

# Synergistic antitumor effect of JAWSII dendritic cells and interleukin 12 in a melanoma mouse model

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**Abstract.** One of the possible ways to augment dendritic cell (DC) efficacy in presentation of tumor antigens to effector T cells is pulsing them with tumor-cell lysates and incubation with certain immunostimulators. We present the results of an immunotherapeutic approach in a murine B78-H1 model using as a vaccine JAWSII DCs in combination with IL-12. Prior to the *in vivo* experiments, phenotypic characterization of JAWSII cells was performed and optimal conditions for stimulation of these cells were established. As no production of IL-12 by JAWSII cells was found, injections of this cytokine were introduced to vaccination protocols. Three vaccination schedules have been tested: i) prophylactic, ii) therapeutic-intratumoral, and iii) therapeutic-systemic. In all the protocols, vaccination with pulsed + stimulated JAWSII cells in combination with IL-12 was superior to the treatment with either agent alone and led to eradication of the tumor in several cases. The results of the study may be helpful in planning optimal DC-based therapeutic protocols in cancer patients.

## Introduction

Dendritic cells (DCs) are the most potent antigen-presenting cells, critical for the induction of specific immune responses (1,2). Despite some differences between the murine and human immune system, DC-based cellular vaccines in mouse tumor models may provide significant information on how to optimize therapeutic antitumor approaches in humans (3). DCs have been proved effective as both prophylactic and therapeutic vaccines in a number of experimental models (4). One of the possible ways to augment DC efficacy in presentation of tumor antigens to effector cells is pulsing them with tumor cell lysates (5). However, DCs pulsed with tumor antigens are not usually able to reach final maturity, which affects their properties to induce antitumor response. Prior to *in vivo* application, incubation of DCs with certain

immunostimulators is therefore necessary. The change in their phenotype greatly enhances their antigen-presenting properties (6,7). It appears that DCs stimulated in such a manner are able to further differentiate to respond to factors secreted by the host immune system.

As the immune response of the host may be suppressed by the tumor, injections of adjuvants/cytokines (such as IL-12) in combination with cellular vaccines may have a crucial effect on overcoming tumor evasion mechanisms (8,9). Systemic injections of IL-12 lead to enhancement of natural killer (NK) and cytotoxic T-cell activity and result in the increase of IFN- $\gamma$  production, as well as in affecting differentiation of antigen-specific Th lymphocytes (10,11). IL-12 was found to effectively inhibit the growth of several experimental tumors in mice, via IFN- $\gamma$  and T cell-dependent pathways, as well as due to anti-angiogenic effects (12,13).

JAWSII cells, immortalized C57BL/6 murine bone marrow-derived DCs, seem to be a promising candidate for use in cancer therapy in a cellular vaccine approach (14,15). In a previous study, we analyzed phenotype and functional properties of JAWSII cells and optimized conditions of their stimulation (16). The purpose of this study was to determine the *in vivo* applicability of JAWSII cells as a component of DC-based cellular vaccine used in combination with IL-12 in a murine B78-H1 melanoma model. Three antitumor approaches of this combination have been tested: prophylactic, local (intratumoral), and systemic therapy.

## Materials and methods

**Cells.** JAWSII cells were purchased from the American Type Culture Collection (CRL-11904). The cells were grown in RPMI-1640 medium (Sigma-Aldrich, St. Louis, MO, USA) supplemented with 10% non-inactivated FCS (Gibco-Invitrogen, Paisley, Scotland, UK), antibiotics (penicillin + streptomycin + amphotericin, Sigma-Aldrich), and 5 ng/ml murine GM-CSF (PeproTech, London, UK). JAWSII cells were maintained in a humidified atmosphere at 37°C and 5% CO<sub>2</sub> and passaged twice a week.

B78-H1 cells, an amelanotic clone of the murine melanoma B16 cell line, were primarily provided by Dr L.H. Graf (Chicago, IL, USA) and were found to be suitable in our previous immunotherapeutic models (17,18). The cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Sigma-Aldrich) supplemented with 10% inactivated FCS and

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antibiotics. Other culturing conditions were similar to those used for JAWSII cells.

For *in vivo* experiments, B78-H1 cells were trypsinized, washed twice in ice-cold PBS and resuspended in PBS at the concentration of  $2 \times 10^5/20 \mu\text{l}$  per mice (or in some experiments  $5 \times 10^5/20 \mu\text{l}$  per mice). Cell viability was determined prior to the inoculation using trypan blue exclusion test and estimated to be >90-95%.

***In vitro* JAWSII stimulatory conditions.** In the present study, we used a protocol of stimulation of JAWSII cells that was previously described (16). In brief, JAWSII cells were pulsed with tumor cell lysates (JAWSII cells to B78-H1 melanoma cells ratio = 1:1) for 3 h followed by a 48-h incubation with polyribinosinic polyribocytidylic acid (poly I:C sodium salt, Sigma-Aldrich, concentration in cultures,  $100 \mu\text{g/ml}$ ) and interferon  $\gamma$  (recombinant mouse IFN- $\gamma$ , BD Pharmingen™,  $10 \text{ ng/ml}$  in cultures).

***Mice.*** For *in vivo* experiments, C57BL/6 mice (8-week-old, bred in a local animal facility and kept in conventional conditions) were used. All the experiments were approved by the Local Ethics Committee.

***Flow cytometric analysis of stimulated JAWSII cell phenotype.*** The following monoclonal antibodies, previously described (16), were used to study surface markers of JAWSII cells: anti-MHC class I, anti-MHC class II, anti-CD11c, anti-CD40, anti-CD80, anti-CD86, anti-CCR7, and anti-CD8 $\alpha$ ; relevant isotype controls were used. Anti-CD80 and anti-CD86 double staining was performed so as to determine simultaneous expression of both CD80 and CD86. Surface markers were analyzed after pulsing JAWSII cells with tumor-cell lysates and incubation with appropriate immunostimulators at concentrations described above. The cells were then collected, resuspended in PBS with 0.5% BSA and 0.05% sodium azide (0.5 million cells in  $50 \mu\text{l}$ ), incubated for 30 min at  $4^\circ\text{C}$  with appropriate mAbs, and analyzed in FACS Scan (Becton-Dickinson). The level of surface marker expression, estimated by the mean fluorescence intensity (MFI), was analyzed using CellQuest software.

***Phagocytosis assay.*** JAWSII cells were stained with 1 mM DiI for 10 min at  $37^\circ\text{C}$  and washed two times with ice-cold PBS and seeded in 12-well plates. The following day, JAWSII cells were stimulated with  $10 \text{ ng/ml}$  IFN- $\gamma$  or/and  $100 \mu\text{g/ml}$  poly I:C. Twenty-four hours later, B78-H1 melanoma cells were stained with  $5 \mu\text{M}$  CFSE for 10 min at  $37^\circ\text{C}$ , washed two times with ice-cold PBS, and placed in a  $25 \text{ cm}^2$  culture flask. After 24 h, B78-H1 cells were lysed by the freeze/thaw method, added in a volume of  $100 \mu\text{l}$  and cocultured with JAWSII cells for an additional 3 h at  $37^\circ\text{C}$  and 5%  $\text{CO}_2$  or at  $4^\circ\text{C}$ . Then, the cells were collected, washed and resuspended in  $300 \mu\text{l}$  of PBS. A total of 10,000 cells were analyzed on a FACS Scan (Becton-Dickinson) using CellQuest Pro Software Version 5.2. Phagocytosis level was measured as a percentage of double positive cells.

***IL-12.*** Recombinant mouse IL-12 (specific activity  $4.6/10^6 \text{ U/mg}$  protein) was a generous gift from the Genetics Institute

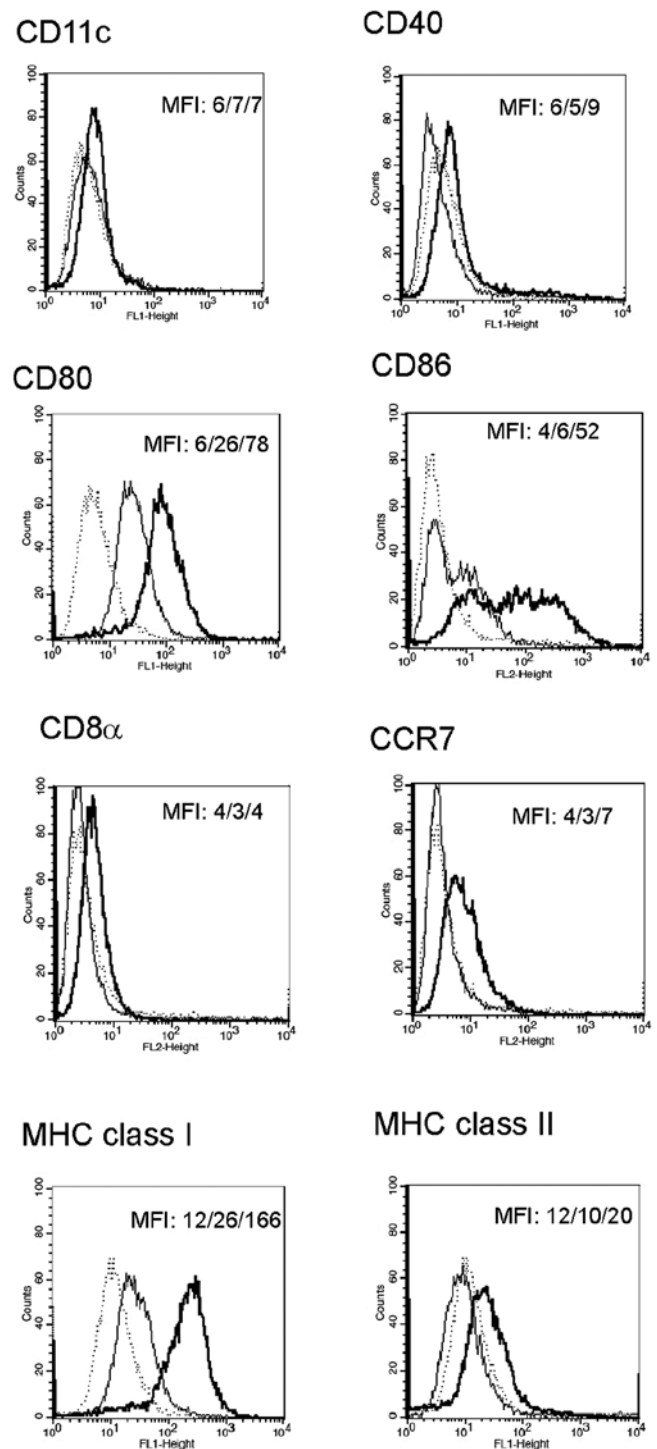
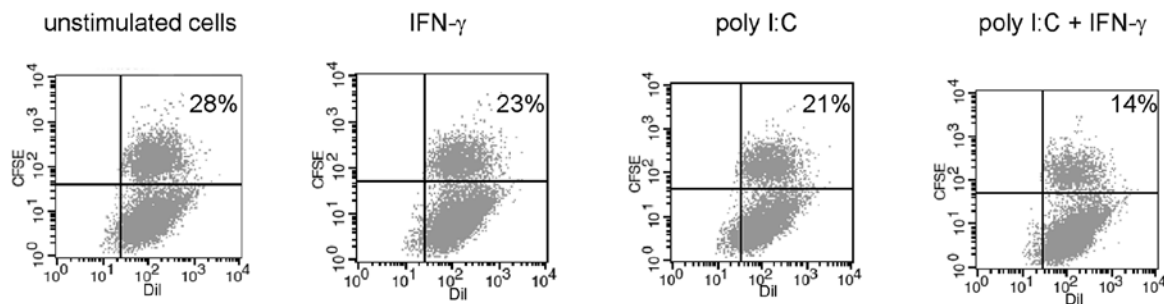


Figure 1. Expression of CD11c, CD40, CD80, CD86, CD8 $\alpha$ , CCR7, and MHC class I and II molecules on JAWSII cells pulsed with tumor cell lysates (B78-H1) and then incubated with poly I:C ( $100 \mu\text{g/ml}$ ) and IFN- $\gamma$  ( $10 \text{ ng/ml}$ ). Thick lines represent specific staining of stimulated cells. Relevant isotype controls are shown as dotted lines and specific staining of unstimulated cells is presented as thin lines. JAWSII cells were pulsed with tumor cell lysates and then incubated with both immunostimulators, followed by washing and immunostaining as described in Materials and methods. MFI = mean fluorescence intensity of isotype control/unstimulated JAWSII cells/stimulated JAWSII cells.

(Cambridge, MA, USA) (18). For *in vivo* experiments, the cytokine was diluted with 0.1% BSA (Sigma Chemicals, St. Louis, MO, USA).

## A - 37°C



## B - 4°C

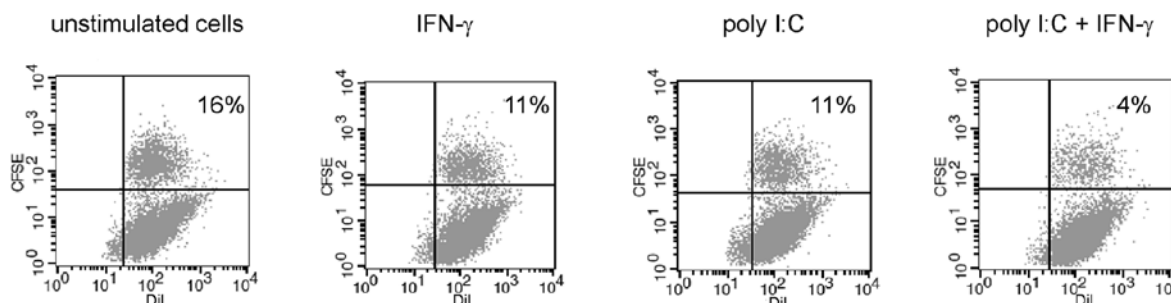


Figure 2. Phagocytosis of frozen-thawed B78-H1 melanoma cells by JAWSII cells at 37°C (A) and at 4°C (B). The JAWSII cells were stained with 1 mM DiI. For the following 48 h, JAWSII cells were stimulated with poly I:C (100  $\mu$ g/ml) and IFN- $\gamma$  (10 ng/ml) alone or in combination. Twenty-four hours after staining the JAWSII cells, tumor B78 cells were stained with 5  $\mu$ M CFSE and cultured for the next 24 h. Then, frozen-thawed B78-H1 melanoma cells were added to the JAWSII cultures and incubated for 3 h prior to FACS analysis. The JAWSII cells that phagocytosed lysed B78-H1 cells (double positive cells) are presented in the upper right quadrant (percentage).

*In vivo protocols used in the B78-H1 melanoma model.* Three variants of experiments with JAWSII cells were performed: a) prophylactic, b) therapeutic - intratumoral, and c) therapeutic - systemic schemes.

In the prophylactic scheme, randomly selected mice were injected into the footpad of the left hind limb with stimulated JAWSII on Day -7. On days: -6, -5 and -4 mice were administered with IL-12 or diluent (0.1% BSA in PBS). On Day 0, all the mice were inoculated with B78-H1 melanoma cells in the right hind limb.

In the intratumoral therapeutic scheme, all the mice were inoculated with B78-H1 melanoma cells into the footpad of the right hind limb. Next, on Day +3, mice were injected into the same site with stimulated JAWSII cells. On Days +4, +5 and +6, mice were injected with IL-12 or diluent (0.1% BSA in PBS).

In the systemic therapeutic scheme, mice were inoculated with B78-H1 melanoma cells into the footpad of the right hind limb. Next, on Day +3, mice were injected into the contralateral hind limb with stimulated JAWSII or PBS. On Days: +4, +5, and +6 mice were treated with IL-12 or diluent (0.1% BSA in PBS).

*Statistical analysis.* Results of experiments on FACS are presented on histograms together with MFI or percentage values. Statistical analysis of the *in vivo* experiments was performed with the Mann-Whitney-Wilcoxon test (tumor diameter in mm measured seven weeks after tumor cell inoculation) and the log-rank test (percentage of mice without tumor) using Statistica® software. For clarity, 3 levels of statis-

tical significance were used:  $p < 0.05$ ;  $p < 0.01$  and  $p < 0.001$  that were related to the control group (unless otherwise indicated).

## Results

*Expression of surface markers in the stimulated JAWSII cells.* Prior to the *in vivo* experiments in a murine melanoma model, JAWSII cells were pulsed with B78-H1 lysates followed by 48 h of incubation with poly I:C + IFN- $\gamma$ . Then, the influence of this treatment on the expression of surface markers of the cells was determined. In some experiments, FACS analysis of the stimulated JAWSII cells that were additionally incubated with IL-12 for the following 24 h was performed. As presented in Fig. 1, the cells pulsed with tumor cell lysates and incubated with poly I:C + IFN- $\gamma$  were characterized by high expression of CD80, CD86, and MHC class I molecules. Moreover, there was a moderate increase in the expression of CD40, CCR7, and MHC class II molecules in the cells.

A supplementary incubation with IL-12 (for 24 h), after pulsing with tumor lysates and a 48-h incubation with poly I:C + IFN- $\gamma$ , did not result in a change of surface markers when compared to the cells analyzed without IL-12 (data not shown).

*Phagocytosis of the frozen-thawed B78-H1 melanoma cells by JAWSII cells.* The ability of JAWSII cells to phagocytose B78-H1 lysates was determined in the experiment on FACS with CFSE dye. It was found that JAWSII cells phagocytosed frozen-thawed B78-H1 melanoma cells effectively. As

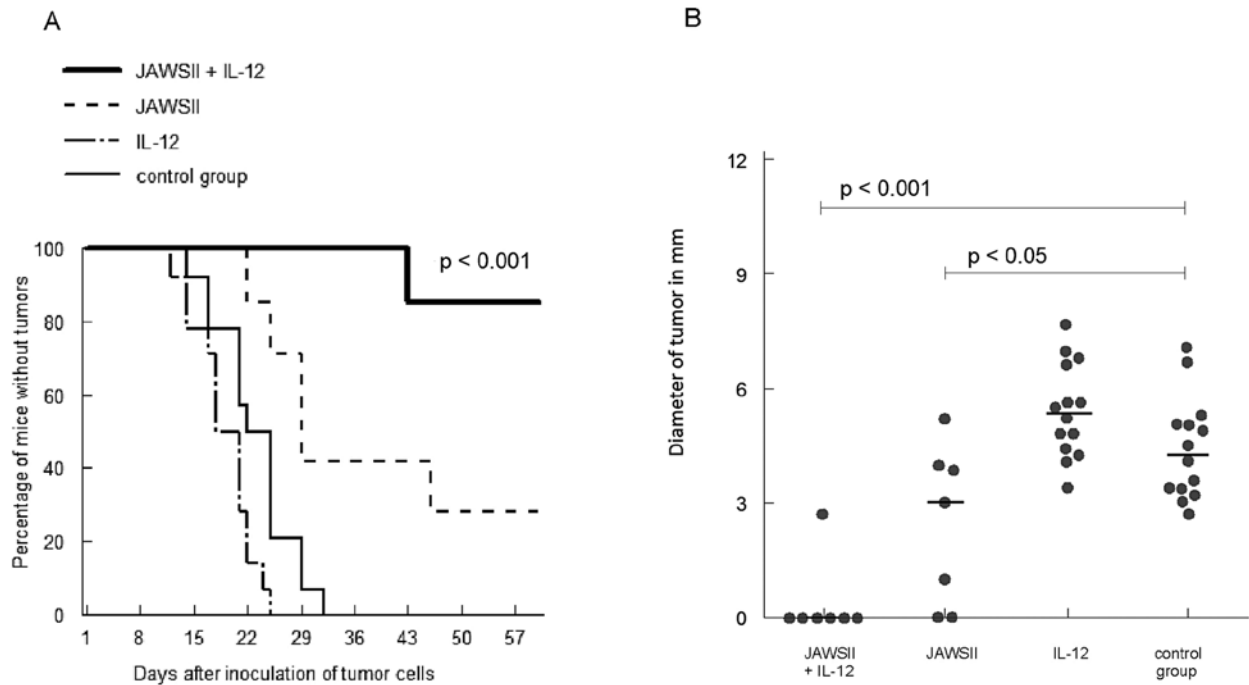


Figure 3. Effects of administration of JAWSII cells and/or IL-12 on the development of B78-H1 melanoma - prophylactic scheme. JAWSII cells (pulsed with B78-H1 cell lysates and incubated with poly I:C and IFN- $\gamma$  as described in Materials and methods) were injected into the footpad of the left hind limb on Day -7, followed by injection of IL-12 on Days -6, -5 and -4. On Day 0, tumor cells were inoculated into the contralateral limb. P denotes probability vs. control. (A) Percentage of mice without tumor. (B) Tumor diameter in mm measured seven weeks after tumor-cell inoculation. Horizontal lines denote medians.

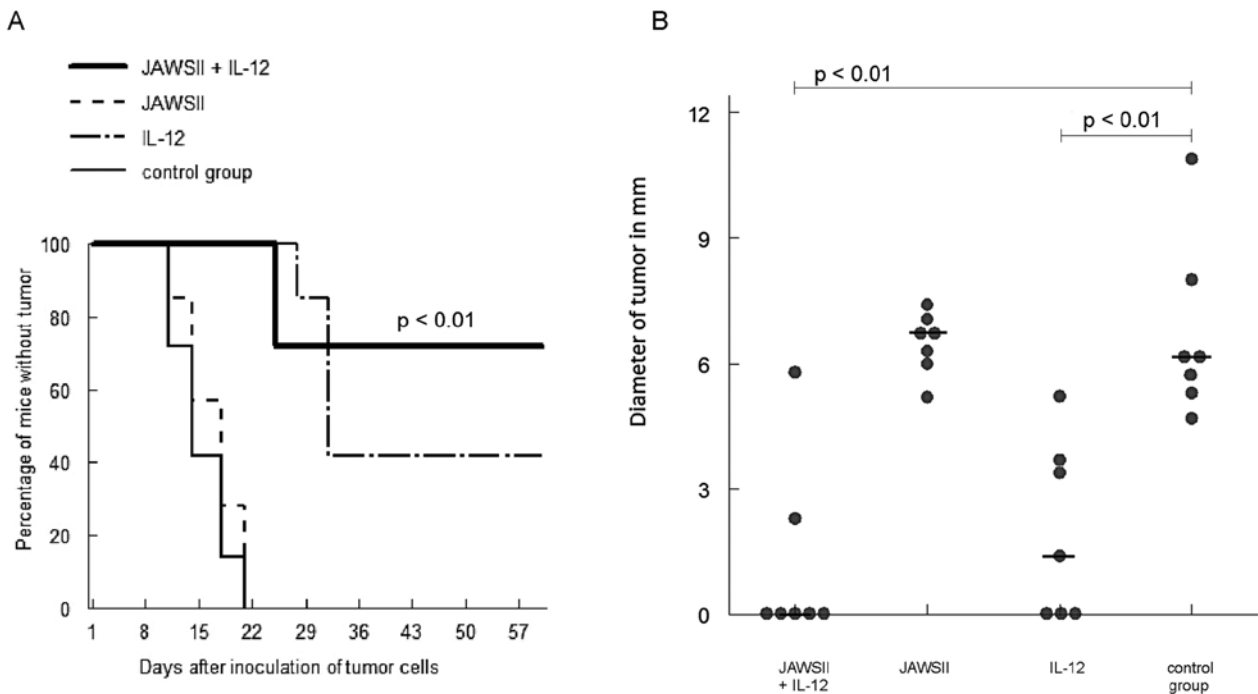


Figure 4. Effects of administration of JAWSII cells and/or IL-12 on the development of B78-H1 melanoma - therapeutic intratumoral scheme. JAWSII cells (pulsed with B78-H1 cell lysates and incubated with poly I:C and IFN- $\gamma$  as described in Materials and methods) were injected on Day +3 into the same footpad of the hind limb in which tumor cells were inoculated (Day 0), followed by injection of IL-12 on Days +4, +5 and +6. P denotes probability vs. control. (A) Percentage of mice without tumor. (B) Tumor diameter in mm measured seven weeks after tumor-cell inoculation. Horizontal lines denote medians.

presented in Fig. 2, the phagocytic capacity of JAWSII cells was decreased after stimulation with poly I:C (100  $\mu$ g/ml) and IFN- $\gamma$  (10 ng/ml), particularly in combination of these two agents.

**Production and secretion of IL-12 by JAWSII cells.** IL-12 was not present in supernatants from unstimulated JAWSII cultures and JAWSII lysates, using an ELISA kit (Mouse IL-12p70 ELISA MAX<sup>TM</sup> Set Deluxe, Biolegend, Inc., San Diego, CA,



of CD40, although some authors argue against the presence of the CD40 molecule in this cell line (21,22). Markedly, further incubation of the cells with IL-12 did not affect the expression of the studied surface markers. In the study of Rossowska *et al*, pulsing bone marrow-derived DCs with tumor cell lysates did not alter the levels of the surface markers (6). However, a slight decrease in expression of CD86, MHC class I and II molecules was observed when compared with untreated cells (6). In our previous study (16), the decreased ability of endocytosis of JAWSII cells (reflecting more mature stage of these cells) in cultures incubated with LPS or poly I:C was described, while in the present study we demonstrated maturation-inducing potential of combination of poly I:C and IFN- $\gamma$ . JAWSII cells are able to phagocytose lysed B78-H1 melanoma cells efficiently and the combination of immunostimulators diminishes that ability (Fig. 2). The impact of appropriate immunostimulators, mostly microbial elements, is crucial for the ability of DCs to promote anticancer response *in vivo* (6,7). They induce changes in the phenotype of DCs, which start to express high levels of costimulatory and MHC molecules (6). Furthermore, stimulated in that manner, semi-mature DCs retain properties for further maturation *in vivo* after the contact with the factors secreted by the host's immune system, mainly cytokines. As the immune response is usually suppressed by the tumor, coadministration of IL-12 with the cellular vaccine may induce stronger antitumor mechanisms (9).

In the prophylactic scheme of therapy in our murine melanoma model, JAWSII cells pulsed with tumor lysates and incubated with poly I:C + IFN- $\gamma$  demonstrated significant antitumor effect, which was augmented in the group of mice treated additionally with IL-12 (Fig. 3). It should be noted that the cytokine alone was found to be ineffective. By contrast, in the intratumoral therapeutic scheme, injections of stimulated JAWSII cells alone showed no antitumor effects while administration of IL-12 induced eradication of tumors in some mice (Fig. 4). The best treatment option was, as in the prophylactic protocol, application of JAWSII cells in combination with IL-12; most mice were cured. Despite induction of similar antitumor effects, it is possible that different mechanisms operated in the prophylactic versus the intratumoral therapeutic protocol. As we have shown in our previous studies, injection of activated and tumor lysate-fed JAWSII cells was associated with specific induction of effector cytotoxic T lymphocytes in regional lymph nodes (16). It is quite probable that supplementation of IL-12 enhanced this specific response in the prophylactic scheme (9,23). In the intratumoral therapeutic schedule, non-specific, NK cell-dependent mechanisms were able to contribute to the overall antitumor effect in the combination treatment, since IL-12 is the strong activator of these cells (24,25). The antiangiogenic effect of IL-12 may also be helpful in the eradication of tumors (26,27). Activation of both specific and non-specific mechanisms of immunity in the combination therapy (JAWSII cell vaccine + IL-12) was strong enough to induce, as shown in Fig. 5, antitumor responses in the most challenging systemic therapeutic scheme, mimicking clinical situation.

IL-12 was used in immunotherapy as a single agent in a number of models of murine cancers (28). However, it should be stressed that although some authors described a decrease in

the rate of tumor growth after injection with IL-12, superior effects were noted when the cytokine was used in combination with other forms of therapy (12,29,30). IL-12 may improve stimulating properties of bone marrow-derived DCs *in vitro* (31). In the study of Tatsumi *et al*, the use of IL-12 in combination with DCs pulsed with tumor lysates was more effective in comparison with DCs alone in a murine liver cancer model, leading to an induction of specific antitumor responses (8). Furthermore, Fallarino *et al* observed specific response of cytotoxic lymphocytes after administration of IL-12 and pulsed DCs in nearly 100% of mice (9). In one clinical study, peripheral blood mononuclear cells (PBMCs) loaded with tumor antigenic peptides and IL-12 induced CD8<sup>+</sup> T-cell response (23). However, methodological differences (using PBMCs in selected HLA-A2<sup>+</sup> patients, lack of IL-12 alone-treated controls) makes comparison of this study with our model unreliable.

In conclusion, in the present study we presented antitumor effects of the JAWSII cell-based cellular vaccine used in combination with IL-12 in a murine model of B78-H1 melanoma. The findings of this study may aid in the planning optimal DC-based therapeutic protocols in cancer patients.

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