

MicroRNA-520c inhibits glioma cell migration and invasion by the suppression of transforming growth factor- β receptor type 2

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Abstract. Dysregulation of microRNAs (miRNAs) is actively involved in the development and progression of glioma. miR-520c was previously found to inhibit glioblastoma cell migration. However, the clinical significance of miR-520c and its biological function in glioma remain largely unknown. In the present study, we found that miR-520c expression in glioma tissues was significantly decreased compared to adjacent non-cancerous tissues. The low level of miR-520c was prominently correlated with advanced World Health Organization (WHO) grade and decreased overall survival of glioma patients. Overexpression of miR-520c in U251 cells significantly decreased the migration and invasion of the cancer cells, while miR-520c silencing promoted U87 cell migration and invasion *in vitro*. Mechanistically, miR-520c inversely regulated transforming growth factor- β receptor type 2 (TGFBRII) abundance in the glioma cells. Herein, TGFBRII was found to be a downstream target of miR-520c in glioma. Furthermore, an inverse correlation between TGFBRII and miR-520c expression was observed in the glioma cases. In contrast, restoration of TGFBRII expression abrogated the effects of miR-520c overexpression in U251 cells with increased cell migration and invasion. In addition, miR-520c overexpression blocked TGF- β 1-induced cell migration and invasion in U251 cells. Collectively, miR-520c may serve as a prognostic predictor and a therapeutic target for glioma patients.

Introduction

MicroRNAs (miRNAs), a group of small non-coding RNAs interacting with the 3'-untranslated region (3'-UTR) of targeted

mRNAs, inhibit the expression of target genes by contributing to the degradation or translational inhibition of target mRNAs (1). They have been found to be actively involved in various biological processes including cell proliferation, apoptosis, differentiation and movement (2,3). Emerging studies have shown that abnormal expression and function of miRNAs play important roles in the initiation and progression of human malignancies (4-6). In addition, miRNAs have been demonstrated to be promising biomarkers and therapeutic targets of glioma (7-9). Investigating the expression and biological function of miRNAs in glioma may contribute to the identification of novel biomarkers and therapeutic targets for glioma patients.

Recently, microRNA-520c (miR-520c) was found to play important roles in various human cancers including diffuse large B cell lymphoma (10), breast cancer (11), fibrosarcoma (12) and hepatocellular carcinoma (HCC) (13). A study on breast cancer showed that miR-520c promoted cancer cell migration and invasion *in vitro* and *in vivo* by suppression of CD44 (14). In addition, expression of miR-520c increased proliferation, migration and invasion of HCC cells *in vitro* (13). However, miR-520c abrogated both *in vitro* cell invasion and *in vivo* intravasation of highly invasive MDA-MB-231 cells (11), indicating that miR-520c functions as a tumor-suppressor in estrogen receptor negative breast cancer. Previously, Lu *et al* reported that miR-520c inhibits glioblastoma U87GM cell migration with decreased activation of proMMP2 *in vitro* (15). However, the clinical significance and biological role of miR-520c in glioma remain practically unknown.

In the present study, we confirmed that the expression of miR-520c was decreased in glioma tissues. Low expression of miR-520c was correlated with poor clinicopathological features and decreased survival of glioma patients. Our data showed that miR-520c inhibited the metastatic ability of glioma cells *in vitro*. Moreover, transforming growth factor- β receptor type 2 (TGFBRII) was identified as a downstream target of miR-520c in glioma.

Materials and methods

Clinical tissues. Fresh human glioma and adjacent non-cancerous tissues were obtained from 60 patients who underwent therapeutic removal of gliomas at Taihe Hospital, Hubei University of Medicine. Patients did not receive any

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chemotherapy or radiotherapy before surgical treatment. All clinical specimens were collected and used after obtaining informed consent from each patient enrolled in the present study. All specimens were stored in liquid nitrogen for further investigation. The protocol which involved clinical specimens in the present study was approved by the Research Ethics Committee of Hubei University of Medicine.

Cell culture and transfection. Human glioma cell lines including U87 and U251, and human kidney epithelial cell line (293T) were obtained from the American Type Culture Collection (ATCC; Manassas, VA, USA). These cells were cultured in Dulbecco's modified Eagle's medium (DMEM) with 10% fetal bovine serum (FBS), 100 U/ml penicillin and 100 mg/ml streptomycin. All cell cultures were maintained in a humidified cell incubator with 5% CO₂ at 37°C.

miR-520c and negative control (NC) mimics, miR-520c and NC inhibitors (anti-NC) were obtained from Genecopoeia (Guangzhou, China), and were then transfected into glioma cells with Lipofectamine 2000 following the manufacturer's protocol. Retroviral vectors pMMP-TGFBR2 were generated by inserting the TGFBR2 cDNA into pMMP. Retrovirus packaging and transduction were previously described (16).

Quantitative real-time RT-PCR (qRT-PCR). Total RNA from glioma cells was extracted by miRNeasy Mini kit (Qiagen, Hilden, Germany) and total RNA from glioma tissues was extracted with TRIzol reagent (Ambion, Thermo Scientific, Shanghai, China). miR-520c levels in these samples were assayed using TaqMan MicroRNA assays based on the manufacturer's protocol (Applied Biosystems, Carlsbad, CA, USA). Real-time PCR of TGFBR2 was performed using an UltraSYBR Mixture (CW0957; CWBIO, Beijing, China) and an LC480 PCR System (Roche, Indianapolis, IN, USA). The primers for miR-520c and U6, TGFBR2 and GAPDH were obtained from Genecopoeia. The expression levels of mRNA and miR-520c were normalized to the endogenous controls GAPDH and U6 respectively, using the 2^{-ΔΔCt} method.

Luciferase reporter assay. To investigate whether miR-520c interacted with the 3'-UTRs of TGFBR2, wild-type (wt) 3'-UTR of TGFBR2 predicted to interact with miR-520c was amplified. Then, the wt 3'-UTR of TGFBR2 and the corresponding vectors (NC, miR-520c mimic, anti-NC, miR-520c inhibitor or mutant miR-520c) were co-transfected into 293T cells by Lipofectamine 2000. Forty-eight hours after co-transfection, the cells were lysed and assayed using Dual-Luciferase[®] Reporter Assay kit (Promega, Madison, WI, USA) based on the manufacturer's instructions.

Wound healing assay. Glioma cells transfected with the corresponding vectors were seeded in 6-well plates to form a single confluent cell layer. Wounds were made in the confluent cell layer using 100-μl tips. Following wound scratching (0 and 12 h), the width of the wounds were photographed with a phase-contrast microscope.

Migration and invasion assays. The migratory and invasive abilities of glioma cells were evaluated with Transwell chambers (BD Biosciences, Franklin Lakes, NJ, USA). Glioma

cells (5-10x10⁴) suspended in 100 μl serum-free medium were seeded into the upper chamber, and the lower chamber was filled with 20% FBS to induce glioma cell migration or invasion through the membrane. Matrigel (1:6 dilution; Becton-Dickinson Labware, Bedford, MA, USA) was added to the upper chamber for the invasion assay. Twenty-four hours later, cells that migrated or invaded the Transwell membrane were stained with crystal violet and the number of cells was counted under a microscope. TGF-β1 (2.5 ng/ml; R&D Systems, Minneapolis, MN, USA) was used for inducing glioma cell migration and invasion.

Western blotting. Before protein extraction, glioma cells were washed with phosphate-buffered saline (PBS) to remove the culture media. Cellular proteins were obtained from glioma cells using RIPA lysis buffer and the protein concentrations were assessed using the BSA method. Cellular proteins (30 μg) were separated by 10% SDS-PAGE, and were transferred to polyvinylidene fluoride (PVDF) membranes (Millipore, Bedford, MA, USA). After blocking with 5% non-fat milk at room temperature for 1 h, the membranes were probed with primary antibodies at 4°C overnight. Then, the membranes were incubated with the corresponding secondary antibodies at room temperature for 2 h. The primary antibodies used in the present study included: TGFBR2 (K105) (#3713; Cell Signaling Technology, Danvers, MA, USA), CD44 (WL0140; Wanleibio, Shanghai, China) and GAPDH (G8140; US Biological, Swampscott, MA, USA).

Immunohistochemistry (IHC). Before IHC staining, glioma tissues were fixed with 4% formalin and embedded with paraffin. Then, the embedded tissues were cut into 4-μm thick sections and visualized by IHC staining following the standard protocol in order to evaluate the expression level of TGFBR2 (Cell Signaling Technology) in glioma tissues.

Statistical analysis. All quantitative data are presented as the mean ± the standard error of the mean (SEM). Statistical analyses including Pearson Chi-squared test, a two-tailed Student's t-test, Kaplan-Meier method, the log-rank test and Spearman's correlation analysis were performed with GraphPad Prism 6 (GraphPad Software Inc., San Diego, CA, USA). P<0.05 was considered to indicate a statistically significant result.

Results

Clinical significance of miR-520c expression in glioma specimens. To examine the expression status of miR-520c in glioma, qRT-PCR was performed for 60 pairs of glioma and adjacent non-tumor tissues. Our results showed that glioma tissues had significant decreased expression levels of miR-520c compared with adjacent non-tumor tissues (P<0.05; Fig. 1A). To clarify the clinical significance and prognostic value of miR-520c in glioma, all patients were divided into two groups (miR-520c low-expressing and miR-520c high-expressing groups) according to the median level of miR-520c expression. As shown in Table I, patients with low expression of miR-520c had advanced World Health Organization (WHO) stage (P=0.024). Furthermore, Kaplan-Meier analysis showed that patients with low expression of miR-520c exhibited

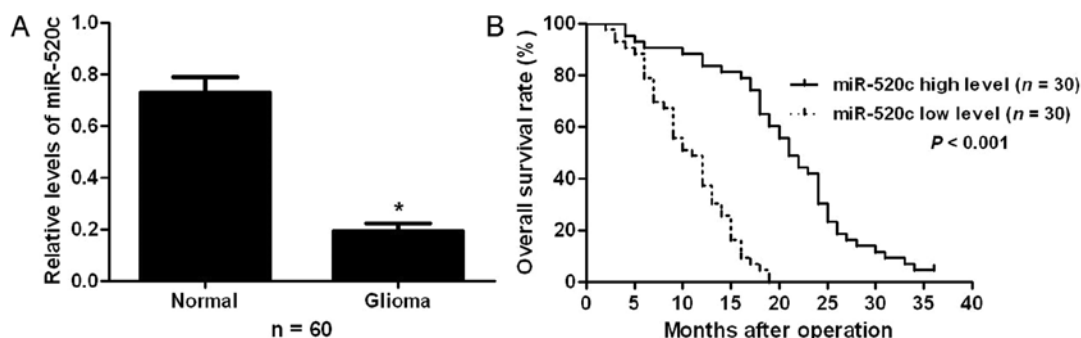


Figure 1. Expression and prognostic value of miR-520c in glioma. (A) Significantly decreased expression of miR-520c was observed in glioma tissues compared to adjacent non-cancerous tissues; * $P < 0.05$. (B) Compared with the patients with high miR-520c expression level, patients with low miR-520c expression had significantly decreased overall survival.

Table I. Correlation between the clinicopathological characteristics and expression of miR-520c in gliomas.

Characteristics	Total no. of pts. (n=60)	No. of patients		P-value
		Low miR-520c	High miR-520c	
Age (years)				0.301
<50	28	12	16	
≥ 50	32	18	14	
Gender				0.592
Male	38	18	20	
Female	22	12	10	
Tumor size (cm)				0.417
<5	21	9	12	
≥ 5	39	21	18	
Histologic tumor type				0.698
Astrocytic	42	22	20	
Oligodendroglial	7	3	4	
Oligoastrocytic	11	5	6	
WHO grade				0.024 ^a
I+II	18	5	13	
III+IV	42	25	17	

WHO, World Health Organization; pts., patients. ^aStatistically significant.

a significantly decreased overall survival rate ($P < 0.001$; Fig. 1B). These results indicate that miR-520c probably plays an oncogenic role in glioma.

miR-520c inhibits the migration and invasion of glioma cells. Next, we explored whether miR-520c modulates the migration and invasion of glioma cells. Transfection of miR-520c mimic into U251 cells significantly increased the expression level of miR-520c ($P < 0.05$; Fig. 2A). The wound healing assays showed that the migration of U251 cells was significantly decreased after miR-520c overexpression ($P < 0.05$; Fig. 2B). In addition, Transwell assays demonstrated that overexpression of miR-520c prominently suppressed the migration and invasion

of U251 cells ($P < 0.05$, respectively; Fig. 2C). In contrast, miR-520c inhibitor significantly decreased the expression level of miR-520c in U87 cells ($P < 0.05$; Fig. 3A). Subsequently, miR-520c silencing significantly facilitated the migration and invasion of U87 cells ($P < 0.05$, respectively; Fig. 3B and C). Thus, miR-520c reversed the migratory and invasive abilities of the glioma cells.

TGFBRII is a downstream target of miR-520c. To disclose the underlying molecular mechanisms of the biological function of miR-520c in glioma cells, TargetScanHuman 7.1 (<http://www.targetscan.org>) was used to search for the downstream target of miR-520c. TGFBRII, a critical regulator of the TGF- β pathway in glioma (17), was recognized as a potential downstream target of miR-520c. U251 cells that were transduced with corresponding vectors were subjected to immunoblotting for TGFBRII and CD44, a known downstream target of miR-520c (14). Notably, miR-520c overexpression decreased the levels of TGFBRII and CD44 protein, while miR-520c silencing increased the expression of these proteins (Fig. 4A). Furthermore, qRT-PCR also confirmed that miR-520c overexpression downregulated the mRNA levels of CD44 and TGFBRII ($P < 0.05$; Fig. 4B). However, miR-520c knockdown showed no significant effect on mRNA expression (Fig. 4B). As shown in Fig. 4C, the 3'-UTR of TGFBRII contained two putative binding sites for miR-520c. Subsequently, we performed luciferase assay to investigate whether miR-520c could bind to the putative binding sites in the 3'-UTR of TGFBRII. Alteration of miR-520c inversely regulated the luciferase activity of TGFBRII 3'-UTR ($P < 0.05$; Fig. 4D), while mutant miR-520c did not exhibit any influence on the luciferase activity of TGFBRII 3'-UTR (Fig. 4D). Therefore, these data indicate that TGFBRII is a direct downstream target of miR-520c in glioma.

An inverse correlation between TGFBRII and miR-520c is observed in glioma specimens. Glioma and non-tumor tissues were subjected to qRT-PCR for TGFBRII mRNA. Our data indicated that the expression levels of TGFBRII mRNA in glioma tissues were significantly higher than those in adjacent non-cancerous tissues ($P < 0.05$; Fig. 5A). Spearman's correlation analysis indicated that miR-520c was strongly correlated with TGFBRII mRNA expression in the glioma specimens ($r = -0.910$, $P < 0.001$; Fig. 5B). Moreover,

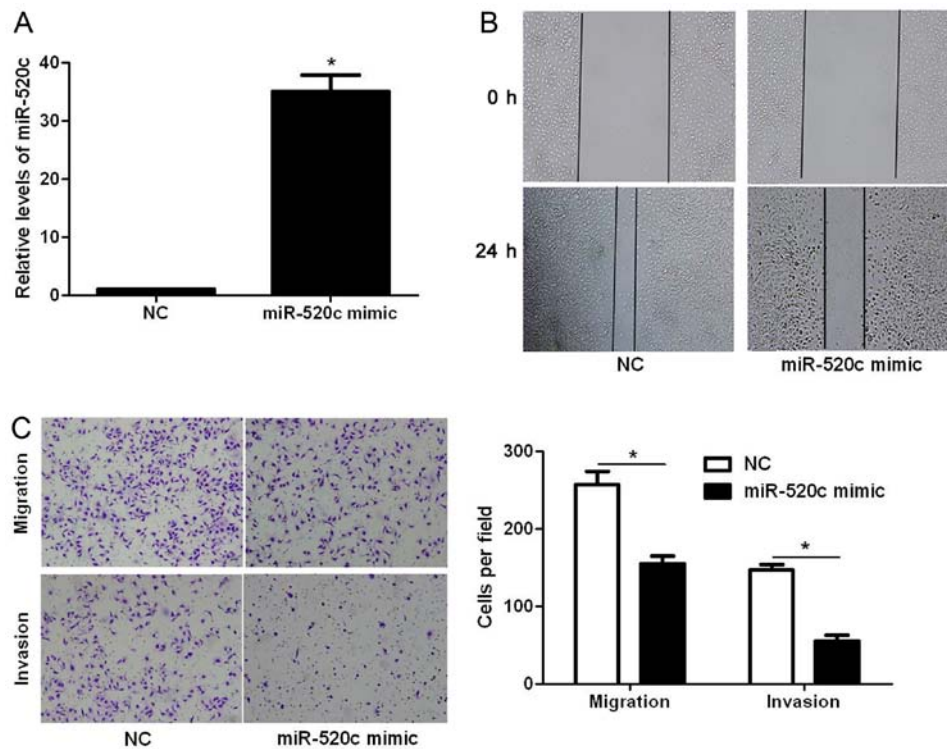


Figure 2. miR-520c mimic inhibits the migration and invasion of U251 cells. (A) miR-520c mimic significantly increased the level of miR-520c expression in U251 cells; * $P < 0.05$. (B) miR-520c overexpression reversed the migration of U251 cells. (C) miR-520c overexpression inhibited the migration and invasion of U251 cells; * $P < 0.05$.

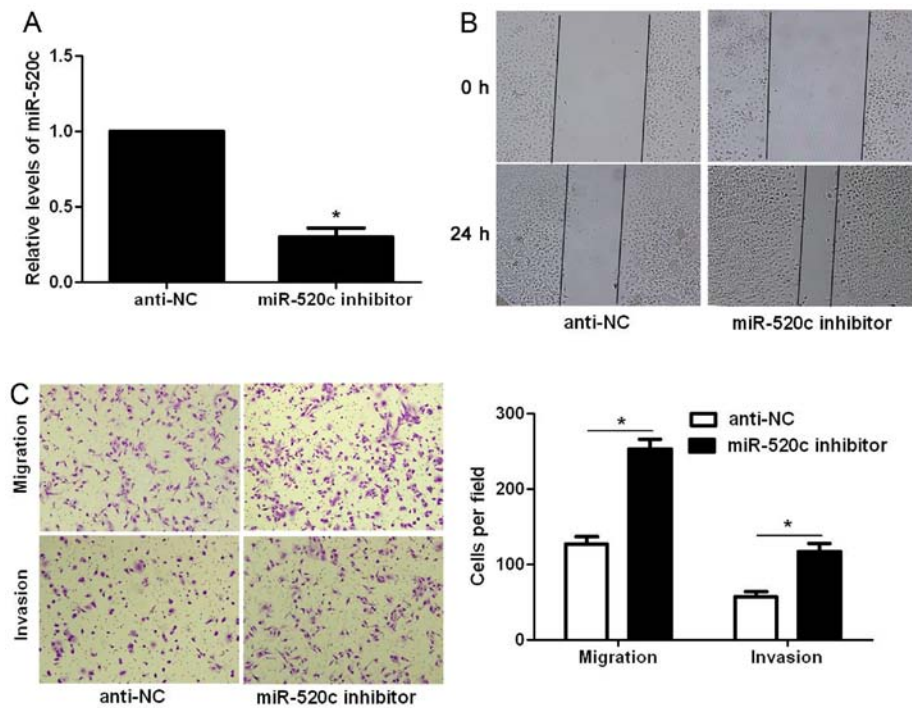


Figure 3. miR-520c inhibitor facilitates the migration and invasion of U87 cells. (A) miR-520c inhibitor significantly decreased the expression of miR-520c in U87 cells; * $P < 0.05$. (B) miR-520c knockdown significantly increased the migration of U87 cells. (C) miR-520c silencing significantly promoted the migration and invasion of U87 cells; * $P < 0.05$.

immunohistochemical staining indicated that miR-520c high-expressing tumors showed weak staining of TGFBR2, while miR-520c low-expressing tumors showed strong staining of TGFBR2 ($P < 0.05$; Fig. 5C and D).

TGFBR2 mediates the functions of miR-520c in glioma cells. Next, U251 cells with miR-520c overexpression were infected with empty vector (EV) or TGFBR2 retroviruses. Restoration of TGFBR2 expression was confirmed by western

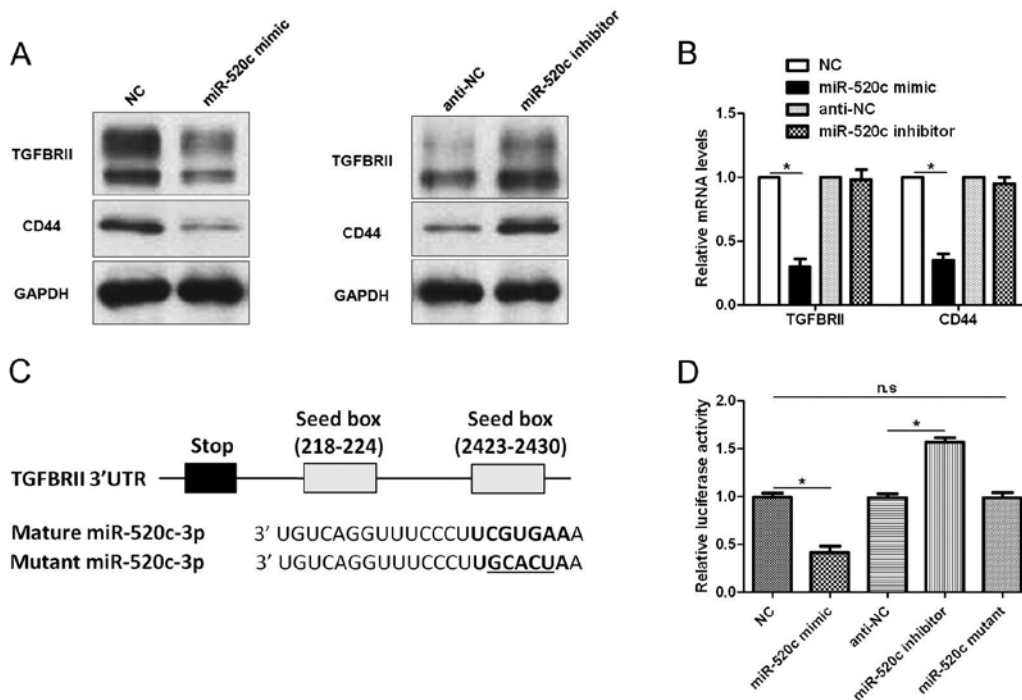


Figure 4. miR-520c binds to the complementary sequence in the 3'-UTR of TGFBR2. (A) miR-520c overexpression reduced while miR-520c silencing increased the level of TGFBR2 and CD44 protein in U251 and U87 cells, respectively. (B) miR-520c overexpression decreased the levels of TGFBR2 and CD44 mRNA while miR-520c knockdown exhibited no effect on mRNA levels; *P<0.05. (C) The binding sites for miR-520c in 3'-UTR of TGFBR2. (D) Overexpression of miR-520c decreased and miR-520c silencing increased the luciferase activity of the 3'-UTR of TGFBR2, while mutant miR-520c had no effect on the luciferase activity of 3'-UTR of TGFBR2 in 293T cells; *P<0.05. n.s., not significantly different.

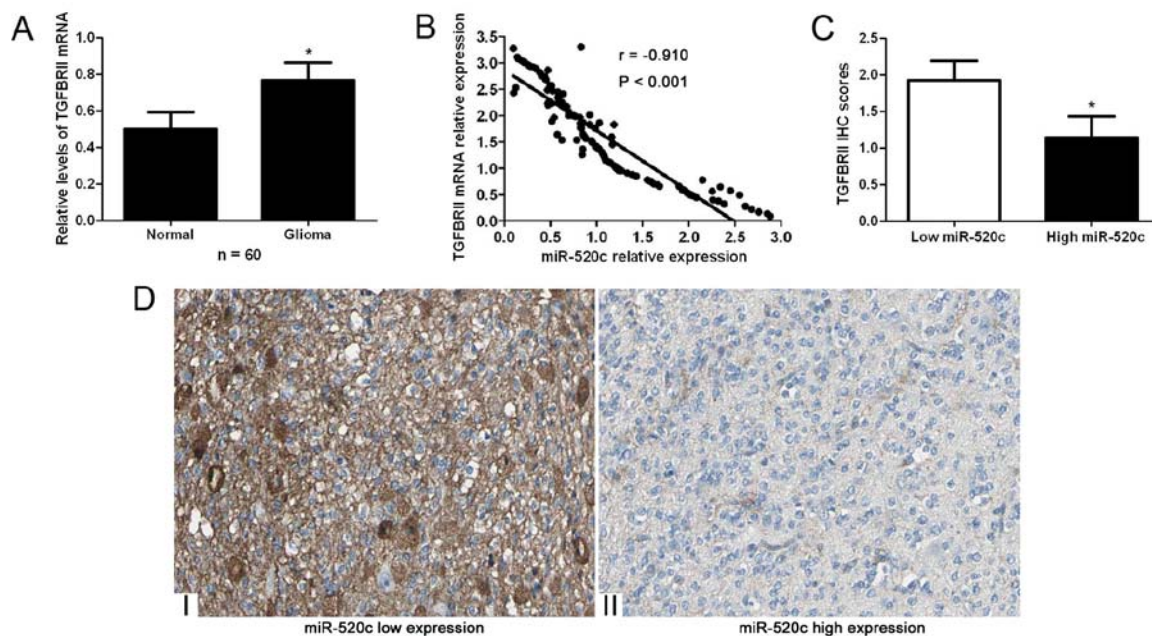


Figure 5. Correlation between TGFBR2 and miR-520c in glioma tissues. (A) The expression of TGFBR2 mRNA was upregulated in glioma compared to non-tumor tissues; *P<0.05. (B) An inverse correlation between TGFBR2 and miR-520c expression was observed in glioma tissues. (C) Quantitative data arising from IHC scores indicated that the expression of TGFBR2 in miR-520c low-expressing tumors was evidently higher than that in miR-520c high-expressing cases; *P<0.05. (D) Representative immunohistochemical staining of TGFBR2 in glioma tissues. miR-520c high-expressing tumors showed weak staining of TGFBR2 (II), while miR-520c low-expressing tumors showed strong staining of TGFBR2 (I).

blotting (P<0.05; Fig. 6A). Wound healing and Transwell assays indicated that TGFBR2 restoration abrogated the effects of miR-520c overexpression in U251 cells with increased migratory and invasive abilities of glioma cells (P<0.05, respectively;

Fig. 6B and C). Since TGFBR2 is a target of miR-520c, and TGFBR2 is an important component in TGF-β1-induced migration and invasion, miR-520c-overexpressing U251 cells were treated with TGF-β1 (2 μg/ml). As expected, the induction

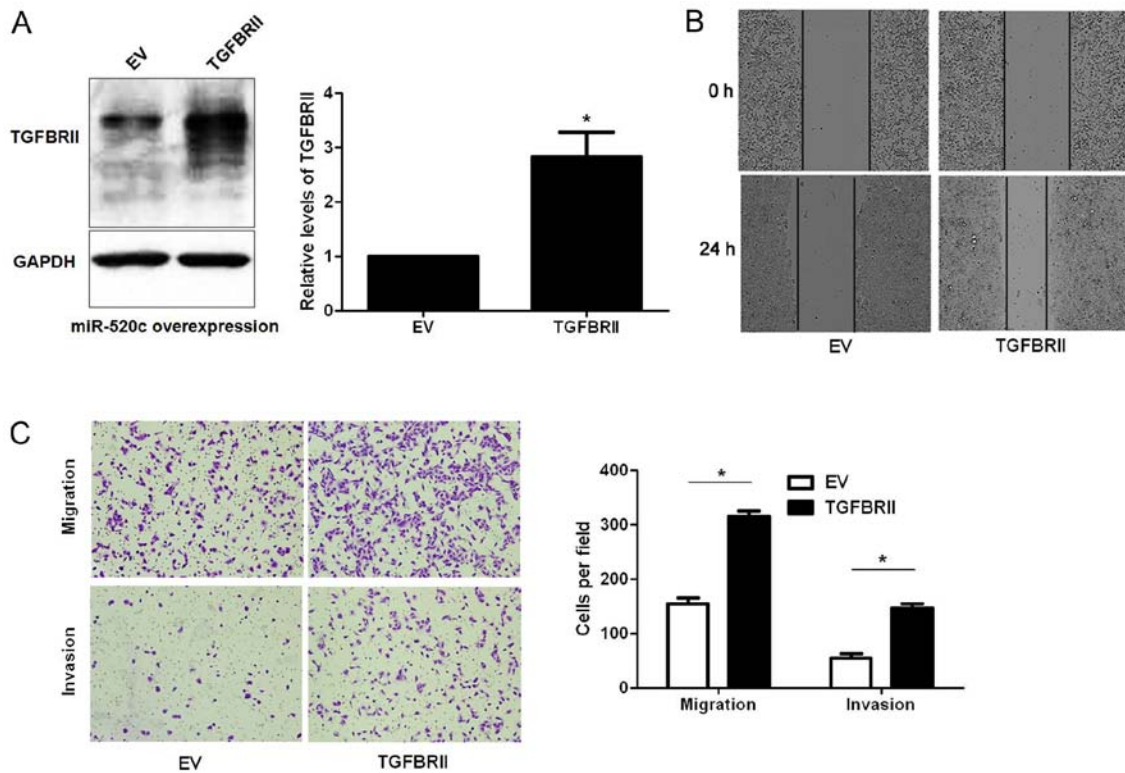


Figure 6. TGFBR2 restoration reverses the effects of miR-520c overexpression in U251 cells. (A) miR-520c-overexpressing U251 cells that were transfected with empty vector (EV) or TGFBR2 retroviruses were subjected to immunoblotting for TGFBR2; * $P < 0.05$. (B) TGFBR2 restoration facilitated the migration of miR-520c-overexpressing U251 cells. (C) TGFBR2 restoration promoted the migration and invasion of miR-520c-overexpressing U251 cells; * $P < 0.05$.

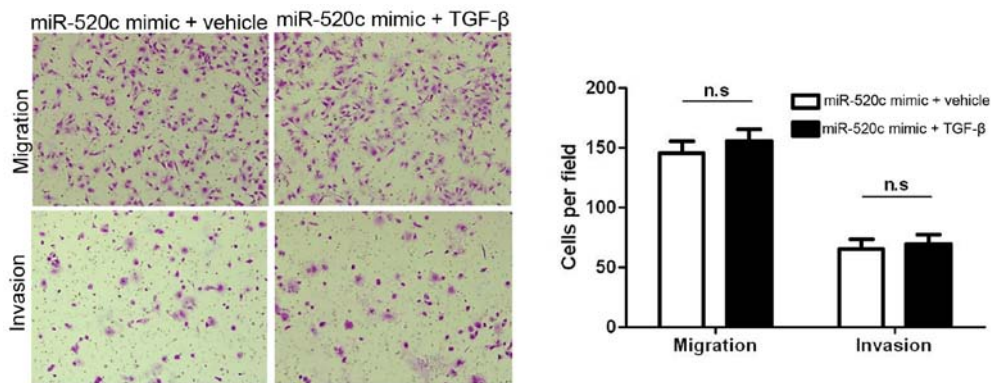


Figure 7. miR-520c overexpression blocks the pro-metastatic effects of TGF- β 1 on U251 cells. U251 cells that were transfected with miR-520c mimics were treated with vehicle and TGF- β 1 (2.5 ng/ml). Transwell assays indicated that miR-520c overexpression blocked TGF- β 1-induced migration and invasion in U251 cells. n.s., not significantly different.

of TGF- β 1 showed no effect on miR-520c-overexpressing U251 cells (Fig. 7). Collectively, TGFBR2 is not only a target but also a functional mediator of miR-520c.

Discussion

Emerging evidence has confirmed that miRNAs are actively involved in the pathogenesis of glioma (18). In addition, miRNAs have been found to be critical regulators of the metastasis and epithelial-mesenchymal transition of glioma cells (19). Due to the important roles of miRNAs in glioma, miRNAs have been proposed as promising biomarkers and therapeutic targets of glioma (20). In the present study, miR-520c was found to be

significantly downregulated in glioma tissues. In addition, the low expression of miR-520c in glioma tissues conferred advanced WHO stage. More importantly, low expression of miR-520c was correlated with decreased overall survival of glioma patients. Therefore, miR-520c plays a tumor-suppressive role in glioma and potentially serves as a promising biomarker for the prognosis of glioma patients.

Systemic metastasis is the cause for the unsatisfactory prognosis of glioma patients (21). Increased migratory and invasive abilities of glioma cells underlie the systemic metastasis of glioma (22). Therefore, it is of great importance to elucidate the molecular mechanisms involved in the metastasis of glioma cells. In the present study, we found that

miR-520c inhibited the migration and invasion of glioma cells *in vitro*, suggesting that miR-520c exerted an antimetastatic role in glioma. The TGF- β signaling pathway has been reported to be implicated in tumor metastasis in various human malignancies (23). The fundamental role of the TGF- β pathway in promoting cancer progression in multiple stages of the metastatic process, including epithelial-to-mesenchymal transition (EMT), is also becoming increasingly clear (23). Notably, the TGF- β -SMAD pathway confers poor prognosis to glioma patients and promotes cell proliferation (24). TGFBR2 is a critical component of the TGF- β signaling pathway and TGFBR2 silencing blocked the effects of TGF- β 1 in glioma cells (25). In the present study, we found that miR-520c negatively regulated the abundance of TGFBR2 in glioma cells. In addition, the expression of TGFBR2 in glioma tissues was inversely correlated with miR-520c expression. Moreover, we found that miR-520c directly interacted with the 3'-UTR of TGFBR2 using a luciferase reporter assay. These data indicate that TGFBR2 is a direct downstream target of miR-520c in glioma. Importantly, restoration of TGFBR2 abolished the effects of miR-520c overexpression in glioma cells and miR-520c overexpression blocked TGF- β 1-induced migration and invasion of glioma cells. Thus, TGFBR2 is a functional mediator of miR-520c in glioma.

In conclusion, the present study demonstrated that miR-520c expression was significantly decreased in glioma tissues. Decreased expression of miR-520c was found to be correlated with adverse clinical features and poor prognosis of glioma patients. In addition, miR-520c inhibited migration and invasion of glioma cells. Furthermore, TGFBR2 is a direct functional target of miR-520c in glioma. These data may result in the discovery of therapeutic candidates of glioma.

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