

# Transcriptional network in ovarian cancer cell line SKOV3 treated with *Pinellia pedatisecta* Schott extract

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**Abstract.** Ovarian cancer is the most lethal disease among the malignant tumors of female reproductive organs. Few successful therapeutic options exist for patients with ovarian cancer. The common therapeutic methods are surgical operation, chemotherapy, radiotherapy, and combination of these treatments. In recent years, studies have indicated that *Pinellia pedatisecta* Schott (PPS), a traditional Chinese medicine, could inhibit tumor growth. In this study, we demonstrated that PPS extract could induce apoptosis in SKOV3 cells in a dose- and time-dependent manner. We further conducted transcriptome sequencing on PPS extract-treated SKOV3 cells along with controls, and identified 1,754 transcripts whose expression differs at least 3-fold over the controls. These differentially expressed transcripts include the apoptosis-related genes such as the caspase family members, and were significantly enriched in steroid biosynthesis in the KEGG pathway database compared with the transcriptome background. Most of the differentially expressed transcripts from this pathway were upregulated in PPS extract-treated cell line, indicating that PPS extract-induced apoptosis was accompanied by increased steroid biosynthesis (e.g. zymosterol). These results suggest that PPS extract could be a new cytostatic therapeutic agent for ovarian cancer.

## Introduction

Epithelial ovarian cancer (EOC), the most common ovarian malignancy, continues to be the leading cause of death among gynecological malignancies (1). The poor survival from EOC is due to the high percentage of patients diagnosed at an advanced stage and a lack of effective treatment (2). Currently the standard treatment protocol used in the initial management of advanced-stage ovarian cancer is primary cytoreductive surgery, followed by platinum/taxane combination chemotherapy. Approximately 70% of patients will have a complete clinical response to this initial therapy, indicated by an absence of clinically detectable residual disease based on clinical exam, radiologic imaging, or measurement of serum tumor marker CA125 (3). Unfortunately, the majority of these patients will eventually develop recurrence or persistent platinum-resistant disease (4). Therefore, more effective treatments for advanced ovarian cancer are urgently required (5).

*Pinellia pedatisecta* Schott (PPS), a traditional Chinese medicine (TCM), has been used, based on many TCM books, to treat several disorders. In the 1970s, *Pedate Pinellia Rhizome* was used in the Obstetrics and Gynecology Hospital of Fudan University to treat 247 cervical cancer patients with a total effective rate of 81.5%. Following, Li *et al* found that a novel lipid-soluble extract (PE) from PPS can induce apoptosis in human cervical cancer cells with few side effects on normal cells, suggesting that PPS extract could be used as a potential drug for cervical cancer treatment (6). In addition, Sun *et al* found that total PPS extract can inhibit mouse sarcoma-180 growth *in vivo* by direct intraperitoneal injection (7). However, the pharmacological mechanism of this action has not been established.

In this study, we first examined whether the PPS extract component could induce apoptosis and exert proliferation inhibition on ovarian cancer cell line SKOV3. To determine how PPS extract affects apoptosis and tumor inhibition, we performed high throughput mRNA sequencing and bioinformatics analysis to determine transcriptome difference between PPS-treated SKOV3 cells and controls.

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**Key words:** ovarian cancer, SKOV3, *Pinellia pedatisecta* Schott, apoptosis, transcriptome sequencing, differential gene expression

## Materials and methods

**Cell line.** The ovarian cancer cell line SKOV3 was purchased from Shanghai Institutes for Biological Sciences and maintained as recommended by the supplier (American Type Culture Collection, Rockville, MD, USA). Cells were cultured in RPMI-1640 supplemented with 10% fetal bovine serum (FBS), 1% sodium pyruvate, and 1% nonessential amino acids. All tissue culture reagents were obtained from Sigma-Aldrich (St. Louis, MO, USA).

**Preparation of total extract from PPS.** Fresh PPS was collected from Xuchang, Henan Province, China. It was washed and peeled, and ground to generate a homogenate with normal saline. The homogenate was centrifuged at 3000 rpm and 4°C for 20 min and the supernatant was retrieved. Crude extract of PPS was obtained by two times gradient sedimentation with acetone (V/V). The sediments was dissolved into 0.9% NaCl (m/m) and followed by dialysis in distilled water at 4°C for 48 h. After high-speed centrifugation, the supernatant was extracted as the final total extract. Extract protein concentrations were measured by bicinchoninic acid (BCA) method. Detection of phosphorus, saccharide and lipid protein in the total extract was measured by ammonium molybdate spectrophotometry, Molisch's test and Sudan black B staining method, respectively.

**Cytotoxicity assay.** SKOV3 cells at logarithmic growth phase were seeded at 5000 cells per well in 96-well culture plates and cultured for 24 h. The PPS extract protein at increasing concentrations (0.8, 4, 20, 100, 200, 300, 400 µg/ml) was added to the wells. After 72 h, CCK-8 (10% V/V) was added to the culture and incubated for 1 h. With 630 nm wavelength as the reference, the absorption values at 450 nm were collected for analysis in an automatic ELISA detection instrument. Medium alone was used as the blank control. Cells without drug treatment or with cisplatin (DDP) treatment were used as negative or positive controls, respectively. All the experiments were replicated six times.

**Flow cytometry analysis.** SKOV3 cells in logarithmic growth phase were seeded in 6-well plates at approximately  $8 \times 10^4$  cells per well and cultured for 24 h. Increasing concentrations of the protein extract of PPS (0, 20, 50, 100, 200, 400 µg/ml) were added to the wells and cells were cultured for an additional 72 h at 37°C. The cells were trypsinized, washed twice with ice-cold PBS extract, adjusted to  $1 \times 10^6$  cells/ml and fixed with 70% ethanol. Then, 100 µl cell suspension aliquot was taken to each labeled tube, followed by adding 10 µl Annexin V-FITC and 10 µl PI (20 µg/ml) (Annexin V/FITC kit: BD Biosciences). After 20 min incubation in darkness at room temperature, 400 µl PBS binding buffer was then added and cell apoptosis was detected immediately by flow cytometry (BD Biosciences Clontech, San Jose, CA, USA) (within 30 min after adding PBS binding buffer). Samples in triplicates were examined. SKOV3 cells without treatment were used as the reference. Data analysis was conducted using the Cell Quest software.

**Transcriptome sequencing.** SKOV3 cells and those treated with 200 µg/ml PPS extract for 72 h were subjected to mRNA-Seq library construction and transcriptome sequencing.

Approximately  $10^8$  cells from each group were collected after PBS washes. Oligo(dT) beads were used to capture poly(A) mRNA after total RNA was extracted. Each poly(A)-enriched RNA sample was chemically fragmented into small pieces using divalent cations at 94°C for 5 min. The fragmented RNA was reverse-transcribed into cDNA using random hexamer primers containing a tagging sequence at the 3' end and a SuperScript III Double-Stranded cDNA synthesis kit according to the manufacturer's protocol (Invitrogen, Carlsbad, CA, USA). The double-stranded cDNA was subjected to end-repair and further 3' terminal tagging by the addition of 5' DNA adaptors and T4 DNA ligase with overnight incubation at 16°C for 16 h. The targeted di-tagged cDNA was purified by polyacrylamide gel electrophoresis and gel excision ( $200 \pm 25$  bp). The clean di-tagged cDNA was enriched by 18 PCR cycles with primer pairs annealed to the tagging sequences of the di-tagged cDNA. Library purification by PAGE removed any residual nucleotides, PCR primers and small amplicons. Two paired-end libraries were sequenced using the HiSeq2000 platform (Illumina). The initial short reads from this study are being submitted to the NCBI sequence read archive database and the accessions will be released soon.

**Bioinformatics analysis.** The raw Illumina/Solexa reads of SKOV3 cells and PPS extract-treated cells were subjected to adapter trimming and low quality filtering using the Trimmomatic program (8). The high quality clean reads were aligned to the human genome using TopHat (9). Human genome (hg19) sequence and gene annotation were obtained from the UCSC Genome Browser (<http://genome.ucsc.edu/>). Cuffdiff was used to perform differential gene expression profiling with default parameters (10,11). The significantly differentially expressed transcripts (DETs) between SKOV3 and PPS extract-treated cell line were selected using the following criteria: i) if the FPKM value for a certain transcript in both samples was  $>1$ , the difference between them should be at least 3-fold. ii) If the FPKM value for a certain transcript in one sample was  $<1$ , the FPKM value for this gene in the paired sample should be  $>3$ . Mapping transcripts to the KEGG pathway were conducted using the BLASTX and followed by custom scripts (12). KEGG pathway enrichment analysis of the DETs compared with transcriptome background (Cuffdiff detected transcript status as 'OK') was performed by hypergeometric distribution testing using the Phyper function of the R software package (<http://www.r-project.org/>). Bonferroni correction was used to adjust P-value for each pathway.

**Quantitative real-time PCR validation of mRNA-Seq data.** The mRNAs which have been used to perform mRNA-Seq experiments from two cell lines were used for qPCR validation. Total RNA from each sample was treated with DNase I before reverse transcription by SuperScript III double-stranded cDNA synthesis kit. Ten intron-spanning target gene primers and two endogenous control (18s rRNA and GAPDH) primers were designed using Primer 5.0 for qPCR experiment. SYBR Green I based qPCR was performed according to the routine protocols (13) on a 7500 Real-time PCR System (Applied Biosystems, Foster City, CA, USA) and three technical replicates were performed for all the genes in each sample. Gene

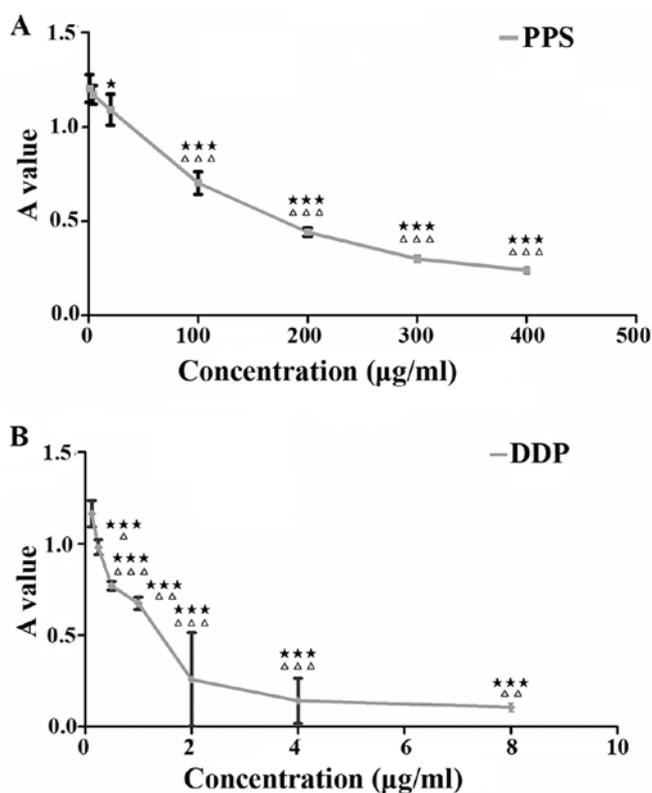


Figure 1. Effect of PPS extract and cisplatin on the proliferation inhibition of SKOV3 cells. (A) Cells were cultured in 10% FBS medium and treated with PPS (0.8, 4, 20, 100, 200, 300, 400  $\mu\text{g/ml}$ ) for 72 h. (B) Cisplatin (DDP) (0.125, 0.25, 0.5, 1.0, 2.0, 4.0, 8.0  $\mu\text{g/ml}$ ) was used as the positive control. Each group was replicated six times. Absorption value was expressed as mean  $\pm$  SD from six independent experiments. Black pentangle indicates statistical difference between experimental group and blank control group (\* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001). White triangle indicates statistical difference between certain concentration group and the next higher concentration group treated with PPS extract or cisplatin ( $\Delta$  $P$ <0.05,  $\Delta\Delta$  $P$ <0.01,  $\Delta\Delta\Delta$  $P$ <0.001).

expression difference between two lines was calculated by the  $2^{-\Delta\Delta\text{Ct}}$  method (14).

**Statistical methods.** All data were first subjected to normality test and expressed as mean  $\pm$  standard deviation. Independent-sample t-test was used to explore the apoptosis rate and/or proliferation inhibition differences between PPS extract-treated group and control group, cisplatin-treated group and control group, 150  $\mu\text{g/ml}$  PPS extract treatment for 48 and 72 h, and 1  $\mu\text{g/ml}$  cisplatin treatment for 48 and 72 h. Multiple concentrations of PPS extract-treated SKOV3 cells within the same group were compared using one-way ANOVA after homogeneity of variance was tested. LSD method was used to analyze the difference in apoptosis rate between two samples if homogeneity of variance existed. Otherwise, Dunnet's T3 testing was adopted. Data analysis was performed using SPSS software version 16.0.

## Results

**Effect of PPS extract on the proliferation of SKOV3 cells.** We evaluated the total extract obtained from the PPS. The absorption value of total extract at 660 nm wavelength was entirely identical to the blank control ( $A_{660}=0$ ) indicating that

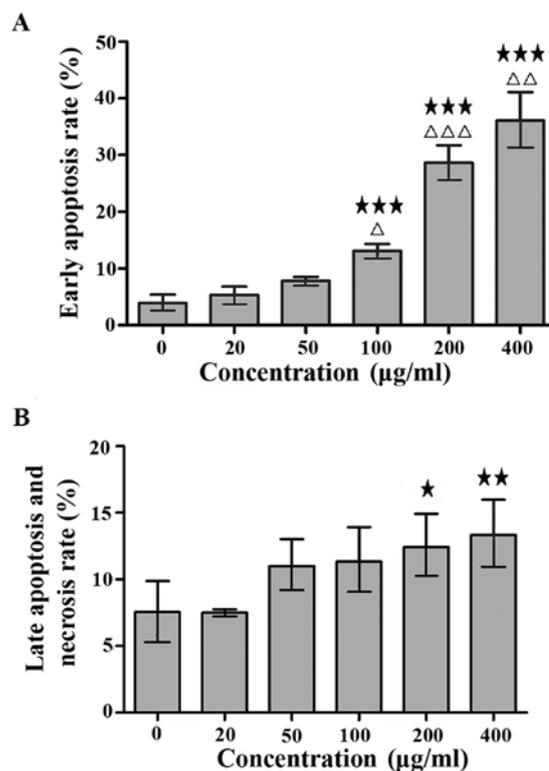


Figure 2. Effects of PPS extract on early apoptosis rate, late apoptosis rate and necrosis rate in SKOV3 cells. Cells were cultured in 10% FBS medium and treated with PPS (0, 20, 50, 100, 200, 400  $\mu\text{g/ml}$ ) for 72 h. Apoptosis rates are expressed as mean  $\pm$  SD from three independent experiments. (A) Early apoptosis rate of SKOV3 cells treated with PPS. (B) Late apoptosis rate and necrosis rate of SKOV3 cells treated with PPS. Black pentangle indicates statistical difference between experimental group and blank control group (\* $P$ <0.05, \*\* $P$ <0.01, \*\*\* $P$ <0.001). White triangle indicates statistical difference between certain concentration group and the next higher concentration group treated with PPS extract ( $\Delta$  $P$ <0.05,  $\Delta\Delta$  $P$ <0.01,  $\Delta\Delta\Delta$  $P$ <0.001).

there was no phosphorus presence. In addition, total extract could not be stained by Sudan dye suggested the absence of lipoprotein. Moreover, total extract can react with  $\alpha$ -naphthol to generate purple compound indicating the presence of glycoprotein in the extract. The productivity of total protein extracted from fresh PPS was 1% (9.97 g total extract from 1000 g PPS). We found that 20  $\mu\text{g/ml}$  of the total extract of PPS significantly inhibited the growth of SKOV3 cells *in vitro*. As the concentration of the PPS extract increased, the inhibition rate was enhanced, showing a significant dose-effect relationship. However, 0.8  $\mu\text{g/ml}$  and 4  $\mu\text{g/ml}$  PPS extract had no obvious effect on cell proliferation inhibition. When the extract protein concentration reached 100  $\mu\text{g/ml}$  or higher, the inhibitory effect of PPS on SKOV3 proliferation was significantly different between the two adjacent concentrations ( $F=407.084$ ,  $P$ <0.001, Fig. 1A). Cisplatin group also showed a significant dose-effect relationship. The inhibitory effect of cisplatin on SKOV3 cells was significantly different between different concentrations ( $F=799.621$ ,  $P$ <0.001, Fig. 1B).

**Effect of the different concentrations of the PPS extract on cell apoptosis rate.** We observed that PPS protein extract at higher than 100  $\mu\text{g/ml}$  significantly increased the early apoptosis rate of SKOV3 cells compared with that of the control group (Fig. 2A). PPS extract induced apoptosis of SKOV3 cells in a

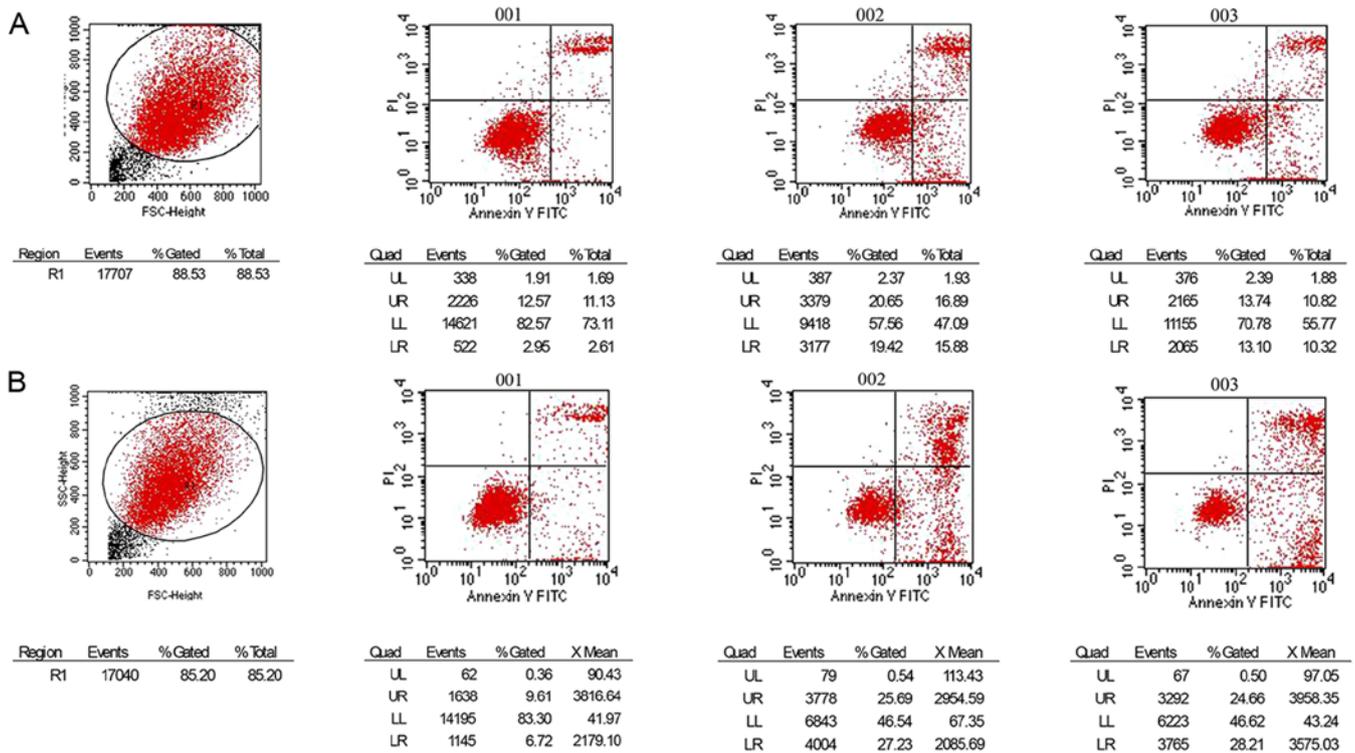


Figure 3. Time-dependent effect of PPS and DDP on the apoptosis rate in SKOV3 cells. (A) Apoptosis rate of SKOV3 cells treated with PPS or DDP for 48 h. (B) Apoptosis rate of SKOV3 cells treated with PPS or DDP for 72 h. 01, Control group (medium only); 002, PPS (150  $\mu\text{g}/\text{ml}$ ) group; 003, DDP (1  $\mu\text{g}/\text{ml}$ ) group; UR, late apoptosis and necrosis; LR, early apoptotic.

dose-dependent manner. When the protein extract concentration increased from 20  $\mu\text{g}/\text{ml}$  to 50  $\mu\text{g}/\text{ml}$ , the early apoptosis rate was slightly increased. However, this rate was significantly increased at extract concentration of 100  $\mu\text{g}/\text{ml}$  or higher when compared with the next higher concentration group (Fig. 2A). Only the high concentrations (200  $\mu\text{g}/\text{ml}$ , 400  $\mu\text{g}/\text{ml}$ ) of PPS extract had significant effects on late cell apoptosis rate and necrosis rate, whereas the low concentrations of PSS protein had no obvious effects (Fig. 2B).

*Time-dependent effect of PPS extract on cell apoptosis rate.* After the treatment with a relatively high concentration of the PPS extract, SKOV3 cells showed obvious apoptosis, and the apoptosis rate increased with time extension (Fig. 3). The early apoptosis rate in the cells treated with 150  $\mu\text{g}/\text{ml}$  PPS protein extract for 48 h was  $14.860 \pm 1.590\%$ , significantly different from the control group ( $t=14.224$ ,  $P<0.001$ ). Similarly, the early apoptosis rate reached  $24.426 \pm 4.149\%$  after 72 h treatment with the same extract concentration and the difference between the control group and the treated group was also statistically significant ( $t=8.461$ ,  $P=0.001$ ). The apoptosis rates of cells treated with 150  $\mu\text{g}/\text{ml}$  PPS extract were significantly different between the 48 and 72 h treatment groups ( $t=3.729$ ,  $P=0.02$ ).

As a classical chemotherapeutic drug, cisplatin has been widely used to treat various cancers. After the treatment with 1  $\mu\text{g}/\text{ml}$  cisplatin for 48 and 72 h, the early apoptosis rates of SKOV3 cells reached  $10.033 \pm 2.834\%$  and  $22.470 \pm 6.913\%$ , respectively, both significantly different from the blank control group ( $t=5.116$ ,  $P=0.007$ ;  $t=4.629$ ,  $P=0.01$ , respectively). The

difference between 48 and 72 h treatment was also statistically significant ( $t=2.877$ ,  $P=0.045$ ). These data indicated that PPS extract plays a significant role in apoptosis induction of ovarian cancer cell line SKOV3 (Fig. 3).

*Transcriptome profiling of SKOV3 and PPS extract-treated SKOV3 and qPCR validation.* To reveal how the PPS extract alters the gene expression of SKOV3 cells after 72 h, the transcriptomes of SKOV3 cells and PPS extract-treated SKOV3 cells were sequenced using Illumina/Solexa technology. In total, 44,576,818 and 52,295,034 raw reads with 100 bp in length were obtained from SKOV3 and PPS extract-treated cell lines, respectively. After filtering to exclude low quality, adaptor contamination and ambiguous base containing reads, 43,071,352 and 50,530,232 high quality clean reads remained, representing approximately 4.3 and 5.0 Giga base. We mapped the reads from two cell lines to hg19 genome for quantitative measurement of gene expression and found that 17,217 transcripts were detected as expression. Differential gene expression analysis between two samples showed that 1,754 transcripts were significantly differentially expressed, of which 840 were upregulated and 914 were downregulated in PPS extract-treated group as compared with the untreated group.

To determine in which biochemical pathways these differentially expressed transcripts (DETs) were enrich, total expressed transcripts (17,217) were mapped to the KEGG pathway. Transcripts (14,135) were assigned to 283 pathways representing 23,308 functional occurrences, of which 1,341 annotated transcripts corresponding to 2,365 frequency of use in our identified DETs. Enrichment analyses were performed

Table I. Transcript expression difference between SKOV3 and PPS extract-treated SKOV3 cells involved in steroid biosynthesis identified by mRNA-Seq.

Transcript accession	Protein symbol	Enzyme code	FPKM-SKOV3	FPKM-PPS	Fold change
NM_005891	ACAT2	EC:2.3.1.9	9.06	119.50	13.19
NM_002130	HMGCS1	EC:2.3.3.10	10.94	67.12	6.13
NM_000859	HMGCR	EC:1.1.1.34	7.56	18.69	2.47
NM_000431	MVK	EC:2.7.1.36	5.69	17.43	3.06
NM_006556	PMVK	EC:2.7.4.2	32.00	35.38	1.11
NM_002461	MVD	EC:4.1.1.33	12.85	37.67	2.93
NM_001135822	FDPS	EC:2.5.1.10	39.68	128.57	3.24
NM_001287748	FDFT1	EC:2.5.1.21	2.68	70.79	26.38
NM_003129	SQLE	EC:1.14.99.7	17.99	68.63	3.82
NM_002340	LSS	EC:5.4.99.7	0.0025	10.95	>1000
NM_000786	CYP51A1	EC:1.14.13.70	4.39	18.04	4.11
NM_015922	NSDHL	EC:1.1.1.170	26.82	46.66	1.74
NM_001017369	MSMO1	EC:1.14.13.72	5.13	25.47	4.97
NM_006745	MSMO1	EC:1.14.13.72	18.84	65.15	3.46
NM_001129765	NSDHL	EC:1.1.1.170	1.36	4.70	3.46
NM_016371	HSD17B7	EC:1.1.1.270	3.35	7.55	2.25
NM_006579	EBP	EC:5.3.3.5	10.77	3.36	0.31

Table II. Primers used in the qPCR.

Gene	Primer name	Sequence (5'→3')
18s rRNA	18s rRNA_F	GGAGCCTGCGGCTTAATTT
	18s rRNA_R	CAACTAAGAACGGCCATGCA
GAPDH	GAPDH_F	CAAATTCCATGGCACC GTCA
	GAPDH_R	GGCAGAGATGATGACCCTTT
ACAT2	ACAT2_F	CGGCAGGAGAAGCAAGATGA
	ACAT2_R	GCTGCCAAGACATGTCCAAA
HMGCS1	HMGCS1_F	GCTGCTGTCTTCAATGCTGT
	HMGCS1_R	TTTGGCCCAATTAGCAGAGC
FDPS	FDPS_F	AACTACTCGACCCACAGAGC
	FDPS_R	CTCAGTCAGCACCCCTAACGA
SQLE	SQLE_F	GTTCGCCCTCTTCTCGGATA
	SQLE_R	TGTTGATGTACAGGCAGCTG
LSS	LSS_F	CCAGCCGGATACAGAGAAGA
	LSS_R	CCACAGCACCCCTTTCTTG
HSD17B7	HSD17B7_F	CGTTTGCTTCACTGCTTGGA
	HSD17B7_R	GCTTCTGCCTTGCTCATGTT
MVK	MVK_F	CCTGAAGTACAACGCCTCCT
	MVK_R	TGCTACCTTGCCATGTACCA
MVD	MVD_F	CGATGAAGAGCTGGTTCTGC
	MVD_R	CCATCCCGTGAGTTCTCTCC
CYP51A1	CYP51A1_F	ACCCTCAGCCTGGTCTACC
	CYP51A1_R	GCATCACTCCCCAGAAGGTA
MSMO1	MSMO1_F	TGGAACACCTGGCGAGTC
	MSMO1_R	TCCCCATGTTGCAATCTGGA

using hypergeometric distribution testing followed by multiple hypothesis testing. Result showed that DETs associated with steroid biosynthesis ( $q < 0.01$ ) were over-represented as 11 of 2,365 (0.47%) DETs could be mapped to this metabolism pathway, whereas the counterparts in the transcriptome background were 32 and 23,308 (0.14%), respectively. Table I showed the gene expression differences between two groups of cell involved in the steroid biosynthesis.

To evaluate high throughput sequencing data, ten DETs involved in the steroid biosynthesis pathway were selected for qPCR assay. Table II summarized the gene primers used in this qPCR assay. 18s rRNA and GAPDH were used as internal reference to normalize gene expression between two samples. We found that all genes were upregulated in PPS extract-treated cell line compared with untreated SKOV3, which is consistent with the mRNA-Seq data although the exact fold changes for these genes between the two techniques within two cell lines are slightly different (Table III). The correlation coefficients (Pearson) of differential expression ratio between two techniques were 0.895 and 0.896 when 18s rRNA and GAPDH were used as internal control for qPCR analysis, respectively (Fig. 4).

## Discussion

*Pedate Pinellia Rhizome*, the rhizome of PPS, is a common TCM found in the mid-region of China. It has been reported in many TCM books to possess efficacy in dispelling wind, relieving convulsion, drying dampness to eliminate phlegm, eliminating stagnation, and has long been used to cure thanatophidia bite, nameless swelling and toxicum. It has also been shown to have antitumor properties without apparent toxic effects on noncancerous cells (6,15). In this study, total extract obtained from PPS was used to investigate apoptosis

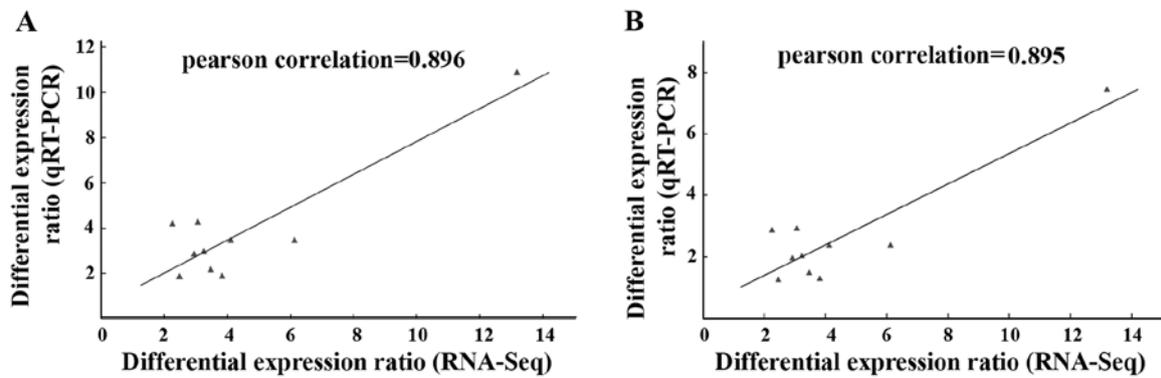


Figure 4. Pearson correlation coefficient of differential expression ratios between mRNA-Seq and qRT-PCR. Correlation was significant at the 0.01 level. GAPDH was used as internal control. (B) 18s rRNA was used as internal control.

Table III. qPCR validation of ten DETs involved in the steroid biosynthesis pathway.

Endogenous control	Relative fold difference of transcript abundance between SKOV3 and PPS protein-treated SKOV3									
	ACAT2	HMGCS1	FDPS	SQLE	LSS	HSD17B7	MVK	MVD	CYP51A1	MSM01
18s rRNA	7.46	2.40	2.05	1.31	1.97	2.89	2.95	1.97	2.40	1.51
GAPDH	10.90	3.50	2.99	1.92	2.87	4.22	4.30	2.88	3.50	2.20

Table IV. Transcript expression difference between SKOV3 and PPS extract-treated SKOV3 cells involved in apoptosis identified by mRNA-Seq.

Transcript accession	Protein symbol	FPKM-SKOV3	FPKM-PPS	Fold change
NM_032991	CASP3 variant $\beta$	2.7	8.85	3.27
NM_033356	CASP8 variant C	0	3.11	Inf
NM_001229	CASP9 variant $\alpha$	0.0008	3.23	>1000
NM_001267056	CASP7 variant e	14.77	4.18	0.28
NM_001227	CASP7 variant a	0.66	6.58	9.96
NM_032974	CASP10 variant 2	0.36	4.53	12.72
NM_001204401	XIAP variant 2	0.0022	4.14	>1000
NM_001012270	BIRC5 variant 2	1.54	5.42	3.52
NM_001171625	VEGFA variant 3	8.71	0.09	0.01
NM_001243733	VEGFB-167	12.6	0	0
NM_198156	VHL variant 2	0.13	3.65	28.1
NM_004465	FGF10	6.11	0	0

induction and proliferation inhibition ability against ovarian cancer cell line SKOV3. Cisplatin, the most commonly used chemotherapy drug in ovarian cancer, was used as the positive control. Results showed that only low concentrations of PPS protein extract (20  $\mu\text{g/ml}$ ) had obvious inhibitory effect on cell proliferation, whereas high concentrations of PPS extract ( $\geq 100 \mu\text{g/ml}$ ) significantly increased the early apoptosis rate of SKOV3 cells. In addition, the early apoptosis rate was also significantly increased when the treatment time was extended from 48 to 72 h under the same extract concentration (150  $\mu\text{g/ml}$ ). These results indicated that the extract of PPS could

induce apoptosis in ovarian cancer cell SKOV3 in a dose- and time-dependent manner.

Transcriptome sequencing and quantitative gene expression analysis identified 1,754 DETs between SKOV3 and PPS extract-treated cells. These DETs were significantly enriched in the steroid biosynthesis pathway. In this pathway, we found that most of the enzymes responsible for the synthesis from farnesyl-diphosphate to 3-keto-4-methylzymosterol were significantly upregulated in response to PPS treatment (Tables I and III). Of note, many genes involved in the synthesis of farnesyl-diphosphate, the original substrate of the

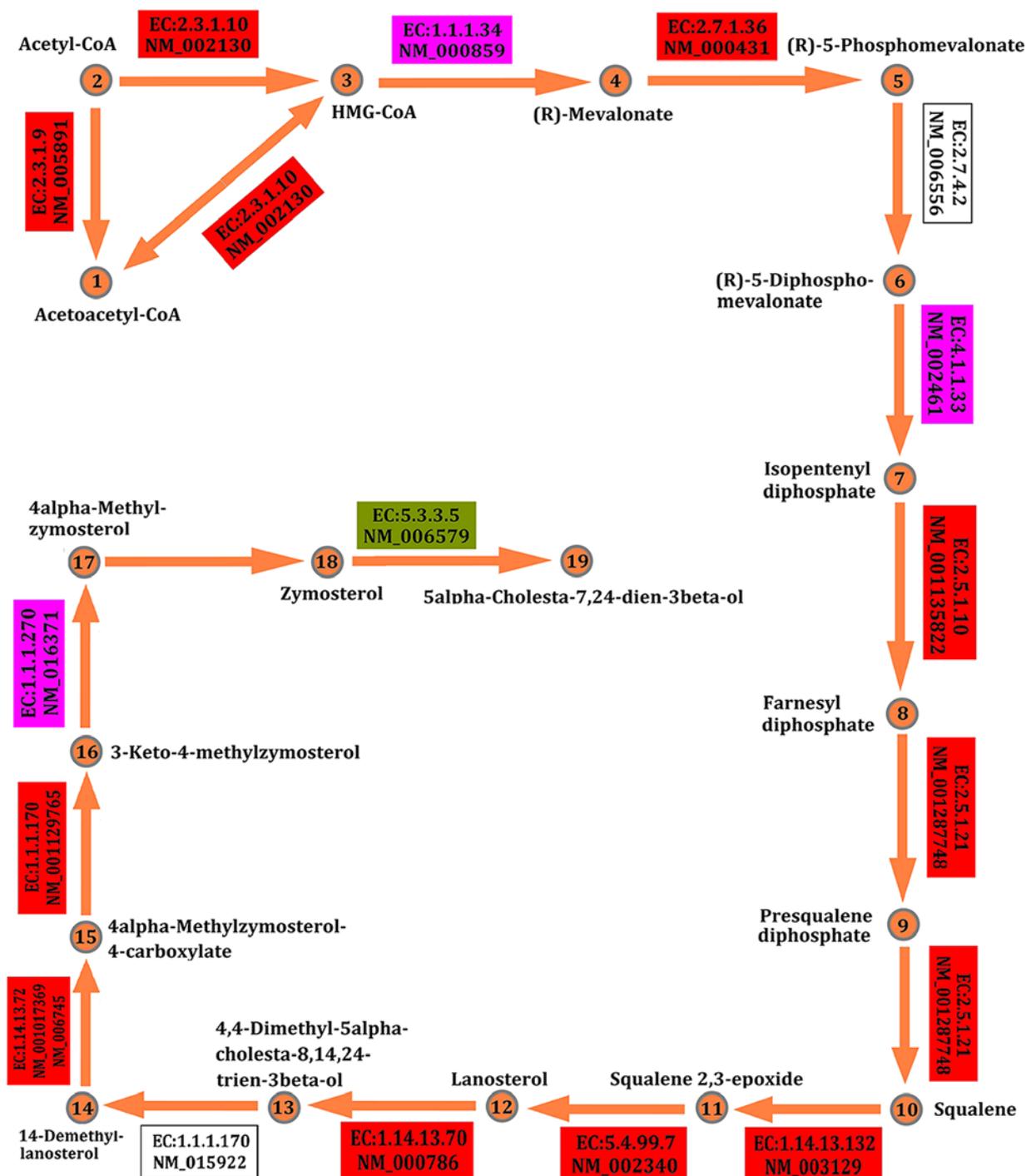


Figure 5. Differentially expressed transcripts (DETs) between SKOV3 and PPS extract-treated SKOV3 cells involved in zymosterol biosynthesis. Red, significantly upregulated transcripts (>3-fold) in PPS extract-treated group compared with control group. Pink, 2-3-fold upregulated transcripts in PPS extract-treated group. Green, significantly downregulated transcripts (<1/3-fold) in PPS extract-treated group.

steroid biosynthesis, were significantly upregulated, although not enriched, in the terpenoid backbone biosynthesis pathway. These upregulated DETs constituted a relatively intact reaction chain associated with the synthesis of acetyl-CoA to 3-keto-4-methylzymosterol, indicative of increased accumulation of zymosterol in the PPS extract-treated cells. Validation of transcriptome sequencing data by using qPCR also showed that genes associated with the zymosterol biosynthesis were significantly upregulated after treated with PPS extract.

In addition, the cholesterol delta-isomerase, which is enzymatically involved in the catalysis of zymosterol to 5alpha-Cholesta-7,24-dien-3beta-ol, was downregulated, further suggesting the retained accumulation of zymosterol in PPS extract-treated cells (Fig. 5). Furthermore, the expression of genes encoding the down-stream enzymes in the steroid biosynthesis pathway were not significantly differential, suggesting that the final product for majority of farnesyl-diphosphate may not be cholesterol or other secosteroid, such

as vitamin D2, but zymosterol or its precursor. Zymosterol, an intermediate precursor of cholesterol, was located in the plasma membrane of human cells and has been considered to play an important role in regulating membrane fluidity and permeability (16-18). Previous studies have demonstrated that compounds derived from the isoprenoid/cholesterol biosynthetic pathway possessed novel biological activities, such as regulating transcriptional and post-transcriptional events that in turn affect lipid synthesis, meiosis, apoptosis, developmental patterning, protein cleavage, as well as protein degradation (19). The upregulated expression of enzymes encoding genes involved in the biosynthesis of zymosterol may be closely related to the increase of cell apoptosis rate after PPS extract treatment.

To date, two classical apoptosis signaling pathways (the extrinsic and intrinsic pathways) have been well established (20,21). The extrinsic pathway is initiated by the extracellular hormones or agonists that can be recognized by the cell surface death receptors (22). The activation of intrinsic pathway can be initiated by the release of cytochrome c or SMAC/DIABLO from mitochondria (21). Programmed cell death (PCD) from both signaling pathways can be finally ascribed to caspase activation, including the initiator, such as caspase-8, -9, -10, and the executioner, such as caspase-3 and -7. Since treatment of SKOV3 cells with 100  $\mu\text{g/ml}$  or higher PPS extract significantly increased the apoptosis rate, it is possible that the expression level of genes participated in PCD might have changed. As expected, the expression of the caspase family members, including caspase-3 $\beta$ , -7a, -8c, -9 $\alpha$  and caspase-10 variant 2, were significantly upregulated in PPS extract-treated group, except for caspase-7e, which was downregulated (Table IV). These upregulated caspases may well explain the increase of cell apoptosis rates.

Controversially, the inhibitors of apoptosis (IAP) protein, such as BIRC5 (Survivin) and XIAP, which inhibit caspase activity by directly binding to the enzyme, were also significantly upregulated (23,24). The overexpressed IAP family members in PPS group might also result from the cell anti-apoptotic response, which in turn antagonized the upregulated caspase activity. However, the enhanced XIAP activity in PPS-treated cells might not be antagonized by the second mitochondrial-derived activator of caspase (SMAC/DIABLO), because the expression of SMAC-encoded transcript variants was not upregulated. In addition, few cytosolic pro-apoptotic Bcl-2 family members that can promote cytochrome c release from mitochondria to cytosol, such as Bid, Bax, Bak, Bad, were upregulated. Taken together, the PPS extract-induced PCD in SKOV3 cells may initially be mediated by the extrinsic apoptosis signaling pathway.

Vascular endothelial growth factor (VEGF) has been suggested to be a key regulator of angiogenesis in many types of cancer, including ovarian cancer (25,26). Due to alternative splicing events, a number of VEGF mRNA variants have been detected and all variants are derived from the same locus in human genome (27). VEGF induces endothelial cell proliferation presumably through binding to the endothelial cell receptor. Elevated VEGF expression across a wide range of cancers has been observed, such as breast cancer and ovarian cancer (28,29). We found that VEGFA variant 3 and VEGFB-167 were moderately expressed in SKOV3 cell line.

However, the expression of these two variants was extremely downregulated in PPS extract-treated cell line, where FPKM values were almost undetectable (Table IV). This may be due to the enhanced von Hippel-Lindau tumor suppressor (VHL) expression in PPS extract-treated group. VHL possesses ubiquitin ligase E3 activity and is a well-characterized VEGF transcriptional repressor, which inhibits VEGF expression through direct ubiquitination and subsequent degradation of its transcription factor (30,31), hypoxia-inducible-factor (HIF). For other hormones, FGF10 was moderately expressed in SKOV3 cells, whereas its expression was not detected in PPS-treated cells. FGF10 has been proven to play an important role in tumor growth and invasion in a variety of cancers, such as breast cancer and pancreatic cancer (32,33). The downregulated FGF10 expression was consistent with the low proliferation ability in response to PPS extract treatment.

In conclusion, total extract obtained from PPS can inhibit epithelial derived ovarian cancer cell line SKOV3 proliferation and further induce apoptosis of these cells in a dose- and time-dependent manner, indicative of PPS extract component as a potential drug for ovarian cancer treatment in the future. Transcriptome sequencing and differential gene expression analysis revealed that the expression of a number of caspase family members were upregulated after PPS extract treatment. This result is mechanistically consistent with the obvious apoptotic phenotype and increased apoptosis rate of SKOV3 cells measured by flow cytometry.

In addition, many differentially expressed transcripts involved in the synthesis of steroid were significantly upregulated and over-represented, suggesting that cell apoptosis induced by extract of PPS might be accompanied by the increased biosynthesis of steroid, probably zymosterol. Therefore, this steroid could be considered as a novel regulator during ovarian cancer cell apoptosis after treated with PPS extract.

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