

Microbiota in pancreatic cancer: Roles in tumor initiation and progression (Review)

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Abstract. Pancreatic cancer is a highly aggressive malignancy with limited therapeutic options and poor survival outcomes, highlighting the need for an improved understanding of its underlying biology. Advances have positioned the human microbiome as a critical regulator of the initiation and progression of pancreatic cancer. Microbial communities across the oral-gut-tumor axis contribute to tumor initiation through coordinated mechanisms, including the induction of genotoxic stress, chronic inflammation and activation of oncogenic signaling pathways. During tumor progression, microbiota dynamically shape the tumor microenvironment by modulating immune responses, metabolic reprogramming and stromal remodeling. Notably, microbial influences are bidirectional, as tumor-promoting and tumor-suppressive taxa exert opposing effects that converge on shared regulatory pathways within the tumor ecosystem. The present study reviews the current understanding of microbiome involvement in pancreatic cancer, focusing on its mechanistic roles in tumor initiation and progression. Furthermore, the key challenges in the field are discussed, and the emerging opportunities for therapeutic intervention are highlighted. These insights provide a conceptual framework for integrating microbiome research into precision oncology for pancreatic cancer.

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1. Introduction

Pancreatic cancer represents one of the most lethal oncological malignancies, characterized by a poor prognosis that has remained largely unchanged over the past several years (1-3). According to GLOBOCAN 2022, pancreatic cancer accounted for ~510,566 new cases and 467,005 deaths worldwide, ranking 12th in incidence and 6th in mortality among all cancers globally. The age-standardized incidence rate (ASR) was 5.5 per 100,000 in men and 4.0 per 100,000 in women, while the corresponding mortality rates were 5.0 and 3.5 per 100,000, respectively (4).

The global burden of pancreatic cancer exhibits substantial geographic variation. The highest ASRs per 100,000 individuals of both sexes are observed in Uruguay (11.4), Hungary (10.4), Japan (9.8), Austria (9.4) and metropolitan France (9.4), whereas comparatively lower rates persist across a number of regions of South-Central Asia and sub-Saharan Africa (5). In terms of absolute case numbers, China (118,672 new cases), the United States (60,127) and Japan (47,627) bear the greatest disease burden, reflecting the combined influence of population size, population aging and the prevalence of established risk factors, such as pancreatitis, cigarette smoking, heavy alcohol consumption, diabetes and obesity (5). Mortality-to-incidence ratios remain high across all regions, with the leading mortality ASRs per 100,000 individuals reported in Germany (8.2), Japan (8.0) and metropolitan France (8.0) (5). In the United States, pancreatic cancer is estimated to account for ~3.2% of all newly diagnosed cancer cases with 67,530 new cases projected for 2026 (3). Moreover, the mortality burden is projected to increase further and pancreatic cancer is projected to become the second leading cause of cancer-related death by 2030 (6).

The poor clinical outcomes associated with pancreatic cancer are fundamentally linked to its biological behavior and diagnostic challenges. Only 15% of cases are identified at the localized stage, when the 5-year survival rate is 43.6%; however, the majority of patients present with distant metastatic disease, for which the 5-year survival rate reduces to 3.2% (7).

The aforementioned survival statistics reflect the advanced stage at which most patients are diagnosed, as pancreatic cancer typically exhibits minimal or no symptoms during its early phases (8), and the near-equivalence between incidence and mortality rates underscores the aggressive nature of the disease and the limited effectiveness of current therapeutic interventions (9).

Surgical resection remains the only potentially curative treatment modality for pancreatic cancer; however, only patients with localized disease are eligible for pancreatectomy at the time of diagnosis. Even among those who undergo successful margin-negative resection, the 5-year survival rate is 17-21% and up to 86% experience disease recurrence despite complete tumor removal (8,10). Therefore, pharmacological therapy serves a central role in the comprehensive management of pancreatic cancer and as a critical component throughout the entire disease course. Current systemic therapeutic approaches, including FOLFIRINOX and gemcitabine-based regimens, have achieved only modest improvements in overall survival. For advanced disease, the median overall survival time remains at <12 months, with median progression-free survival time at <8 months (11). Pancreatic ductal adenocarcinoma (PDAC), which accounts for >90% of pancreatic malignancies, is characterized by a dense desmoplastic stroma that forms a substantial physical barrier, thereby limiting drug delivery and contributing to systemic therapeutic resistance (8,12). In addition, the hypovascular nature of the tumor microenvironment (TME) further restricts the perfusion-dependent delivery of systemic agents (12). The low frequency of KRAS G12V mutations and the immunosuppressive TME, characterized by limited T-cell infiltration, explain the disappointing efficacy of target therapy and immunotherapy (13). These persistent challenges have intensified research efforts to identify novel therapeutic targets and innovative treatment paradigms.

Microbiota has emerged as a critical determinant of cancer pathogenesis, progression and therapeutic response. Over the past decade, accumulating evidence has fundamentally transformed understanding of the TME, revealing that both local and distant microbial communities actively participate in modulating oncogenic processes across multiple cancer types (14,15). The microbiota influences tumor biology through diverse mechanisms, including the induction of host DNA damage, modulation of oncogenic signaling pathways such as WNT- β -catenin, NF- κ B and PI3K-AKT cascades, and orchestration of local and systemic inflammatory responses (15-19). These microbe-host interactions are bidirectional: Specific microbes can promote pro-tumorigenic niches by suppressing cytotoxic immune cells and enhancing immunosuppressive populations, whereas microbial components may also function as neoantigens or generate metabolites that potentiate antitumor immunity (20).

The gastrointestinal tract, which harbors the largest reservoir of human microbiota, is closely connected to the pancreas through the gut-pancreas axis, suggesting a potential route for microbial influence on pancreatic biology (21). Dysbiosis, characterized by perturbations in microbial composition and function, has been associated with alterations in the pancreatic tumor immune microenvironment, the promotion of chronic inflammation and the facilitation of immune evasion (22,23). As a digestive system malignancy,

PDAC appears to interact with microbial communities across multiple anatomical sites, including the oral cavity, gastrointestinal tract and tumor tissue, indicating that the microbiome may serve as a candidate source of diagnostic biomarkers and therapeutic targets (24).

The present review aimed to systematically summarize current knowledge on the role of the human microbiota in pancreatic cancer, with a focus on its mechanistic contributions to tumorigenesis and therapeutic response. Microbial influences across the oral, gut and intratumoral compartments are examined, and how dysbiosis shapes the tumor immune microenvironment through immune reprogramming, metabolic interactions and microbe-microbe crosstalk is discussed. Furthermore, the potential of microbiota-derived biomarkers for early diagnosis and risk stratification, as well as emerging microbiome-targeted therapeutic strategies, is evaluated.

2. Microbiota in pancreatic cancer initiation

Pancreatic cancer initiation is a multistep process involving the accumulation of genetic and epigenetic alterations together with changes in the TME. Recent studies have suggested that microbial communities across the oral, gastrointestinal and pancreatic compartments may contribute to early carcinogenesis through mechanisms such as genomic instability, inflammation, immune modulation and metabolic reprogramming (16,25). PDAC develops over a prolonged latency period, during which precursor lesions, including pancreatic intraepithelial neoplasia, undergo stepwise progression to invasive carcinoma (26). During this phase, microbial dysbiosis has been associated with alterations in oncogenic signaling, stromal activation and the establishment of an immunosuppressive microenvironment that may facilitate neoplastic progression (21). These observations indicate a potential role for microbiota in pancreatic cancer initiation and support further investigation into their value in early detection and prevention.

Oral and intratumoral microbiome. The oral cavity harbors a diverse microbial community, some members of which have been associated with oncogenic processes. Epidemiological studies have reported consistent associations between periodontal disease and increased pancreatic cancer risk (27,28), and prospective cohort analyses have identified specific oral taxa, particularly members of the 'red' and 'orange' complexes, as being linked to disease development (28). These microorganisms may disseminate to pancreatic tissue via hematogenous or lymphatic routes, and are hypothesized to contribute to pro-tumorigenic microenvironments through virulence-associated mechanisms (27,29). *Porphyromonas gingivalis* has been frequently implicated in this context; its gingipains have been shown to regulate MMP-9 expression through NF- κ B signaling, which may influence extracellular matrix remodeling and tumor invasion (30,31). In addition, the production of nucleoside diphosphate kinase enables hydrolysis of extracellular ATP and inhibition of P2X7-mediated apoptosis, a mechanism that may permit the persistence of damaged cells (32). Following cellular invasion, *P. gingivalis* has been reported to modulate ERK1/2-Ets1, p38/HSP27, JAK/STAT and AKT signaling pathways, which

may collectively contribute to enhanced cell survival under oncogenic conditions (27,33,34). *Aggregatibacter actinomycetemcomitans* produces cytolethal distending toxin, the CdtB subunit of which exhibits phosphatase activity that alters PIP3 signaling and may enhance PI3K pathway activation (35-38). Given the central role of PI3K in KRAS signaling, this mechanism may be relevant to early tumorigenesis (39-41).

The intrapancreatic microbiome is increasingly considered a distinct, low-biomass ecosystem that may become altered and metabolically active during PDAC progression (42). Although previously considered sterile, the pancreas has been shown to harbor microbial communities predominantly composed of Proteobacteria, Firmicutes, Bacteroidetes and Actinobacteria (24,43). The fungal component of the microbiome has also been implicated, with PDAC tissues exhibiting substantially increased fungal burden and distinct composition compared with normal pancreas (44). *Malassezia* species have been associated with activation of the mannose-binding lectin-complement pathway, whereby fungal glycans may induce C3 cleavage and C3a-mediated signaling, potentially promoting tumor cell proliferation and survival (44).

Gut microbiome. The gastrointestinal microbiome is increasingly recognized as a potential contributor to pancreatic cancer initiation through the gut-pancreas axis (45). Studies using metagenomic profiling have identified distinct gut microbial features in patients with PDAC, including reduced α diversity and shifts in phylum-level composition, such as increased *Bacteroides* and decreased Proteobacteria (46,47). These changes have been associated with increased levels of pro-inflammatory cytokines, including TNF- α , and may contribute to the establishment of a microenvironment permissive to tumor development (24,48). Chronic pancreatitis, a well-established risk factor for pancreatic cancer, has also been associated with gut microbiota dysbiosis, with reduced abundance of *Enterococcus faecalis* and other commensals suggesting a potential role for decreased microbial diversity in disease progression (49). In addition, small intestinal bacterial overgrowth may induce lipopolysaccharide-mediated inflammation and has been proposed to contribute to KRAS activation, particularly in the context of sustained inflammatory signaling (50).

Alterations in microbial metabolism may provide an additional mechanistic link between dysbiosis and pancreatic carcinogenesis. Reduced levels of commensal taxa such as *Faecalibacterium* and *Akkermansia* have been associated with disruptions in bile acid and tryptophan metabolic pathways, which may activate pro-inflammatory signaling and promote epithelial-mesenchymal transition (EMT) (51,52). Secondary bile acids may act as damage-associated molecular patterns, inducing reactive oxygen species (ROS) production and NF- κ B activation, while microbial metabolism of tryptophan yields kynurenine derivatives that activate the aryl hydrocarbon receptor, promoting regulatory T-cell differentiation and suppressing antitumor immunity, thereby establishing an immunosuppressive environment conducive to tumor initiation (53). *Helicobacter pylori* has also been implicated in this context; chronic infection has been associated with alterations in gastric endocrine function, including reduced somatostatin secretion and increased secretin activity, which may contribute

to pancreatic ductal hyperplasia (54). In addition, *H. pylori* overgrowth may promote N-nitrosamine formation and DNA damage, while the CagE protein has been reported to exhibit helicase activity and regulate DNA methylation, suggesting a potential role in mutagenesis (27). Together, these findings suggest that the gut microbiome may contribute to pancreatic cancer initiation through interconnected metabolic and immune mechanisms.

3. Microbiota in pancreatic cancer progression

The progression of pancreatic cancer from localized disease to invasive and metastatic stages is increasingly understood to be influenced by complex interactions among cancer cells, the TME and microbial communities (55). Emerging evidence has suggested that specific microbial taxa may exert dichotomous effects during disease progression, with some species potentially promoting malignant advancement and metastatic dissemination through mechanisms such as immune suppression, metabolic reprogramming and stromal remodeling, while others may inhibit tumor progression by enhancing immune activation and maintaining homeostatic barriers (56). These observations provide a conceptual framework for distinguishing metastasis-promoting and metastasis-inhibiting microbes, and highlight the importance of elucidating species-specific mechanisms to inform microbiome-based therapeutic strategies.

Tumor-promoting microbial species. Accumulating evidence has indicated that tumor-promoting microbiota facilitate pancreatic cancer progression through convergent oncogenic mechanisms, despite taxonomic diversity (15,16). *Fusobacterium nucleatum* represents a prototypical example, integrating inflammatory signaling, extracellular matrix remodeling and immune evasion (57). Through Fap2-mediated adhesion, it induces pro-tumorigenic cytokine (IL-8 and CXCL1) secretion (58), while activation of p38 MAPK signaling enhances MMP-dependent matrix degradation (47). In parallel, *F. nucleatum* suppresses antitumor immunity via TIGIT engagement and promotes metastatic communication through extracellular vesicles, collectively driving aggressive disease phenotypes (59).

A second mechanistic axis involves protease-driven signaling and cytoskeletal remodeling, exemplified by *P. gingivalis*. Notably, gingipains from *P. gingivalis* activate protease-activated receptor (PAR)2 and downstream NF- κ B signaling while simultaneously enhancing p38 pathway activity, leading to increased MMP activation and invasion (60). Moreover, *P. gingivalis* modulates cytoskeletal dynamics via protein citrullination, facilitating tumor cell motility (47,61). Other bacteria, including *Klebsiella* species and *E. faecalis*, reinforce these processes by sustaining chronic inflammation, inducing DNA damage, activating inflammasomes and promoting immune escape, thereby amplifying tumor progression (45,62,63).

In addition to signaling and inflammatory pathways, microbial regulation of tumor metabolism and the microenvironment has emerged as a critical determinant of disease progression. *Pseudomonas* species are implicated in amino acid metabolism and the induction of autophagy, potentially linking

microbial activity to PI3K/AKT/mTOR signaling and tumor progression (64). *Streptococcus anginosus* further contributes through MAPK activation and proliferative signaling (65). *P. gingivalis* accelerates pancreatic cancer progression by fostering a neutrophil-dominated proinflammatory TME through enhanced secretion of neutrophilic chemokines (e.g., Cxcl1, Cxcl2, Cxcr2) and neutrophil elastase (66). These findings suggest that distinct microbial taxa converge on shared oncogenic programs, underscoring the microbiome as an integral regulator of pancreatic cancer progression.

Tumor-suppressing microbial species. In contrast to tumor-promoting microbiota, tumor-suppressive microbial species exert protective effects through convergent mechanisms centered on immune activation, metabolic regulation and maintenance of host-microbe homeostasis (67-69). *Lactobacillus* species, particularly *Lactobacillus reuteri*, exemplify this paradigm by enhancing antitumor immunity. Through promoting natural killer (NK)-cell recruitment and activation, as well as inducing M1 macrophage polarization via inhibition of Toll-like receptor (TLR)4 signaling, *Lactobacillus* strengthens innate immune responses within the TME (56,70). Additionally, microbial metabolites such as acetate further potentiate T-cell and NK-cell function, reinforcing systemic and local antitumor immunity (70).

Another key protective microbe, *Akkermansia muciniphila*, contributes to pancreatic cancer suppression through modulation of host immunity and preservation of mucosal barrier integrity (71). This mucin-degrading bacterium is enriched in long-term survivors with pancreatic cancer and is associated with a T cell-inflamed TME (72). *A. muciniphila* enhances antitumor immunity by promoting M1-like macrophage polarization, activating the NLRP3 inflammasome and increasing T-cell infiltration into tumors (73). Notably, in patient-based studies, restoration of *A. muciniphila* has been associated with improved responses to chemotherapy (71,74). However, its effects appear to be context-dependent, as excessive colonization may compromise barrier function and induce systemic inflammation (75).

Metabolic regulation constitutes an additional layer of tumor suppression. *Faecalibacterium prausnitzii* and *Bifidobacterium* species, both reduced in PDAC, produce short-chain fatty acids (SCFAs) such as butyrate that inhibit tumor proliferation, induce apoptosis and enhance immune activation (76-78). Loss of these beneficial microbes leads to impaired barrier function, reduced SCFA production and increased susceptibility to pro-tumorigenic inflammation (78). Collectively, these findings illustrate that tumor-suppressive microbiota converge on key host pathways to counteract pancreatic cancer progression, providing a conceptual framework for microbiome-based therapeutic strategies.

Microbiota-immune crosstalk in the PDAC TME. The oral-gut-tumor microbial axis may shape PDAC progression through coordinated effects on adaptive immunity, myeloid-cell function, innate sensing pathways and metabolite-mediated immune regulation. Emerging evidence has suggested that gut microbiome dysbiosis contributes to T-cell exclusion and dysfunction, whereas microbiota depletion is associated with enhanced intratumoral T-cell infiltration, reduced

myeloid-derived suppressor cell (MDSC) accumulation and suppression of tumor growth in preclinical models (22,79). IL-17 appears to represent an important mechanistic link, as IL-17 neutralization abrogates the tumor-suppressive effect of antibiotic treatment, suggesting a connection between commensal-driven T helper 17 (Th17) responses and PDAC immune evasion (80,81). In parallel, PDAC-derived CCL2 recruits monocytic MDSCs and promotes acquisition of a suppressive phenotype characterized by increased ROS and arginase production, a process that may be partially dependent on MAPK signaling (82).

Microbial regulation of the PDAC immune landscape also involves tumor-intrinsic immunosuppressive programs, innate immune sensing pathways and metabolite-dependent signaling. Thrombin-PAR1 signaling has been linked to reduced cytotoxic T-cell infiltration and increased tumor-associated macrophage accumulation through upregulation of Csf2 and Ptg2 (83). In addition, microbial products (e.g., lipopolysaccharides, flagellins) engage TLRs, cGAS-STING and NOD-like receptors, generating context-dependent effects that range from chronic tumor-promoting inflammation to type I interferon-mediated antitumor responses (22,80,84). Microbial metabolites, including SCFAs, secondary bile acids and tryptophan catabolites, may regulate inflammatory activity, chromatin accessibility, NK-cell function and T-cell differentiation (80,85-87). Furthermore, disruption of gut barrier integrity may facilitate systemic endotoxemia, promoting myeloid skewing and stromal remodeling (88).

Collectively, these observations suggest that microbiota-mediated immune regulation converges on several key pathways, including IL-17/Th17 signaling, CCL2-dependent MDSC recruitment and PAR1-associated myeloid programming. These findings raise the possibility that targeted modulation of microbial communities or their metabolites could help reprogram the PDAC TME toward more durable antitumor immunity.

4. Obesity, diabetes and the microbiome in PDAC

Impact of obesity and diabetes on PDAC. Obesity and type 2 diabetes mellitus (T2DM) are consistently associated with an elevated risk of PDAC and adverse outcomes (8). Pancreatic cancer and T2DM exhibit a bidirectional relationship, in which diabetes acts as both a risk factor and a metabolic consequence of occult pancreatic malignancy (89,90). In available epidemiological analyses, T2DM has been associated with an increased annual incidence rate of PDAC, with a standardized incidence ratio of 1.54 (95% CI, 1.45-1.64) (91). Major contributors to this association include hyperglycemia, hyperinsulinemia, pancreatitis, dyslipidemia, insulin resistance and inflammatory processes within the TME (92). Obesity may further amplify this axis by increasing circulating insulin levels, and altering leptin and adiponectin signaling (93-95).

Obesity and diabetes promote pancreatic carcinogenesis through a convergent immunometabolic program (96). Excess nutrient availability and metabolite accumulation support carcinogenesis by amplifying mutagenic and genotoxic stress, while obesity- and diabetes-associated alterations in gastrointestinal and sex-hormone signaling interact with microbiome dysfunction to foster an immunosuppressive TME (96,97).

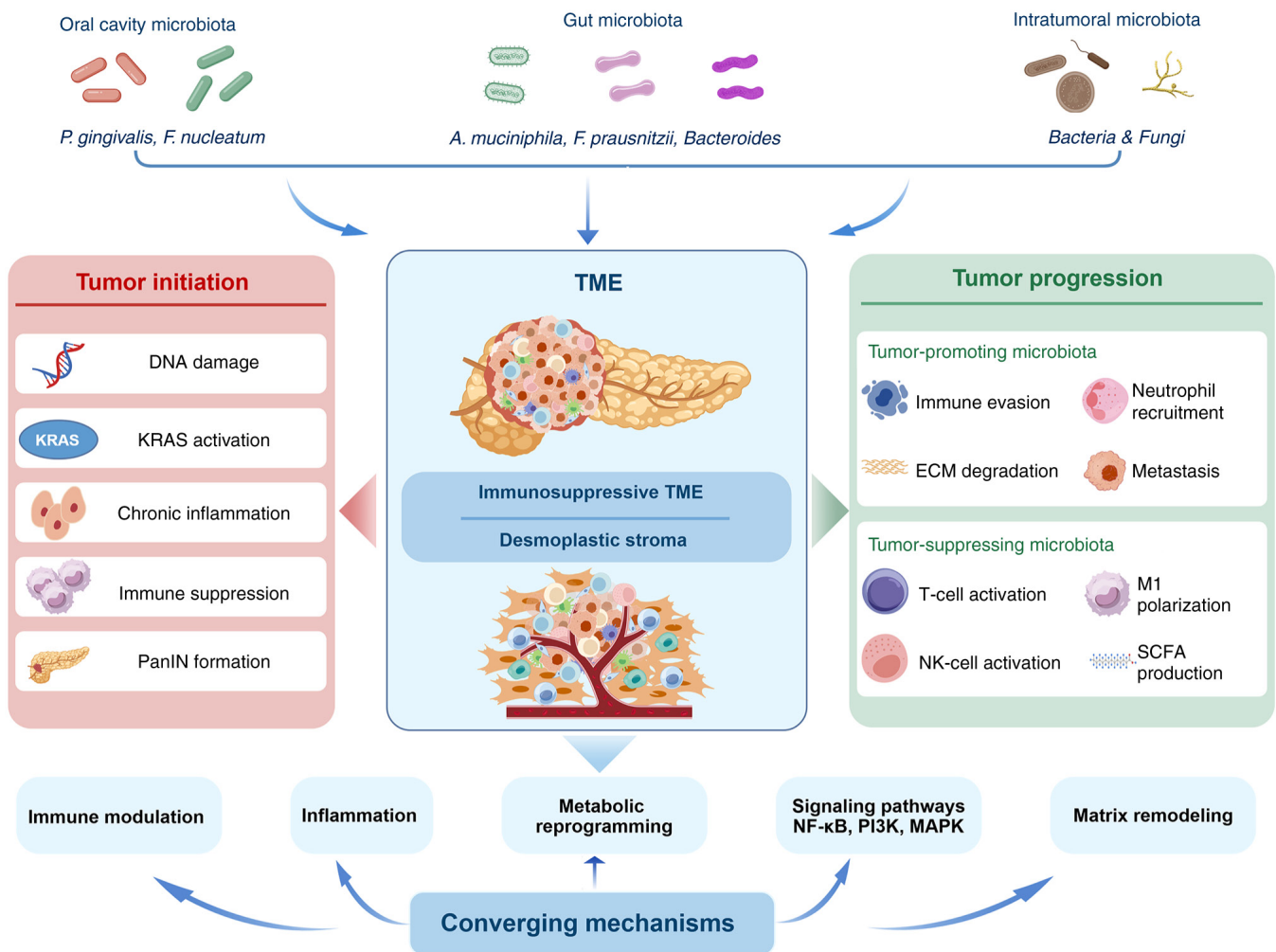


Figure 1. Microbiome-tumor interactions in pancreatic cancer initiation and progression. Oral, gut and intratumoral microbiota may promote tumor initiation through persistent activation of NF- κ B signaling, induction of genotoxic stress by microbial toxins and inflammatory mediators, and activation of oncogenic pathways such as PI3K and MAPK. During disease progression, microbial metabolites and microbial-associated molecular patterns reshape the tumor micro-environment by enhancing inflammation, promoting recruitment of myeloid-derived suppressor cells and tumor-associated macrophages, impairing cytotoxic T-cell function, and facilitating immune evasion and desmoplastic stromal remodeling. Conversely, selected microbial taxa may support antitumor immunity through immune activation and maintenance of tissue homeostasis. This figure was created with BioGDP.com. ECM, extracellular matrix; NK, natural killer; PanIN, pancreatic intraepithelial neoplasia; SCFA, short-chain fatty acid; TME, tumor microenvironment.

Adipokines regulate metabolic homeostasis, inflammatory cascades and immune surveillance through direct effects on the TME. Among these mediators, adiponectin is generally considered tumor-suppressive, whereas leptin promotes proliferative, angiogenic and pro-inflammatory signaling (98). The net effect of adipokine dysregulation in obesity therefore shifts the tumor ecology toward a PDAC-permissive state (98). These changes are reinforced by autophagy dysregulation, endoplasmic reticulum stress, oxidative stress, EMT and exosome secretion, all of which contribute to malignant transformation and tumor evolution (99-102). These observations support earlier pancreatic cancer screening in selected high-risk individuals, particularly those with new-onset diabetes or unexplained metabolic deterioration.

Obesity paradox in immunotherapy. The so-called obesity paradox has attracted growing interest in cancer immunotherapy. Although obesity is generally associated with an increased risk of PDAC and a chronic pro-inflammatory state (8), several clinical studies have reported comparable

or, in some settings, improved responses to immune checkpoint inhibitors among obese patients (103,104). While the clinical relevance of this phenomenon remains incompletely understood, emerging evidence has suggested that obesity-associated alterations in the gut microbiome may interact with systemic and intratumoral immune pathways through multiple interconnected mechanisms. At the metabolic level, gut microbiota-derived SCFAs regulate colonic macrophage function and Treg cell homeostasis, while influencing host leptin secretion, a hormone that correlates with PD-1 expression on CD8⁺ T cells and promotes pro-inflammatory macrophage polarization through TNF- α and IL-6 production (105). Immunologically, specific commensal bacteria such as *Bacteroides fragilis* modulate the balance between CD4⁺ T cell subsets by mediating regulatory T cell conversion, whereas *Lactobacillus* species translocate to secondary lymphoid organs to influence Th1/Th17 responses (105). Notably, *Bifidobacterium* can stimulate antitumor immunity via T cell cross-reactivity between bacterial surface epitopes and tumor neoantigens (105). These microbiota-mediated immune

Table I. Emerging microbial- and vaccine-based therapies in PDAC.

Type	Therapy	NCT number	Phase	Status/results
Microbial therapy	FMT during colonoscopy and then FMT oral capsules	NCT04975217	Early phase I	Suspended
Personalized mRNA vaccine	Autogene cevumeran + atezolizumab + mFOLFIRINOX vs. mFOLFIRINOX alone	NCT05968326	Phase II	Recruiting
mKRAS peptide vaccine	ELI-002 2P	NCT04853017	Phase I	Completed; ~84% of patients developed mKRAS-specific T-cell responses; vaccine-induced immunity was associated with improved relapse-free survival and overall survival, and 67% of patients demonstrated antigen spreading
Personalized neoantigen vaccine	XH001 combined with immune checkpoint inhibition and chemotherapy	NCT06353646	Not applicable	Recruiting
Personalized mRNA vaccine	XP-004 plus PD-1 inhibitor	NCT06496373	Early phase I	Recruiting
Personalized mRNA vaccine	mRNA-0217/S001 with or without pembrolizumab	NCT05916261	Early phase I	Recruiting
Personalized mRNA vaccine	Personalized tumor vaccine combined with a PD-L1 inhibitor	NCT06156267	Early phase I	Not yet recruiting
KRAS-targeted mRNA vaccine	mRNA-5671/V941 with or without pembrolizumab	NCT03948763	Phase I	Terminated; no detailed PDAC-specific efficacy data have been reported publicly
Personalized mRNA vaccine	Personalized neoantigen mRNA vaccine	NCT03468244	Not applicable	Unknown status
PPV	PPV, alone or combined with imiquimod, pembrolizumab or sotigalimab	NCT02600949	Phase I	Recruiting

FMT, fecal microbiota transplantation; mFOLFIRINOX, modified FOLFIRINOX; PD-1, programmed cell death protein 1; PDAC, pancreatic ductal adenocarcinoma; PD-L1, programmed death-ligand 1; PPV, personalized peptide vaccine.

modulations collectively shape the tumor microenvironment, which is characterized in obese patients by increased infiltration of exhausted CD8⁺ T cells with elevated PD-1 expression, enhanced natural killer cell activity and a pro-inflammatory M1-like macrophage phenotype, features that may underlie the improved therapeutic responses to immune checkpoint inhibitors observed in this patient population (105).

Evidence from diet-induced mice obesity models indicates that CD8⁺ tumor-infiltrating lymphocytes exhibit impaired effector function, characterized by reduced expression levels of *Ifng*, *Prf1* and *Gzmb*, together with a metabolic shift from glycolysis toward oxidative phosphorylation. Notably, this dysfunctional state is not accompanied by increased expression of canonical exhaustion markers (106). Diet-induced weight loss, but not semaglutide-induced pharmacological weight

normalization, has been shown to restore T-cell effector function and improve responses to immunotherapy (106). These findings suggest that metabolic health, rather than body mass index alone, may influence antitumor immune competence and treatment efficacy. They are also consistent with the possibility that obesity-associated microbial and inflammatory states contribute to shaping immune fitness and drug responsiveness.

Although some studies have reported an association between obesity and improved survival in patients with pancreatic cancer, the obesity paradox in PDAC should currently be viewed as a hypothesis-generating concept rather than an established clinical paradigm (107). Several non-mutually exclusive mechanisms may contribute to the observed heterogeneity, including differences in body composition (visceral adiposity vs. sarcopenia), metabolically healthy

vs. unhealthy obesity phenotypes, treatment dose normalization and obesity-associated microbiome configurations that may influence immune priming (105,106). Future prospective studies incorporating metabolic, body-composition and microbiome-based stratification may help clarify causality, and facilitate the development of personalized immunotherapy strategies and microbiome-targeted co-interventions.

5. Conclusion

The emerging body of evidence positions the microbiome as an integral component of pancreatic cancer biology, orchestrating tumor initiation and progression through complex and interconnected mechanisms. Microbial communities across the oral-gut-pancreas axis actively contribute to carcinogenesis by inducing genotoxic stress, sustaining chronic inflammation and shaping the tumor immune microenvironment. During disease progression, distinct microbial taxa exert opposing effects; however, these influences converge on shared oncogenic pathways, including immune suppression, metabolic reprogramming and extracellular matrix remodeling. This conceptual framework highlights the microbiome as a dynamic and bidirectional regulator of pancreatic cancer rather than a passive ecological feature (Fig. 1).

The rapidly expanding landscape of microbiome-targeted, immune-modulatory and vaccine-based therapeutic strategies in PDAC is summarized in Table I, together with representative clinical-trial registration numbers (obtained from ClinicalTrials.gov). Increasing evidence suggests that the microbiome-immune axis may represent a therapeutically actionable component of the PDAC TME (23,108). Current investigational approaches encompass multiple complementary strategies, including microbiome modulation through fecal microbiota transplantation, probiotics, defined microbial consortia and dietary interventions (70); myeloid-targeted therapies aimed at disrupting CCL2-CCR2 signaling and MDSC recruitment (82,109); combination immunotherapy approaches designed to overcome T-cell exclusion and myeloid-dominant immune suppression (110); and therapeutic cancer vaccines intended to enhance tumor antigen presentation and T-cell priming (111,112). In parallel, emerging modalities such as oncolytic viruses, KRAS-targeted therapies combined with immune checkpoint blockade, and metabolic interventions in obese or diabetic patients are being explored for their potential to reshape antitumor immunity.

Among these approaches, personalized neoantigen vaccines have generated particular interest. Early clinical studies of autogene cevumeran have demonstrated durable neoantigen-specific CD8⁺ T-cell responses and prolonged persistence of vaccine-induced T-cell clonotypes, supporting the feasibility of individualized immunotherapy in PDAC (112). Nevertheless, most microbiome-directed and immune-modulatory strategies remain in early-phase clinical development, and their long-term efficacy, optimal patient selection criteria and mechanisms of response require further investigation. Collectively, these ongoing studies highlight the growing translational potential of targeting the microbiome-immune axis and provide a framework for the development of more effective precision immunotherapy strategies in PDAC.

Notwithstanding these advances, critical gaps remain. The field is constrained by technical challenges in low-biomass microbiome analysis, limited ability to infer causality and insufficient understanding of temporal dynamics during tumor evolution. Moreover, the high degree of inter-individual variability and the influence of external factors complicate the identification of reproducible microbial signatures. The interactions between bacterial and fungal communities, as well as their collective impact on tumor biology, represent an additional layer of complexity that remains largely unexplored.

Future efforts should move toward a more integrative and translational paradigm. Comprehensive multi-omics profiling, coupled with longitudinal human cohorts and advanced experimental models, will be essential to dissect causal mechanisms and identify actionable targets. In parallel, the development of microbiome-based biomarkers for early detection and patient stratification, as well as therapeutic strategies aimed at reshaping microbial ecosystems, holds considerable promise. Ultimately, integrating microbiome science into precision oncology may redefine current approaches to pancreatic cancer, offering novel options to improve diagnosis, treatment and patient survival.

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YL and YF performed the literature review and wrote the manuscript. Data authentication is not applicable. Both authors read and approved the final manuscript.

Ethics approval and consent to participate

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Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Use of artificial intelligence tools

During the preparation of this work, AI tools (ChatGPT; GPT-5.5; OpenAI; <https://chatgpt.com>) were used to improve the readability and language of the manuscript,

and subsequently, the authors revised and edited the content produced by the AI tools as necessary, taking full responsibility for the ultimate content of the present manuscript.

References

1. Siegel RL, Giaquinto AN and Jemal A: Cancer statistics, 2024. *CA Cancer J Clin* 74: 12-49, 2024.
2. Siegel RL, Kratzer TB, Giaquinto AN, Sung H and Jemal A: Cancer statistics, 2025. *CA Cancer J Clin* 75: 10-45, 2025.
3. Siegel RL, Kratzer TB, Wagle NS, Sung H and Jemal A: Cancer statistics, 2026. *CA Cancer J Clin* 76: e70043, 2026.
4. Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I and Jemal A: Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 74: 229-263, 2024.
5. World Cancer Research Fund: Pancreatic cancer statistics, 2025. <https://www.wcrf.org/preventing-cancer/cancer-statistics/pancreatic-cancer-statistics/>
6. Hesami Z, Olfatifar M, Sadeghi A, Zali MR, Mohammadi-Yeganeh S, Habibi MA, Ghadir MR and Hourii H: Global trend in pancreatic cancer prevalence rates through 2040: An illness-death modeling study. *Cancer Med* 13: e70318, 2024.
7. National Cancer Institute: SEER Cancer Stat Facts: Pancreatic Cancer. In: Surveillance, Epidemiology, and End Results (SEER) Program, Bethesda, MD, 2025. <https://seer.cancer.gov/stat-facts/html/pancreas.html>
8. Stoop TF, Javed AA, Oba A, Koerkamp BG, Seufferlein T, Wilmink JW and Besselink MG: Pancreatic cancer. *Lancet* 405: 1182-1202, 2025.
9. Park W, Chawla A and O'Reilly EM: Pancreatic cancer: A review. *JAMA* 326: 851-862, 2021.
10. Zhang L, Sanagapalli S and Stoita A: Challenges in diagnosis of pancreatic cancer. *World J Gastroenterol* 24: 2047-2060, 2018.
11. Leng Q, Zhou J, Wang X, Zhang P, Xu H and Cao D: HRS-4642 combined with nimotuzumab in the treatment of recurrent or metastatic pancreatic ductal adenocarcinoma: Study protocol of a single-arm, prospective phase Ib/II trial. *Front Pharmacol* 16: 1562481, 2025.
12. Zhang X, Qi B and Chen J: Clinical application and drug resistance mechanism of gemcitabine. *Front Cell Dev Biol* 13: 1702720, 2025.
13. Zou L, Chen J, Bai X, Wang Y, Lu C, Wang Q, Tuerhong S, Li M, Zheng Q, Meng F and Du J: Tumor-penetrating peptide boosts bispecific T-cell engager antitumor efficacy for the pancreatic cancer. *Front Immunol* 16: 1693755, 2025.
14. Zhang W, Xiang Y, Ren H, Liu Y, Wang Q, Ran M, Zhou W, Tian L, Zheng X, Qiao C, *et al*: The tumor microbiome in cancer progression: Mechanisms and therapeutic potential. *Mol Cancer* 24: 195, 2025.
15. Cao Y, Xia H, Tan X, Shi C, Ma Y, Meng D, Zhou M, Lv Z, Wang S and Jin Y: Intratumoral microbiota: A new frontier in cancer development and therapy. *Signal Transduct Target Ther* 9: 15, 2024.
16. Yang L, Li A, Wang Y and Zhang Y: Intratumoral microbiota: Roles in cancer initiation, development and therapeutic efficacy. *Signal Transduct Target Ther* 8: 35, 2023.
17. Jiang Y, Huang Y, Hu Y, Yang Y, You F, Hu Q, Li X and Zhao Z: Banxia Xiexin Decoction delays colitis-to-cancer transition by inhibiting E-cadherin/ β -catenin pathway via *Fusobacterium nucleatum* FadA. *J Ethnopharmacol* 328: 117932, 2024.
18. Zhang L, Leng XX, Qi J, Wang N, Han JX, Tao ZH, Zhuang ZY, Ren Y, Xie YL, Jiang SS, *et al*: The adhesin RadD enhances *Fusobacterium nucleatum* tumour colonization and colorectal carcinogenesis. *Nat Microbiol* 9: 2292-2307, 2024.
19. Stein-Thoeringer CK, Saini NY, Zamir E, Blumenberg V, Schubert ML, Mor U, Fante MA, Schmidt S, Hayase E, Hayase T, *et al*: A non-antibiotic-disrupted gut microbiome is associated with clinical responses to CD19-CAR-T cell cancer immunotherapy. *Nat Med* 29: 906-916, 2023.
20. Leonov G, Starodubova A, Makhnach O, Goldshtein D and Salikhova D: Intratumoral Microbiome: Impact on cancer progression and cellular immunotherapy. *Cancers (Basel)* 18: 100, 2025.
21. Cheng H, Guo H, Wen C, Sun G, Tang F and Li Y: The dual role of gut microbiota in pancreatic cancer: New insights into onset and treatment. *Ther Adv Med Oncol* 17: 17588359251324882, 2025.
22. Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, Mohan N, Aykut B, Usyk M, Torres LE, *et al*: The pancreatic cancer microbiome promotes oncogenesis by induction of innate and adaptive immune suppression. *Cancer Discov* 8: 403-416, 2018.
23. Cruz MS, Tintelnot J and Gagliani N: Roles of microbiota in pancreatic cancer development and treatment. *Gut Microbes* 16: 2320280, 2024.
24. Bautista J, Bedón-Galarza R, Martínez-Hidalgo F, Masache-Cruz M, Benítez-Núñez M, Valencia-Arroyo C and López-Cortés A: Decoding the microbial blueprint of pancreatic cancer. *Front Med (Lausanne)* 13: 1737582, 2026.
25. Yuan Y, Zhang W and Wang D: Targeting the gut-pancreatic axis: Microbial modulation of immunotherapy in pancreatic cancer. *Front Immunol* 17: 1682390, 2026.
26. Lyman MR, Mitchell JT, Raghavan S, Kagohara LT, Huff AL, Haldar SD, Shin SM, Guinn S, Barrett B, Longway G, *et al*: Spatial proteomics and transcriptomics reveal early immune cell organization in pancreatic intraepithelial neoplasia. *JCI Insight* 10: e191595, 2025.
27. Sun Z, Xiong C, Teh SW, Lim JCW, Kumar S and Thilakavathy K: Mechanisms of oral bacterial virulence factors in pancreatic cancer. *Front Cell Infect Microbiol* 9: 412, 2019.
28. Meng Y, Wu F, Kwak S, Wang C, Usyk M, Freedman ND, Huang WY, Um CY, Gonda TA, Oberstein PE, *et al*: Oral bacterial and fungal microbiome and subsequent risk for pancreatic cancer. *JAMA Oncol* 11: 1331-1340, 2025.
29. Jiang H, Li L, Bao Y, Cao X and Ma L: Microbiota in tumors: New factor influencing cancer development. *Cancer Gene Therapy* 31: 1773-1785, 2024.
30. Toraya S, Uehara O, Hiraki D, Harada F, Neopane P, Morikawa T, Takai R, Yoshida K, Matsuoka H, Kitaichi N, *et al*: Curcumin inhibits the expression of proinflammatory mediators and MMP-9 in gingival epithelial cells stimulated for a prolonged period with lipopolysaccharides derived from *Porphyromonas gingivalis*. *Odontology* 108: 16-24, 2020.
31. Maquera-Huacho PM, Spolidorio DP, Manthey J and Grenier D: Effect of hesperidin on barrier function and reactive oxygen species production in an oral epithelial cell model, and on secretion of macrophage-derived inflammatory mediators during *Porphyromonas gingivalis* infection. *Int J Mol Sci* 24: 10389, 2023.
32. Wei W, Li CX, Li MQ, Tan XR and Gong ZC: *Porphyromonas gingivalis* promotes oral carcinogenesis through the NDK-ATP-P2X7 signal axis: An in vitro experimental study. *Arch Oral Biol* 177: 106332, 2025.
33. Inaba H, Sugita H, Kuboniwa M, Iwai S, Hamada M, Noda T, Morisaki I, Lamont RJ and Amano A: *Porphyromonas gingivalis* promotes invasion of oral squamous cell carcinoma through induction of proMMP9 and its activation. *Cell Microbiol* 16: 131-145, 2014.
34. Gao Z, Weng X, Yu D, Pan Z, Zhao M, Cheng B and Li Z: *Porphyromonas gingivalis*-derived lipopolysaccharide promotes glioma cell proliferation and migration via activating Akt signaling pathways. *Cells* 11: 4088, 2022.
35. Yaghoobi H, Bandehpour M and Kazemi B: Apoptotic effects of the B subunit of bacterial cytolethal distending toxin on the A549 lung cancer cell line. *Asian Pac J Cancer Prev* 17: 299-304, 2016.
36. Shenker BJ, Boesze-Battaglia K, Scuron MD, Walker LP, Zekavat A and Dlakić M: The toxicity of the *Aggregatibacter actinomycetemcomitans* cytolethal distending toxin correlates with its phosphatidylinositol-3,4,5-triphosphate phosphatase activity. *Cell Microbiol* 18: 223-243, 2016.
37. Shenker BJ, Dlakić M, Walker LP, Besack D, Jaffe E, LaBelle E and Boesze-Battaglia K: A novel mode of action for a microbial-derived immunotoxin: The cytolethal distending toxin subunit B exhibits phosphatidylinositol 3,4,5-triphosphate phosphatase activity. *J Immunol* 178: 5099-5108, 2007.
38. Jinadasa RN, Bloom SE, Weiss RS and Duhamel GE: Cytolethal distending toxin: A conserved bacterial genotoxin that blocks cell cycle progression, leading to apoptosis of a broad range of mammalian cell lineages. *Microbiology (Reading)* 157: 1851-1875, 2011.
39. Ma J, Fu S, Tan J, Han Y, Chen Y, Deng X, Shen H, Zeng S, Peng Y and Cai C: Mechanistic foundations of KRAS-driven tumor ecosystems: Integrating Crosstalk among immune, metabolic, microbial, and stromal microenvironment. *Adv Sci (Weinh)* 12: e02714, 2025.

40. Diehl AC, Hannan LM, Zhen DB, Coveler AL, King G, Cohen SA, Harris WP, Shankaran V, Wong KM, Green S, *et al*: KRAS mutation variants and Co-occurring PI3K pathway alterations impact survival for patients with pancreatic ductal adenocarcinomas. *Oncologist* 27: 1025-1033, 2022.
41. McDavid WJ, Wilson L, Adderley H, Martinez-Lopez A, Baker MJ, Searle J, Ginn L, Budden T, Aldea M, Marinello A, *et al*: The PI3K-AKT-mTOR axis persists as a therapeutic dependency in KRAS(G12D)-driven non-small cell lung cancer. *Mol Cancer* 23: 253, 2024.
42. Saba E, Farhat M, Daoud A, Khashan A, Forkush E, Menahem NH, Makkawi H, Pandi K, Angabo S, Kawasaki H, *et al*: Oral bacteria accelerate pancreatic cancer development in mice. *Gut* 73: 770-786, 2024.
43. Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, Quesada P, Sahin I, Chandra V, San Lucas A, *et al*: Tumor microbiome diversity and composition influence pancreatic cancer outcomes. *Cell* 178: 795-806.e712, 2019.
44. Aykut B, Pushalkar S, Chen R, Li Q, Abengozar R, Kim JJ, Shadaloey SA, Wu D, Preiss P, Verma N, *et al*: The fungal microbiome promotes pancreatic oncogenesis via activation of MBL. *Nature* 574: 264-267, 2019.
45. Arikath K, Batra SK and Ponnusamy MP: Unveiling the gut-pancreas axis: Microbial influence on stemness and tumor microenvironment of PDAC. *Stem Cells* 44: sxaf064, 2026.
46. Half E, Keren N, Reshef L, Dorfman T, Lachter I, Kluger Y, Reshef N, Knobler H, Maor Y, Stein A, *et al*: Fecal microbiome signatures of pancreatic cancer patients. *Sci Rep* 9: 16801, 2019.
47. Guo X and Shao Y: Role of the oral-gut microbiota axis in pancreatic cancer: A new perspective on tumor pathophysiology, diagnosis, and treatment. *Mol Med* 31: 103, 2025.
48. Sun J, Chen F and Wu G: Potential effects of gut microbiota on host cancers: Focus on immunity, DNA damage, cellular pathways, and anticancer therapy. *ISME J* 17: 1535-1551, 2023.
49. Wu C, Li M and Chen W: Characteristics of gut microbiota in cerulein-induced chronic pancreatitis. *Diabetes Metab Syndr Obes* 14: 285-294, 2021.
50. Yang Q, Zhang J and Zhu Y: Potential roles of the gut microbiota in pancreatic carcinogenesis and therapeutics. *Front Cell Infect Microbiol* 12: 872019, 2022.
51. He Y, Shaoyong W, Chen Y, Li M, Gan Y, Sun L, Liu Y, Wang Y and Jin M: The functions of gut microbiota-mediated bile acid metabolism in intestinal immunity. *J Adv Res* 80: 351-370, 2026.
52. Nagata N, Nishijima S, Kojima Y, Hisada Y, Imbe K, Miyoshi-Akiyama T, Suda W, Kimura M, Aoki R, Sekine K, *et al*: Metagenomic identification of microbial signatures predicting pancreatic cancer from a multinational study. *Gastroenterology* 163: 222-238, 2022.
53. Tang W, Li F, Zheng H, Zhou S, Li C, Xu X and Fu J: Unveiling hidden players: The role of intratumoral microbiota in gastrointestinal cancer dynamics. *J Cancer Res Clin Oncol* 152: 15, 2025.
54. Kunovsky L, Dite P, Jabandziev P, Dolina J, Vaculova J, Blaho M, Bojkova M, Dvorackova J, Uvirova M, Kala Z and Trna J: *Helicobacter pylori* infection and other bacteria in pancreatic cancer and autoimmune pancreatitis. *World J Gastrointest Oncol* 13: 835-844, 2021.
55. Fakruddin M, Chowdhury Z, Suprova SN, Ul Islam B, Sultana Jime J, Bulbul N, Anam MB, Bin Mannan S and Shishir MA: Microbial influences: The Microbiome's impact on pancreatic cancer development and progression. *Curr Pharm Biotechnol: Apr* 18, 2025 doi: 10.2174/011389201035642250407143257 (Epub ahead of print).
56. Zheng SH, Li KZ, Feng G, Wang YT, Wang JN, Li SQ and Sun YD: Gut microbiota reshaping the pancreatic cancer immune microenvironment: New avenues for immunotherapy. *Mol Cancer* 24: 313, 2025.
57. D'Antonio DL, Zenoniani A, Umme S, Piattelli A and Curia MC: Intratumoral *Fusobacterium nucleatum* in pancreatic cancer: Current and future perspectives. *Pathogens* 14: 2, 2024.
58. Udayasuryan B, Ahmad RN, Nguyen TTD, Umaña A, Monét Roberts L, Sobol P, Jones SD, Munson JM, Slade DJ and Verbridge SS: *Fusobacterium nucleatum* induces proliferation and migration in pancreatic cancer cells through host autocrine and paracrine signaling. *Sci Signal* 15: eabn4948, 2022.
59. Pignatelli P, Nuccio F, Piattelli A and Curia MC: The role of *Fusobacterium nucleatum* in oral and colorectal carcinogenesis. *Microorganisms* 11: 2358, 2023.
60. Wang B, Deng J, Donati V, Merali N, Frampton AE, Giovannetti E and Deng D: The roles and interactions of *Porphyromonas gingivalis* and *Fusobacterium nucleatum* in oral and gastrointestinal carcinogenesis: A narrative review. *Pathogens* 13: 93, 2024.
61. Wan Jiun T, Taib H, Majdiah Wan Mohamad W, Mohamad S and Syamimee Wan Ghazali W: Periodontal health status, *Porphyromonas gingivalis* and anti-cyclic citrullinated peptide antibodies among rheumatoid arthritis patients. *Int Immunopharmacol* 124: 110940, 2023.
62. Fiordaliso M, Pala B, Marincola G, Piscione M, Savino L, Mazzone M, Di Marcantonio MC and Mincione G: Biliary stents can modify the microbiota and promote the progression of pancreatic cancer. *Front Oncol* 16: 1633611, 2026.
63. Shimosaka M, Kondo J, Sonoda M, Kawaguchi R, Noda E, Nishikori K, Ogata A, Takamatsu S, Sasai K, Akita H, *et al*: Invasion of pancreatic ductal epithelial cells by *Enterococcus faecalis* is mediated by fibronectin and enterococcal fibronectin-binding protein A. *Sci Rep* 15: 2585, 2025.
64. Luo D, Chen Q, Li Y, Yang J, Tao Y, Ji L and Gong X: Microbiome-metabolome interplay in pancreatic cancer progression: Insights from multi-omics analysis. *Mol Cancer* 24: 240, 2025.
65. Xia K, Zhou Y, Wang W and Cai Y: *Streptococcus anginosus*: The potential role in the progression of gastric cancer. *J Cancer Res Clin Oncol* 151: 143, 2025.
66. Tan Q, Ma X, Yang B, Liu Y, Xie Y, Wang X, Yuan W and Ma J: Periodontitis pathogen *Porphyromonas gingivalis* promotes pancreatic tumorigenesis via neutrophil elastase from tumor-associated neutrophils. *Gut Microbes* 14: 2073785, 2022.
67. Zhang H, Tian Y, Xu C, Chen M, Xiang Z, Gu L, Xue H and Xu Q: Crosstalk between gut microbiotas and fatty acid metabolism in colorectal cancer. *Cell Death Discov* 11: 78, 2025.
68. Zhang C, Wang Y, He M, Wang C, Cao K, Zhong Y, Wang X, Yang M, Zhang G, Lu J, *et al*: Mannose enhances immunotherapy efficacy in ovarian cancer by modulating gut microbial metabolites. *Cancer Res* 85: 2468-2484, 2025.
69. Gao Y, Xu P, Sun D, Jiang Y, Lin XL, Han T, Yu J, Sheng C, Chen H, Hong J, *et al*: *Faecalibacterium prausnitzii* abrogates intestinal toxicity and promotes tumor immunity to increase the efficacy of Dual CTLA4 and PD-1 checkpoint blockade. *Cancer Res* 83: 3710-3725, 2023.
70. Liang Y, Du M, Li X, Gao J, Li Q, Li H, Li J, Gao X, Cong H, Huang Y, *et al*: Upregulation of *Lactobacillus spp.* in gut microbiota as a novel mechanism for environmental eustress-induced anti-pancreatic cancer effects. *Gut Microbes* 17: 2470372, 2025.
71. Chen X, Li Y, Wei G, Zheng Z, Liu W, Li M, Dai X, Liu B, Zhong R and Ye J: The role of *Akkermansia muciniphila* in cancer: Mechanisms, therapeutic potential, and challenges. *IMetaOmics* 2: e70010, 2025.
72. Liu Y, Li Y, Ma H, Deng S and Cheng C: Mechanistic insights into pancreatic cancer progression from circadian rhythm disruption and gut microbiota dysbiosis (Review). *Int J Mol Med* 57: 73, 2026.
73. Fan L, Xu C, Ge Q, Lin Y, Wong CC, Qi Y, Ye B, Lian Q, Zhuo W, Si J, *et al*: *A. Muciniphila* suppresses colorectal tumorigenesis by inducing TLR2/NLRP3-mediated M1-like TAMs. *Cancer Immunol Res* 9: 1111-1124, 2021.
74. Li N, Bai C, Zhao L, Sun Z, Ge Y and Li X: The relationship between gut microbiome features and chemotherapy response in gastrointestinal cancer. *Front Oncol* 11: 781697, 2021.
75. Gubernatorova EO, Gorshkova EA, Bondareva MA, Podosokorskaya OA, Sheynova AD, Yakovleva AS, Bonch-Osmolovskaya EA, Nedospasov SA, Kruglov AA and Drutskaya MS: *Akkermansia muciniphila*-friend or foe in colorectal cancer? *Front Immunol* 14: 1303795, 2023.
76. Kharofa J, Haslam D, Wilkinson R, Weiss A, Patel S, Wang K, Esslinger H, Olowokure O, Sohal D, Wilson G, *et al*: Analysis of the fecal metagenome in long-term survivors of pancreas cancer. *Cancer* 129: 1986-1994, 2023.
77. Tavanaean S, Feizabadi MM, Falsafi S, Aghdaei HA and Hour H: Oral and fecal microbiome alterations in pancreatic cancer: Insights into potential diagnostic biomarkers. *BMC Microbiol* 25: 624, 2025.
78. Li S, Duan Y, Luo S, Zhou F, Wu Q and Lu Z: Short-chain fatty acids and cancer. *Trends Cancer* 11: 154-168, 2025.
79. Sethi V, Kurtom S, Tarique M, Lavania S, Malchiodi Z, Hellmund L, Zhang L, Sharma U, Giri B, Garg B, *et al*: Gut Microbiota promotes tumor growth in mice by modulating immune response. *Gastroenterology* 155: 33-37.e36, 2018.

80. Halle-Smith JM, Pearce H, Nicol S, Hall LA, Powell-Brett SF, Beggs AD, Iqbal T, Moss P and Roberts KJ: Involvement of the gut microbiome in the local and systemic immune response to pancreatic ductal adenocarcinoma. *Cancers (Basel)* 16: 996, 2024.
81. McAllister F, Bailey JM, Alsina J, Nirschl CJ, Sharma R, Fan H, Rattigan Y, Roeser JC, Lankapalli RH, Zhang H, *et al*: Oncogenic Kras activates a hematopoietic-to-epithelial IL-17 signaling axis in preinvasive pancreatic neoplasia. *Cancer Cell* 25: 621-637, 2014.
82. Gu H, Deng W, Zheng Z, Wu K and Sun F: CCL2 produced by pancreatic ductal adenocarcinoma is essential for the accumulation and activation of monocyctic myeloid-derived suppressor cells. *Immun Inflamm Dis* 9: 1686-1695, 2021.
83. Schweickert PG, Yang Y, White EE, Cresswell GM, Elzey BD, Ratliff TL, Arumugam P, Antoniaki S, Mackman N, Flick MJ, *et al*: Thrombin-PAR1 signaling in pancreatic cancer promotes an immunosuppressive microenvironment. *J Thromb Haemost* 19: 161-172, 2021.
84. Zambirinis CP, Levie E, Nguy S, Avanzi A, Barilla R, Xu Y, Seifert L, Daley D, Greco SH, Deutsch M, *et al*: TLR9 ligation in pancreatic stellate cells promotes tumorigenesis. *J Exp Med* 212: 2077-2094, 2015.
85. Temel HY, Kaymak Ö, Kaplan S, Bahcivanci B, Gkoutos GV and Acharjee A: Role of microbiota and microbiota-derived short-chain fatty acids in PDAC. *Cancer Med* 12: 5661-5675, 2023.
86. Nogal A, Louca P, Zhang X, Wells PM, Steves CJ, Spector TD, Falchi M, Valdes AM and Menni C: Circulating levels of the short-chain fatty acid acetate mediate the effect of the gut microbiome on visceral fat. *Front Microbiol* 12: 711359, 2021.
87. Hezaveh K, Shinde RS, Klötgen A, Halaby MJ, Lamorte S, Ciudad MT, Quevedo R, Neufeld L, Liu ZQ, Jin R, *et al*: Tryptophan-derived microbial metabolites activate the aryl hydrocarbon receptor in tumor-associated macrophages to suppress anti-tumor immunity. *Immunity* 55: 324-340.e8, 2022.
88. Bautista J, Lamas-Maceiras M, Hidalgo-Tinoco C, Guerra-Guerrero A, Betancourt-Velarde A and López-Cortés A: Gut microbiome-driven colorectal cancer via immune, metabolic, neural, and endocrine axes reprogramming. *NPJ Biofilms Microbiomes* 12: 21, 2026.
89. Gapstur SM, Gann PH, Lowe W, Liu K, Colangelo L and Dyer A: Abnormal glucose metabolism and pancreatic cancer mortality. *JAMA* 283: 2552-2558, 2000.
90. Li D: Diabetes and pancreatic cancer. *Mol Carcinog* 51: 64-74, 2012.
91. Shen B, Li Y, Sheng CS, Liu L, Hou T, Xia N, Sun S, Miao Y, Pang Y, Gu K, *et al*: Association between age at diabetes onset or diabetes duration and subsequent risk of pancreatic cancer: Results from a longitudinal cohort and mendelian randomization study. *Lancet Reg Health West Pac* 30: 100596, 2023.
92. Sapoor S, Nageh M, Shalma NM, Sharaf R, Haroun N, Salama E, Pratama Umar T, Sharma S and Sayad R: Bidirectional relationship between pancreatic cancer and diabetes mellitus: A comprehensive literature review. *Ann Med Surg (Lond)* 86: 3522-3529, 2024.
93. Jiménez-Cortegana C, López-Saavedra A, Sánchez-Jiménez F, Pérez-Pérez A, Castiñeiras J, Virizuela-Echaburu JA, de la Cruz-Merino L and Sánchez-Margalet V: Leptin, Both bad and good actor in cancer. *Biomolecules* 11: 913, 2021.
94. Jiang J, Fan Y, Zhang W, Shen Y, Liu T, Yao M, Gu J, Tu H and Gan Y: Adiponectin suppresses human pancreatic cancer growth through attenuating the β -catenin signaling pathway. *Int J Biol Sci* 15: 253-264, 2019.
95. Kern PA, Di Gregorio GB, Lu T, Rassouli N and Ranganathan G: Adiponectin expression from human adipose tissue: Relation to obesity, insulin resistance, and tumor necrosis factor-alpha expression. *Diabetes* 52: 1779-1785, 2003.
96. Ruze R, Song J, Yin X, Chen Y, Xu R, Wang C and Zhao Y: Mechanisms of obesity- and diabetes mellitus-related pancreatic carcinogenesis: A comprehensive and systematic review. *Signal Transduct Target Ther* 8: 139, 2023.
97. Ho WJ, Jaffee EM and Zheng L: The tumour microenvironment in pancreatic cancer-clinical challenges and opportunities. *Nat Rev Clin Oncol* 17: 527-540, 2020.
98. Kounatidis D, Vallianou NG, Karampela I, Grivakou E and Dalamaga M: The intricate role of adipokines in cancer-related signaling and the tumor microenvironment: Insights for future research. *Semin Cancer Biol* 113: 130-150, 2025.
99. Chang HH, Moro A, Takakura K, Su HY, Mo A, Nakanishi M, Waldron RT, French SW, Dawson DW, Hines OJ, *et al*: Incidence of pancreatic cancer is dramatically increased by a high fat, high calorie diet in KrasG12D mice. *PLoS One* 12: e0184455, 2017.
100. Chen S, Zhang J, Chen J, Wang Y, Zhou S, Huang L, Bai Y, Peng C, Shen B, Chen H and Tian Y: RER1 enhances carcinogenesis and stemness of pancreatic cancer under hypoxic environment. *J Exp Clin Cancer Res* 38: 15, 2019.
101. Dai E, Han L, Liu J, Xie Y, Kroemer G, Kliksky DJ, Zeh HJ, Kang R, Wang J and Tang D: Autophagy-dependent ferroptosis drives tumor-associated macrophage polarization via release and uptake of oncogenic KRAS protein. *Autophagy* 16: 2069-2083, 2020.
102. Otto L, Rahn S, Daunke T, Walter F, Winter E, Möller JL, Rose-John S, Wesch D, Schäfer H and Sebens S: Initiation of pancreatic cancer: The interplay of hyperglycemia and macrophages promotes the acquisition of malignancy-associated properties in pancreatic ductal epithelial cells. *Int J Mol Sci* 22: 5086, 2021.
103. Alifano M, Daffré E, Iannelli A, Brouchet L, Falcoz PE, Le Pimpech Barthes F, Bernard A, Pages PB, Thomas PA, Dahan M and Porcher R: The reality of lung cancer paradox: The impact of body mass index on long-term survival of resected lung cancer. A french nationwide analysis from the Epithor database. *Cancers* 13: 4574, 2021.
104. Petrelli F, Cortellini A, Indini A, Tomasello G, Ghidini M, Nigro O, Salati M, Dottorini L, Iaculli A, Varricchio A, *et al*: Association of obesity with survival outcomes in patients with cancer: A systematic review and Meta-analysis. *JAMA Netw Open* 4: e213520, 2021.
105. Delaye M, Rousseau A, Mailly-Giacchetti L, Assoun S, Sokol H and Neuzillet C: Obesity, cancer, and response to immune checkpoint inhibitors: Could the gut microbiota be the mechanistic link? *Pharmacol Ther* 247: 108442, 2023.
106. Piening A, Ebert E, Gottlieb C, Khojandi N, Kuehm LM, Hoft SG, Pyles KD, McCommis KS, DiPaolo RJ, Ferris ST, *et al*: Obesity-related T cell dysfunction impairs immunosurveillance and increases cancer risk. *Nat Commun* 15: 2835, 2024.
107. Lee B, Han HS, Yoon YS, Park Y, Kang M and Kim J: 'Obesity paradox' as a new insight from long-term survivors in pancreatic cancer patients. *HPB (Oxford)* 27: 922-929, 2025.
108. Mirji G, Worth A, Bhat SA, El Sayed M, Kannan T, Goldman AR, Tang HY, Liu Q, Auslander N, Dang CV, *et al*: The microbiome-derived metabolite TMAO drives immune activation and boosts responses to immune checkpoint blockade in pancreatic cancer. *Sci Immunol* 7: eabn0704, 2022.
109. Nywening TM, Wang-Gillam A, Sanford DE, Belt BA, Panni RZ, Cusworth BM, Toriola AT, Nieman RK, Worley LA, Yano M and Fowler KJ: Targeting tumour-associated macrophages with CCR2 inhibition in combination with FOLFIRINOX in patients with borderline resectable and locally advanced pancreatic cancer: A single-centre, open-label, dose-finding, non-randomised, phase 1b trial. *Lancet Oncol* 17: 651-662, 2016.
110. Carpenter E, Nelson S, Bednar F, Cho C, Nathan H, Sahai V, di Magliano MP and Frankel TL: Immunotherapy for pancreatic ductal adenocarcinoma. *J Surg Oncol* 123: 751-759, 2021.
111. Thomas AM, Santarsiero LM, Lutz ER, Armstrong TD, Chen YC, Huang LQ, Laheru DA, Goggins M, Hruban RH and Jaffee EM: Mesothelin-specific CD8(+) T cell responses provide evidence of in vivo cross-priming by antigen-presenting cells in vaccinated pancreatic cancer patients. *J Exp Med* 200: 297-306, 2004.
112. Sethna Z, Guasp P, Reiche C, Milighetti M, Ceglia N, Patterson E, Lihm J, Payne G, Lyudovik O, Rojas LA, *et al*: RNA neoantigen vaccines prime long-lived CD8⁺ T cells in pancreatic cancer. *Nature* 639: 1042-1051, 2025.
113. Jiang S, Li H, Zhang L, Mu W, Zhang Y, Chen T, Wu J, Tang H, Zheng S, Liu Y, *et al*: Generic diagramming platform (GDP): A comprehensive database of high-quality biomedical graphics. *Nucleic Acids Res* 53: D1670-D1676, 2025.