

Endogenous Leu332Gln mutation in p53 disrupts the tetramerization ability in a canine mammary gland tumor cell line

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Abstract. Mutations in the *p53* gene are associated with more than half of all human cancers. These mutations often cause a disruption of the tumor-suppressor function of p53 and induce genomic instabilities. Wild-type p53 requires tetramerization to function as an initiator of cell cycle arrest and apoptosis. Although alterations in p53 tetramerization caused by mutation have been well studied, there are few cell lines containing an endogenous mutation in the tetramerization domain of p53. Here, we report the discovery of a canine mammary gland tumor cell line CTB-m2, which contains the Leu332Gln (L332Q) mutation corresponding to Leu344 in the tetramerization domain of human p53. Although CTB-m2 cells are genetically heterozygous for the Leu332Gln mutation, the mutant mRNA was almost exclusively expressed.

CTB-m2 cells showed enhanced cell proliferation compared to wild-type p53-expressing CTB-m cells of the same lineage. A p53 tetramerization reporter assay showed that the ability of the p53 mutant to form tetramers was significantly lower than that of wild-type p53. An immunoblot analysis of cross-linked p53 oligomerized forms demonstrated that the L332Q mutant lacked the ability to form tetramers but retained the ability to form dimers. These data suggest that the p53 mutant cell line CTB-m2 could be a useful tool for analyzing the precise tetramerization mechanisms of p53 and verifying the effects of therapeutic agents against tumors expressing p53 mutants that lack the ability to tetramerize.

Introduction

The tumor suppressor protein p53 plays a central role in the cell cycle and maintenance of genomic integrity (1-3). The p53 gene is frequently mutated in human cancers, and germline mutations are the underlying cause of Li-Fraumeni syndrome (4,5). The human p53 protein is comprised of an N-terminal transactivation domain, a proline-rich region, a structured DNA-binding domain connected to a tetramerization domain (TD) via a flexible linker, and a C-terminal regulatory domain (6). The p53 protein is active as a homotetramer which adopts a dimer of dimer topology (7). The TD in human p53 (amino acid residues 326-356) exhibits a dihedral symmetry of dimers (8). Two monomers interact with the β -strands (Glu326-Arg333) to form a dimer, and the two dimers then interact as a α -helix bundle (Arg335-Gly356) to form the tetramer (9). Nine residues in the TD in human p53 (Phe328, Leu330, Ile332, Arg337, Phe338, Met340, Phe341, Leu344 and Leu348) are critical determinants in stabilizing the p53 tetramer (10). Leu344 mutants lacking the ability to

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dimerize (L344P), or tetramerize (L344R), have been previously reported (11), and notably, germline L344P mutation has been found in a family with Li-Fraumeni syndrome (12). Leu344 is located in the α -helix which forms the hydrophobic core of the tetramer interface.

Although, numerous cell lines containing mutated p53 genes have been established from cancer tissues, there are few cell lines which contain a mutated p53 tetramerization domain (13,14). Uyama *et al* established and characterized four pairs of canine mammary tumor cell lines derived from either primary and metastatic origin (15). One of these cell lines, CTB-m, was passaged 50 times in our laboratory to obtain a line containing a spontaneous L332Q mutation in p53 (corresponding to human L344); we designated this new cell line as CTB-m2. In this study, we assessed the oligomerization abilities of this canine p53 L332Q mutant and performed a tetramerization reporter assay.

Materials and methods

Cell culture and transfection. The CIP-p, CIP-m, and CTB-m cell lines were kindly provided by Dr N. Sasaki of the University of Tokyo. CIP-p, CIP-m, CTB-m and CTB-m2 cells were cultured in RPMI-1640 medium (Wako, Osaka, Japan) and HeLa cells (ATCC, Rockville, MD, USA) were cultured in DMEM (Wako). All media were supplemented with 10% fetal bovine serum (FBS) and penicillin-streptomycin (Wako) and cells were cultured under a humidified atmosphere of 5% CO₂ at 37°C. Cell transfection was performed using FuGENE HD (Promega, Madison, WI, USA), as previously reported (16).

Establishment of the CTB-m2 cell line. CTB-m2 cells were spontaneously established from the CTB-m line after 50 passages in our laboratory.

Sequencing analysis and cloning of the canine p53 gene. Total RNA from CTB-m and CTB-m2 cells was obtained using a High Pure RNA Tissue Kit (Roche Diagnostics, Mannheim, Germany), and reverse transcription conducted using a Reverse Transcription System (Promega) according to the manufacturer's protocol. Full length canine p53 cDNA was amplified by PCR using the following primers: 5'-CTCGAGGACCACCATGCAAGAGCCACAGTCAGAGC-3' and 5'-GAATTCCCGTCTGAGTCGGGCCCTTCTCTC-3'. Following digestion with *Xho*I and *Eco*RI, the PCR product was cloned into the *Xho*I and *Eco*RI sites of the plasmid pMACS KK HA(C) (Miltenyi Biotec, Bergisch Gladbach, Germany). Samples containing genomic DNA from cells were prepared using the ZR Genome DNATM Tissue Miniprep (Zymo Research, Irvine, CA, USA). Exon 10-11 of p53 containing the sequence encoding L332 was amplified by PCR using the following primers; 5'-GCACTTACACCTTAGTCTGAG-3', and 5'-CGGAATAGGTGTGCTCAAGC-3'. The amplified canine p53 cDNA and exon 10-11 were directly sequenced using an ABI 3730 system (Applied Biosystems, Waltham, MA, USA).

Cell proliferation analysis. A total of 20,000 cells were plated into each well of a 24-well plate, and counted every 24 h up until 72 h using a TC20™ Automated Cell Counter (Bio-Rad, Hercules, CA, USA).

p53 tetramerization reporter assay. The p53 response element in p21WAF (GenBank accession no. U24170, nucleotides 2303-2321) and human cytomegalovirus in the pTet-splice vector (nucleotides 318-446; Clontech Laboratories Inc., Palo Alto, CA, USA) (17) was inserted between the *Xho*I and *Hind*III sites in pNL1.1[Nluc] (p53RE-pNL1.1) (Promega). To measure the endogenous p53 tetramerization activities, CTB-m and CTB-m2 cells were transfected with the p53RE-pNL1.1 reporter plasmid. The cells were harvested 48 h after transfection, and luciferase activity was measured using the Nano-Glo Dual-Luciferase Reporter Assay System (Promega). The luciferase activity was normalized to the value of the luc2 activity from co-transfected pGL4.51[luc2/CMV/Neo] (Promega). To measure the exogenous p53 tetramerization activities, HeLa cells were co-transfected with p53RE-pNL1.1 and either WT canine p53 or its L332Q mutant in pMACS KK HA (C).

Sample preparation and cross-linking procedure. HeLa cells in 6-well plates were transfected with either WT or L332Q p53 expression plasmids (1 μ g/well). After 48 h of transfection, the cells were treated with doxorubicin at a final concentration of 0.5 μ M for 6 h. Cells were lysed with mammalian lysis buffer (Promega) supplemented with a protease inhibitor cocktail (Promega). After lysis, the samples were centrifuged (15,000 \times g for 15 min at 4°C) to obtain the supernatant. Total protein levels were measured using BCA (Nacalai Tesque, Kyoto, Japan). Equal amounts of proteins (100 μ g/condition) were incubated with glutaraldehyde at different concentrations (0, 0.02 or 0.04%) and incubated on ice for 30 min. To make a working solution of glutaraldehyde, the commercially available 25% glutaraldehyde solution was diluted in PBS and discarded after use. To quench the reaction, sample buffer was added to obtain the following final concentrations: 250 mM Tris-HCl, pH 8.5; 2% lithium dodecyl sulfate; 100 mM DTT; 0.4 mM EDTA; 10% glycerol; 0.2 mM bromophenol blue. Samples were then separated by polyacrylamide gel electrophoresis (18).

Electrophoresis and immunoblotting. Samples were resolved on a 5-12.5% Tris-glycine polyacrylamide gradient gel using a Perfect Cell B from DRC (Tokyo, Japan) (ΔV = 150 V, 50 min at 25°C), transferred to a PVDF membrane using a standard semi-dry apparatus from Bio-Rad (ΔV = 15 V, 1 h at 25°C), and probed with an anti-HA specific mouse monoclonal antibody (M180-3, MBL, Aichi, Japan), and then incubated with HRP-conjugated secondary anti-mouse IgG (7076; Cell Signaling Technology, Beverly, MA, USA). Immunoreactive bands were visualized with a gel documentation system (LAS-4000 mini; Fujifilm, Tokyo, Japan).

Crystal structure modeling. We retrieved the crystal structure of the human p53 tetramerization domain from the Research Collaboratory for Structural Bioinformatics Protein Data Bank at <http://www.rcsb.org/> (PDB ID: 3FAK) and analyzed it using the University of California, San Francisco (UCSF) Chimera software (<http://www.cgl.ucsf.edu/chimera/>) (18).

Microsatellite genotyping of canine cell lines by polymerase chain reaction (PCR) and amplified fragment length

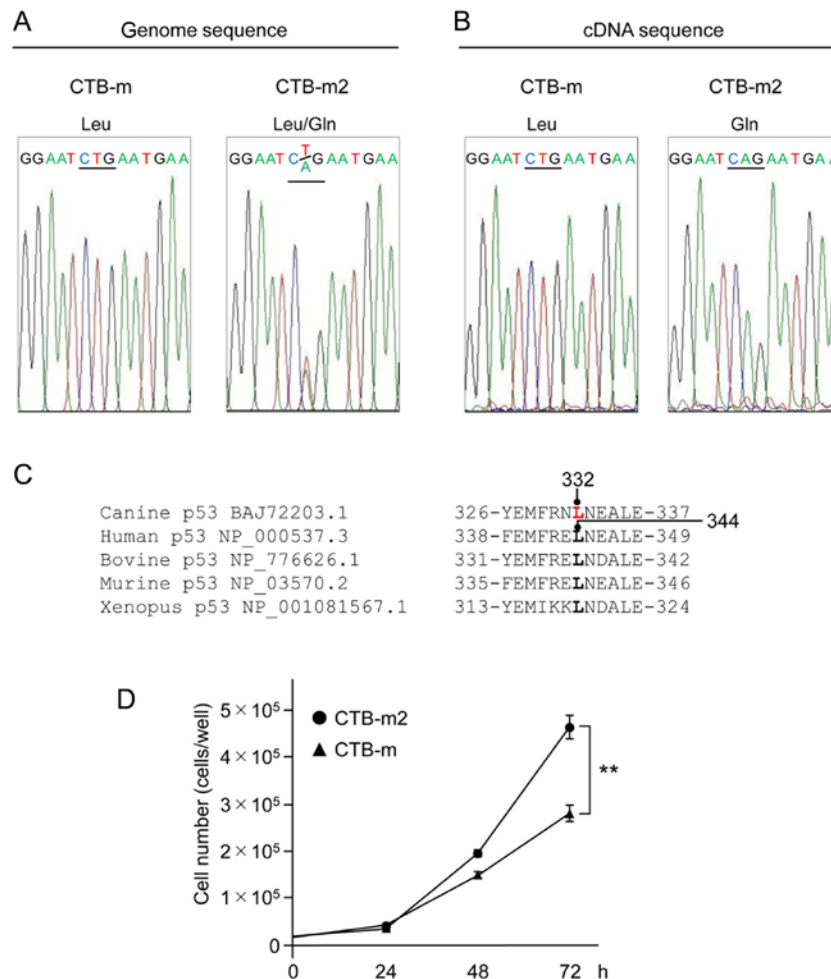


Figure 1. Mutation and cell proliferation analysis of the canine p53 L332Q mutant cell line CTB-m2. (A) The electropherogram shows the nucleotide substitutions at position c.1151T>A in the canine *p53* gene (GenBank accession: AB537893.1) amplified from CTB-m2 genomic DNA (right) and the same position amplified from CTB-m genomic DNA (left). In each case, the nucleotide sequences 1150-1152 encoding Leu332 (CTB-m) or Leu/Gln (CTB-m2) are underlined. (B) Homozygous electropherograms of the *p53* gene amplified from either CTB-m or CTBm-2 cDNAs. In each case, the nucleotide sequences 1150-1152 encoding Leu332 (CTB-m) or Gln332 (CTB-m2) are underlined. (C) Amino acid sequence alignments of canine, human bovine, murine and *Xenopus* p53 proteins (GenBank accession: BAJ72203.1; NP_000537.3; NP_776626.1; NP_03570.2; NP_001081567.1). Canine p53 Leu 332 is homologous to human p53 Leu 344 and other species. The bold font indicates the homology positions. (D) Cell proliferation was assessed in CTB-m or CTB-m2 cells every 24 h after passage using a TC20™ Automated Cell Counter. Data were obtained from three independent experiments. Values are shown as mean \pm SD. **P<0.01 (Student's t-test).

polymorphism analysis. Thirteen microsatellite loci were compared between the CTB-m and CTB-m2 or CIP-p and CIP-m cell lines by PCR and amplified fragment length polymorphism analysis in a genetic diagnosis laboratory (<http://www.kahotechno.co.jp/clinic/index.html>, Kahotechno, Fukuoka, Japan).

Statistical analysis. Data are expressed as mean \pm standard deviation (SD). Differences between two groups were analyzed using the Student's t-test. Analysis of variance (ANOVA) with a Tukey's post hoc test was used when multiple comparisons were required. Significance was assessed at the 0.05 (or lower) level for all tests.

Results

p53 in CTB-m2 cells contains a heterozygous mutation at L332Q. Sequencing analysis of genomic *p53* from the canine mammary gland tumor cell line CTB-m2 showed the presence of a heterozygous missense mutation L332Q, compared to

CTB-m cells, which expressed homozygous L332 (Fig. 1A). However, the sequence of *p53* mRNA derived from CTB-m2 cells showed almost exclusively the mutated allele (L332Q) (Fig. 1B). A comparison of a part of the tetramerization domain of the canine p53 protein (GenBank accession: BAJ72203.1) with human, bovine, murine and *Xenopus* p53 protein (NP_000537.3; NP_776626.1; NP_03570.2; NP_001081567.1) showed that canine Leu332 corresponds to human Leu344, and that this sequence is completely conserved in other species (Fig. 1C).

Cell viability of CTB-m2 cells is higher than CTB-m. A cell proliferation analysis was performed using an automatic cell counter every 24 h after cultivation. Seventy-two hours after cultivation, CTB-m2 cell proliferation was found to be significantly higher than CTB-m cell proliferation (Fig. 1D).

L332Q mutant in p53 reduces the tetramerization ability compared with WT p53. The p53 tetramer response element, along with the human cytomegalovirus minimal promoter sequence, were cloned upstream of the sequence encoding

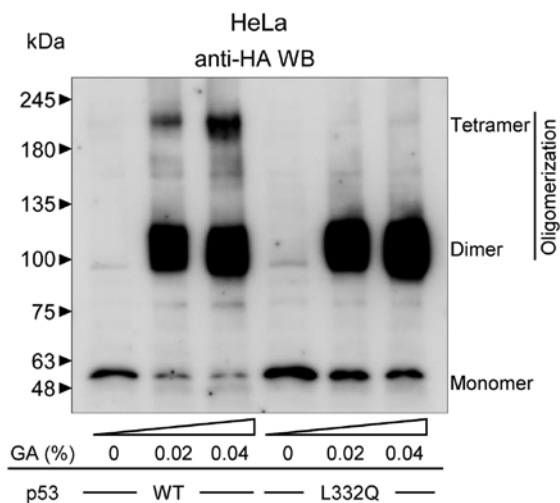


Figure 3. Glutaraldehyde cross-linking analysis of WT and the p53 L332Q mutant. HeLa cells were transfected with plasmids encoding either HA-tagged canine WT or L332Q p53. Cell lysates from the transfected cells were incubated on ice in the presence of the indicated glutaraldehyde (GA) concentrations for 30 min, following which the lysates were separated by polyacrylamide gel electrophoresis and immunoblotted with an anti-HA antibody. Both p53 (WT and L332Q) showed dimer formation after being treated with GA, but tetramerization was not observed for the p53 L332Q mutant. WT, wild-type.

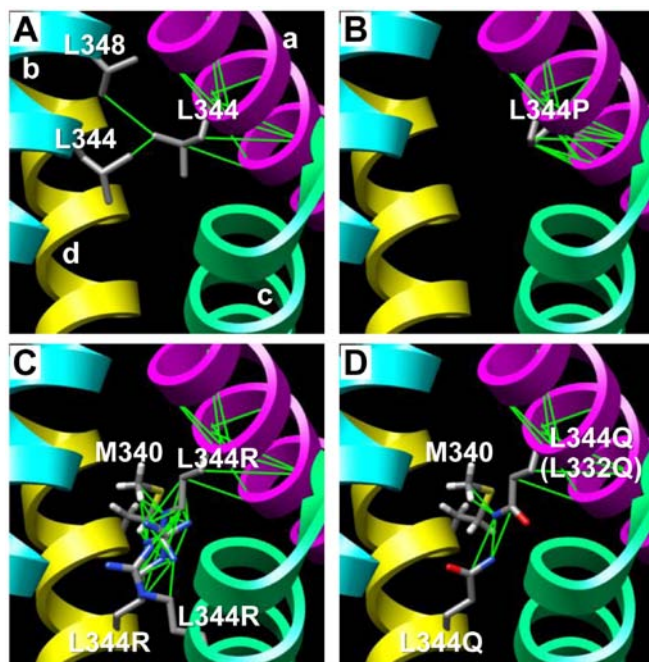


Figure 4. Effect of the L332Q mutation of canine p53 on its oligomerization. The crystal structure of the human p53 tetramerization domain (PDB ID: 3FAK) was visualized using the Chimera software from UCSF. L344 (corresponding to L332 in canine p53) was mutated to Pro (P), Arg (R), or Gln (Q) residues using the Dunbrack backbone-dependent rotamer library algorithm in the Chimera software from UCSF. Contacts between the residue at position 344 in chain a were calculated and are depicted for (A) L344, (B) L344P, (C) L344R, and (D) L344Q. Solid green lines signify stable contacts, as determined by the Chimera program.

Microsatellite analysis of cell lines established from the same origins. Thirteen microsatellite loci were compared among the CTB or CIP canine mammary gland tumor cell lines.

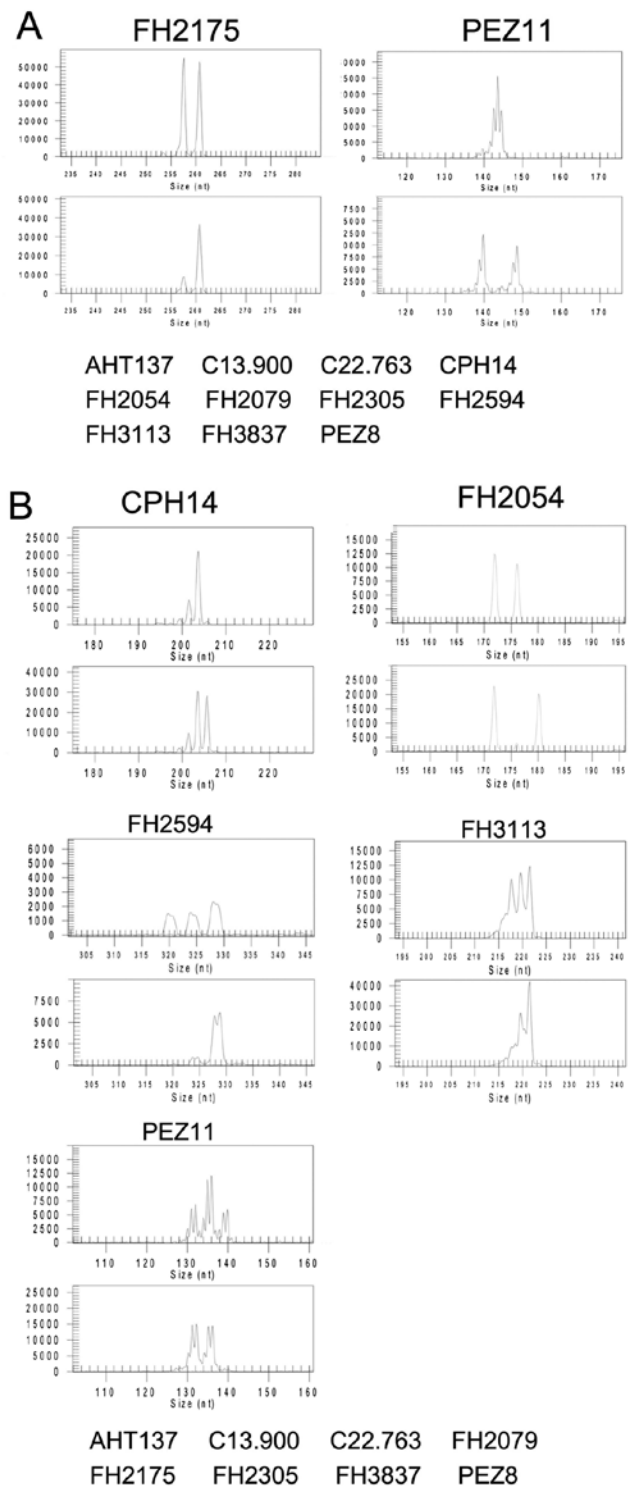


Figure 5. Microsatellite analysis of cell lines established from the same origins. (A) Thirteen microsatellite loci were compared between CTB-m (upper panel) and CTB-m2 (lower panel) cells. Two loci (FH2175 and PEZ11) were different, but the other 11 loci shown under the panels were identical between these two cell lines. (B) Comparison between CIP-p (upper panel) and CIP-m (lower panel). Five (CPH14, FH2054, FH2594, FH3113, and PEZ11) of the 13 loci were found to be different between these two cell lines. The other 8 loci shown under the panels were identical between these two cell lines.

Two microsatellite loci between the CTB-m and CTB-m2 cell lines were different, while the other 11 were identical. Five microsatellite loci between the CIP-p and CIP-m cell lines were different, while the other 8 were identical (Fig. 5A and B).

Discussion

The functions of the tetramerization domain in the p53 protein and its inhibitory and stabilizing ligands have been well investigated using biochemical, cell biological and *in silico* analysis (20–22). However, there were no suitable models expressing a mutant of p53 containing a dysfunction in the tetramerization domain. The NCI-H1299 cell line, which does not express the p53 protein, has typically been used to investigate the effect of exogenous p53 mutant expression (23,24). Here, we established the cell line CTB-m2 which expresses the p53 L332Q (which corresponds to L344 in human p53) mutant. Microsatellite analysis using 11 markers showed that in CTB-m cells, which are the parental cells for CTB-m2, are nearly identical compared with another pair of canine mammary gland tumor cell lines, namely CIP-p and CIP-m (Fig. 5A and B) (15). These data suggest that the L332Q mutation in CTB-m2 is a post-oncogenic transformation. Despite being genetically heterozygous at the nucleotides encoding L332, CTB-m2 cells expressed almost only the mutated L332Q p53 mRNA suggesting that in these cells the wild-type p53 allele is inactive. The expression of p53 mutants has been shown to be heterogenous and gene methylation has also been observed in human cancers (25,26). CTB-m2 cells may therefore have a unique expression status among p53 mutant alleles. Leu332 in canine p53 corresponds to Leu344 in human p53, which is widely recognized as being an essential residue for tetramerization (10). A cell proliferation analysis of the mutated p53 showed enhanced viability of CTB-m2 cells compared with CTB-m cells. The p53 R280T mutant, which is located in the DNA binding domain also promoted cell proliferation in a human glioma cell line (27). Ablation of p53 tetramerization as a result of the L332Q mutation would be expected to change the conformation of the DNA binding domains, which may then result in enhancement of cell proliferation (28).

We examined the tetramerization ability of the p53 L332Q mutant using a functional reporter assay. Two p53 response elements and the minimal human cytomegalovirus promoter sequence, cited by a previous study (17), were modified and cloned into a vector containing a luciferase reporter. The large difference in luciferase activities observed between CTB-m and CTB-m2 cells, which reflects the endogenous tetramerization abilities of p53 in these two cells, suggest that the L332Q mutation is critical for tetramerization, and also confirms that almost only the mutated allele is expressed in CTB-m2 cells. Although there was a significant attenuation in tetramerization for the p53 L332Q mutant, the dimer forming ability of this mutant was retained, as shown by electrophoresis and immunoblotting of p53 cross-linked with glutaraldehyde. The *in silico* simulation of amino acid substitution supports this retention of dimerization ability, and L332Q (L344Q) mutant showed a greater degree of similarity to L344R, which dimerizes, than to L344P, which can only form monomers. The mutated residues in the L332Q (L344Q), L344P, and L344R mutants could not form bonds with Leu344 and Leu348 of the neighboring chain in the tetramerization domain. However, both L332Q (L344Q) and L344R mutants showed contacts with Met340 and L344Q (L344R) of the oblique neighboring chain.

In summary, we established and characterized a new cell line, CTB-m2, which expresses p53 L332Q, a mutant in the p53 tetramerization domain. As a result of this mutation, p53 L332Q lacked tetramerization but not dimerization ability. These CTB-m2 cells can be used to investigate p53 pathogenesis and to evaluate new strategies to restore p53 function (29).

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Availability of data and materials

The analyzed data sets generated during the study are available from the corresponding author upon reasonable request.

Authors' contributions

KO, MM1 and TO designed the research. KO and DA mainly did the research. HH, SK, MM2, AE and TS also performed some experiments. TN and MW analyzed the data. All authors read and approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the study are appropriately investigated and resolved.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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