

# Rapid decrease of intracellular pH associated with inhibition of Na<sup>+</sup>/H<sup>+</sup> exchanger precedes apoptotic events in the MNK45 and MNK74 gastric cancer cell lines treated with 2-aminophenoxazine-3-one

HIROYUKI NAGATA<sup>1\*</sup>, XIAO-FANG CHE<sup>2\*</sup>, KEISUKE MIYAZAWA<sup>2</sup>, AKIO TOMODA<sup>2</sup>, MASATO KONISHI<sup>3</sup>, HIDEYUKI UBUKATA<sup>1</sup> and TAKAFUMI TABUCHI<sup>1</sup>

<sup>1</sup>Fourth Department of Surgery, Tokyo Medical University, Ibaragi Medical Center, 3-20-1 Chuo, Ami, Inashiki, Ibaraki 300-0395; Departments of <sup>2</sup>Biochemistry, <sup>3</sup>Cell Physiology, Tokyo Medical University, 6-1-1 Shinjuku, Tokyo 160-8402, Japan

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**Abstract.** The effects of Phx-3 on changes in intracellular pH (pH<sub>i</sub>) in the MKN45 and MKN74 human gastric cancer cell lines were evaluated in order to determine the mechanism for the proapoptotic effects of 2-aminophenoxazine-3-one (Phx-3) on these cells. Phx-3 (100 μM) reduced pH<sub>i</sub> in MKN45 from 7.45 to 5.8, and in MKN74 from 7.5 to 6.2 within 1 min of engagement with these cells. Such a decrease of pH<sub>i</sub> was closely correlated with the dose of this phenoxazine and continued for 4 h. The activity of Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 (NHE1), which is involved in H<sup>+</sup> extrusion from the cells, was dose-dependently suppressed by Phx-3 in these cells, and was greatly suppressed in the presence of 100 μM Phx-3. This result indicates that the decrease of pH<sub>i</sub> in MKN45 and MKN74 cells is closely associated with the inhibition of NHE1 in these cells. The morphology of these cells at 24 h after treatment with Phx-3 indicated shrinkage of the cells and condensation of the nuclear chromatin structure, which are characteristic of the apoptotic events in these gastric cancer cells. Cytotoxicity of Phx-3 against MKN45 and MKN74 cells was extensive because almost all MKN45 cells lost viability at 24 h in the presence of 20 μM Phx-3, and nearly 50% of the MKN74 cells lost viability in the presence of 50 μM Phx-3. These results suggest that rapid and extensive decrease of pH<sub>i</sub> in human gastric cancer MKN45

and MKN74 cells caused by Phx-3 might disturb intracellular homeostasis, leading to apoptotic and cytotoxic events in these cells. Phx-3 is a good candidate for therapeutics of gastric cancer that is intractable to conventional chemopreventive therapies.

## Introduction

It has been suggested that a decrease of intracellular pH (pH<sub>i</sub>) precedes apoptotic events in cancer cells (1,2). Therefore, drugs to induce pH<sub>i</sub> reduction have been considered good candidates for treating cancer and causing programmed cell death (3). For example, cyclohexamide, etoposide, and camptothecin decrease pH<sub>i</sub> in cancer cells, leading to apoptosis of the cells (3-5); however, these drugs often exert significant adverse effects on the body (6,7).

The oxidative phenoxazines [e.g., 2-amino-4,4a-dihydro-4a,7-dimethyl-3H-phenoxazine-3-one (Phx-1) and 2-aminophenoxazine-3-one (Phx-3)] exert anticancer effects on various species of cancer cells *in vitro* (8-10) and *in vivo*, without marked adverse effects (11-13). Recently, Che *et al* (14) reported that apoptotic events in KB-3-1 cells (human epidermoid carcinoma cell line) and K562 cells (human chronic myeloid leukemia cell line) are preceded by a decrease in pH<sub>i</sub> in cells treated with Phx-3, a phenoxazine produced by the reaction of *o*-aminophenol with bovine hemoglobin. However, the detailed mechanism for pH<sub>i</sub> decrease in these cancer cells with Phx-3 remains unclear.

Regarding the regulation of pH<sub>i</sub>, it has been suggested that among Na<sup>+</sup>/H<sup>+</sup> exchangers, Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 (NHE1) is ubiquitously present in the plasma membrane in the cells and plays a pivotal role in regulating pH<sub>i</sub> (15-17). Therefore, it is important to investigate whether a decrease in pH<sub>i</sub> precedes apoptosis in cancer cells treated with Phx-3, and whether a decrease in Phx-3-inducible pH<sub>i</sub> is associated with the activity of NHE1 that is primarily involved in pH<sub>i</sub> regulation in cancer cells.

*Correspondence to:* Dr Akio Tomoda, Department of Biochemistry, Tokyo Medical University, 6-1-1 Shinjuku, Tokyo 160-8402, Japan  
E-mail: tomoda@tokyo-med.ac.jp

\*Contributed equally

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Kasuga *et al* (18) reported that gastric cancer cells such as MKN45 and MKN74 are susceptible to the treatment of Phx-3, i.e., Phx-3 greatly inhibited the growth of these cells and the cell cycle and consequently induced the programmed death of these cells. However, the effects of Phx-3 on changes in  $pH_i$  of these cells have not been investigated. Thus, we investigated the effect of Phx-3 on these gastric cancer cells in terms of the  $pH_i$  regulation associated with NHE1 and the causal relationship of  $pH_i$  decrease and apoptotic events. The present study is the first report on the inhibitory effects of Phx-3 on NHE1 in human gastric cancer cells, including MKN45 and MKN74 cells, leading to a drastic decrease in  $pH_i$  in these cells, and finally to cellular apoptosis.

## Materials and methods

**Phx-3 and other reagents.** For this study, Phx-3 was obtained by the reaction of *o*-aminophenol (Wako Pure Chemicals, Osaka, Japan) with bovine erythrocytes and was purified using column chromatography, as described by Nakachi *et al* (19). The chemical structure of Phx-3 is presented in Fig. 1. Phx-3 was dissolved in a mixture of dimethylsulfoxide (DMSO) and ethyl alcohol (3:1) as a vehicle to make 20 mM solution.

This solution was diluted appropriately with an isotonic saline and was used for the experiments with MKN45 and MKN74 cells.

**Cell line and culture condition.** Human gastric cancer cell lines MKN45 and MKN74 were obtained from the Health Science Research Resources Bank (Osaka, Japan). MKN45 is a poorly differentiated adenocarcinoma, and MKN74 is a moderately differentiated tubular adenocarcinoma. These cells were cultured in Eagle's minimum essential medium  $\alpha$  (MEM $\alpha$ ) (Kohjin Bio, Saitama, Japan) supplemented with 10% heat inactivated fetal calf serum (Equitech-Bio, Kerrville, TX) at 37°C in a 5% CO<sub>2</sub> humidified atmosphere.

**Measurement of  $pH_i$  in MKN45 and MKN74 cells treated with Phx-3.** The  $pH_i$  of MKN45 and MKN74 cells was determined according to the method described by Litman *et al* (20). Briefly, these gastric cancer cells ( $4 \times 10^7$ /ml) were incubated with a pH-sensitive fluorescent probe (2,7)-biscarboxyethyl-5(6)-carboxyfluorescein-AM (BCECF-AM) (3  $\mu$ M) (Dojin Chemical, Kumamoto, Japan) in HEPES buffer (142 mM NaCl, 3 mM KCl, 1 mM MgSO<sub>4</sub>, 1 mM CaCl<sub>2</sub>, 0.2% albumin, 10 mM glucose, 20 mM HEPES, pH 7.4) at 37°C for 30 min. After being washed once with HEPES buffer, the cells were resuspended in HEPES buffer. The cells ( $3 \times 10^6$ ) were treated with or without various concentrations of Phx-3 for 0, 1, 2, 5, 10, 20 and 30 min. Fluorescence was measured at an excitation wavelength of 500 nm and an emission wavelength of 530 nm, using a FP-6200 spectrofluorometer (Jasco Co. Ltd, Tokyo). To calibrate fluorescence, BCECF-AM-loaded cells ( $3 \times 10^6$ ) were suspended in pH 6.6, 7.2, 7.4, 7.8 and 8.2 calibration buffer (130 mM KCl, 10 mM NaCl, 1 mM MgSO<sub>4</sub>, 10 mM Na-MOPS) and 10  $\mu$ g/ml nigericin was added to equilibrate the external and internal pH, which enabled determining the unknown  $pH_i$ . A linear calibration curve for  $pH_i$  was obtained

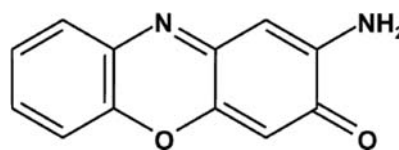


Figure 1. Chemical structure of Phx-3.

(data not shown). Therefore, it was possible to estimate the  $pH_i$  of MKN45 or MKN74 cells loaded with BCECF-AM.

**Measurement of NHE activity in MKN45 and MKN74 cells, and the inhibitory effects of Phx-3 on NHE.** NHE activity was determined by ammonium chloride (NH<sub>4</sub>Cl)-prepulse technique (21), monitoring the changes in fluorescence of BCECF (an excitation wavelength of 500 nm and an emission wavelength of 530 nm) in MKN45 and MKN74 cells, as described above. Briefly, cells were washed twice after BCECF-AM loading, and were poured in a 2 ml quartz cell. NH<sub>4</sub>Cl from 2 M stock solution (approximately pH 7.3) was added to cell suspensions at a final concentration of 50 mM. The  $pH_i$ , which was monitored by the changes in fluorescence at 530 nm, rapidly increased as neutral NH<sub>3</sub> entered the cells. This increase was followed by a slow decline that was due to an influx of NH<sub>4</sub><sup>+</sup>, accompanied by its partial dissociation. After 3 min, the cells were centrifuged at 400 x g for 30 sec and resuspended in HEPES buffer (pH 7.4), which caused a rapid acidification as the NH<sub>4</sub><sup>+</sup> ions trapped inside the cells crossed the membrane in the form of neutral NH<sub>3</sub>, leaving protons behind. Total elapsed time to centrifuge and resuspend was ~2 min. The sample was poured in a 2 ml quartz cell, again. Final concentration of 0, 20, 50 or 100  $\mu$ M Phx-3 was quickly added to the sample, and was mixed well. The fluorescence spectra were taken to obtain a record of  $pH_i$ , using FP-6200 spectrofluorometer as described above.

**Morphological assessment of culture cells.** For morphological assessment, the cell suspension was taken out at the indicated time and was sedimented in a Shandon Cytospin II (Shandon, Pittsburgh, PA). The preparations were then stained with May-Grünwald-Giemsa solution. Morphological changes were assessed by digital microscopy using BZ-8000 (Keyence Co., Osaka, Japan).

**Estimating the viability of gastric cancer cells treated with various concentrations of Phx-3.** MKN45 and MKN74 cells (3,000/well) in 100  $\mu$ l of culture medium were incubated with or without various concentrations of Phx-3 for 24, 48 and 72 h in 96-well plates. The number of viable cells was assessed using a cell viability assay kit, Cell Titer Blue (Promega Co. Ltd., Madison, WI), with fluorescence measurements at 570 nm for excitation and 590 nm for emission using Multi-detection microplate reader, Powerscan HT (Dainippon Pharmaceutical, Osaka, Japan).

## Results

**Changes in  $pH_i$  in MKN45 and MKN74 cells treated with Phx-3.** Human gastric cancer cell lines MKN45 and MKN74

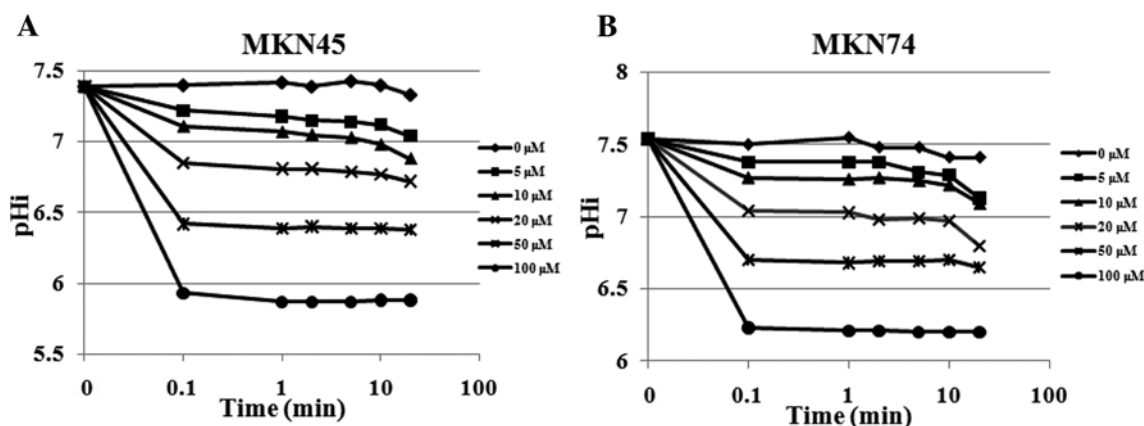


Figure 2. Time course of  $pH_i$  change in human gastric cancer MKN45 or MKN74 cells after the administration of various concentrations of Phx-3. (A) MKN45 cells. (B) MKN74 cells. The value at zero time indicates  $pH_i$  in these cells without Phx-3.

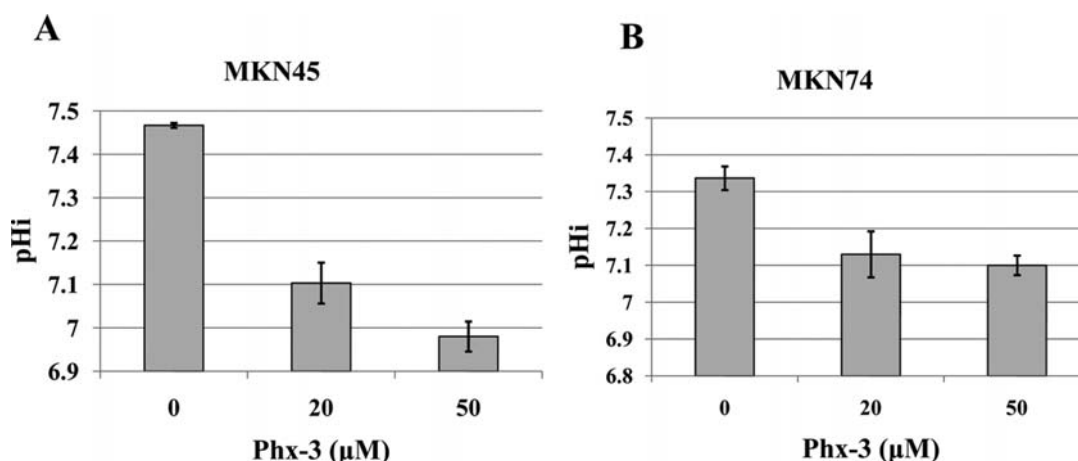


Figure 3. Level of  $pH_i$  in MKN45 or MKN74 cells 4 h after administering 0, 20, or 50  $\mu M$  Phx-3. (A) MKN45 cells. (B) MKN74 cells.

were treated with different concentrations of Phx-3 in order to examine changes in  $pH_i$ . At the resting state,  $pH_i$  in MKN45 was 7.45, and that in MKN74 was 7.5 (Fig. 2), which was higher than the medium pH (7.4) and extensively higher than  $pH_i$  in normal cells [e.g.,  $pH_i$  is 7.0 in normal hematopoietic cells (17) and 7.2 in human erythrocytes (22)]. When MKN45 and MKN74 cells were treated with 100  $\mu M$  Phx-3,  $pH_i$  was decreased to 5.8 in MKN45 and 6.2 in MKN74 within 1 min (Fig. 2). The decreased level of  $pH_i$  was closely dependent on the concentrations of Phx-3 added to these cells. Such intracellular acidification continued for 4 h in both MKN45 and MKN74 cells, dependent on the dose of Phx-3 (Fig. 3).

**Effect of Phx-3 on the activity of  $Na^+/H^+$  exchanger isoform 1 in MKN45 and MKN74 cells.** Among  $Na^+/H^+$  exchangers, the ubiquitous NHE1 is the membrane transporter primarily involved in regulating  $pH_i$  (15-17). We studied the effects of Phx-3 on the activity of NHE1 in MKN45 and MKN74 cells according to the method described by Coates *et al* (21). Fig. 4 depicts changes in  $pH_i$  that were caused by  $NH_4Cl$  pulse loading, washing out  $NH_4Cl$ , and administration of different

concentrations of Phx-3 in MKN45 and MKN74 cells. When MKN45 cells were treated with  $NH_4Cl$  alone,  $pH_i$  increased instantly from 7.38 to 7.65, and then decreased gradually for 3 min (Fig. 4A). When  $NH_4Cl$  was washed out from the cells,  $pH_i$  rapidly decreased from 7.4 to 6.3 for 2 min, and then began to increase (0  $\mu M$  Phx-3). This recovery process is assumed to be due to proton extrusion by NHE1 (21,23).

When 20  $\mu M$  Phx-3 was administered to MKN45 cells, 2 min after washing-out  $NH_4Cl$ , the recovery of  $pH_i$  was extensively suppressed. Such inhibition of the recovery of  $pH_i$  after washing out  $NH_4Cl$  was complete when 50 or 100  $\mu M$  Phx-3 was administered to the cells (Fig. 4A). These results indicate that Phx-3 inhibited the activity of NHE1, which is involved in proton extrusion from the cells (21,23). Similar results were obtained for MKN74 cells treated with different concentrations of Phx-3 (Fig. 4B). When the cells were treated with 100  $\mu M$  Phx-3, the recovery of  $pH_i$  in MKN74 cells was completely suppressed, indicating that NHE1 activity was suppressed by Phx-3 in MKN74 cells.

**Effect of Phx-3 on apoptosis and viability in MKN45 and MKN74 cells.** Kasuga *et al* (18) previously demonstrated that

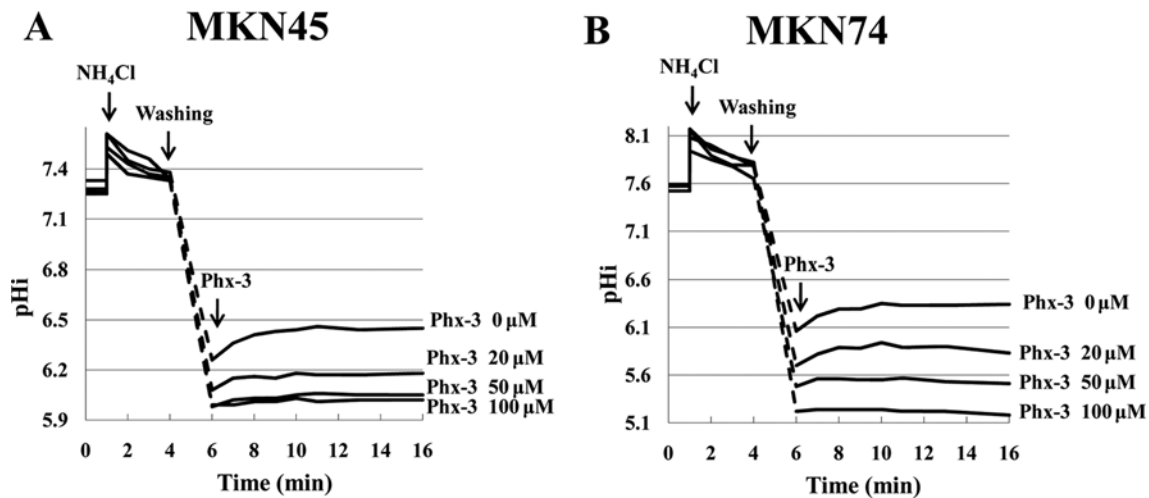


Figure 4. Changes in  $pH_i$  in MKN45 or MKN74 cells after  $NH_4Cl$  pulse loading, washing-out, and administering various concentrations of Phx-3 (0, 20, 50, and 100  $\mu M$  Phx-3). (A) MKN45 cells. (B) MKN74 cells.

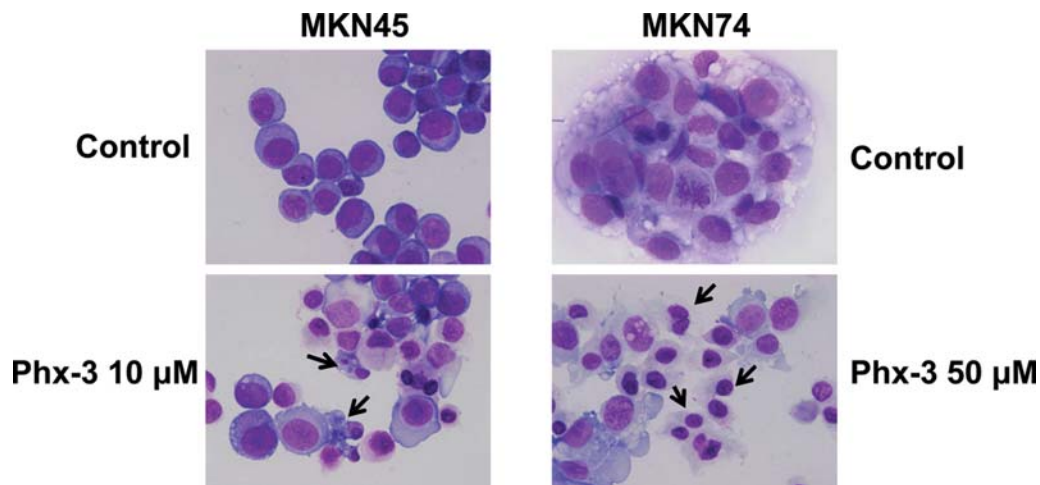


Figure 5. Morphological changes in MKN45 or MKN74 cells with or without Phx-3. MKN45 cells were treated with 10  $\mu M$  Phx-3, and MKN74 cells, with 50  $\mu M$  Phx-3. The control designates the cells without treatment of Phx-3. The arrows denote cells with typical apoptotic events. Original magnification,  $\times 800$ .

apoptosis occurs in MKN45 and MKN74 cells in response to Phx-3 engagement. Thus, we examined whether the morphological changes that are characteristic of apoptotic events may occur in these cells when treated with 10 or 50  $\mu M$  Phx-3 (Fig. 5). Results indicated that MKN45 treated with 10  $\mu M$  Phx-3 and MKN74 treated with 50  $\mu M$  Phx-3 for 24 h exhibited features of apoptosis, including cell shrinkage and condensation of nuclear chromatin.

Since Cell Titer Blue is reduced by mitochondria in viable cells, we evaluated the viability of MKN45 cells and MKN74 cells after treatment with various concentrations of Phx-3, using Cell Titer Blue. Fig. 6 illustrates changes in the viability of MKN45 and MKN74 cells during 72 h of incubation with different concentrations of Phx-3. Almost all the MKN45 cells lost viability 24 h after treatment with  $>20 \mu M$  Phx-3 (Fig. 6A). However, at lower concentrations of Phx-3 (0.5 to 5  $\mu M$ ), resulting in slight decrease of  $pH_i$  (Fig. 2A), the number of viable MKN45 cells significantly increased, though the

detailed mechanism is unclear. Relatively fewer cytotoxic effects of Phx-3 were observed in MKN74 cells than in MKN45 cells during 72 h, but loss of viability of MKN74 occurred at different concentrations (Fig. 6B). In particular, at 50  $\mu M$  Phx-3, 85% of the MKN74 cells lost viability at 72 h.

## Discussion

It has been recognized that  $pH_i$  is strictly regulated in normal cells and even in cancer cells, in order to maintain their cellular homeostasis (1,2,15-17). In normal cells,  $pH_i$  is usually kept lower than the pH of the extracellular medium (17,22). However,  $pH_i$  in cancer cells is maintained higher than extracellular pH ( $pH_e$ ) (3-5,17). Such higher  $pH_i$  in cancer cells seems to be suitable for proliferation, and oncogene transformation of the cells (24,25) and increases tumorigenesis (16). Rotin *et al* (26) indicated that  $Na^+/H^+$

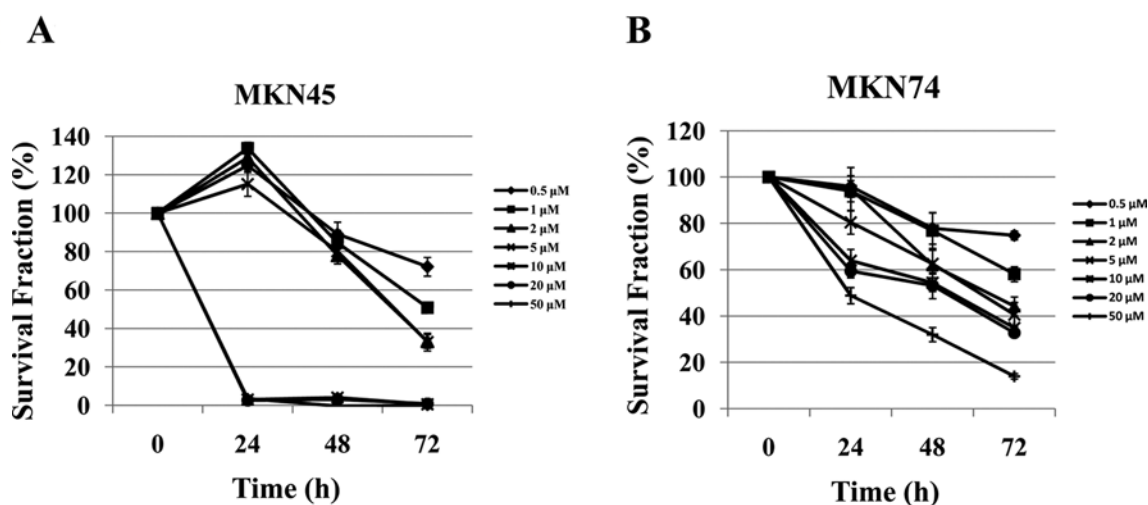


Figure 6. Effects of various concentrations of Phx-3 on the viability of MKN45 cells or MKN74 cells during 74 h. MKN45 or MKN74 cells were treated with various concentrations of Phx-3 (0.5, 1, 2, 5, 10, 20, and 50  $\mu$ M Phx-3), and the viability of these cells was evaluated by an Cell Titer Blue method, which estimates the capacity of mitochondria in the cells, to reduce Cell Titer Blue.

exchanger is indispensable for keeping  $pH_i$  higher in cancer cells. On the other hand, suppression of cellular proliferation and induction of apoptotic cell death have been known to be promoted by intracellular acidification (3-5). In particular, endonuclease II, which is responsible for digestion of genomic DNA, inducing programmed cell death, and is activated at acidic pH lower than pH 6.8 (27). This enzyme operates when cancer cells are treated with drugs to cause cellular acidification. Drugs that cause intracellular acidification may thus induce apoptosis of cancer cells (1,2,17), and are considered potential anticancer agents (3-5).

In this study, we found that  $pH_i$  in human gastric cancer cell lines MKN45 and MKN74 is higher than  $pH_e$  ( $pH_i$  of 7.48 for MKN45 and 7.5 for MKN74 cells, versus medium pH of 7.4) at a resting state. When these cells were treated with Phx-3,  $pH_i$  decreased rapidly, dependent on the dose of Phx-3 (Fig. 2), and such intracellular acidification was sustained for at least 4 h (Fig. 3). In particular,  $pH_i$  decreased by 1.6 pH units in MKN45 cells and 1.2 pH units in MKN74 cells immediately after administration of 100  $\mu$ M Phx-3 (Fig. 2). Such rapid and drastic changes in  $pH_i$  may be explained by the inhibition of NHE1 caused by administering Phx-3, as described later.

The decrease of  $pH_i$  in MKN45 and MKN74 cells was sustained for 4 h (Fig. 3) and may perturb intracellular homeo-stasis and metabolism in these cells, and finally cause apoptotic cell death during 72 h (Fig. 5). Kasuga *et al* (18) reported that MKN45 and MKN74 cells underwent apoptosis, accompanying cell growth inhibition and cell cycle arrest, when treated with Phx-3. Our morphological findings (Fig. 5) are in good agreement with the data of Kasuga *et al* (18) because the cellular shrinkage and DNA condensation characteristic of apoptotic cells were indicated in these cells treated with 10 or 50  $\mu$ M Phx-3 for 24 h. Additionally, these gastric cancer cells lost viability during 72 h of incubation with Phx-3 (Fig. 6). These results confirm that a decrease in  $pH_i$  preceded apoptotic events in MKN45 and MKN74 cells treated with Phx-3.

NHE1 is most likely to be involved in the mechanism for  $pH_i$  decrease in MKN45 and MKN74 because it has been demonstrated that among  $Na^+/H^+$  exchangers, NHE1 plays the most important role in regulating  $pH_i$  in various cells (15-17). NHE1 exists ubiquitously in the plasma membranes of normal and cancer cells (15,16), and its major role is to extrude intracellular proton to the extracellular medium, thereby regulating  $pH_i$ . Drugs administered to inhibit the activity of NHE1 (e.g., amiloride and methylamilorides) decrease  $pH_i$  by inhibiting the extrusion of intracellular proton (17,23). Thus, we studied the effects of Phx-3 on the activity of NHE1 in MKN45 and MKN74 cells (Fig. 4) and found that this phenoxazine compound exerted inhibitory effects on NHE1 in these cells, dependent on concentrations of Phx-3, consistent with the decrease of  $pH_i$  (Fig. 2). Thus, the decrease of  $pH_i$  in MKN45 and MKN74 cells caused by Phx-3 may be associated with inhibition of the activity of NHE1 by this phenoxazine compound. The mechanism for inhibiting NHE1 by Phx-3 is still unclear but may be clarified by the findings of Hendrich *et al* (28) that phenoxazine molecules are located close to the polar/apolar interface of lipid bilayers and weakly interact with lipid bilayers, altering the lipid phase properties of the cellular membranes. It is possible that rapid changes in the conditions of the plasma membranes induced by Phx-3 extensively affect the activity of NHE1 existing in the cellular membranes.

Gastric cancer is a common malignancy in adults but is often refractory to chemotherapy alone (29,30). Thus, it is crucial to develop drugs exerting anticancer effects but with low adverse reactions. In the present study, we demonstrated that Phx-3 causes apoptosis of human gastric cancer cells *in vitro*. Phx-3 has proapoptotic effects against various cancer cells *in vitro* (8-10) and anticancer effects *in vivo*, with few adverse effects on mice (11-13). Moreover, Kohno *et al* (31) reported that oral administration of 500-1,500 mg/kg Phx-3 to ddY mice did not cause gastrointestinal injury and that repeated oral administration of 10 mg/kg Phx-3 to mice for four weeks caused no diarrhea. Therefore, Phx-3 holds promise as an agent to treat human gastric cancer in the future.

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## References

- Gottlieb RA, Nordberg J, Skowronski E and Babior BM: Apoptosis induced in Jurkat cells by several agents is preceded by intracellular acidification. *Proc Natl Acad Sci USA* 93: 654-658, 1996.
- Yamagata M and Tannock IF: The chronic administration of drugs that inhibit the regulation of intracellular pH in vitro and anti-tumor effects. *Br J Cancer* 73: 1328-1334, 1996.
- Perez-Sala D, Collado-Escobar D and Mollinedo F: Intracellular alkalization suppresses lovastatin-induced apoptosis in HL-60 cells through the inactivation of a pH-dependent endonuclease. *J Biol Chem* 270: 6235-6242, 1995.
- Barry MA, Reynold JE and Eastman A: Etoposide-induced apoptosis in human HL-60 cells is associated with intracellular acidification. *Cancer Res* 53: 2349-2357, 1993.
- Goossens J-F, Henichrt J-P, Dassonneville L, Facompre M and Bailly C: Relation between intracellular acidification and camptothecin-induced apoptosis in leukemia cells. *Eur J Pharm Sci* 10: 125-131, 2000.
- Walgren RA: Systemic therapy for cancer. *Cancer: Principles and Practice of Oncology*. DeVita VT, Lawrence TS and Rosenberg SA (eds). 8th edition, Lippincott Williams & Wilkin, PA, pp114-127, 2008.
- Hsiang Y-H, Liu LF, Wall ME, Wani MC, Nicholas AW, Manikumar G, Kirschenbaum S, Silber R and Potmesil M: DNA topoisomerase I-mediated DNA cleavage and cytotoxicity of camptothecin analogues. *Cancer Res* 49: 4385-4389, 1989.
- Kato S, Shirato K, Imaizumi K, Toyota H, Mizuguchi J, Odawara M, Che X-F, Akiyama S, Abe A and Tomoda A: Anticancer effects of phenoxazine derivatives combined with tumor necrosis factor-related apoptosis-inducing ligand on pancreatic cancer cell lines, KLM-1 and MIA-PaCa-2. *Oncol Rep* 15: 843-848, 2006.
- Takasaka A, Hanyu H, Iwamoto T, Shirato K, Izumi R, Toyota H, Mizuguchi J, Miyazawa K and Tomoda A: Mitochondrial depolarization and apoptosis associated with sustained activation of c-jun-N-terminal kinase in the human multiple myeloma cell line U266 induced by 2-aminophenoxazine-3-one. *Mol Med Rep* 2: 199-203, 2009.
- Zheng C-L, Che X-F, Akiyama S, Miyazawa K and Tomoda A: 2-Aminophenoxazine-3-one induces cellular apoptosis by causing rapid intracellular acidification and generating reactive oxygen species in human lung adenocarcinoma cells. *Int J Oncol* 36: 641-650, 2010.
- Mori H, Honda K, Ishida R, Nohira T and Tomoda A: Antitumor activity of 2-amino-4,4a-dihydro-4a,7-dimethyl-3H-phenoxazine-3-one against Meth A tumor transplanted into BALB/c mice. *Anticancer Drugs* 11: 653-657, 2000.
- Shimamoto T, Tomoda A, Ishida R and Ohyashiki K: Antitumor effects of a novel phenoxazine derivative on human leukemia cell lines in vitro and in vivo. *Clin Cancer Res* 7: 704-708, 2001.
- Miyano-Kurosaki N, Kurosaki K, Hayashi M, Takaku H, Hayafune M, Shirato K, Kasuga T, Endo T and Tomoda A: 2-Aminophenoxazine-3-one suppresses the growth of mouse malignant melanoma B16 cells transplanted into C57BL/6Cr Slc mice. *Biol Pharm Bull* 29: 2197-2201, 2006.
- Che X-F, Akiyama S and Tomoda A: Suppression of the proliferation of cancer cell lines, KB-3-1 and K562 cells preceded by a decrease in intracellular pH caused by phenoxazine derivatives. *Oncol Rep* 19: 1253-1258, 2008.
- Tse C-M, Levine SA, Cris Yun CH, Brant SR, Nath S, Pouyssegur J and Donowitz M: Molecular properties, kinetics and regulation of mammalian Na<sup>+</sup>/H<sup>+</sup> exchangers. *Cell Physiol Biochem* 4: 282-300, 1994.
- Putney LK, Denker SP and Barber DL: The changing face of the Na<sup>+</sup>/H<sup>+</sup> exchanger, NHE1: structure, regulation, and cellular actions. *Ann Rev Pharmacol Toxicol* 42: 527-552, 2002.
- Rich IN, Worthington-White D, Garden OA and Musk P: Apoptosis of leukemic cells accompanies reduction in intracellular pH after targeted inhibition of the Na<sup>+</sup>/H<sup>+</sup> exchanger. *Blood* 95: 1427-1434, 2000.
- Kasuga T, Tabuchi T, Shirato K, Imaizumi K and Tomoda A: Caspase-independent cell death revealed in human gastric cancer cell lines, MKN45 and KATO III treated with phenoxazine derivatives. *Oncol Rep* 17: 409-415, 2007.
- Nakachi T, Tabuchi T, Takasaki A, Arai S, Miyazawa K and Tomoda A: Anticancer activity of phenoxazines produced by bovine erythrocytes on colon cancer cells. *Oncol Rep* 23: 1517-1522, 2010.
- Litman T, Pedersen SF, Karamhoft B, Skovsgaard T and Hoffmann EK: pH regulation in sensitive multidrug resistant Ehrlich ascites tumor cells. *Cell Physiol Biochem* 8: 37-50, 1998.
- Coates G, Nissim I, Battarbee H and Welbourne T: Glitazones regulate glutamine metabolism by inducing a cellular acidosis in MDCK cells. *Am J Physiol* 282: E729-E737, 2002.
- Tomoda A, Tsuda-Hirota S and Minakami S: Glycolysis of red cells suspended in solutions of impermeable solutes. *J Biochem* 81: 697-701, 1977.
- Owen CS, Pooler PM, Wahl ML, Coss RA and Leeper DB: Altered proton extrusion in cells adapted to growth at low extracellular pH. *J Cell Physiol* 173: 397-405, 1997.
- Lagadic-Gossmann D, Huc L and Lecœur V: Alteration of intracellular pH homeostasis in apoptosis: origins and roles. *Cell Death Differ* 11: 953-961, 2004.
- Shrode LD, Tapper H and Grinstein S: Role of intracellular pH in proliferation, transformation and apoptosis. *J Bioenerg Biomembr* 29: 293-299, 1997.
- Rotin D, Steele-Norwood D, Grinstein S and Tannock I: Requirement of the Na<sup>+</sup>/H<sup>+</sup> exchanger for tumor growth. *Cancer Res* 49: 205-211, 1989.
- Barry MA and Eastman A: Endonuclease activation during apoptosis: The role of cytosolic Ca<sup>2+</sup> and pH. *Biochem Biophys Res Commun* 186: 782-789, 1992.
- Hendrich AB, Stanczak K, Komorowska M, Motohashi N, Kawase M and Kichalak K: A study on the perturbation of model lipid membranes by phenoxazines. *Bioorg Med Chem* 14: 5948-5954, 2006.
- Crew KN and Neugut AI: Epidemiology of gastric cancer. *World J Gastroenterol* 12: 354-362, 2006.
- Smith MG, Hold GL, Tahara E and El-Omar EM: Cellular and molecular aspects of gastric cancer. *World J Gastroenterol* 12: 2979-2990, 2006.
- Kohno K, Miyake M, Sano O, Tanaka-Kataoka M, Yamamoto S, Koya-Miyata S, Arai N, Fujii M, Watanabe H, Ushio S, Iwaki K and Fukuda S: Anti-inflammatory and immunomodulatory properties of 2-amino-3-phenoxazin-3-one. *Biol Pharm Bull* 31: 1938-1943, 2008.