

Vasodilator-stimulated phosphoprotein regulates osteosarcoma cell migration

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Abstract. Vasodilator-stimulated phosphoprotein (VASP) has been reported to play an important role in the process of cell migration and tumor metastasis. However, to date, no study has examined VASP expression and its function in osteosarcoma cells. In this study, we analyzed the effect of VASP on osteosarcoma cell migration and the signal transduction pathways involved. We used two osteosarcoma cell strains (Mg-63 and Saos-2 cells) with different metastatic potential. Silencing of VASP gene expression was carried out using RNA interference in these cells. Knockdown of expression at the transcriptional or translational level was determined by RT-PCR or Western blot analysis, respectively. The metastatic potential of the tumor cells was determined by a wound healing migration assay. VASP mRNA expression was also determined in 30 human osteosarcoma samples. Furthermore, Rac1 was determined as a regulator of VASP function. RT-PCR and Western blotting showed that Mg-63 cells had a significantly higher VASP expression at both the transcriptional and translational levels compared to Saos-2 cells. The wound healing assay revealed that Mg-63 cells had more migratory potential compared to Saos-2 cells. The effect was found to be reversible when VASP was knocked down by siRNA in Mg-63 cells. Specimens from human patients with metastases had higher VASP expression compared to specimens of patients without metastases. Knockdown of Rac1 resulted in inhibition of VASP expression in sarcoma cells. These results suggest that VASP

protein regulates osteosarcoma cell migration and metastasis. Rac1 and VASP interaction may be a potential target for osteosarcoma treatment.

Introduction

Osteosarcoma is the most prevalent primary malignant bone tumor in children and young adults. Although the life expectancy of osteosarcoma patients has been prolonged due to improvement of neoadjuvant chemotherapy and surgical resection during the past three decades, the long-term survival and quality of life is still unsatisfactory. Poor outcome of osteosarcoma is mainly caused by metastatic disease, and previous studies have demonstrated that more than 30% of patients already display distant metastasis without clinical detectable signs at the time of diagnosis (1,2).

Vasodilator-stimulated phosphoprotein (VASP), a member of the Ena/VASP family, has been implicated in regulating key cellular functions, such as shape change, adhesion and migration, which is mainly due to its ability to modify dynamic action cytoskeleton (3). In addition to its role in normal cell growth (4,5), embryonic development (6) and homeostasis (7), VASP plays an essential role in many diseases, such as cancer metastasis (8,9), thrombosis (10), arteriosclerosis (11) and nephritis (12).

Expression of VASP has been reported to be dysregulated in several types of malignancies and is relevant to tumor metastasis. For example, its level was found to be significantly increased in lung adenocarcinomas and gastric carcinomas compared to normal tissue, and this increase was correlated with the metastatic ability of these tumors (8,9). Depletion of VASP in breast cancer MDA-MB-231 cells resulted in decreased cell migration (13) and, conversely, enhanced VASP in breast cancer MDA-231 cells led to increased cell mobility (14). These findings suggest that VASP acts as a positive regulator of cell motility and tumor metastasis. However, there is a debate concerning its role in cell migration. When VASP was overexpressed in fibroblasts, cell mobility was decreased in a dose-dependent manner, while depletion of VASP enhanced cell movement (15,16). Thus, how VASP affects cell mobility

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and the related underlying mechanism of its action are not fully known.

Rac1, a member of the Rho family of small GTPase, is also implicated in the processes of cell migration, cytoskeleton reorganization and formation of lamellipodia. Activation of Rac has been demonstrated to play an important role in processes induced by growth factor (17) and integrin (18). During cell migration, Rac1 translocates to the plasma membrane (19), interacts with WAVE via an adaptor protein, the insulin receptor substrate Irs53, and then stimulates Arp2/3-mediated actin polymerization (20).

Since previous studies suggest that VASP plays an essential role in the process of cell migration and tumor metastasis, we reasoned that it is valuable to explore the role of VASP in osteosarcoma since no studies to date have been carried out in both osteosarcoma cells *in vitro* or human tissues *in vivo*. Here, we analyzed VASP expression in different osteosarcoma cell lines and human clinical samples. Furthermore, we demonstrated that the silencing of VASP affects osteosarcoma cell migration and tumor metastasis, and this action involves the small GTPase Rac1 signal transduction pathway.

Materials and methods

Cell culture and transfection. In the present study we used human osteosarcoma Mg-63 cells (characterized by a high metastatic potential *in vivo*) and Saos-2 cells (characterized by a low metastatic potential) (21,22). These two cell lines were purchased from ATCC (LGC Promochem, Wesel, Germany), and were cultured in DMEM medium (Sigma, USA) supplemented with 10% FBS (Amresco, USA) and 1% penicillin/streptomycin.

For transient transfection, the Rac1-directed siRNA vector was transfected using Lipofectamine 2000 (Invitrogen, USA) as specified in the manufacturer's instructions. Rac1 was silenced using specific siRNA (synthesized by Bibobio Co., China) which are as follows: sense 5'-UGAAGAAGAGGAAGAGAAAdTdT-3' and anti-sense 3'-dTdTACUUCUUCUCCUUCUCUUU-5'. For establishing stable clones of the VASP-siRNA plasmid and empty vector, VASP-siRNA and vector were transfected using Lipofectamine 2000. Stable clones were selected using blasticidin at 5 µg/ml for 3 weeks.

Tissue specimens. We recruited 44 human patients diagnosed with osteosarcoma. The study was carried out according to the Chinese National Ethical Guidelines ('Code for Proper Secondary Use of Human Tissue', Chinese Federation of Medical Scientific Societies). A written informed consent was obtained from each subject prior to enrollment in the study. The detailed clinical data were described in our previous study (23). Tissues were stored at -80°C until further use. Specimens were dissected into samples 0.5 mm in diameter for RNA isolation for the RT-PCR analysis.

Wound healing assay. A wound healing assay was used to determine the migratory potential of the cells. For this study, 5x10⁵ cells were plated into each well of a 6-well plate overnight. On the following day, cells were serum-starved and parallel wounds were made using a 1000-µl pipette tip. Images were collected at various times for the same spots. The cell

migration distance was evaluated by measuring the width of the wound divided by two and subtracting this value from the initial half-width of the wound. Average velocity was calculated based on different observation times.

Isolation of RNA and RT-PCR. Total RNA was extracted from cells or tumor specimens using the TRIzol reagent (Invitrogen, USA). First-strand complementary DNA (cDNA) was synthesized from total RNA in a final volume of 20 µl using Superscript II (Invitrogen, USA). cDNA was amplified by PCR using specific primers corresponding to mRNA encoding human gene products. Annealing temperatures for VASP and Rac1 were 65 and 56°C. Amplification products were resolved by electrophoresis on a 1.5% w/v agarose gel. Primers used are listed as follows: VASP, sense 5'-GCGCCTGGTACCATGACGCAAGTTGGGGAGAAAACC-3' and antisense 5'-GCGCCTGGATCCCAGGGAGAACCCCGCTTCC-3'; Rac1, sense 5'-AGACGGAGCTGTAGGTAATAA-3' and antisense 5'-GCAGGACTCACAAAGGGA-3'.

Protein extraction and Western blot analysis. For determining the translation expression of VASP or Rac1, cells were lysed in lysis buffer [50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 1% Triton X-100, 0.25% deoxycholate, 1.5 mM MgCl₂, 1 mM EGTA, 1 mM phenylmethylsulfonyl fluoride (PMSF), 10 mM NaF, 10 mM pervanadate, 10 µg/ml leupeptin and 10 µg/ml aprotinin], and the protein concentration in the cell lysates was measured using the BCA protein assay kit (Sangon Biotech, China). Equal amounts of protein were separated on 12% (v/v) SDS-PAGE and transferred to a nitrocellulose membrane. After blocking with 5% milk, the membrane was incubated with polyclonal mouse antibody against VASP (1:2000; Immuno Globe, Germany) or Rac1 (1:500; Cytoskeleton). After three washings, the membrane was incubated with horseradish peroxidase (HRP)-conjugated goat anti-mouse IgG (1:300; Santa Cruz Biotechnology) for 1 h at room temperature. The ECL immune enzymic reaction kit (Pierce Biotechnology) was used for immunodetection. The band intensity was analyzed using Quantity One software (Bio-Rad).

Results

Increased expression of VASP correlates with the migratory potential of cells. Cell migratory ability is considered to be well correlated with the metastatic potential of the cells and is one of the first steps in the process of tumor metastasis. Mg-63 cells had a much higher metastasis potential when compared to the Saos-2 cells as determined by the wound healing assay. Mg-63 cells showed a 3.8-fold higher migratory potential compared to the Saos-2 cells, which was determined to be statistically significant (p<0.01) (Fig. 1).

We observed a significantly increased transcriptional level of VASP in the Mg-63 cells as compared to the Saos-2 cells (p<0.01) (Fig. 2A). Similarly, augmented VASP expression at the protein level was also observed in Mg-63 cells when compared to the Saos-2 cells using Western blot analysis (Fig. 2B and C).

Knockdown of VASP results in the down-regulation of migratory potential in osteosarcoma cells. In the literature,

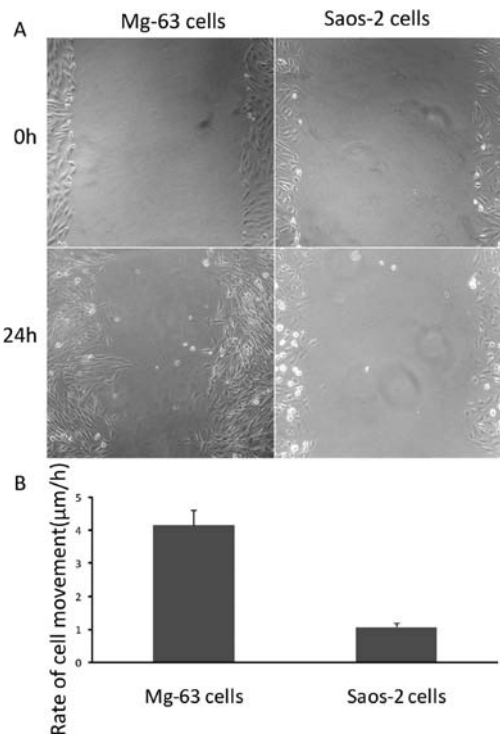


Figure 1. Rate of Mg-63 cell movement is higher than that of the Saos-2 cells. (A) After serum-starvation for 12 h, a wound was made using a 1000- μ l pipette tip. Images were captured 24 h after scratching; (B) Mg-63 cells moved faster than the Saos-2 cells. The average velocity was calculated using the migration distance divided by the observation time (24 h). The difference between the velocities of these two cells was significant ($p < 0.01$). Results are the mean \pm SD of six measurements at the same time.

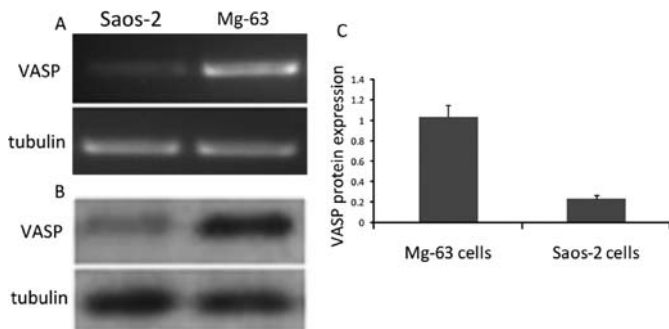


Figure 2. VASP expression in Mg-63 cells is higher than that in Saos-2 cells. (A) RT-PCR indicated that VASP mRNA was higher in the Mg-63 cells than that in the Saos-2 cells. Tubulin was used as a loading control. (B) VASP expression was confirmed by Western blot analysis. Tubulin was used as a loading control. (C) VASP protein in the Mg-63 cells was significantly higher than that in the Saos-2 cells ($p < 0.01$). Shown are the mean values \pm SD of results from 4 independent experiments.

a positive correlation between cell migration/movement and expression of VASP has been documented (4,9,10,13-15). Our results are in line with these observations as we also observed a higher competency of cell migration and VASP expression in Mg-63. To gain more insight, we silenced the VASP using specific siRNA in the Mg-63 cells. Silencing of VASP in the Mg-63 cells was confirmed by RT-PCR and Western blot analysis (Fig. 3).

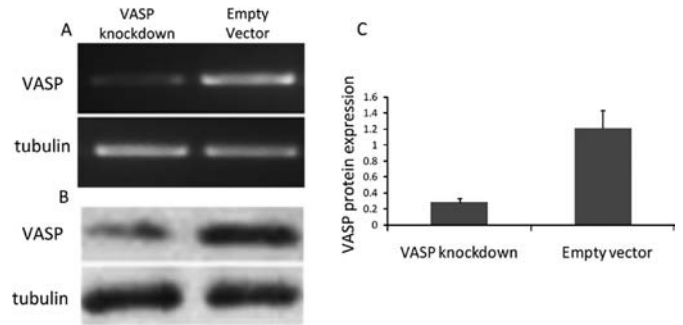


Figure 3. Characterization of siRNA-mediated VASP knockdown. (A) Two pools of stable clones of Mg-63 cells were established, VASP mRNA level was determined by RT-PCR. Tubulin was used as a loading control. (B) VASP protein expression was confirmed by Western blot analysis. Tubulin was used as a loading control. (C) Approximately 77% inhibition of the VASP protein was achieved by the VASP-siRNA plasmid compared to the empty vector ($p < 0.01$). Bars indicate the mean \pm SD of triplicate experiments.

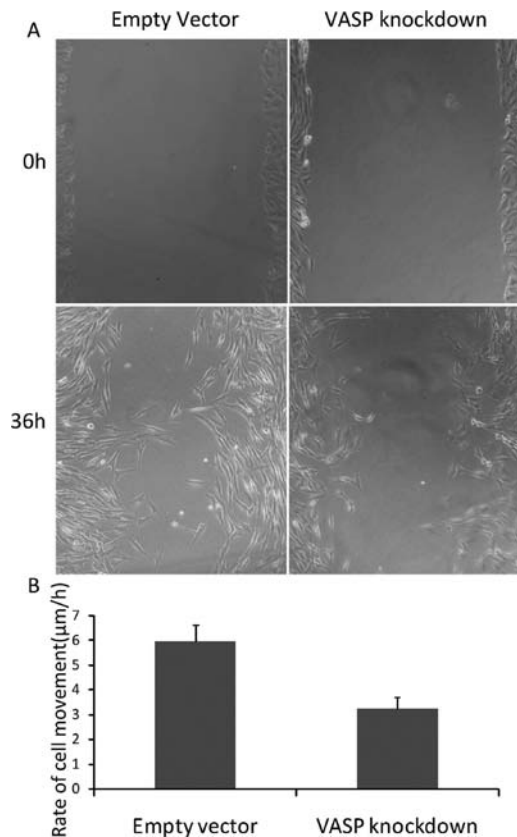


Figure 4. Depletion of VASP inhibits cell migration. (A) Two pools of stable clones were established using the VASP-siRNA plasmid and empty vector. Cell mobility was determined using a wound healing assay. Images were captured 36 h after scratching. (B) VASP knockdown resulted in a decreased mobility compared to the empty vector group ($p < 0.01$). Results are the mean \pm SD of six measurements at the same time.

After silencing of VASP in the Mg-63 cells, the effect on the migratory potential of these cells was determined using a wound healing assay at 36 h. As depicted in Fig. 4, VASP-silenced Mg-63 cells lost their migratory capability by 55% when compared to the cells transfected with the empty vector alone. In contrast, the Saos-2 cells which had lower VASP expression than Mg-63 showed reduced cell migration. Taken

Table I. Clinical features of the osteosarcoma cases.

Features	Total no.	Percentage
Age (years)		
<12	1	3.33
13-20	11	36.67
21-30	8	26.67
31-40	8	26.67
>41	2	6.67
Gender		
Male	22	73.33
Female	8	26.67
Location of the primary tumor		
Distal femur	9	30.00
Proximal tibia	7	23.33
Humerus	3	10.00
Tibia diaphysis	2	6.67
Femur diaphysis	7	23.33
Other	2	6.67
Histological type		
Osteoblastic	18	60.00
Small cell	1	3.33
Chondroblastic	4	13.33
Round cell	2	6.67
Fibroblastic	4	13.33
Mixed	1	3.33
Metastasis		
No	13	43.33
Lung	15	50.00
Other	2	6.67

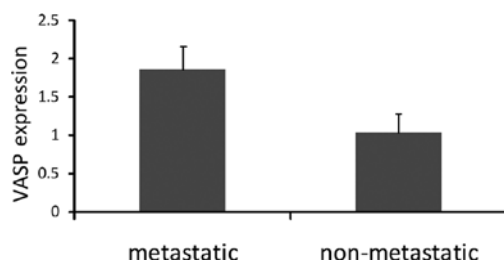


Figure 5. VASP mRNA is expressed higher in patients with metastasis than in patients without metastasis ($p < 0.01$). Seventeen patients with distant metastasis and 13 without metastasis were analyzed by RT-PCR. Results are the mean \pm SD of samples.

together, these results suggest that VASP is at least partly responsible for regulating the motility of osteosarcoma cells.

Transcriptional (mRNA) expression of VASP in human tissue samples. We isolated RNA samples from 30 human osteosarcoma patients. Table I depicts the anthropometric and clinical features of the subjects recruited with osteosarcoma in the present study. These 30 subjects were divided into two groups

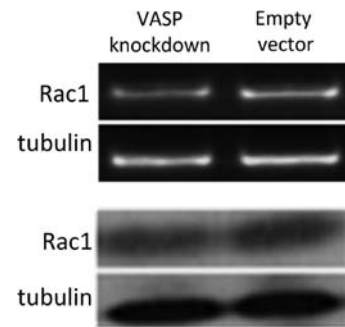


Figure 6. VASP depletion results in no detectable change in Rac1 expression. Rac1 expression was tested in the two established stable pools of Mg-63 cells. RT-PCR and Western blot analysis showed that depletion of VASP did not affect Rac1 expression. Tubulin was used as a loading control.

on the basis of the presence (Group I; $n=17$) and absence (Group II; $n=13$) of metastasis.

We determined the mRNA expression of VASP in these tissue samples using semi-quantitative RT-PCR. A significantly increased mRNA expression was observed in Group I subjects compared to the subjects in Group II ($p < 0.01$) (Fig. 5). Increased expression of VASP in subjects with metastasis suggests that VASP may be involved in regulating osteosarcoma metastasis *in vivo*.

Involvement of the Rac1 pathway. To determine whether there is any involvement of activation of the Rac1 signal transduction pathway in osteosarcoma cells, we determined the expression of Rac1 in VASP-silenced Mg-63 cells. We observed no significant difference in the expression of Rac1 both at the mRNA and protein level in the Mg-63 and VASP-silenced Mg-63 cells (Fig. 6).

We also examined expression of VASP in the Rac1-silenced Mg-63 cells. Silencing of Rac1 was carried out using Rac1-directed siRNA and a negative siRNA control (as described in Materials and methods). Rac1-siRNA oligonucleotides demonstrated an efficient inhibitory effect on Rac1 expression both at the mRNA and protein level compared to the negative control (Fig. 7A and B). A significant decreased expression of VASP was found in the Rac1-silenced cell lines when compared to the controls (Fig. 7). In addition, as depicted in Fig. 8, a significant inhibition in migration was observed in the Rac1-silenced Mg-63 cells compared to the control cells. Our results demonstrated that VASP moderated osteosarcoma cell migration via the Rac1 pathway and VASP may act as a downstream effector of Rac1.

Discussion

Cancer metastasis is a complicated process involving the invasion of the host matrix, intravasation and extravasation of blood or lymphatic vessels, and dissemination and proliferation in distant organs (24). Metastatic potential of a tumor is primarily dependent on the migratory ability of the cells (25,26).

In this study, we demonstrated that VASP expression is associated with the metastatic potential of cells using two different cellular models of osteosarcoma. Mg-63 cells, which

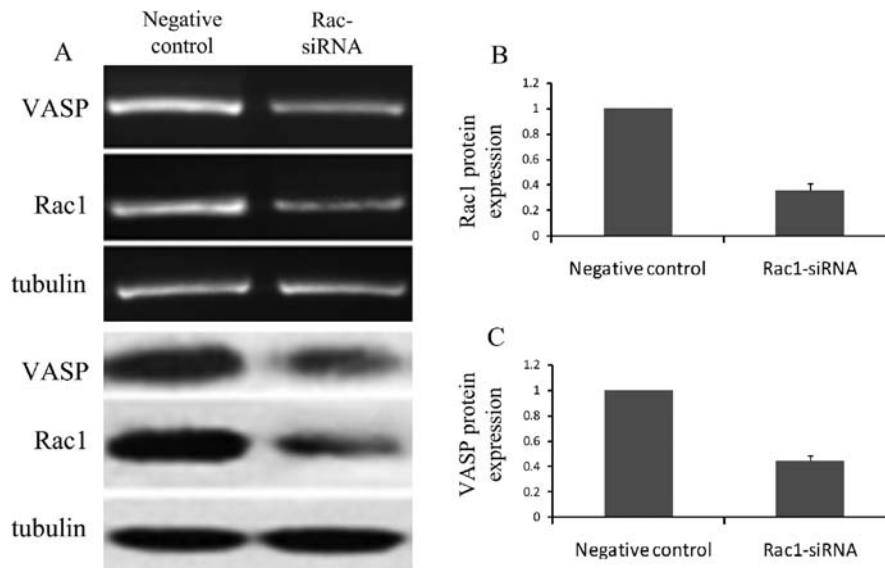


Figure 7. Depletion of Rac1 inhibits VASP expression. (A and B) Characterization of siRNA oligos against Rac1. Mg-63 cells were transiently transfected using Rac1-directed siRNA oligos and negative control oligos. Forty-eight hours after transfection, total mRNA or protein was isolated for analyzing Rac1 expression using RT-PCR or Western blot analysis. Expression level of Rac1 in Rac1-depleted cells was normalized to that in cells treated with negative siRNA control. Rac1-siRNA showed the effective inhibition of Rac1 in the Mg-63 cells, with a reduced Rac1 level (36% to the control). Tubulin was used as a loading control. (A and C) Expression of VASP gradually decreased with depletion of Rac1. After cells were transfected with Rac1-siRNA, mRNA or protein was isolated for VASP expression. Rac1-siRNA resulted in a significant decrease in VASP.

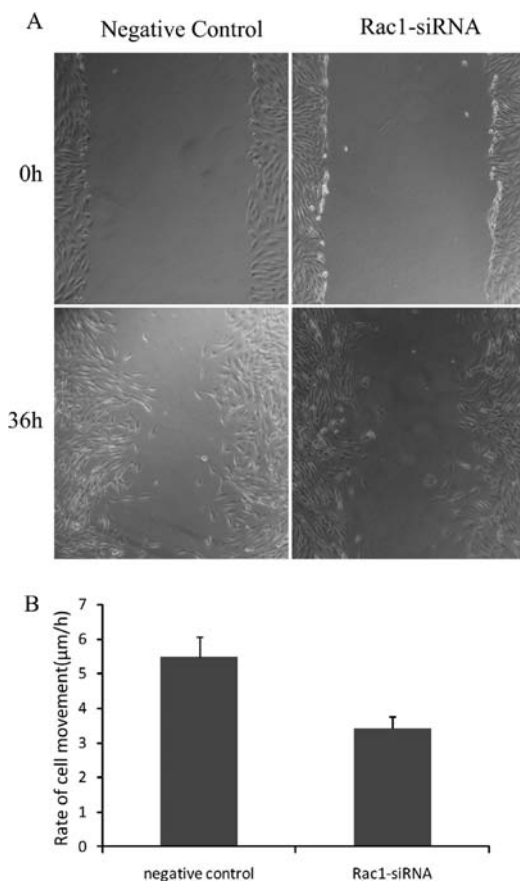


Figure 8. Depletion of Rac1 inhibits migration. (A) Mg-63 cells were transiently transfected using Rac1-siRNA and negative siRNA, respectively. Twenty-four hours after transfection, cells were detached and plated into a 6-well plate at a concentration of 5×10^5 cells/well. Twelve hours later, parallel wounds were made. Images were captured right after scratching or 36 h after scratching. (B) Rac1 knockdown resulted in a decreased mobility compared to the negative siRNA group ($p < 0.01$). Results are the mean \pm SD of six measurements.

expressed higher levels of VASP, moved much faster than Saos-2 cells. Our observations are in accordance with various reports presented in the literature for other types of cancer cells (8,9,13,14). Furthermore, we silenced VASP using specific siRNA in Mg-63 cells. We observed that silenced cells had a decreased migratory potential when compared to the control. Taken together, our data indicate that VASP may be a positive regulator of osteosarcoma cell migration.

We also determined VASP mRNA expression in tumors obtained from human osteosarcomas. We observed that all of the human osteosarcoma samples had VASP expression. However, a significant higher expression was observed in patients with metastasis. To our knowledge, this is the first report to demonstrated that the mRNA expression of VASP is correlated with the progression of osteosarcoma in patients. Thus, VASP may be a promising biomarker in determining metastasis of osteosarcoma.

In the process of cancer metastasis, VASP has been documented to act as an actin anti-capping protein (27,28). This allows actin filament elongation and clustering at the leading edge toward the membrane, which can regulate formation of membrane protrusions such as lamellipodia and filopodia, consequently resulting in modified cell motility and tumor metastasis (3). Rottner *et al* demonstrated that, in a B16 melanoma cell line, VASP was localized in a line running along the tips of the protruding membrane and its expression was proportional to the protrusion rate (29). Depletion of VASP proteins in growth cones of neurons resulted in a reduction in long filaments, and an increase in short action filament has been demonstrated in another study (30). In contrast, Bear *et al* demonstrated that enrichment of VASP in fibroblasts promotes elongation of actin filament (16). In addition, VASP was able to counteract capping protein (CP), delaying termination of actin filament

elongation. In addition to their anti-capping ability, several other mechanisms exist by which VASP proteins regulate actin-based membrane protrusion, including depression of actin-filament branching (31) and recruitment of profilin (32).

As discussed above, the role of VASP proteins in facilitating actin-filament and membrane protrusions has been well-established. However, the global cell motility affected by VASP is not fully determined and seems to be cell-type dependent. In the present study, Mg-63 cells with higher expression of VASP had higher migratory potential as compared to Saos-2 cells. Inhibition of cell migration after silencing VASP in Mg-63 cells clearly suggests that VASP plays a positive role in regulating osteosarcoma cell motility. Our observations are in line with other studies carried on *Listeria* (33,34), macrophages (35), and several types of cancer cells (8,9,14,36,37). However, there have been reports of an opposite effect of VASP. For example, studies on neurocytes (38), platelets (7) and fibroblasts (15) found that VASP negatively regulated cell migration. Since cell movement is the result of the co-ordination of four major processes which include extension of membrane protrusion, stabilization of protrusion by attachment via focal adhesion, translocation of the cell body and detachment of the rear region (39), this contradictory phenomenon implies that cell movement is a complicated process.

Rac1 is a member of the Rho family of small GTPase, which has been implicated in actin cytoskeleton organization in diverse cellular functions including cell motility. Rac1 is widely expressed in many tissues, while Rac2 and Rac3 were restrictively expressed in the hematopoietic and nervous systems, respectively (40). Rac1 was found to be required in growth factor- (17) and integrin-mediated (18) cytoskeleton remodeling and migration. Other studies in the literature also showed that Rac1 plays a crucial role in tumor cell migration (9,13,36). Given the importance of both Rac1 and VASP in the actin-based process, we tried to determine whether there is a relationship between these two proteins.

The molecular mechanisms that lead to a harmonious interplay between VASP proteins and Rac proteins are poorly understood. Therefore, we silenced Rac1 in Mg-63 cells and observed a significant decrease in the expression of VASP both at the transcription and translational levels. In contrast, when VASP was silenced in Mg-63 cells, we did not observe any change in the expression of Rac1 either at the transcriptional or translational levels. These results clearly suggest that VASP may act as a downstream effector of Rac1.

One of the possible factors that may contribute to the cytoskeletal effect of VASP is serine/threonine p21-activated kinase (PAK). PAK was generally identified as a direct substrate of Rac. PAK comprises five proline-rich regions, while VASP contains an N-terminal Ena/VASP homology 1 (EVH1) domain which can bind to a specific proline-rich motif (41). After serine 21 phosphorylation of PAK by Rac, the association of VASP/PAK was found to be enhanced while Nck/PAK was inhibited (42). Consistent with our result here, in breast cancer cells, inhibition of Rac1 also decreased the expression of VASP, and cGMP-dependent protein kinase (PKG) was suspected to play a crucial role to bridge the relationship between Rac and VASP (13,36,37). In a novel Rac/PAK/GC/cGMP pathway, Rac, using its downstream effector PAK, was found to activate transmembrane guanylyl cyclases

(GCs), resulting in elevated cGMP. Then as a major target of cGMP, activated PKG regulated VASP via the cGMP/PKG/VASP pathway (43). Therefore, therapeutic tools, via targeting Rac1 and VASP interaction (13,36,37), represent a promising drug candidate for cancer treatment.

In conclusion, our study demonstrates that VASP regulates cell migration and metastasis in osteosarcoma. Furthermore, VASP is a downstream effector of Rac1 and thus may serve as a potential drug target for osteosarcoma treatment.

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