

Downregulation of hypoxia-inducible factor-1 α inhibits growth, invasion, and angiogenesis of human salivary adenoid cystic carcinoma cells under hypoxia

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Abstract. Surgical and medical treatments usually fail to completely remove the primary lesions of salivary adenoid cystic carcinoma (SACC), resulting in local recurrence due to its strong infiltration, hematogenous metastasis and other unique biological behaviors. Targeted gene therapy, including hypoxia-inducible factor-1 α (HIF-1 α) therapy, sheds new light on the treatment of salivary gland tumors. However, the mechanisms underlying the downregulation of HIF-1 α expression in SACC have not been well studied. The present study aimed to determine the effects of HIF-1 α downregulation on the proliferation, invasion, apoptosis, cell cycle and angiogenesis of hypoxic SACC-83 cells, and to elucidate the molecular mechanisms underlying these effects. Western blot analysis was used to evaluate the protein expression levels of HIF-1 α and matrix metalloproteinase-2 (MMP-2). Angiogenesis and the protein expression of vascular endothelial growth factor (VEGF) were detected by tube formation assay of human umbilical vein endothelial cells and ELISA, respectively. It was revealed that short hairpin RNA was considerably downregulated following

overexpression of HIF-1 α . In addition, downregulation of HIF-1 α inhibited the expression of VEGF and MMP-2, as well as the proliferation, invasion, migration, and tube formation of hypoxic SACC-83 cells, while inducing apoptosis in these cells. These results suggest that the downregulation of HIF-1 α may be a novel targeted therapy for SACC.

Introduction

Salivary adenoid cystic carcinoma (SACC) is one of the most common salivary epithelium-derived malignant tumor types, and is associated with persistent slow growth, perineural invasion, a high recurrence rate, and distant metastases (1). SACC accounts for ~18% of all salivary gland malignancies in China (2). Due to neurotropic invasion and lung metastasis, surgery, radiotherapy and chemotherapy are often unsuccessful, and the recurrence rate is 16-85% (3). Furthermore, the rate of lung metastasis may be up to 40%, with the 5-10-year survival rate as low as 37.4-41.8% (3). As the basic molecular mechanism of SACC carcinogenesis remains unclear, it is important to elucidate the malignant biological behaviors involved in SACC invasion and metastasis.

In human tissues, the normal oxygen partial pressure is 30-60 mmHg, while in the areas surrounding tumors, oxygen partial pressure is often <5 mmHg (4,5). When the tumor volume exceeds 3 mm³, a necrotic tumor center (hypoxic zone) is usually formed as a result of the inadequate oxygen, nutrients and energy supplied by new blood vessels (6). Hypoxia preferentially stimulates the overexpression of hypoxia-inducible factors (HIFs) and induces the expression of downstream genes via signal transduction pathways (7,8). Thus, the overexpression of HIFs allows cells to adapt to hypoxic conditions and proliferate, resulting in high levels of invasion, metastasis and tolerance to radiation and chemotherapy (9-11). HIF-1 is a protein found in the nuclear extracts of the hepatocarcinoma cell line Hep-3B when cultured under hypoxic conditions. It specifically binds to the hypoxia response element (HRE) of the erythropoietin (*EPO*) gene (12-14). HIF-1 is a heterodimer consisting of α and β subunits (15), and growing evidence indicates that the α unit (HIF-1 α) is the functional subunit that is the unique limiting factor in oxygen regulation (16). The

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Abbreviations: DMEM, Dulbecco's modified Eagle's medium; ECM, extracellular matrix; ELISA, enzyme-linked immunosorbent assay; FBS, fetal bovine serum; GFP, green fluorescent protein; HIF-1 α , hypoxia-inducible factor-1 α ; HUVECs, human umbilical vein endothelial cells; MMP-2, matrix metalloproteinase-2; PI, propidium iodine; PVDF, polyvinylidene fluoride; SACC, salivary adenoid cystic carcinoma; SEM, standard error of the mean; shRNA, short hairpin RNA; TBS, Tris-buffered saline; VEGF, vascular endothelial growth factor

Key words: salivary adenoid cystic carcinoma, HIF-1 α , proliferation, invasion, angiogenesis

blockade of the degradation of HIF-1 α leads to the accumulation and overexpression of HIF-1 α under hypoxia, promoting tumor growth, neovascularization, distant metastases and chemoradiation resistance (17,18). In addition, HIF-1 α is frequently overexpressed in various solid tumor tissues, and is associated with tumor invasion and poor prognosis (6). Thus, blocking HIF-1 α expression is a potentially important strategy for targeting anoxic tumor cells. Stoeltzing *et al* (9) constructed a *HIF1 α* -carrying plasmid (pHIF-1 α) that, when transfected into gastric cancer cells, was associated with decreased tumor growth. Sun *et al* (19) reported that transfection with a plasmid containing antisense *HIF1 α* downregulated HIF-1 α expression, resulting in the inhibition of vascular endothelial growth factor VEGF expression and a decrease in tumor microvessel density. However, the mechanisms underlying the suppression of cancer growth by HIF-1 α are complicated, and the effect of HIF-1 α expression on SACC has not previously been discussed.

The present study aimed to investigate the effects of HIF-1 α on the proliferation, invasion, metastasis, apoptosis, and angiogenesis of SACC-83 cells under hypoxia. The results revealed that the downregulation of HIF-1 α inhibited the proliferation, growth, invasion, metastasis and angiogenesis of SACC-83 cells, while inducing morphological changes and apoptosis in these cells.

Materials and methods

Cells and reagents. SACC-83 cell lines were cultured in Dulbecco's modified Eagle's medium (DMEM; HyClone Laboratories, Inc., Logan, UT, USA) containing 10% fetal bovine serum (FBS; Biological Industries, Kibbutz Beit Haemek, Israel) and antibiotics (100 U/ml penicillin and 100 μ g/ml streptomycin) in a 5% CO₂ incubator at 37°C. Following subculture, cells were placed in an anoxic incubator (5% CO₂, 5% O₂, and 90% N₂; Biospherix, Parish, NY, USA) for hypoxic treatment. Lipofectamine 2000 and OPTI-MEM were purchased from Thermo Fisher Scientific, Inc. (Invitrogen; Waltham, MA, USA). PBS, DMSO and bovine serum albumin (BSA) were purchased from Beijing Solarbio Science & Technology Co., Ltd. (Beijing, China). The Annexin V/FITC kit and Matrigel were purchased from BD Biosciences (Franklin Lakes, NJ, USA). The human VEGF ELISA (cat. no. EHC108) kit was purchased from NeoBioscience (Shenzhen, China). Crystal violet staining solution was purchased from Beyotime Institute of Biotechnology (Haimen, China). Anti-HIF-1 α (cat. no. ab16066), anti- β -actin (cat. no. 8457), and anti-matrix metalloproteinase-2 (MMP-2; cat. no. ab92536) antibodies were purchased from Abcam (Cambridge, UK), while horseradish peroxidase-conjugated anti-mouse IgG (cat. no. ZB-2305) and anti-rabbit IgG (cat. no. ZB-2301) secondary antibodies were purchased from OriGene Technologies, Inc. (Beijing, China).

Transfection. The plasmid pGPU6/GFP/Neo, containing short hairpin RNA (shRNA) complementary to *HIF1 α* , was used to examine the function of HIF-1 α . The pGPU6/GFP/Neo-HIF-1 α -homo plasmid (Shanghai GenePharma, Shanghai, China) contained a promoter-driven

green fluorescent protein (GFP) reporter. The most effective *HIF1 α* -targeted small interfering RNA (siRNA) sequence (5'-GCAGCTACTACATCACTTTCT-3') was transformed into shRNA (stem-loop-stem structure) and cloned into the pGPU6/GFP/Neo plasmid. A non-targeting sequence (5'-GTTCTCCGAACGTGTCACGT-3') was used to generate a negative control plasmid. SACC-83 cells were cultured in 12-well plates (2x10⁵ cells/well) with DMEM containing 10% FBS for 24 h at 37°C in 5% CO₂ to 80% confluence. Subsequently, cells were transfected with *HIF1 α* shRNA plasmid or empty plasmid using Lipofectamine 2000 reagent according to the manufacturer's instructions. Cells were transfected with 4 μ g/ml *HIF1 α* shRNA plasmid and 4 μ g/ml empty plasmid in OPTI-MEM. The culture medium was changed from OPTI-MEM to fresh DMEM supplemented with 10% FBS 6 h after transfection. The number of cells expressing GFP was determined using a fluorescence microscope (Olympus IX71; Olympus Corporation, Tokyo, Japan) 24 h after transfection, and the total cell number in the same field of view was determined under brightfield illumination. The transfected cell rate (%) was estimated as the number of GFP-positive cells/total cell number x100. GFP-positive clones with G418 resistance were screened after 6-8 weeks of culture by selecting cells transfected with the sequence resulting in the best interference effect. G418-resistant cells were further cultured and used in subsequent experiments.

Western blot analysis. Cells from the three groups were collected after hypoxic culture for 48 h and washed twice with cold PBS. Total protein was isolated from cells using ice-cold RIPA for cell lysis (1% Triton X-100, 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.1% sodium dodecyl sulfate, 1 mM phenylmethylsulfonyl fluoride and 1 mM ethylenediaminetetraacetic acid). Each mixture was centrifuged at 12,000 x g, for 20 min at 4°C, and the supernatant was collected and boiled for 10 min at 100°C for immunoblotting. The concentration of proteins was assessed using Bradford's method (20). Proteins (30 μ g) were separated by 10% SDS-polyacrylamide gel electrophoresis at 100 V for 2 h under reducing conditions and transferred to a polyvinylidene fluoride (PVDF) membrane (Beijing Solarbio Science & Technology Co., Ltd.) using the electrophoresis apparatus. Membranes were blocked in 5% nonfat milk in Tris-buffered saline containing 0.1% Tween-20 (TBST) for 1 h at room temperature. Subsequently, PVDF membranes were incubated with antibodies against HIF-1 α (1:500), MMP-2 (1:1,000), and β -actin (1:2,000) overnight at 4°C. Then, the membranes were washed with TBST for 10 min three times and incubated with the appropriate secondary antibody as aforementioned (1:5,000) for 1 h at room temperature. Immunoreactive proteins were detected using the SuperSignal™ West Pico Chemiluminescent substrate (Thermo Fisher Scientific, Waltham, MA, USA) followed by exposure to Bio-Rad ChemiDoc XRS+ image analyzer (Hercules, CA, USA). Western blot signals were semi-quantified by densitometry using Image-Pro Plus 6.0 software (Media Cybernetics Inc., Bethesda, MD, USA). All western blot experiments were repeated at least three times and then were used to analyze the densitometry data.

Cell proliferation assay. Cell proliferation was determined by MTT assay (Sigma-Aldrich; Merck KGaA, Darmstadt,

Germany). SACC-83 cells were plated on 96-well plates (2×10^3 cells/well) and cultured for 7 days under hypoxic conditions. Cell proliferation was documented every 24 h according to the manufacturer's protocol. Briefly, MTT solution (20 μ l, 2.5 mg/ml) was added to each well and incubated at 37°C. After 4 h, the medium was removed, and DMSO (150 μ l) was added to each well to dissolve the formazan. Absorbance in each well was measured at 490 nm using an automatic multi-well spectrophotometer. The experiment was repeated at least three times.

Cell apoptosis analysis. Cells were cultured under hypoxic conditions for 48 h. Subsequently, cells were collected and washed in ice-cold PBS prior to staining with Annexin V/FITC and propidium iodide (PI) solution for 15 min in the dark at room temperature. Cell apoptosis was examined using a FACSCanto flow cytometer (BD Biosciences). All experiments were repeated three times.

Cell cycle analysis. SACC-83 cells were seeded in 60-mm culture plates at a density of 1×10^5 cells/well for observing the cell cycle distribution. Cells were cultured under hypoxic conditions for 48 h and were then harvested by trypsinization, fixed with 70% ethanol and stored at 4°C overnight. After washing with PBS, cells were incubated with 100 mg/ml RNase A and 50 mg/ml PI at room temperature for 30 min in the dark. The distribution of the cells was detected using a FACSCanto flow cytometer and analyzed using FlowJo 10.0.7 software (TreeStar, Inc., Ashland, OR, USA).

ELISA. The concentration of VEGF in SACC-83 cell culture medium was determined using human VEGF ELISA kit. SACC-83 cells were cultured for 48 h to 90% confluency under hypoxic conditions. Following treatment, the culture medium was collected and stored at -80°C for use in the ELISA. All experiments were performed at least three times, and the absorbance was measured at 450 nm.

Tube formation assay. SACC-83 cells were seeded in 60-mm plates. Following adhesion, cells were cultured overnight in DMEM only and placed in the hypoxic incubator for an additional 24 h. Human umbilical vein endothelial cells (HUVECs; 3×10^4 cells/well) were seeded on Matrigel-coated 96-well plates and incubated at 37°C. Serum-free supernatant from shControl, shNC, and shHIF-1 α cells were collected and centrifuged at 716 x g, for 5 min at 25°C. The supernatants were then incubated with HUVECs at 37°C for 24 h under hypoxic conditions. The enclosed capillary structure of the tubes formed by HUVECs was observed, and three different fields per well were photographed (400x magnification) using an inverted microscope (DMI1; Leica, Wetzlar, Germany). The total tube length of the tubes in each field of view was measured using ImageJ 1.48 software (National Institutes of Health, Bethesda, MD, USA).

Wound healing assay. The migration of SACC-83 cells was measured using wound healing assays. SACC-83 cells (5×10^5 cells/well) were seeded into a 6-well plate for 24 h. Next, 10- μ l sterile pipette tips were used to create linear scratch wounds on the confluent cell monolayers. The three

types of SACC-83 cells (shControl, shNC, and shHIF-1 α) were cultured in serum-free medium. The scratch wounds were observed using an inverted microscope (DMI1; Leica Microsystems GmbH), and images were acquired at 0, 24 and 48 h (x200 magnification). The migration of cells in the wound area was quantified using ImageJ software. The scratch migration rate (%) was estimated as [(Scratch width at 0 h - Scratch width at 24 or 48 h)/Scratch width at 0 h] x 100.

Matrigel invasion assay. A Transwell plate was pre-coated with 30 μ l DMEM containing 30% Matrigel and added to the membrane of the upper chamber. The chamber was incubated at 37°C for 1 h. A suspension (400 μ l) of SACC-83 cells (2×10^5) at logarithmic growth phase in serum-free medium only was seeded in the upper chamber. DMEM (500 μ l) with 10% FBS was added to the lower chamber, and cells were cultured at 37°C in 5% CO₂, 5% O₂, and 90% N₂ for 24 h. The liquid in the upper chamber and non-invasive cells on the membrane surface were removed with a wet cotton swab. The filter was fixed in 90% ethanol for 10 min and stained with crystal violet. The degree of invasion of SACC-83 cells was estimated under a microscope (x400 magnification) in five fields. Invasion was assessed quantitatively by counting the cells invading the lower side of the filter. The experiments were conducted in triplicate.

Statistical analysis. Data were analyzed using GraphPad Prism (version 5.0; GraphPad Software, Inc., La Jolla, CA, USA). Quantitative data are expressed as mean \pm standard error of the mean. Statistical analysis was performed using one-way analysis of variance with Tukey's post hoc test for ≥ 3 groups. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Identification of stable cell lines. Western blot analysis was used to confirm the inhibitory effects of HIF1 α -targeted shRNA on the expression of HIF-1 α in SACC-83 cells. The results showed that the protein expression of HIF-1 α in SACC-83 cells was significantly reduced in the shHIF-1 α group compared with levels in the shControl and shNC groups ($P < 0.01$ and $P < 0.05$, respectively; Fig. 1).

Downregulation of HIF-1 α inhibits SACC-83 cell growth and induces cell apoptosis but has no significant effect on cell cycle progression under hypoxic conditions. The effect of HIF-1 α downregulation on the proliferation of SACC-83 cells was assessed using a MTT assay. As demonstrated in Fig. 2A, the rate of proliferation of cells transfected with shHIF-1 α was significantly lower compared with that of cells transfected with shControl or shNC. The effect of HIF-1 α downregulation on SACC-83 cell apoptosis was also examined (Fig. 2B). Following shHIF-1 α transfection, the percentage of apoptotic SACC-83 cells increased to 13.09%, which was significantly higher compared with the rates in the shControl (6.01%) and shNC (5.13%) groups ($P < 0.05$, Fig. 2C). Finally, the effect of HIF-1 α downregulation on cell cycle progression was examined using flow cytometry with PI DNA staining (Fig. 2D). As shown in Fig. 2E, the cell cycle distribution of the shHIF-1 α

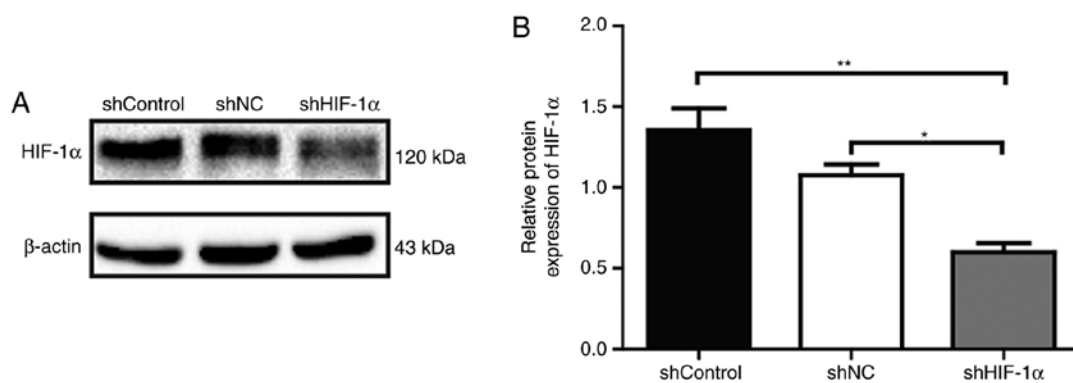


Figure 1. Protein expression of HIF-1 α in SACC-83 cells is inhibited by transfection with *HIF1 α* -targeted shRNA. (A) The inhibition efficiency of the *HIF1 α* -targeted shRNA was determined by western blot analysis. (B) Quantification of HIF-1 α expression. All data are expressed as the mean \pm standard error of the mean. * P <0.05 vs. shNC group; ** P <0.01 vs. shControl group. shControl group, wild-type SACC-83 cells; shNC group, empty plasmid transfection group; shHIF-1 α group, *HIF1 α* -targeted shRNA transfection group; SACC, salivary adenoid cystic carcinoma; shRNA, short hairpin RNA; HIF-1 α , hypoxia-inducible factor-1 α ; NC, negative control.

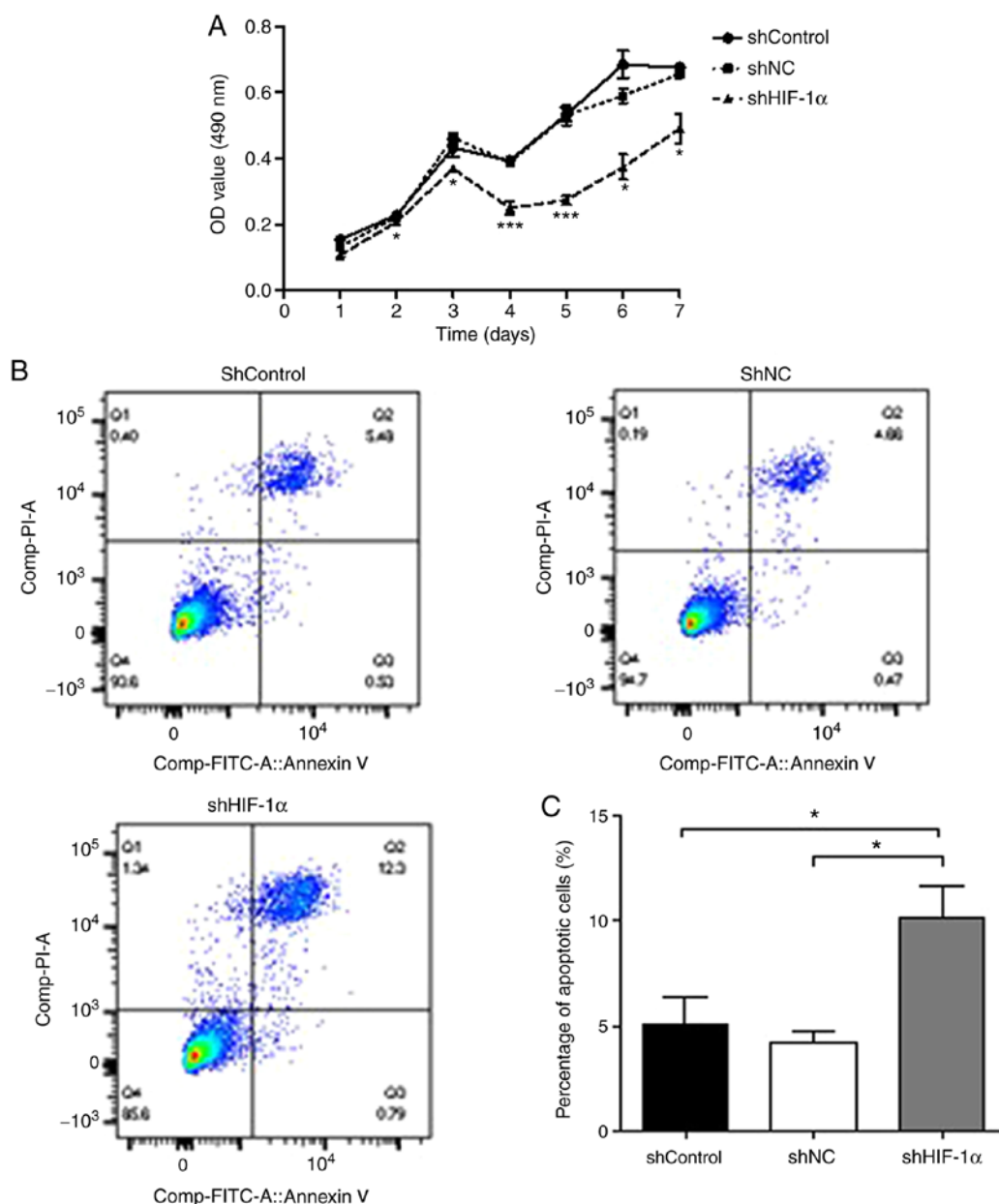


Figure 2. Downregulation of HIF-1 α significantly inhibits the proliferation and apoptosis of SACC-83 cells but does not affect the cell cycle. (A) Growth curves of the three groups of SACC-83 cells under hypoxia. (B) Cell apoptosis rates of different groups under hypoxia. (C) Percentage of apoptotic cells.

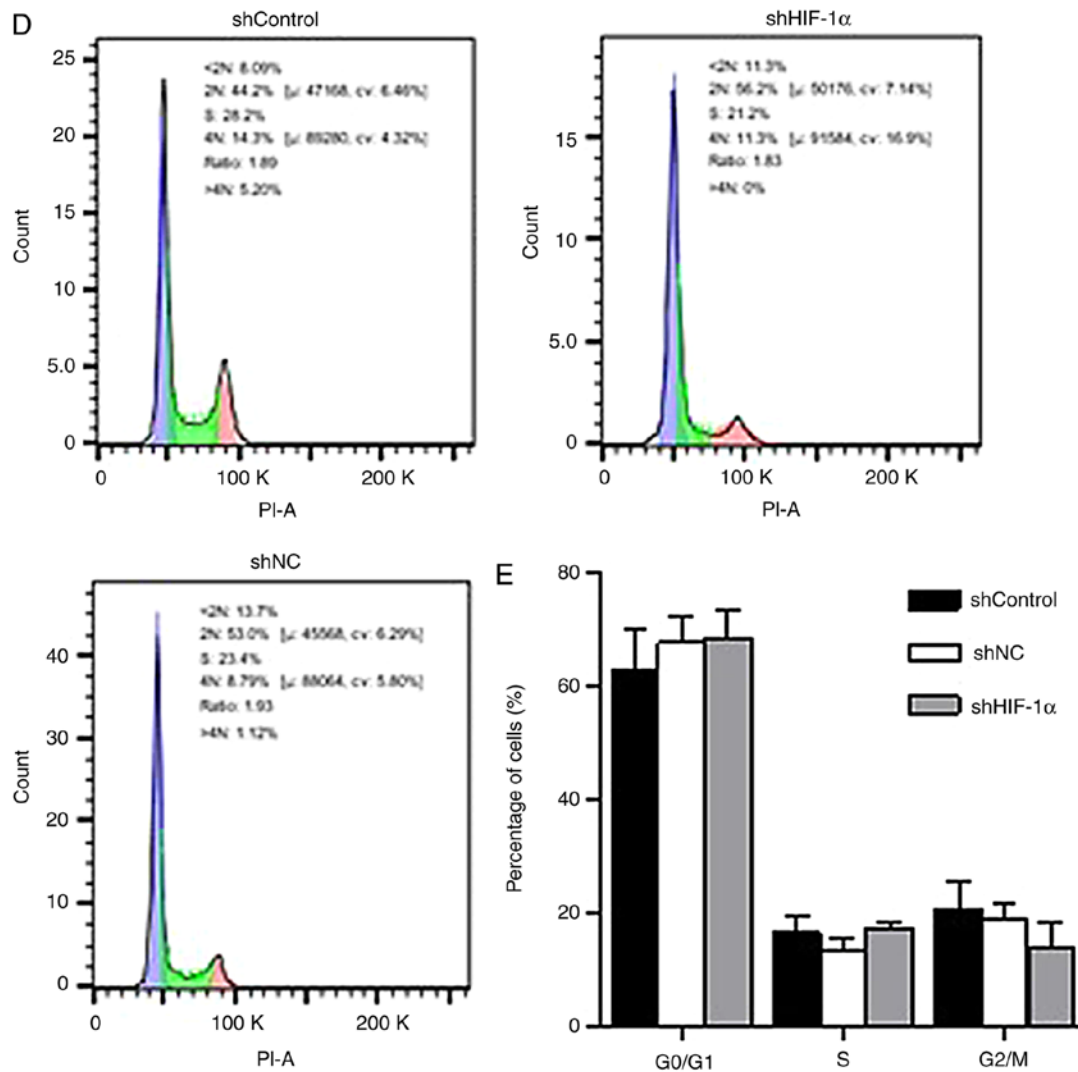


Figure 2. Continued. (D) Distribution of cells in each cell cycle phase. (E) No significant effects on the distribution of G0/G1, S or G2/M cells were noted. All data are expressed as the mean \pm standard error of the mean. * $P < 0.05$ and *** $P < 0.001$ vs. the shControl group and shNC group unless otherwise indicated. shControl group, wild-type SACC-83 cells; shNC group, empty plasmid transfection group; shHIF-1 α group, *HIF1 α* -targeted shRNA transfection group; SACC, salivary adenoid cystic carcinoma; shRNA, short hairpin RNA; HIF-1 α , hypoxia-inducible factor-1 α ; NC, negative control; OD, optical density.

group was not significantly different from those in the shControl and shNC groups ($P > 0.05$).

Downregulation of HIF-1 α decreased MMP-2 expression under hypoxic conditions. Western blot assays were performed to explore the effect of HIF-1 α downregulation on the expression of MMP-2, which is closely linked with tumor invasion and metastasis (21). As shown in Fig. 3A and B, in correspondence with the downregulation of HIF-1 α , the expression of MMP-2 in the shHIF-1 α group was significantly lower compared with levels in the shControl and shNC groups ($P < 0.01$).

Downregulation of HIF-1 α suppresses SACC-83 cell invasion and migration under hypoxic conditions. Cell migration and invasion are important processes in tumor development and metastasis. Thus, the migration and invasion rates of SACC-83 cells under hypoxic conditions were assessed using wound healing and Matrigel Transwell assays. As shown in Fig. 4A and B, the downregulation of HIF-1 α in SACC-83 cells significantly reduced cell migration. The

results indicated that the migration rate was 9.60% at 24 h and 22.08% at 48 h in the shHIF-1 α group, and these rates were significantly lower compared with those in the shControl (23.71% at 24 h and 42.33% at 48 h) and shNC (23.35% at 24 h and 42.87% at 48 h) groups ($P < 0.05$). In addition, the *in vitro* Matrigel Transwell assay showed that the relative number of SACC-83 cells/field was significantly lower in the shHIF-1 α group (51%) compared with in the shControl and shNC groups (Fig. 4C and D; $P < 0.001$), suggesting that the downregulation of HIF-1 α suppressed the mobility and migration of SACC-83 cells.

Downregulation of HIF-1 α reduced angiogenesis and VEGF expression in hypoxic SACC-83 cells. Tumor angiogenesis not only provides adequate nutrition for the tumor, but also provides access for tumor metastasis (22). To study the effects of HIF-1 α downregulation on the angiogenesis of hypoxic SACC-83 cells, HUVECs were treated with cell culture medium derived from each cell type for 24 h and then the tube formation rate was evaluated in each group of

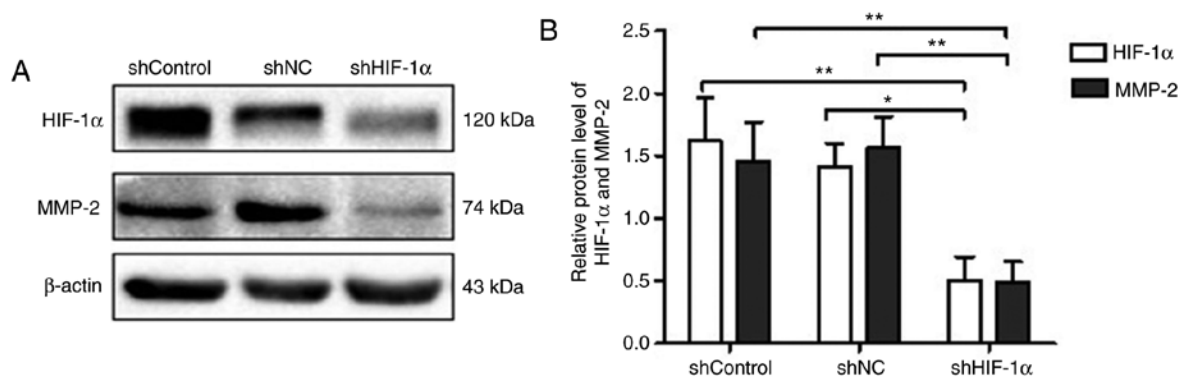


Figure 3. Effects of downregulation of HIF-1 α on MMP-2 expression in SACC-83 cells under hypoxia. (A) Western blot analysis of shControl, shNC and shHIF-1 α groups. (B) Quantification of MMP-2 and HIF-1 α protein expression. All data are expressed as the mean \pm standard error of the mean. * P <0.05, ** P <0.01. shControl group, wild-type SACC-83 cells; shNC group, empty plasmid transfection group; shHIF-1 α group, *HIF1 α* -targeted shRNA transfection group; SACC, salivary adenoid cystic carcinoma; shRNA, short hairpin RNA; HIF-1 α , hypoxia-inducible factor-1 α ; NC, negative control; MMP, matrix metalloproteinase.

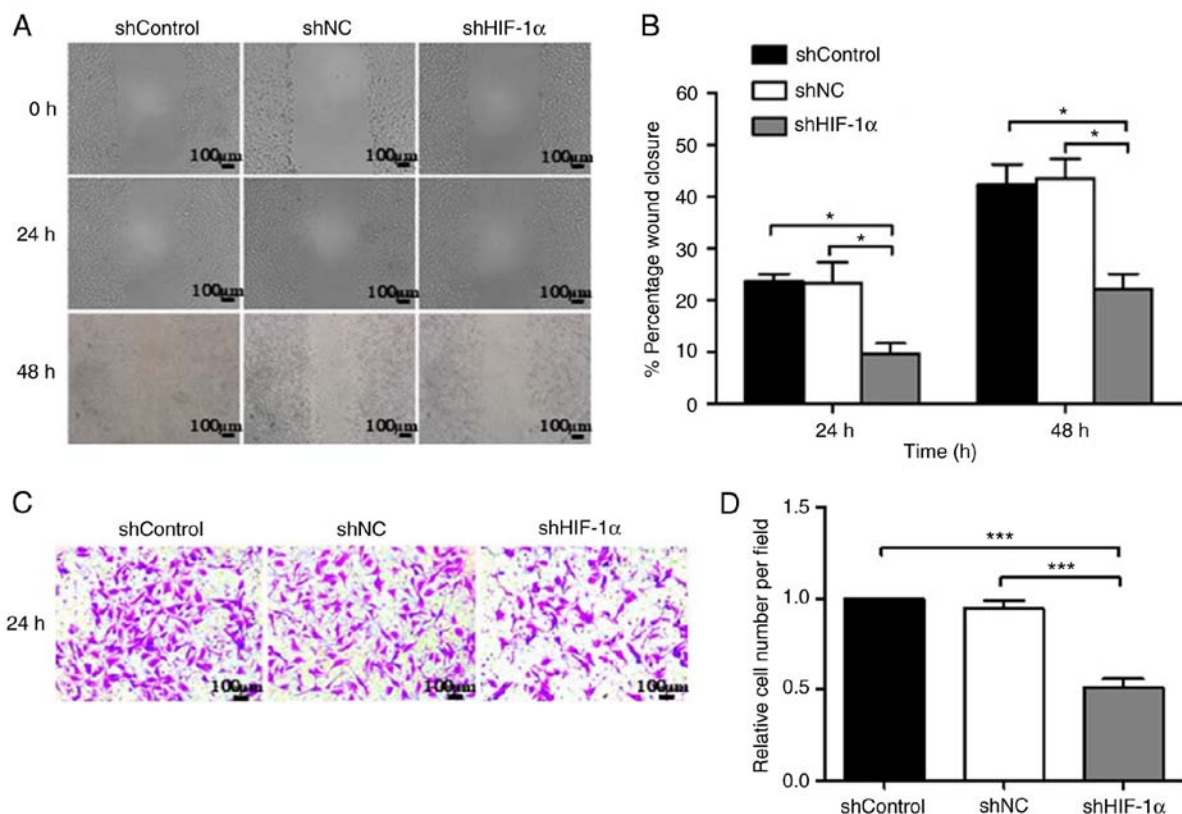


Figure 4. Downregulation of HIF-1 α inhibits the invasion and migration of SACC-83 cells. (A) Wound-healing assay of SACC-83 cells at 0, 24 and 48 h. Scale bar, 100 μ m. (B) Quantification of the effect of HIF-1 α on SACC-83 cell migration in the wound-healing assay. (C) The Transwell assay demonstrated the number of invasive cells in the shControl, shNC and shHIF-1 α groups. Scale bar, 100 μ m. (D) Quantification of the number of invasive cells. The data are expressed as mean \pm standard error of the mean. * P <0.05, *** P <0.001. shControl group, wild-type SACC-83 cells; shNC group, empty plasmid transfection group; shHIF-1 α group, *HIF1 α* -targeted shRNA transfection group; SACC, salivary adenoid cystic carcinoma; shRNA, short hairpin RNA; HIF-1 α , hypoxia-inducible factor-1 α ; NC, negative control.

HUVECs. Compared with the shControl and shNC groups, the shHIF-1 α group exhibited significantly shorter tube lengths (Fig. 5A and B; P <0.001).

A large number of clinical studies have reported that high expression of VEGF is associated with tumor microvessel density, the degree of malignancy and poor patient prognosis (23,24). To investigate whether HIF-1 α downregulation affected the protein expression of VEGF,

double-antibody ELISA was used. The results of the ELISA revealed that the protein expression level of VEGF was lower in the shHIF-1 α group compared with in the shControl and shNC groups (Fig. 5C; P <0.001 and P <0.05, respectively). The optical density values of the shControl and shNC groups were 49.27 and 42.53, respectively, whereas that of the shHIF-1 α group was significantly lower (33.62). Together, these results suggest that a reduction in HIF-1 α expression significantly

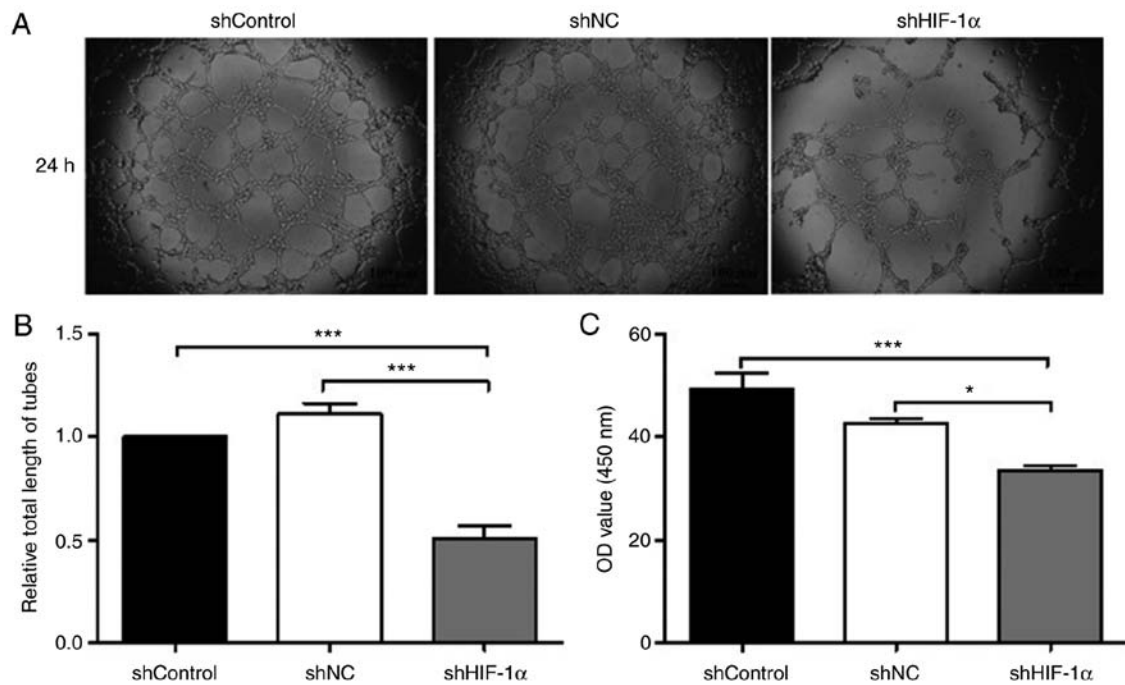


Figure 5. Downregulation of HIF-1 α inhibits tube formation and VEGF expression in SACC-83 cells. (A) Effects of downregulation of HIF-1 α on angiogenesis were verified by human umbilical vein endothelial cell tube-formation assays. Representative images of tube formation (magnification, x400). (B) Relative total length of tubes in each group of SACC-83 cells. (C) VEGF levels were determined by ELISA and the absorbance at 450 nm was measured. All data are expressed as the mean \pm standard error of the mean. * $P < 0.05$, *** $P < 0.001$. shControl group, wild-type SACC-83 cells; shNC group, empty plasmid transfection group; shHIF-1 α group, HIF1 α -targeted shRNA transfection group; SACC, salivary adenoid cystic carcinoma; shRNA, short hairpin RNA; HIF-1 α , hypoxia-inducible factor-1 α ; NC, negative control; VEGF, vascular endothelial growth factor.

weakened tube-formation ability and reduced VEGF expression in SACC-83 cells.

Discussion

SACC is a common malignant tumor of the head and neck that exhibits distinct potential for invasion and distant migration (25). It has long been recognized that tumors are hypoxic, tumor hypoxia is a poor prognostic factor, and established tumors are protected by hypoxia and HIF-1 α -mediated biochemical pathways (26). A previous study showed that the inhibition of HIF-1 α expression using specifically targeted siRNA could increase apoptosis and reduce migration in clear cell renal cell carcinoma 786-O cells (27). Consistent with these results, in the present study, we confirmed that HIF-1 α downregulation induced proliferation and apoptosis in hypoxic SACC cells.

As an important target of HIF-1 α , VEGF is the most potent and selective angiogenesis-promoting factor, increasing microvascular permeability and inducing endothelial cell division, proliferation, and migration (28). HIF-1 α is regulated by the phosphatidylinositol 3-kinase and mitogen-activated protein kinase signaling pathways, which ultimately promote its expression (29). In this study, SACC-83 cells were cultured in a simulated hypoxic environment (5% O₂) and a RNA interference technique was used to downregulate HIF-1 α expression in SACC-83 cells. The results demonstrated that the downregulation of HIF-1 α significantly reduced the expression of VEGF in SACC-83 cells and that VEGF-associated tumor angiogenesis was significantly suppressed in HUVECs. HIF-1 α serves an important role in regulating the VEGF

signal transduction pathway under hypoxic conditions. In the process of early angiogenesis, HIF-1 α not only increases the stability of VEGF mRNA but also increases the transcriptional activity of VEGF, resulting in the transport of more oxygen and nutrients to the tumor and ultimately promoting tumor invasion and distant transfer (30). Reducing the expression of HIF-1 α in tumor cells may thus decrease the production of VEGF and inhibit tumor growth. HIF-1 α also serves a key role in the autocrine VEGF-VEGF receptor (VEGFR) signal transduction pathway, which may be another mechanism by which the downregulation of HIF-1 α inhibits tumor angiogenesis (9,31,32).

HIFs induce a variety of factors and enzymes associated with tumor invasion and metastasis, including MMPs (33). MMP-2 is the most widely distributed proteolytic enzyme in the MMP family, and is associated with tumor invasion and metastasis (34). Certain studies have reported that HIF-1 α promotes the invasion and metastasis of cervical cancer by downregulating E-cadherin expression and upregulating the expression of MMP-2 (35,36). MMP-2 increases the extent of tumor vascularization and is an important component of the angiogenesis initiation system (9,37). In hypoxic environments, tumor cells secrete large amounts of HIF-1 α , inducing the secretion of MMP-2 by endothelial cells (9,37). In the present study, it was demonstrated that the downregulation of HIF-1 α significantly attenuated MMP-2 expression. Furthermore, as expected, the results of wound healing and Transwell assays revealed that HIF-1 α significantly suppressed the migration and invasion of SACC cells.

In summary, the present study demonstrated that the downregulation of HIF-1 α significantly inhibited proliferation,

invasion, migration and angiogenesis in hypoxic SACC-83 cells, and promoted apoptosis in this cell type. Additionally, HIF-1 α downregulation significantly attenuated the expression of MMP-2 and VEGF. The inhibition of angiogenesis may be useful for the treatment of neoplastic lesions and pro-angiogenic therapies may be important for future treatment of ischemic diseases (38). In the future, these findings should be confirmed in xenograft models and the signaling molecules underlying the effects of HIF-1 α on SACC should be elucidated through further research.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

CX, YP, XZ, LW, ZL, SY and HW contributed to the conception, design and completion of the present study. CX and YP contributed equally to the experimental work and drafting of the manuscript. XZ analyzed the data and revised the manuscript critically for important intellectual content. LW and ZL performed the experiments and interpreted the results. SY produced the figures and HW contributed to study design.

Ethics approval and consent

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no conflict of interest.

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