

Aloe-emodin, an anthraquinone, *in vitro* inhibits proliferation and induces apoptosis in human colon carcinoma cells

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Abstract. The present study aimed to investigate the anti-cancer effect of aloe-emodin, an anthraquinone compound present in the leaves of *Aloe vera*, on two human colon carcinoma cell lines, DLD-1 and WiDr. Colon carcinoma cells were treated with various concentrations of aloe-emodin for different durations. Cell viability was measured by sodium 3'-[1-(phenylamino-carbonyl)-3,4-tetrazolium]-bis(4-methoxy-6-nitro)benzene-sulfonic acid hydrate assay. DNA fragmentation was analyzed by agarose gel electrophoresis. Nuclear shrinkage was visualized by Hoechst 33258 staining. Western blotting was used to indicate the release of apoptosis-inducing factor and cytochrome *c* from mitochondria and the phosphorylation of Bid. Caspase-3 and casein kinase II activities were measured by the respective assays. Cell viability analyses showed that aloe-emodin induced cell death in a dose- and time-dependent manner. Notably, the WiDr cells were more sensitive to aloe-emodin than the DLD-1 cells. Aloe-emodin caused the release of apoptosis-inducing factor and cytochrome *c* from mitochondria, followed by activation of caspase-3 leading to DNA fragmentation, nuclear shrinkage and apoptosis. In addition, exposure of colon carcinoma cells to aloe-emodin suppressed the casein kinase II activity in a time-dependent manner and was accompanied by a reduced phosphorylation of Bid, a downstream substrate of casein kinase II and a pro-apoptotic molecule. These findings showed that the inhibition of casein kinase II activity, the release of apoptosis-inducing factor and cytochrome *c*, and the caspase-3 activation are involved in aloe-emodin-mediated apoptosis in colon carcinoma cells.

Introduction

Aloe-emodin is a natural active compound present in the leaves of *Aloe vera* (1). Some studies have found that aloe-emodin has numerous biological properties including antiviral, antimicrobial and hepatoprotective activities (2). Aloe-emodin has been reported to exhibit anticancer activity on neuroectodermal tumors, lung squamous cell carcinoma and hepatoma cells (3-5). Aloe-emodin has also been shown to inhibit S-phase progression in both a transformed glia and a human glioma cell line, sensitize HeLa cells to As₂O₃ via the generation of reactive oxygen species, and affect the anticancer activity of cisplatin by blocking the activation of extracellular signal-regulated kinase (6-8). However, the effect of aloe-emodin on human colon cancer cells has yet to be investigated.

Apoptosis is an actively regulated process of cell death since its intrinsic pathway involves mitochondria (9). Mitochondrial outer membrane permeabilization in response to cell death triggers (e.g., DNA damage) is an important early step which is regulated by Bcl-2 and controls the release of proteins, such as cytochrome *c*, from the mitochondria to the cytoplasm where they initiate apoptosis, ultimately leading to cell death (10). Apoptosis-inducing factor, another mitochondrial protein that is released into the cytosol and nucleus, induces chromatin condensation and DNA fragmentation (11). Members of the caspase superfamily are produced and activated, a step that hastens the cell death process involving the caspase-dependent apoptotic pathway (10).

Casein kinase II is a conserved and ubiquitous protein serine/threonine kinase engaged in various functions including normal and abnormal cell proliferation (12). Casein kinase II is localized in both the cytoplasm and nuclear compartment of healthy cells, but is predominantly present in the nuclear compartment of cancer cells and has been found to be deregulated in all carcinomas studied thus far (13). Evidence links casein kinase II to apoptosis (14). Casein kinase II plays a role in the modulation of caspase susceptibility with Bid, a pro-apoptotic member of the Bcl-2 family. It was shown that casein kinase II phosphorylates Bid close to the caspase-8 cleavage site, thereby preventing cells from undergoing apoptosis (15). Thus, casein kinase II is an important target for

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the treatment of cancer, as the disruption of casein kinase II activity prevents the phosphorylation of Bid, thereby allowing the cleavage of Bid by caspases, followed by apoptosis (13,16). Notably, an extensive down-regulation of casein kinase II by antisense casein kinase II in prostate cancer xenografts was found to eventually lead to a complete disappearance of the tumor (13).

To evaluate the anticancer effect of aloe-emodin, we employed two distinct human colon carcinoma cell lines, DLD-1 and WiDr. The observation of DNA fragmentation and nuclear morphology, and the examination of the release of apoptosis-inducing factor and cytochrome *c* showed the apoptotic activity of aloe-emodin. The role of casein kinase II in aloe-emodin-induced apoptosis was also investigated. This study reports for the first time that the natural compound aloe-emodin induces apoptosis in human colon carcinoma cells.

Materials and methods

Aloe-emodin. Aloe-emodin [1,8-dihydroxy-3-(hydroxymethyl)-anthraquinone; CAS registry no. 481-72-1, EU no. 2075717, purity $\geq 95\%$] was purchased from Sigma-Aldrich Co. (St. Louis, MO, USA). It was dissolved in dimethylsulfoxide to a concentration of 18.5 mM and stored at -20°C until use.

Cell culture and treatments. Human colon carcinoma cell lines, DLD-1 and WiDr, were obtained from the Food Industry Research and Development Institute (Hsinchu, Taiwan). Cells were cultured in modified Eagle's medium (MEM) (Sigma-Aldrich Co.), supplemented with 10% heat-inactivated fetal bovine serum (Moregate BioTech, Bulimba QLD, Australia), 1% MEM non-essential amino acid, 100 U/ml penicillin G, 100 $\mu\text{g}/\text{ml}$ streptomycin sulfate and 250 ng/ml amphotericin B (all from Sigma-Aldrich Co.). The two cell lines were grown at 37°C in a humidified atmosphere containing 5% CO_2 . Prior to treatment, the cells were grown to 80-90% confluency and starved by incubation in basal medium (MEM + 1% MEM non-essential amino acid) for 24 h. Various concentrations of aloe-emodin (0-0.37 mM in basal medium) and durations (0, 2, 3, 4, 6, 12, 24 and 48 h) were applied.

Cell viability assay. Cell viability was assessed using the XTT [sodium 3'-[1-(phenylamino-carbonyl)-3,4-tetrazolium]-bis(4-methoxy-6-nitro)benzene-sulfonic acid hydrate] assay kit (Sigma-Aldrich Co.), according to the manufacturer's instructions. The assay was conducted three times, independently.

Lactate dehydrogenase activity assay. At the end of the treatment, the culture medium was centrifuged at 250 x g for 10 min, and the supernatant was saved for the lactate dehydrogenase activity assay. The lactate dehydrogenase released from the lysed cells was detected using the CytoTox 96 Non-Radioactive Cytotoxicity assay (Promega, Madison, WI, USA), according to the manufacturer's instructions. The assay was conducted three times, independently.

DNA fragmentation assay. Treated cells were centrifuged and lysed in lysis buffer [10 mM Tris-HCl (pH 8.0), 100 mM NaCl, 1% SDS, 1 mM EDTA and 2 mg/ml proteinase K] for

1 h at 65°C . Following two successive extractions with phenol/chloroform, the DNA samples were precipitated in ethanol. After washing with 70% ethanol, the DNA samples were resuspended in TE buffer and subjected to 2% agarose gel electrophoresis.

Hoechst 33258 staining. Hoechst 33258 staining was performed as described in a previous study (17). Hoechst 33258-positive nuclei were visualized and photographed using an Olympus fluorescence microscope (Olympus, Tokyo, Japan).

Isolation of mitochondria and extraction of mitochondrial proteins. Mitochondria were isolated using a cell mitochondrial isolation kit (Sigma-Aldrich Co.), according to the manufacturer's instructions with minor modifications. Briefly, treated cells were harvested, resuspended in lysis buffer containing a protease inhibitor cocktail (Sigma-Aldrich Co.) and incubated on ice for 1 min. Extraction buffer A (1X) containing a protease inhibitor cocktail was added, and the solution was centrifuged at 600 x g for 10 min at 4°C . The supernatant (containing mitochondria) was carefully transferred to a fresh tube and centrifuged at 11,000 x g for 10 min at 4°C . Then, the cytosolic fraction was carefully transferred to a new tube. Mitochondrial proteins were extracted by suspending the pellet in CellLytic M Cell Lysis Reagent (Sigma-Aldrich Co.) with a protease inhibitor cocktail. The samples were stored at -80°C until use. The protein content of the cytosolic and mitochondrial fractions was determined using a BCA protein assay kit (Pierce Biotechnology, Inc., Rockford, IL, USA) with bovine serum albumin as a standard.

Total protein preparation. Total proteins were extracted with an M-PER mammalian protein extraction reagent (Pierce Biotechnology, Inc.), according to the manufacturer's instructions. The samples were stored at -80°C until use. The protein concentration was measured as described above.

Immunoblotting. Denatured protein samples were subjected to 15% SDS-PAGE. Proteins were transferred to nitrocellulose membranes. Blocked blots were incubated for 1 h at room temperature with the primary antibodies monoclonal anti-apoptosis-inducing factor (E-1), monoclonal anti-cytochrome *c* (A-8) (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) and polyclonal anti-phospho Bid-Ser61 (Bethyl Laboratories, Inc., Montgomery, TX, USA). Monoclonal anti- β -actin (mAbcam 8226) (Abcam Ltd., Cambridge, UK) was used as a control for equal protein loading. Blots were further incubated with secondary antibodies conjugated with alkaline phosphatase for 1 h at room temperature and then incubated with Lumi-Phos WB Chemiluminescent substrate (Pierce Biotechnology, Inc.) and exposed to a Fuji Medical X-ray film (Fuji Photo Film Co., Tokyo, Japan). Image processing was performed using the software Fuji Image Gauge. All experiments were conducted three times, independently.

Caspase-3 activity assay. The caspase-3 activity was determined using a caspase-3 assay kit (Sigma-Aldrich Co.). The caspase-3 inhibitor N-acetyl-Asp-Glu-Val-Asp-CHO (Ac-DEVD-CHO; Biomol Research Laboratories, Plymouth, PA, USA) was co-treated with aloe-emodin at a final concentration of 200 μM

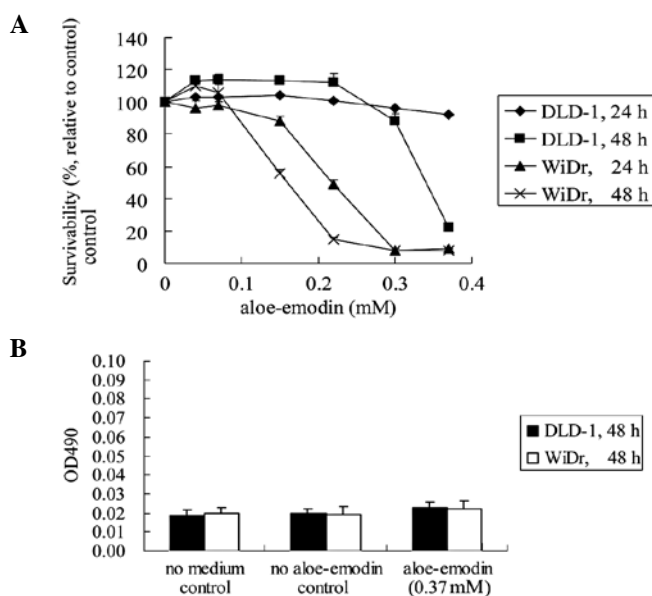


Figure 1. Aloe-emodin is cytotoxic to but does not induce necrosis in the DLD-1 and WiDr colon carcinoma cell lines. The cells were incubated with different concentrations of aloe-emodin for 24 and 48 h. (A) Survival curve of DLD-1 and WiDr cells. The percentage of viable cells was calculated by defining the viability of cells without aloe-emodin treatment as 100%. (B) Lactate dehydrogenase release from DLD-1 and WiDr cells. Data are the results of three independent experiments and are presented as the mean \pm SD.

to inhibit caspase-3 activity in a parallel experiment (18). The assay was conducted three times, independently.

Casein kinase II activity assay. Casein kinase II activity was measured using a casein kinase assay kit (Sigma-Aldrich Co.) combined with the PKLight HTS protein kinase assay kit (Cambrex Bio Science Rockland, Inc., Rockland, MA, USA). Briefly, after aloe-emodin treatment, casein kinase II substrate α -casein and a reaction buffer containing ATP (both from the casein kinase assay kit) were added to the cell lysates and incubated at 37°C for 15 min. The remaining ATP was then consumed by luciferase, and light was generated from ATP and luciferin (luciferase and luciferin were from the PKLight HTS protein kinase assay kit). Bioluminescence was measured using a luminometer (Berthold Detection Systems, Oak Ridge, TN, USA). The PKLight HTS protein kinase assay kit measures the consumption of ATP and is based on the bioluminescent measurement of the remaining ATP present in the samples after kinase activity. The bioluminescent signal is inversely proportional to the activity of the kinase, that is, a higher value is indicative of an increased amount of remaining ATP and a lower casein kinase II activity in the cells. The assay was conducted three times, independently.

Statistical analysis. The unpaired Student's t-test was used to identify means that were significantly different from each other ($P < 0.05$).

Results

Aloe-emodin induced cell death in a dose- and time-dependent manner. The effect of aloe-emodin on the viability

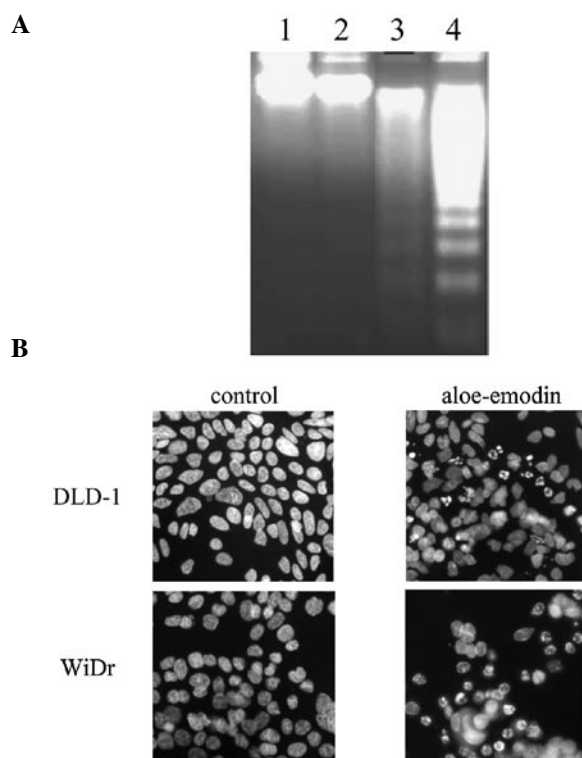


Figure 2. Aloe-emodin induces apoptosis in DLD-1 and WiDr cells. Cells were treated with 0.37 mM aloe-emodin for 24 h. (A) DNA fragmentation was analyzed by 2% agarose gel electrophoresis. Lane 1, untreated DLD-1 cells; lane 2, untreated WiDr cells; lane 3, treated DLD-1 cells and lane 4, treated WiDr cells. (B) Apoptotic cells were detected by Hoechst 33258 staining and examined by fluorescence microscopy. The representative images of three independent experiments are shown (magnification, x400).

of DLD-1 and WiDr colon carcinoma cells was examined using the XTT viability assay. Exposure of DLD-1 cells to various concentrations of aloe-emodin (0.04, 0.07, 0.15, 0.22, 0.30 and 0.37 mM) for 24 or 48 h resulted in a dose- and time-dependent decrease in cell viability relative to control cell cultures (Fig. 1A). The IC_{50} value for 48 h of exposure to aloe-emodin was 0.30-0.37 mM. A similar effect of aloe-emodin was observed after 24 or 48 h of incubation using the WiDr cell line (Fig. 1A). In the latter case, however, the IC_{50} value for a 48-h exposure to aloe-emodin was 0.15-0.22 mM, suggesting that the WiDr cells were more sensitive to aloe-emodin than the DLD-1 cells. Aloe-emodin-induced cell death was significant at a concentration of 0.37 mM. Thus, this concentration was applied for the rest of the experiments.

Aloe-emodin did not induce necrosis in DLD-1 and WiDr cells. Treated cells were evaluated for the presence of necrotic cell death by measuring the lactate dehydrogenase release into the medium. Exposure of DLD-1 and WiDr cells to 0.37 mM aloe-emodin for 48 h did not result in the release of lactate dehydrogenase (Fig. 1B).

Aloe-emodin induced apoptosis in DLD-1 and WiDr cells. To investigate whether the aloe-emodin-mediated cell death in DLD-1 and WiDr cells was due to an apoptotic mechanism, DNA fragmentation and nuclear morphological changes that occurred during aloe-emodin treatment were observed.

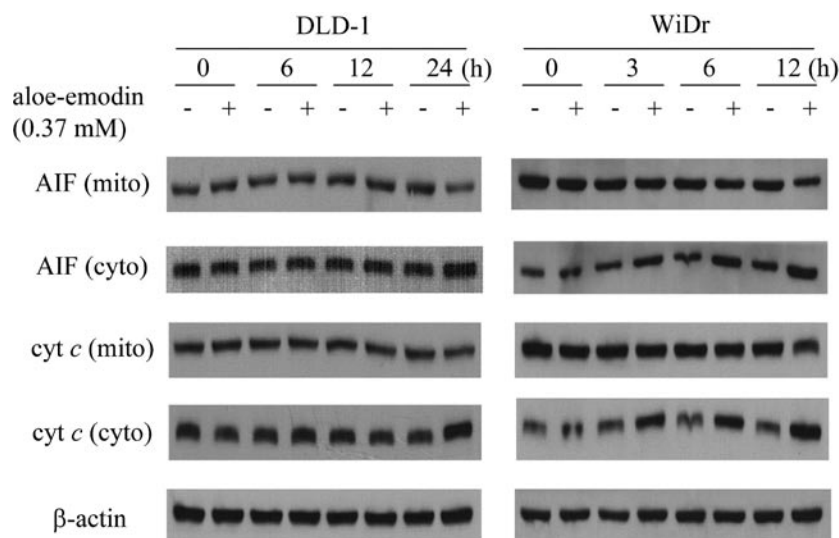


Figure 3. Aloe-emodin induces the release of apoptosis-inducing factor and cytochrome *c* in DLD-1 and WiDr cells. Cells were incubated with or without 0.37 mM aloe-emodin for 0, 3, 6, 12 and 24 h. Mitochondrial (mito) and cytosolic (cyto) fractions were analyzed by 15% SDS-PAGE and probed with primary antibodies as described in Materials and methods. Results are representative of three independent experiments.

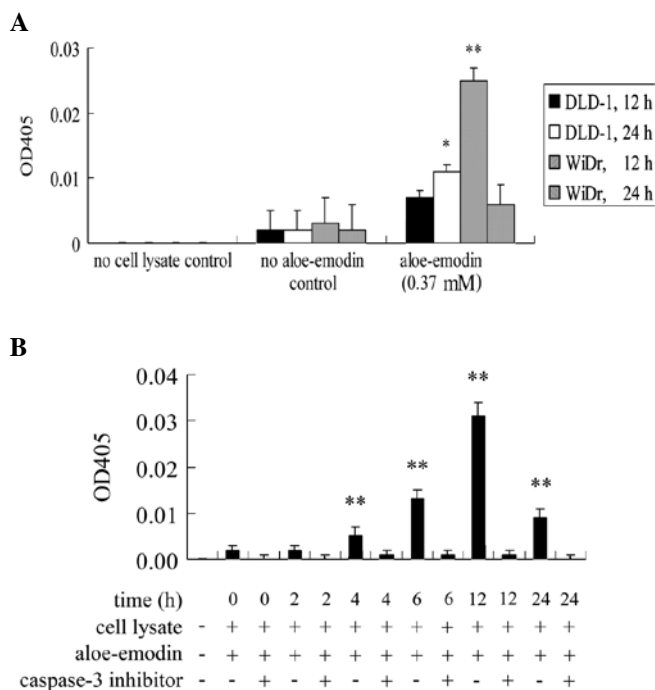


Figure 4. Aloe-emodin activates caspase-3 in DLD-1 and WiDr cells. (A) Cells were incubated with or without 0.37 mM aloe-emodin for 12 and 24 h. (B) WiDr cells were incubated with or without 0.37 mM aloe-emodin for 0, 2, 4, 6, 12 and 24 h. To inhibit caspase-3 activity, a caspase-3 inhibitor, Ac-DEVD-CHO, co-treated with aloe-emodin, was added to WiDr cells at a final concentration of 200 μ M. Each bar indicates the value of caspase-3 activity by measuring the release of p-nitroaniline (OD405). Data are the results of three independent experiments and are presented as the mean \pm SD. Means are significantly different from the respective control values; * P <0.05 and ** P <0.01.

Treatment of DLD-1 and WiDr cells with 0.37 mM aloe-emodin for 24 h resulted in DNA fragmentation (Fig. 2A). Treatment of DLD-1 and WiDr cells with 0.37 mM aloe-emodin for 24 h also resulted in changes in the nuclear

morphology as evidenced by the Hoechst 33258 staining (Fig. 2B), a dye often used to label DNA in living cells and to observe morphological and nuclear changes (17,19,20). More fragmented nuclei were observed upon aloe-emodin treatment, both in the DLD-1 and WiDr cells. These observations indicated that aloe-emodin-induced cell death in DLD-1 and WiDr cells involved a typical apoptotic pathway.

Aloe-emodin induced the release of apoptosis-inducing factor and cytochrome c. The release of apoptosis-inducing factor and cytochrome *c* in DLD-1 and WiDr cells upon treatment with aloe-emodin was characterized because these processes are typically related to apoptosis (14). An immunoblot analysis of the mitochondrial and cytosolic fractions derived from aloe-emodin-treated DLD-1 and WiDr cells showed a significant decrease in the amount of apoptosis-inducing factor and cytochrome *c* in mitochondrial fractions. An increase in apoptosis-inducing factor and cytochrome *c* in the cytosolic fraction for the indicated time intervals was also noted (Fig. 3). Following treatment of the DLD-1 cells, the release of apoptosis-inducing factor and cytochrome *c* commenced at 24 h, while that of WiDr cells started after 3 h of treatment. The results were consistent with the cell viability data and confirm that WiDr cells are more sensitive to aloe-emodin than DLD-1 cells.

Effect of aloe-emodin on the activation of caspase-3. The involvement of caspase-3 in aloe-emodin-induced apoptosis in DLD-1 and WiDr cells was evaluated. The activity of caspase-3 was significantly induced 12 h after aloe-emodin treatment in WiDr cells (Fig. 4A). This effect was also observed in DLD-1 cells albeit only significantly after 24 h of aloe-emodin treatment (Fig. 4A). To illustrate the sequence of caspase-3 activation and the release of apoptosis-inducing factor and cytochrome *c*, a time course of caspase-3 activation after aloe-emodin treatment was performed in WiDr cells. As shown in Fig. 4B, caspase-3 activity was significantly induced after 4 h of aloe-emodin treatment. The results showed that the

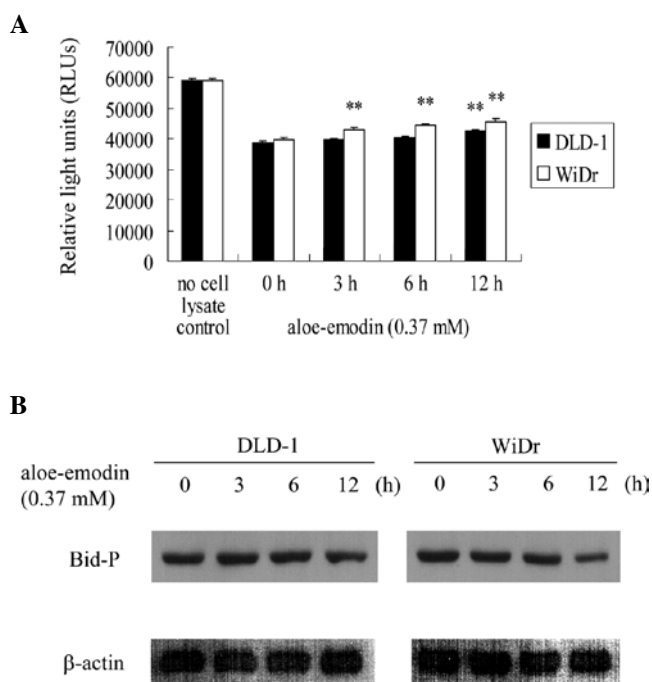


Figure 5. Aloe-emodin inhibits the activity of casein kinase II and the phosphorylation of Bid in DLD-1 and WiDr cells. Cells were incubated with 0.37 mM aloe-emodin for 0, 3, 6 and 12 h. (A) The activity of casein kinase II was measured as described in Materials and methods. Data are the results of three independent experiments presented as the mean \pm SD. Means that are significantly different from the respective control values: ** $P < 0.01$. (B) Cell lysates were analyzed by 15% SDS-PAGE and probed with primary antibodies as described in Materials and methods. Results are representative of three independent experiments.

release of apoptosis-inducing factor and cytochrome *c* started before caspase-3 activation. To demonstrate whether the activation of caspase-3 is essential for aloe-emodin-mediated apoptosis, the influence of the peptidyl inhibitor of caspase-3, Ac-DEVD-CHO, a generally used caspase-3 inhibitor, was analyzed. Aloe-emodin-induced caspase-3 activation in WiDr cells was blocked when co-treated with Ac-DEVD-CHO (Fig. 4B). These data showed that the activation of caspase-3 was an essential step in aloe-emodin-induced apoptosis in the two cell lines. However, the DLD-1 cells appeared to be less sensitive than the WiDr cells (data not shown).

Effect of aloe-emodin on casein kinase II activity and Ser61 phosphorylation of Bid. To investigate the role of casein kinase II in aloe-emodin-induced apoptosis, the activity of casein kinase II was examined, as well as the Ser61 phosphorylation of Bid, a downstream substrate of casein kinase II (Ser61 is one of the phosphorylation sites of Bid). The activity of casein kinase II started to decrease at 3 h of exposure to aloe-emodin and continued to decrease in a time-dependent manner in WiDr cells (Fig. 5A). A similar effect was observed in the DLD-1 cells (Fig. 5A); however, the activity of casein kinase II started to decrease after 12 h of exposure to aloe-emodin. Using anti-phospho Bid-Ser61 polyclonal antibodies, immunoblotting was used to confirm the kinase activity. The aloe-emodin-mediated inhibition of casein kinase II activity caused an inhibition of the Ser61 phosphorylation of Bid

(Fig. 5B). These results suggest that aloe-emodin induces apoptosis in DLD-1 and WiDr cells through inhibition of the phosphorylation of Bid.

Discussion

The present study demonstrated that aloe-emodin exhibits anticancer activity in DLD-1 and WiDr colon carcinoma cells and that the cytotoxic mechanism involves the induction of apoptosis. The latter was characterized by the release of apoptosis-inducing factor and cytochrome *c* from the mitochondria to the cytosol, activation of caspase 3 and inhibition of the activity of casein kinase II, with the subsequent inhibition of the downstream phosphorylation of Bid. It was reported that, using the human lung squamous carcinoma cell line CH27, apoptosis induced by aloe-emodin was associated with changes in the expression of members of the Bcl-2 family apoptosis regulators (i.e., increase in the Bax level and decrease in Bcl-XL). Aloe-emodin was found to cause cytochrome *c* release from mitochondria (21). In addition, it was observed that aloe-emodin-induced apoptosis in human hepatocellular carcinoma cell lines HepG2 and Hep3B was accompanied by the induction of p53 and p21 expression (5). Another report suggests that the decrease in the expression of protein kinase C δ and ϵ isoforms plays a critical role in aloe-emodin-induced apoptosis (21). A similar role of protein kinase C was observed in U-937 glioma cells (6). With the exception of protein kinase C α , most of the protein kinase C isozymes (protein kinase C α , β I, γ , δ , ϵ , θ , ξ and μ) were found to decrease upon aloe-emodin treatment. Notably, two hydrophobic residues unique to casein kinase II, Val66 and Ile174, were found to be essential for the interaction with aloe-emodin; a Val66Ala casein kinase II variant exhibited a substantially affected interaction with aloe-emodin (22,23).

In the present study, aloe-emodin-induced apoptosis in colon carcinoma cells involved inhibition of casein kinase II activity and blockage of the phosphorylation of Bid. We also demonstrated that aloe-emodin resulted in the release of apoptosis-inducing factor and cytochrome *c* from mitochondria, followed by the activation of caspase-3. It is known that phosphorylated Bid is cleaved by caspase-8, and truncated Bid binds to mitochondria to induce the release of apoptosis-inducing factor and cytochrome *c* from mitochondria (10,24,25). It is also known that, in the cytosol, cytochrome *c* mediates the allosteric activation of Apf-1, which is required for the proteolytic maturation of caspase-9 and -3 (10,26). Thus, the inhibition of casein kinase II activity following aloe-emodin treatment may lead to the activation of caspase-3 through the phosphorylation of Bid and release of apoptosis-inducing factor and cytochrome *c* from mitochondria. A similar pathway was described for *Radix Paeoniae Alba* extract-induced apoptosis in HL-60 leukemic cells and for quercetin-mediated cell apoptosis in murine melanoma B16-BL6 cells (27,28). Ahmad *et al* found that the down-regulation of casein kinase II by antisense RNA elicited intracellular hydrogen peroxide production, which was associated with caspase-3 activation (29). Although not relative to aloe-emodin, this is another possible pathway between casein kinase II and caspase-3. Thus, further studies are required to elucidate the relationship between casein kinase II and caspase-3 activities.

Notably, in all of the assays used in the present study, the WiDr colon carcinoma cells were significantly more sensitive to aloe-emodin than the DLD-1 cells. The reason for this difference in sensitivity remains unclear. Controversial data have been analyzed to attempt to correlate differences in response to a given drug with the differentiation state of a given cell line. In an *in vitro* system using epidermal cells, maturing basal cells were found to undergo apoptosis upon drug treatment, whereas stem cells appeared to be resistant (30). In addition, it has been suggested that undifferentiated keratinocyte stem cells are protected from spontaneous apoptosis (31). Similarly, differentiated human neural crest-derived tumor cells exhibit an increased resistance to potassium ionophore-mediated apoptosis. Cells and human keratinocytes that are undifferentiated are more sensitive to H₂O₂-induced cell death than their differentiated counterparts (32,33). However, it was shown that the TNF-related apoptosis-inducing ligand triggers apoptosis in undifferentiated but not in differentiated human keratinocytes (34). Microscopy analyses have demonstrated that DLD-1 cells are less differentiated than WiDr cells (data not shown). Thus, our results strongly suggest that the degree of differentiation of colon carcinoma cells affects the cellular response to aloe-emodin. In addition to the degree of differentiation of culture cells, evidence shows that p53-mutant cell lines are less sensitive to aloe-emodin than their p53 wild-type counterparts (35). In the present study, the amount of mutant p53 in the DLD-1 cells was seven times higher than that in the WiDr cells (1.96 and 0.27 ng/ml cell lysate, respectively). Our data are consistent with the above-mentioned study and suggest that the p53 phenotype affects the cellular response to aloe-emodin.

In conclusion, using two human colon carcinoma cell lines, we investigated the anticancer effect of the natural product, aloe-emodin, isolated from *Aloe vera* leaves. We demonstrated for the first time that aloe-emodin exhibits an anticancer effect against colon carcinoma.

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