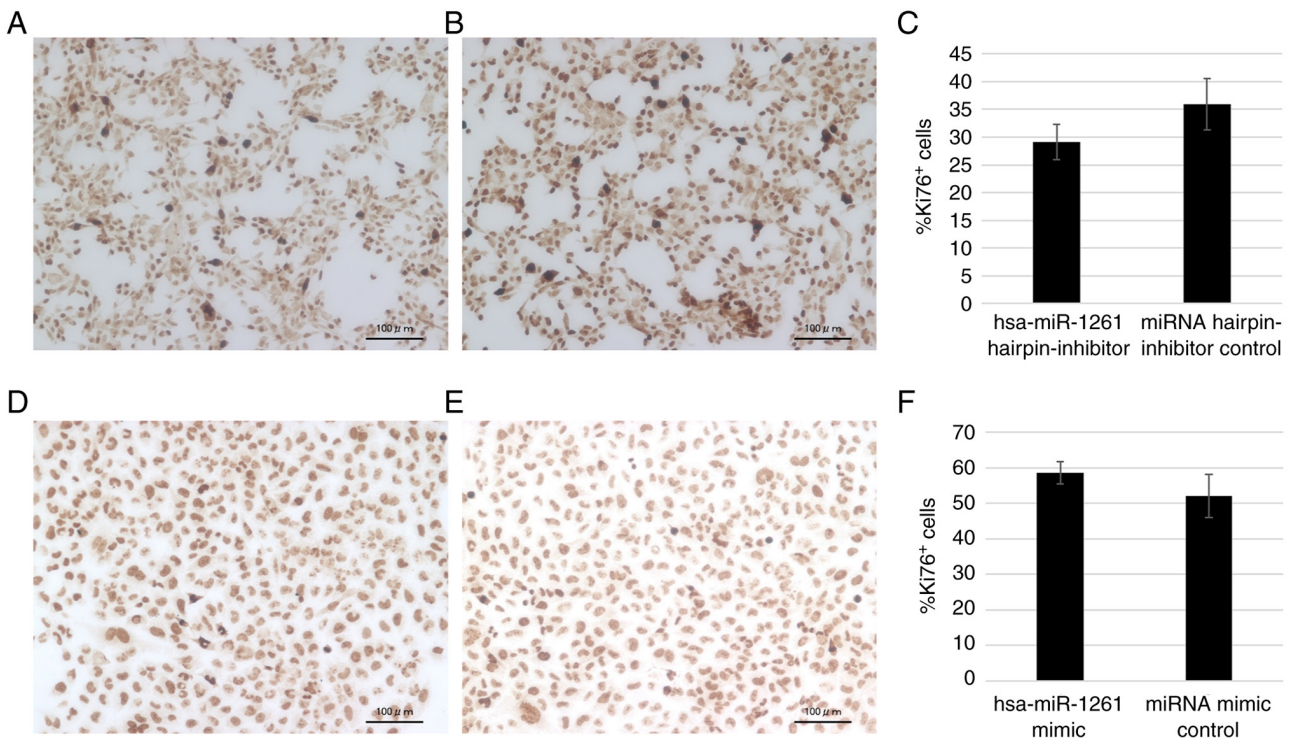


Figure 7. Immunocytochemical staining of Ki-67. (A) 143B cells transfected with hsa-miR-1261 hairpin-inhibitor. (B) 143B cells transfected with miRNA hairpin inhibitor control. (C) Comparison of the Ki-67⁺ level between 143B cells transfected with hsa-miR-1261 hairpin-inhibitor or miRNA hairpin-inhibitor control. (D) HOS cells transfected with hsa-miR-1261 mimic. (E) HOS cells transfected with miRNA mimic control. (F) Comparison of the Ki-67⁺ level between HOS cells transfected with hsa-miR-1261 mimic or miRNA mimic control. miR, microRNA.

cancer, but very few miRNAs are clinically used as specific markers for the disease (49). One reason for this might be due to the diverse functions of the miRNAs, depending on the disease. miR-487b-3p improves chemoresistance or metastasis in osteosarcoma but impairs osteoblastogenesis, and promotes tumorigenesis in colon cancer, prostate cancer and glioma (50-54). miR-1260b promotes tumorigenesis in breast cancer and lung adenocarcinoma and also inhibits bone loss (55-57). miR-4758-3p promotes tumorigenesis in glial tumor, gastric cancer and endometrial tumors (58-60), but its association with osteosarcoma is unknown.

In the human body, normal cells located around tumor cells might absorb some tumor-derived miRNAs that negatively influence cell survival (45). As a result of this, only a small amount of the tumor-derived miRNAs that exist in the stromal tissue enter the bloodstream. Some miRNAs have also been shown to be secreted by normal cells, including inflammatory cells, in response to the presence of tumor cells (46-48). Thus, identifying the origin of serum miRNAs might be difficult, and further investigations are required to confirm the utility of the 13 miRNAs analyzed in the present study as prognostic markers in the clinical setting.

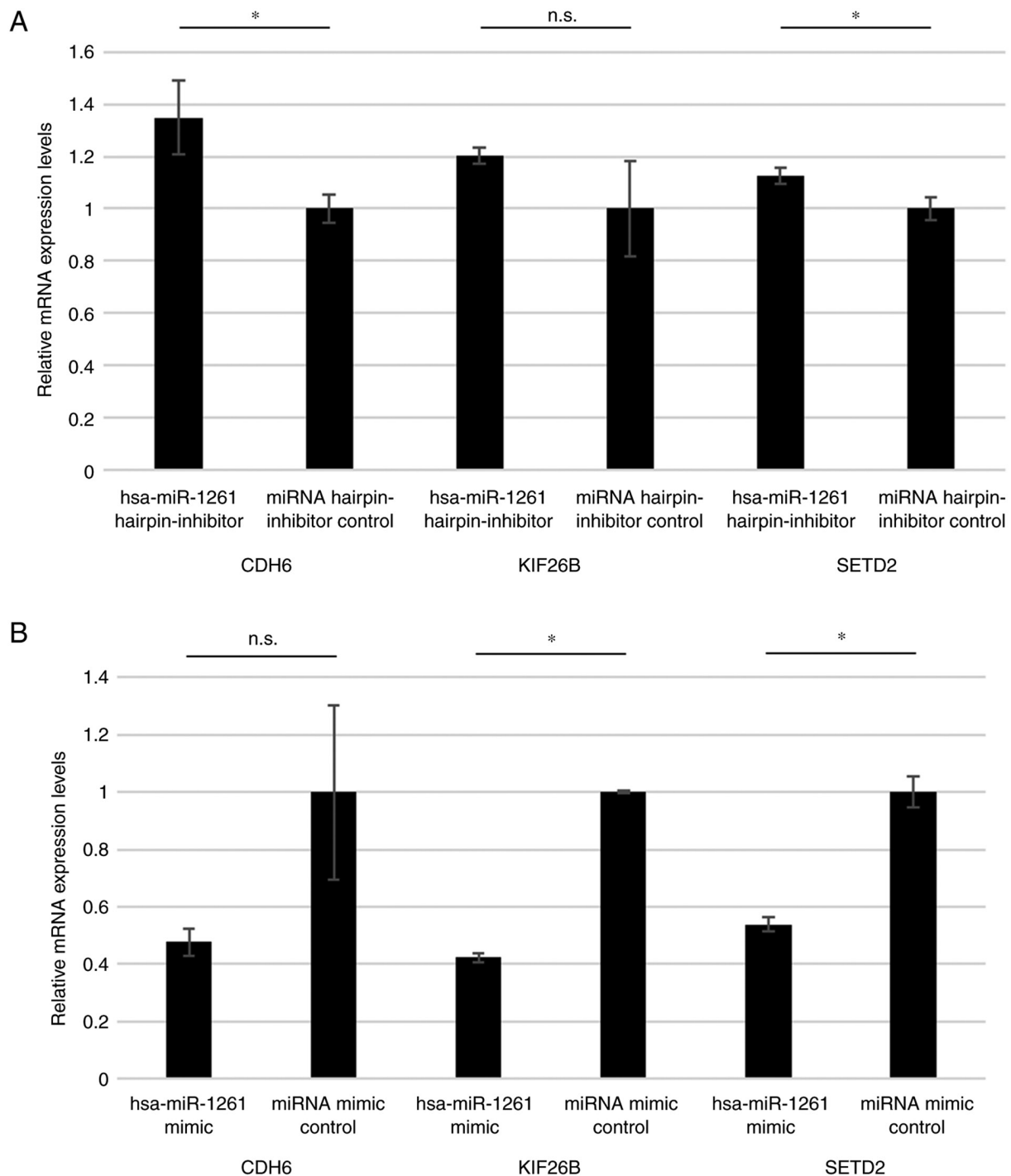


Figure 8. Reverse transcription-quantitative PCR of miR-1261 target genes. (A) mRNA levels of CDH6, KIF26B and SETD2 after the addition of has-miR-1261 hairpin-inhibitor and miRNA hairpin-inhibitor control in 143B cells. (B) mRNA levels of CDH6, KIF26B and SETD2 after the addition of has-miR-1261 mimic and miRNA mimic control in HOS cells. * $P < 0.05$. CDH6, cadherin 6; KIF26B, kinesin family member 26B; miR, microRNA; n.s., no significant difference; SETD2, SET domain containing 2, histone lysine methyltransferase. * $P < 0.05$. miR, microRNA.; n.s., no significant difference.

miR-1261 has been reported to be associated with migration and invasion of prostate cancer, progression of lung adenocarcinoma and as a diagnostic marker of hepatocellular carcinoma (61-64). However, its association with osteosarcoma is unclear. In the present study, the serum levels of miR-1261 in patients with osteosarcoma were significantly higher than those in healthy controls, patients with benign bone tumors and patients with intermediate-grade bone tumors, although there were no significant differences between patients with

osteosarcoma and those of patients with other high-grade bone tumors. The presence of histological high-grade malignancies, including osteosarcoma, might be reflected by the high serum levels of miR-1261 in patients with bone tumors. In the present study, *in vitro* experiments were performed to investigate the effect of miR-1261 on cellular viability and proliferation. Following the transfection of HOS cells with an miR-1261 mimic and 143B cells with an miR-1261 hairpin-inhibitor, the cell viabilities were similar, but cell proliferation appeared to

be associated with miR-1261. Furthermore, the mRNA levels of CDH6, KIF26B and SETD2 [target genes of miR-1261 (32)] increased by inhibiting miR-1261 and, by contrast, these target genes decreased upon the addition of an miR-1261 mimic. The SETD2 gene has been described as a tumor suppressor in various types of cancers, including renal, gastric and lung cancer (65,66). However, there are only a few studies associated with osteosarcoma, and the involvement of miR-1261 and SETD2 has not yet been previously described (35,36,65,66). Thus, miR-1261 is potentially a novel therapeutic target in addition to being useful for differentiating whether a bone tumor is high-grade, although a further investigation is necessary.

Several limitations associated with the present study warrant mention. Firstly, this was a retrospective, small-scale, single-institution study. Secondly, the peripheral blood findings were not compared between the preoperative and postoperative periods. Changes in the levels of miRNA after surgery may indicate tumor-derived miRNAs (21-24), but, in the present study, the postoperative blood findings for miRNAs were not investigated as the timing of surgery differed among patients. Thirdly, the characteristics concerning age and sex in healthy patients and patients with other types of bone tumors were not compared with those in patients with osteosarcoma since there was only a small number of patients, and also because the predominance of age and sex depends on the histological type of bone tumors (12).

In conclusion, regarding the 13 serum miRNAs evaluated in the present study, serum miR-1260a may be a potential prognostic marker for patients with osteosarcoma. Patients with osteosarcoma had higher levels of serum miR-1261 than those with benign or intermediate-grade bone tumors, which may be a potential novel therapeutic target in addition to being useful for differentiating whether or not a bone tumor is high-grade. A larger investigation is required to clarify the utility of these miRNAs in the clinical setting.

Acknowledgements

The authors would like to thank Dr Akihiko Yoshida (Department of Diagnostic Pathology, The National Cancer Center Hospital, Tokyo, Japan) for performing histological examination of all the specimens.

Funding

This research was supported by The JSPS Fujita Memorial Fund for Medical Research (2021) and Grants-in-Aid for Scientific Research (KAKENHI) from The Ministry of Education, Culture, Sports, Science and Technology (MEXT; grant no. 22K16712).

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

HT, NY, AK, NA and YAr conceived and designed the study. YAr carried out data acquisition. AK and NA provided

assistance for data acquisition. AK and NA managed the patients for the appropriate treatment and observed them at the follow-up outpatient clinic. YAr and TY performed the *in vitro* experiment. YAr, TY, RH, KH, AT, SMi, KI, TH, KA, HY, SMo and YAs analyzed and interpreted all the patient's data. HT, NY, JM, TO and YT contributed to the analysis of the data and critical appraisal. YAr, TY, NA and NY confirm the authenticity of all the raw data. YAr wrote the manuscript. KH, AT, SMi, KI, TH, KA, HY, SMo and YAs supervised the analysis. All authors read and approved the final manuscript.

Ethics approval and consent to participate

All patients underwent standard treatment during the follow-up period at The National Cancer Center Hospital in Tokyo, Japan. The present study was conducted in accordance with the 1975 Declaration of Helsinki. It was approved by The NCCCH Institutional Review Board (Tokyo, Japan; approval nos. 2004-050, 2013-111 and 2015-266). Written informed consent was obtained from each participant and/or their family.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Saliminejad K, Khorram Khorshid HR, Soleymani Fard S and Ghaffari SH: An overview of microRNAs: Biology, functions, therapeutics, and analysis methods. *J Cell Physiol* 234: 5451-5465, 2019.
2. Rupaimoole R, Calin GA, Lopez-Berestein G and Sood AK: miRNA deregulation in cancer cells and the tumor microenvironment. *Cancer Discov* 6: 235-246, 2016.
3. Sun Z, Shi K, Yang S, Liu J, Zhou Q, Wang G, Song J, Li Z, Zhang Z and Yuan W: Effect of exosomal miRNA on cancer biology and clinical applications. *Mol Cancer* 17: 147, 2018.
4. Pontecorvi G, Bellenghi M, Puglisi R, Carè A and Mattia G: Tumor-derived extracellular vesicles and microRNAs: Functional roles, diagnostic, prognostic and therapeutic options. *Cytokine Growth Factor Rev* 51: 75-83, 2020.
5. Tang Z, Li D, Hou S and Zhu X: The cancer exosomes: Clinical implications, applications and challenges. *Int J Cancer* 146: 2946-2959, 2020.
6. Chen Q, Li Y, Liu Y, Xu W and Zhu X: Exosomal Non-coding RNAs-Mediated crosstalk in the tumor microenvironment. *Front Cell Dev Biol* 9: 646864, 2021.
7. McGuire A, Brown JA and Kerin MJ: Metastatic breast cancer: The potential of miRNA for diagnosis and treatment monitoring. *Cancer Metastasis Rev* 34: 145-155, 2015.
8. Ishikawa D, Yoshikawa K, Takasu C, Kashihara H, Nishi M, Tokunaga T, Higashijima J and Shimada M: Expression level of microRNA-449a predicts the prognosis of patients with gastric cancer. *Anticancer Res* 40: 239-244, 2020.
9. Amankwah EK, Devidas M, Teachey DT, Rabin KR and Brown PA: Six candidate miRNAs associated with early relapse in pediatric B-cell acute lymphoblastic leukemia. *Anticancer Res* 40: 3147-3153, 2020.
10. Yu H, Guan Z, Cuk K, Brenner H and Zhang Y: Circulating microRNA biomarkers for lung cancer detection in Western populations. *Cancer Med* 7: 4849-4862, 2018.
11. Zeng Z, Li Y, Pan Y, Lan X, Song F, Sun J, Zhou K, Liu X, Ren X, Wang F, *et al*: Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. *Nat Commun* 9: 5395, 2018.

12. WHO Classification of Tumours Editorial Board: WHO Classification of Tumours: Soft Tissue and Bone Tumours. 5th edition. IARC, Lyon, pp 403-409, 2020.
13. Reed DR, Hayashi M, Wagner L, Binitie O, Steppan DA, Brohl AS, Shinohara ET, Bridge JA, Loeb DM, Borinstein SC and Isakoff MS: Treatment pathway of bone sarcoma in children, adolescents, and young adults. *Cancer* 123: 2206-2218, 2017.
14. Tsukamoto S, Errani C, Angelini A and Mavrogenis AF: Current treatment considerations for osteosarcoma metastatic at presentation. *Orthopedics* 43: e345-e358, 2020.
15. Araki Y, Yamamoto N, Hayashi K, Takeuchi A, Miwa S, Igarashi K, Higuchi T, Abe K, Taniguchi Y, Yonezawa H, *et al*: Delayed initiation of treatment is associated with metastasis of malignant bone tumor. *Anticancer Res* 41: 2993-2999, 2021.
16. Hao H, Chen L, Huang D, Ge J, Qiu Y and Hao L: Meta-analysis of alkaline phosphatase and prognosis for osteosarcoma. *Eur J Cancer Care (Engl)* 26: e12536, 2017.
17. Araki Y, Yamamoto N, Hayashi K, Takeuchi A, Miwa S, Igarashi K, Higuchi T, Abe K, Taniguchi Y, Yonezawa H, *et al*: Pretreatment neutrophil count and platelet-lymphocyte ratio as predictors of metastasis in patients with osteosarcoma. *Anticancer Res* 42: 1081-1089, 2022.
18. Ding WZ, Liu K, Li Z and Chen SR: A meta-analysis of prognostic factors of osteosarcoma. *Eur Rev Med Pharmacol Sci* 24: 4103-4112, 2020.
19. Araki Y, Yamamoto N, Hayashi K, Takeuchi A, Miwa S, Igarashi K, Higuchi T, Abe K, Taniguchi Y, Yonezawa H, *et al*: The number of osteoclasts in a biopsy specimen can predict the efficacy of neoadjuvant chemotherapy for primary osteosarcoma. *Sci Rep* 11: 1989, 2021.
20. Wang J, Liu S, Shi J, Li J, Wang S, Liu H, Zhao S, Duan K, Pan X and Yi Z: The Role of miRNA in the diagnosis, prognosis, and treatment of osteosarcoma. *Cancer Biother Radiopharm* 34: 605-613, 2019.
21. Raimondi L, De Luca A, Gallo A, Costa V, Russelli G, Cuscino N, Manno M, Raccosta S, Carina V, Bellavia D, *et al*: Osteosarcoma cell-derived exosomes affect tumor microenvironment by specific packaging of microRNAs. *Carcinogenesis* 41: 666-677, 2020.
22. Huang C, Sun Y, Ma S, Vadamotoo AS, Wang L and Jin C: Identification of circulating miR-663a as a potential biomarker for diagnosing osteosarcoma. *Pathol Res Pract* 215: 152411, 2019.
23. Fujiwara T, Uotani K, Yoshida A, Morita T, Nezu Y, Kobayashi E, Yoshida A, Uehara T, Omori T, Sugiu K, *et al*: Clinical significance of circulating miR-25-3p as a novel diagnostic and prognostic biomarker in osteosarcoma. *Oncotarget* 8: 33375-33392, 2017.
24. Huang C, Wang Q, Ma S, Sun Y, Vadamotoo AS and Jin C: A four serum-miRNA panel serves as a potential diagnostic biomarker of osteosarcoma. *Int J Clin Oncol* 24: 976-982, 2019.
25. Asano N, Matsuzaki J, Ichikawa M, Kawauchi J, Takizawa S, Aoki Y, Sakamoto H, Yoshida A, Kobayashi E, Tanzawa Y, *et al*: A serum microRNA classifier for the diagnosis of sarcomas of various histological subtypes. *Nat Commun* 10: 1299, 2019.
26. Araki Y, Aiba H, Yoshida T, Yamamoto N, Hayashi K, Takeuchi A, Miwa S, Igarashi K, Nguyen TD, Ishii K, *et al*: Osteosarcoma-Derived small extracellular vesicles enhance tumor metastasis and suppress osteoclastogenesis by miR-146a-5p. *Front Oncol* 11: 667109, 2021.
27. National Comprehensive Cancer Network: NCCN Guidelines Bone Cancer. <https://www.nccn.org/guidelines/guidelines-detail?category=1&id=1418>. Accessed April 1, 2023.
28. Amin MB, Edge SB, Greene FL, Compton CC, Gershenwald JE, Broolland RK, Meyer L, Gress DM, Byrd DR and Winchester DP: AJCC Cancer Staging Manual. 8th edition. Springer, Chicago, IL, pp471-486, 2017.
29. Livak KJ and Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 25: 402-408, 2001.
30. Ihaka R and Gentleman R: R: A language for data analysis and graphics. *J Comput Graph Stat* 5: 299-314, 1996.
31. Kanda Y: Investigation of the freely available easy-to-use software 'EZ' for medical statistics. *Bone Marrow Transplant* 48: 452-458, 2013.
32. miRDB: There are 437 predicted targets for hsa-miR-1261 in miRDB. <https://mirdb.org/cgi-bin/search.cgi?searchType=miRNA&searchBox=hsa-miR-1261&full=1> Accessed April 1, 2023.
33. Ji Q, Xu X, Song Q, Xu Y, Tai Y, Goodman SB, Bi W, Xu M, Jiao S, Maloney WJ and Wang Y: miR-223-3p inhibits human osteosarcoma metastasis and progression by directly targeting CDH6. *Mol Ther* 26: 1299-1312, 2018.
34. Pu Y, Yi Q, Zhao F, Wang H, Cai W and Cai S: MiR-20a-5p represses multi-drug resistance in osteosarcoma by targeting the KIF26B gene. *Cancer Cell Int* 16: 64, 2016.
35. Sakthikumar S, Elvers I, Kim J, Arendt ML, Thomas R, Turner-Maier J, Swofford R, Johnson J, Schumacher SE, Alföldi J, *et al*: SETD2 is recurrently mutated in whole-exome sequenced canine osteosarcoma. *Cancer Res* 78: 3421-3431, 2018.
36. Jiang C, He C, Wu Z, Li F and Xiao J: Histone methyltransferase SETD2 regulates osteosarcoma cell growth and chemosensitivity by suppressing Wnt/ β -catenin signaling. *Biochem Biophys Res Commun* 502: 382-388, 2018.
37. Yurtsever A, Yoshida T, Badami Behjat A, Araki Y, Hanayama R and Fukuma T: Structural and mechanical characteristics of exosomes from osteosarcoma cells explored by 3D-atomic force microscopy. *Nanoscale* 13: 6661-6677, 2021.
38. Xu X, Yu H and Xu Y: Ras-ERK1/2 signaling promotes the development of osteosarcoma by regulating H2BK12ac Through CBP. *Cancer Manag Res* 11: 9153-9163, 2019.
39. Boele J, Persson H, Shin JW, Ishizu Y, Newie IS, Søkilde R, Hawkins SM, Coarfa C, Ikeda K, Takayama K, *et al*: PAPD5-mediated 3' adenylation and subsequent degradation of miR-21 is disrupted in proliferative disease. *Proc Natl Acad Sci USA* 111: 11467-11472, 2014.
40. Faversoni A, Favero C, Dioni L, Pesatori AC, Bollati V, Montoli M, Musso V, Terrasi A, Fusco N, Nosotti M, *et al*: An EBC/Plasma miRNA Signature discriminates lung adenocarcinomas from pleural mesothelioma and healthy controls. *Front Oncol* 11: 643280, 2021.
41. Chen L, Wang K, Li L, Zheng B, Zhang Q, Zhang F, Chen J and Wang S: Plasma exosomal miR-1260a, miR-7977 and miR-192-5p as diagnostic biomarkers in epithelial ovarian cancer. *Future Oncol* 18: 2919-2931, 2022.
42. Mancini M, Grasso M, Muccillo L, Babbio F, Precazzini F, Castiglioni I, Zanetti V, Rizzo F, Pistore C, De Marino MG, *et al*: DNMT3A epigenetically regulates key microRNAs involved in epithelial-to-mesenchymal transition in prostate cancer. *Carcinogenesis* 42: 1449-1460, 2021.
43. Latchana N, DiVincenzo MJ, Regan K, Abrams Z, Zhang X, Jacob NK, Gru AA, Fadda P, Markowitz J, Howard JH and Carson WE III: Alterations in patient plasma microRNA expression profiles following resection of metastatic melanoma. *J Surg Oncol* 118: 501-509, 2018.
44. Wu D, Chang X, Tian J, Kang L, Wu Y, Liu J, Wu X, Huang Y, Gao B, Wang H, *et al*: Bone mesenchymal stem cells stimulation by magnetic nanoparticles and a static magnetic field: Release of exosomal miR-1260a improves osteogenesis and angiogenesis. *J Nanobiotechnology* 19: 209, 2021.
45. Li K, Rodosthenous RS, Kashanchi F, Gingeras T, Gould SJ, Kuo LS, Kurre P, Lee H, Leonard JN, Liu H, *et al*: Advances, challenges, and opportunities in extracellular RNA biology: Insights from the NIH exRNA Strategic Workshop. *JCI Insight* 3: e98942, 2018.
46. Engin A: Dark-Side of Exosomes. *Adv Exp Med Biol* 1275: 101-131, 2021.
47. Robbins PD and Morelli AE: Regulation of immune responses by extracellular vesicles. *Nat Rev Immunol* 14: 195-208, 2014.
48. Sinkovics JG: Molecular biology of oncogenic inflammatory processes. I. Non-oncogenic and oncogenic pathogens, intrinsic inflammatory reactions without pathogens, and microRNA/DNA interactions (Review). *Int J Oncol* 40: 305-349, 2012.
49. Ye H, Hu X, Wen Y, Tu C, Hornicek F, Duan Z and Min L: Exosomes in the tumor microenvironment of sarcoma: From biological functions to clinical applications. *J Nanobiotechnology* 20: 403, 2022.
50. Cheng M, Duan PG, Gao ZZ and Dai M: MicroRNA-487b-3p inhibits osteosarcoma chemoresistance and metastasis by targeting ALDH1A3. *Oncol Rep* 44: 2691-2700, 2020.
51. John AA, Prakash R and Singh D: miR-487b-3p impairs osteoblastogenesis by targeting Notch-regulated ankyrin-repeat protein (Nrap). *J Endocrinol* 241: 249-263, 2019.
52. Yi H, Geng L, Black A, Talmon G, Berim L, Wang J: The miR-487b-3p/GRM3/TGF β signaling axis is an important regulator of colon cancer tumorigenesis. *Oncogene* 36: 3477-3489, 2017.
53. Zhang BL, Dong FL, Guo TW, Gu XH, Huang LY and Gao DS: MiRNAs Mediate GDNF-Induced proliferation and migration of glioma cells. *Cell Physiol Biochem* 44: 1923-1938, 2017.
54. Daniel R, Wu Q, Williams V, Clark G, Guruli G and Zehner Z: A Panel of MicroRNAs as diagnostic biomarkers for the identification of prostate cancer. *Int J Mol Sci* 18: 1281, 2017.

55. Park S, Kim J, Cho Y, Ahn S, Kim G, Hwang D, Chang Y, Ha S, Choi Y, Lee MH, *et al*: Promotion of tumorigenesis by miR-1260b-targeting CASP8: Potential diagnostic and prognostic marker for breast cancer. *Cancer Sci* 113: 2097-2108, 2022.
56. Xia Y, Wei K, Hu LQ, Zhou CR, Lu ZB, Zhan GS, Pan XL, Pan CF, Wang J, Wen W, *et al*: Exosome-mediated transfer of miR-1260b promotes cell invasion through Wnt/ β -catenin signaling pathway in lung adenocarcinoma. *J Cell Physiol* 235: 6843-6853, 2020.
57. Hayashi C, Fukuda T, Kawakami K, Toyoda M, Nakao Y, Watanabe Y, Shinjo T, Sano T, Iwashita M, Yotsumoto K, *et al*: miR-1260b inhibits periodontal bone loss by targeting ATF6 β mediated regulation of ER stress. *Front Cell Dev Biol* 10: 1061216, 2022.
58. Zakrzewska M, Gruszka R, Stawiski K, Fendler W, Kordacka J, Grajkowska W, Daszkiewicz P, Liberski PP and Zakrzewski K: Expression-based decision tree model reveals distinct microRNA expression pattern in pediatric neuronal and mixed neuronal-glioma tumors. *BMC Cancer* 19: 544, 2019.
59. Omura T, Shimada Y, Nagata T, Okumura T, Fukuoka J, Yamagishi F, Tajika S, Nakajima S, Kawabe A and Tsukada K: Relapse-associated microRNA in gastric cancer patients after S-1 adjuvant chemotherapy. *Oncol Rep* 31: 613-618, 2014.
60. Wu YS, Lin H, Chen D, Yi Z, Zeng B, Jiang Y and Ren G: A four-miRNA signature as a novel biomarker for predicting survival in endometrial cancer. *Gene* 697: 86-93, 2019.
61. He JH, Li BX, Han ZP, Wang L, Lv YB, Zhou JB, Cao MR, Li YG and Zhang JZ: Snail-activated long non-coding RNA PCA3 up-regulates PRKD3 expression by miR-1261 sponging, thereby promotes invasion and migration of prostate cancer cells. *Tumour Biol*: Oct 14, 2016 (Epub ahead of print).
62. Lyu N, Zeng Y, Kong Y, Chen Q, Deng H, Ou S, Bai Y, Tang H, Wang X and Zhao M: Ferroptosis is involved in the progression of hepatocellular carcinoma through the circ0097009/miR-1261/SLC7A11 axis. *Ann Transl Med* 9: 675, 2021.
63. Cao F, Liu S, Li Z, Meng L, Sang M and Shan B: Activation of circ_0072088/miR-1261/PIK3CA pathway accelerates lung adenocarcinoma progression. *Thorac Cancer* 13: 1548-1557, 2022.
64. Zhu Y, Cao F, Liu F, Liu S, Meng L, Gu L, Zhao H, Sang M and Shan B: Identification of potential circular RNA biomarkers in lung adenocarcinoma: A bioinformatics analysis and retrospective clinical study. *Oncol Lett* 23: 144, 2022.
65. Chen R, Zhao WQ, Fang C, Yang X and Ji M: Histone methyltransferase SETD2: A potential tumor suppressor in solid cancers. *J Cancer* 11: 3349-3356, 2020.
66. Das S, Idate R, Regan DP, Fowles JS, Lana SE, Thamm DH, Gustafson DL and Duval DL: Immune pathways and TP53 missense mutations are associated with longer survival in canine osteosarcoma. *Commun Biol* 4: 1178, 2021.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.