

Survivin downregulation by siRNA sensitizes human hepatoma cells to TRAIL-induced apoptosis

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Abstract. Survivin, an anti-apoptotic protein, is abundantly expressed in a variety of cancer cells, including hepatoma cells, resulting in the resistance of these cells to various apoptotic stimuli. Tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) is known to induce cancer cell-specific apoptosis, but hepatoma cells are resistant to TRAIL-induced apoptosis. In the present study, we have examined whether the downregulation of survivin by short interfering RNA (siRNA) promotes spontaneous or TRAIL-induced apoptosis in Huh-7 human hepatoma cells. Survivin siRNA transfection downregulated the expression of survivin in Huh-7 cells and reduced cell viability by 20% through inducing spontaneous apoptosis. TRAIL (1 to 2 ng/ml) only slightly induced apoptosis in Huh-7 cells; however, survivin siRNA transfection apparently enhanced TRAIL-induced apoptosis. These results suggest that the level of survivin is linked to the susceptibility of Huh-7 cells to TRAIL. It is possible that survivin downregulation by siRNA combined with TRAIL administration may provide a new therapeutic strategy against hepatoma.

Introduction

Survivin, the smallest mammalian member of the inhibitor of apoptosis (IAP) family, plays a key role, not only in cell division but also in apoptosis inhibition, by repressing caspase activities (1-3). Survivin is abundantly expressed in a variety of cancer cells, including hepatoma cells, but not in normal cells (3-6), resulting in the resistance of cancer cells to various apoptotic stimuli (3,5,7,8). In addition, the overexpression of survivin in cancer is closely related to unfavorable disease

outcome (3,6,9-11). Therefore, survivin is a possible target of cancer therapy. In fact, the downregulation of survivin by anti-sense oligonucleotides, short interfering RNA (siRNA) and dominant-negative mutant induces apoptosis of cancer cells or sensitizes cancer cells to chemotherapeutic agents and irradiation (12).

Tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL), a member of the TNF superfamily, induces apoptosis in a variety of cancer cells with little or no effect on normal cells (13,14). However, several cancer cells, including hepatoma cells, are resistant to TRAIL-induced apoptosis (15). Griffith *et al* have reported that the cellular level of survivin is linked to the resistance against TRAIL-induced apoptosis of renal cell carcinoma cells (16). It has been also reported that the downregulation of survivin by siRNA sensitizes resistant melanoma cells to TRAIL-induced apoptosis (17). Moreover, we previously reported that interferon (IFN)- α downregulated the survivin expression in human hepatoma cells and sensitized these cells to TRAIL-induced apoptosis, and ectopic expression of survivin partially rescued these cells from IFN- α TRAIL-induced apoptosis (18). Therefore, it is possible that survivin plays a role in the resistance to TRAIL-induced apoptosis in human hepatoma cells. To confirm this, in the present study, we examined whether downregulation of survivin by siRNA can sensitize human hepatoma cells to TRAIL-induced apoptosis.

Materials and methods

siRNAs. To select siRNA duplexes for survivin mRNA, we searched for sequences of AA and the N19 (N, any nucleotide) from the open reading frame of human survivin as described previously (19). siRNAs with the following sequences were used for experiments: surA, 5'-GGUAAUUCUCAA CUGCTT-3' (antisense); surB, 5'-GCAAUUUUGUUCUUG GCUCTT-3' (antisense).

siRNA against green fluorescent protein (GFP) was also used as a control: siGFP, 5'-UGCGCUCUGGACGUAGC CTT-3' (antisense). All siRNAs were purchased from Nihon Bioservice (Saitama, Japan).

Cell culture and transfection. Huh-7 human hepatoma cells were maintained in RPMI with 5% fetal bovine serum. Cells were plated on a 96-well plate or Lab-Tek chamber slide at

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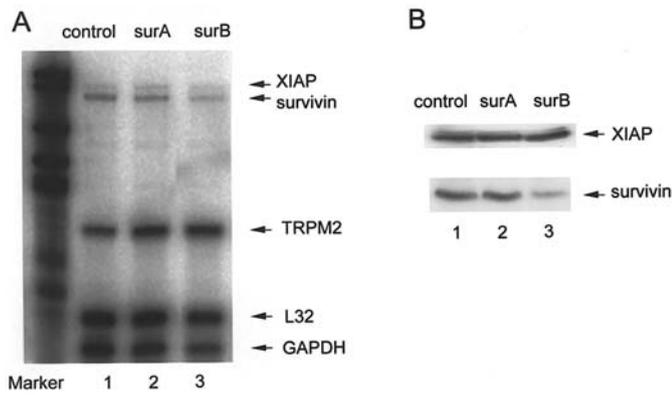


Figure 1. Effect of siRNA transfection on survivin expression in Huh-7 cells. Huh-7 cells were transfected with siGFP as a control (lane 1), surA (lane 2) or surB (lane 3), respectively. (A) The levels of indicated mRNAs were analyzed by RNase protection assay. (B) The expression of XIAP and survivin was analyzed by Western blotting. Results are from one representative experiment from a total of three performed.

an appropriate density. A day later, cells were transfected with each siRNA (4 pmol/10⁴ cells) using oligofectamine (Gibco-Invitrogen, Rockville, MD, USA) according to the instructions provided by the manufacturer. The medium was replaced with fresh medium 6 h after transfection, and the cells were cultured for 36 h and treated with or without varying concentrations of TRAIL for 12 h.

RNase protection assay. The RNase protection assay was performed using a RiboQuant Multi-Probe RNase Protection Assay System (BD Pharmingen, Franklin Lakes, NJ, USA). According to the instructions provided by the manufacturer, an hAPO5c (inhibitors of apoptosis) template set, including an L32 ribosomal protein and a glyceraldehyde-3-phosphate dehydrogenase (GAPDH) template as internal controls, was labeled with [³²P]-UTP using T7 RNA polymerase. The labeled RNA probes were hybridized with 10 μg of total RNA from Huh-7 cells transfected with each siRNA. Samples were digested with RNase to remove single-stranded (non-hybridized) RNA. The remaining probes were resolved on 6% urea-polyacrylamide-bis-acrylamide gels. Gels were dried and analyzed using an image analyzer (BAS; Fuji Film Co., Tokyo, Japan).

Western blot analysis. Cells were lysed by adding lysis buffer (50 mM Tris [pH 8.0], 150 mM NaCl, 0.1% SDS, 1% NP40 and 100 μg/ml PMSF) for 10 min at 4°C and passed several times through a 25-gauge needle. The same amount of protein from each lysate (20 μg/well) was subjected to 12% SDS-PAGE. Proteins were transferred onto nitrocellulose membranes that were then blocked for 1.5 h using 5% non-fat dried milk in PBS containing 0.1% Tween-20 (PBS-T), washed with PBS-T and incubated at room temperature for 1 h in the presence of each antibody (mouse monoclonal anti-human XIAP from MBL, Nagoya, Japan; and rabbit polyclonal anti-human survivin from Alpha Diagnostic International Inc., San Antonio, TX, USA). The membranes were washed with PBS-T and incubated with horseradish peroxidase-conjugated anti-rabbit IgG or anti-mouse IgG for 1 h. Following washing with PBS-T, immunoreactive bands were visualized using the

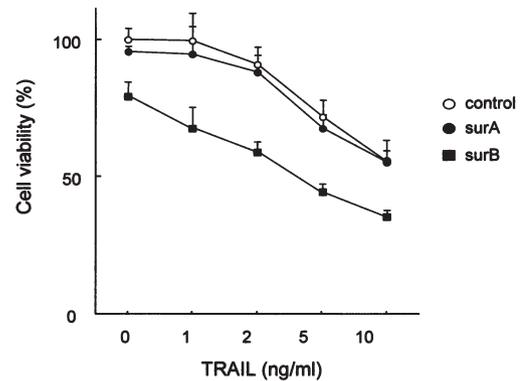


Figure 2. Effect of siRNA transfection with or without TRAIL on the viability of Huh-7 cells. Huh-7 cells were transfected with siGFP as a control (○), surA (●) or surB (■), respectively, and then treated with various concentrations of TRAIL for 12 h. Cell viability was determined by the colorimetric method. Data represent mean ± SD values of the four experiments.

ECL chemiluminescence system (Amersham Life Science, Buckinghamshire, UK).

Analysis of cell viability and apoptosis. Cell viability was determined by the colorimetric method using a Cell Counting kit (Wako Life Science, Osaka, Japan). The absorbance of each well was measured at 405 nm with a microtiter plate reader (Multiskan JX; Thermo BioAnalysis Co., Tokyo, Japan). For the detection of apoptosis, TUNEL assay was performed using a DeadEnd™ Fluorometric TUNEL System (Promega, Madison, WI, USA). Briefly, the cells cultured in Lab-Tek chamber slides were fixed in 4% paraformaldehyde in PBS. The fragmented DNA of apoptotic cells was detected by catalytically incorporating fluorescein-12-dUTP at 3'-OH DNA ends using the terminal deoxynucleotidyl transferase. 1 μg/ml of propidium iodide (PI) was added for nuclear staining. Sample slides were directly visualized by fluorescence microscopy (BX 50; Olympus, Tokyo, Japan) at 520 nm for the green fluorescence of fluorescein and at >620 nm for the red fluorescence of PI.

Results

siRNA transfection downregulates survivin expression. To confirm that siRNA transfection downregulates survivin expression in Huh-7 cells, RNase protection assay and Western blotting were performed (Fig. 1). surA transfection did not repress the level of survivin mRNA, but slightly upregulated the level of XIAP mRNA. In contrast, surB transfection repressed the level of survivin mRNA, but did not upregulate the level of XIAP mRNA (Fig. 1A). The levels of TRPM2, L32 and GAPDH mRNAs were almost unchanged by the transfection of surA and surB. Similarly, surB transfection clearly repressed survivin expression, but did not repress XIAP expression (Fig. 1B).

Survivin downregulation by siRNA enhances TRAIL-mediated cytotoxicity. We examined the effect of survivin siRNA transfection with or without TRAIL on cell viability (Fig. 2). surB transfection alone repressed the viability of Huh-7 cells by almost 20% compared with the control but surA transfection

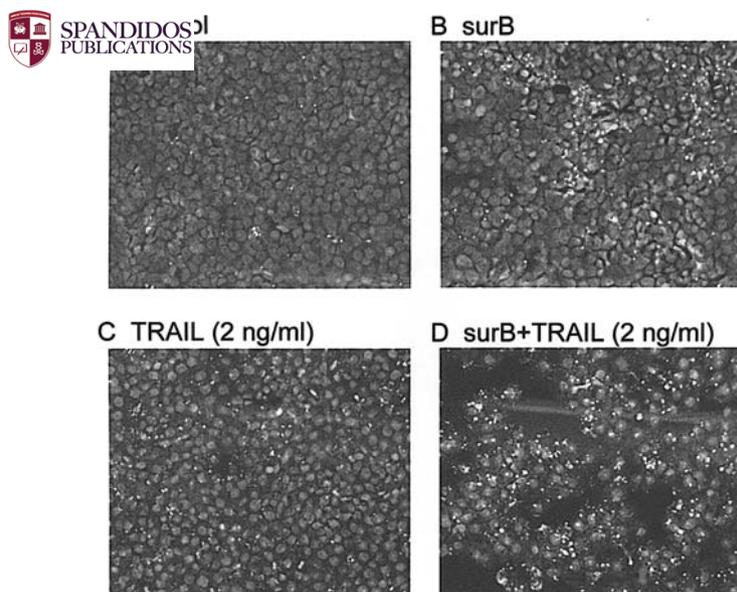


Figure 3. Effect of siRNA transfection with or without TRAIL on apoptosis in Huh-7 cells. Huh-7 cells were transfected with (A) siGFP alone as a control, (B) surB alone, (C) siGFP followed by TRAIL (2 ng/ml) treatment and (D) surB followed by TRAIL (2 ng/ml) treatment. Apoptotic cells were detected by TUNEL assay. Results are from one representative experiment from a total of four performed.

did not. Low concentrations of TRAIL (1 and 2 ng/ml) showed little effect on the viability of control cells, although high concentrations of TRAIL (5 and 10 ng/ml) reduced the viability. On the other hand, surB transfection sensitized Huh-7 cells to TRAIL-mediated cytotoxicity. In fact, the viability of cells transfected with surB and treated with TRAIL (1 or 2 ng/ml) was nearly equal to the viability of control cells treated with TRAIL (5 or 10 ng/ml), respectively.

Survivin downregulation by siRNA enhances TRAIL-induced apoptosis. To clarify whether surB transfection enhances TRAIL-induced apoptosis in Huh-7 cells, TUNEL assay was performed (Fig. 3). TUNEL-positive nuclei, small and bright fluorescence spots, were scattered in the cells either transfected with surB or treated with TRAIL (2 ng/ml), respectively. However, TUNEL-positive nuclei were apparently increased by combination of surB transfection with TRAIL treatment.

Discussion

We previously reported that IFN- α sensitized Huh-7 cells to TRAIL-induced apoptosis, in which IFN- α not only reduced survivin expression but also inhibited NF- κ B activity and increased the expression of DR5, a death receptor of TRAIL (18). However, we could not address which was the key change directly relevant to the susceptibility to TRAIL, although ectopic expression of survivin partially rescued the IFN- α /TRAIL-induced apoptosis. In the present study, the downregulation of survivin by siRNA transfection sensitized Huh-7 cells to TRAIL-induced apoptosis as well as IFN- α , suggesting that downregulation of survivin is the major cause of IFN- α -mediated susceptibility of Huh-7 cells to TRAIL. Therefore, it is possible that the level of survivin is linked to the resistance against TRAIL-induced apoptosis in Huh-7

cells. This is consistent with previous observations in renal cell carcinoma cells and melanoma cells (16,17).

In this study, downregulation of survivin reduced the viability of Huh-7 cells through the induction of apoptosis. It was reported that an adenovirus expressing the dominant negative mutant of survivin caused spontaneous apoptosis in human cell lines of breast, cervical, prostate, lung and colorectal cancer but did not affect the viability of normal human cells, including fibroblasts, endothelium or smooth muscle cells (12,20). It was also reported that transduction of antisense oligonucleotides and siRNA against survivin induced apoptosis in HepG2 (21) and SMMC-7721 (22) human hepatoma cells, respectively. Therefore, it is likely that survivin supports the survival of hepatoma cells through inhibiting spontaneous apoptosis.

Survivin is abundantly expressed in hepatoma cells but not in normal hepatocytes (5,6), and TRAIL specifically induces apoptosis in cancer cells but not in normal hepatocytes (13,14). Therefore, the combination therapy of survivin knockdown and TRAIL administration may provide a new strategy for hepatoma treatment.

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