

Resveratrol inhibited the progression of human hepatocellular carcinoma by inducing autophagy via regulating p53 and the phosphoinositide 3-kinase/protein kinase B pathway

BAICHAO ZHANG¹, XIAONING YIN¹ and SHAOGUANG SUI²

¹General Department, Zuanshiwan Hospital District of The Second Hospital of Dalian Medical University;

²Emergency Department, The Second Hospital of Dalian Medical University, Dalian, Liaoning 116000, P.R. China

Received February 26, 2018; Accepted August 2, 2018

DOI: 10.3892/or.2018.6648

Abstract. Resveratrol, a natural product, has been revealed to exert antitumor effects in multiple types of tumors. However, the antitumor effects of resveratrol on hepatocellular carcinoma (HCC) and its potential underlying mechanisms have not yet been elucidated. The present study demonstrated that resveratrol inhibited viability, proliferation, invasion and migration of HCC cells significantly in a time- and dose-dependent manner, indicating that resveratrol exerted antitumor effects in HCC. Furthermore, relative expression of autophagy-related proteins Beclin1 and LC3 II/I ratio was increased while p62 expression was decreased by resveratrol treatment dose-dependently. The LC3⁺ puncta formation, which represented autophagosome formation was also markedly dose-dependently upregulated by resveratrol treatment, suggesting that resveratrol induced autophagy in HCC cells. In addition, treatment with autophagy inhibitor 3-methyladenine (3-MA) counteracted the inhibitory effect of resveratrol on HCC cell proliferation, invasion and migration, indicating that suppressing autophagy may hamper the antitumor effect of resveratrol in HCC. It was revealed that resveratrol upregulated the expression of p53 while decreasing the ratio of phosphorylated protein kinase B (p-Akt)/Akt in HCC cells. Treatment with p53 inhibitor pifithrin- α and Akt activator insulin-like growth factor-1 decreased the expression of Beclin1 while significantly promoting cell proliferation, invasion and migration compared with the resveratrol treatment group. Taken together, the results of the present study

revealed that resveratrol inhibited the proliferation and mobility of HCC cells through inducing autophagy via activating p53 and inhibiting phosphoinositide 3-kinase/Akt. Enhancing autophagy can augment the antitumor effects of resveratrol in HCC. Therefore, combining resveratrol with an autophagy inducer may be a viable option for treating HCC.

Introduction

Hepatocellular carcinoma (HCC) is a common malignant neoplasm and the fourth leading cause of cancer-related mortality in China (1). HCC is the dominant form of primary liver cancer in patients with chronic liver disease and cirrhosis (2). Patients with early-stage HCC can be treated with surgery to remove part of the liver can undergo or transplantation, which contributes toward a 5-year survival rate of >70% (3). However, for the majority of patients with advanced-stage HCC whose disease is too advanced for them to undergo surgical treatment, the 5-year survival rate is extremely low at ~15% (4). Therefore, improving understanding of the molecular mechanism involved in HCC progression and identifying novel approaches to inhibit HCC carcinogenesis are important.

Over recent decades, natural products have served significant roles in the development of anticancer agents (5,6). Resveratrol (trans-3,4',5-trihydroxystilbene) was originally isolated from the roots of white hellebore at 1940 (7). Resveratrol is often used in traditional Chinese medicine to treat skin inflammations, cardiovascular and liver diseases (8,9). Resveratrol exerted a variety of biological effects, including antioxidation, antiproliferation and chemopreventative effects (10,11). Recently, accumulating evidence has indicated that resveratrol also exerts antitumor effects by inhibiting proliferation while inducing apoptosis and autophagy in tumor cells (12,13). For instance, Szekeres *et al* (9) reported that resveratrol induced cell cycle arrest, apoptosis and autophagy in T-cell acute lymphoblastic leukemia cells through inhibiting the protein kinase B (Akt)/mechanistic target of rapamycin (mTOR)/p70S6K/4E-BP1 and activating p38-mitogen-activated protein kinase signaling pathways. Fabre *et al* (10) indicated that resveratrol could induce autophagic and apoptotic death in drug-resistant oral cancer cells and may become

Correspondence to: Dr Shaoguang Sui, Emergency Department, The Second Hospital of Dalian Medical University, 467 Zhongshan Road, Dalian, Liaoning 116000, P.R. China
E-mail: suishaoguangdl@163.com

Abbreviations: HCC, hepatocellular carcinoma; 3-MA, 3-methyladenine; IGF-1, insulin-like growth factor-1; SD, standard deviation; EMT, epithelial-mesenchymal transition

Key words: resveratrol, hepatocellular carcinoma, autophagy, p53, phosphoinositide 3-kinase/protein kinase B

a novel approach for the treatment of oral cancer in the near future (14). However, the antitumor effect of resveratrol in the tumor progression of HCC remains unclear and requires further investigation.

Autophagy is a highly conserved catabolic mechanism in eukaryotes in which intracellular contents, including large protein complexes and dysfunctional organelles, are transported to lysosomes for degradation and reuse (15). Autophagy can prolong the survival of cancer cells and enhance their resistance to apoptosis, and paradoxically, defective autophagy has been associated with increased tumorigenesis, but the mechanism underpinning this remains unclear (16).

The present study aimed to investigate the antitumor effect of resveratrol in HCC. Resveratrol treatment inhibited cell viability, proliferation, invasion and migration of HCC cells through inducing autophagy via regulating p53, phosphoinositide 3-kinase (PI3K)/Akt. The results of the present study suggested that combined treatment with resveratrol and autophagy inducer may serve as effective agents against HCC.

Materials and methods

Reagents and antibodies. Resveratrol, 3-methyladenine (3-MA) and pifithrin- α were obtained from Sigma-Aldrich; Merck KGaA (Darmstadt, Germany), and were dissolved in dimethyl sulfoxide. Insulin-like growth factor-1 (IGF-1) was purchased from PeproTech, Inc. (Rocky Hill, NJ, USA). Primary antibodies against Beclin1 (1:1,000; cat. no. 3738), LC3 I/II (1:1,000; cat. no. 12741), p62 (1:1,000; cat. no. 88588), p53 (1:1,000; cat. no. 2524), p-Akt (1:2,000; cat. no. 4060), Akt (1:1,000; cat. no. 4685) and GAPDH (1:1,000; cat. no. 5174), as well as horseradish-peroxidase (HRP)-conjugated secondary anti-rabbit (1:1,000; cat. no. 4877) and anti-mouse antibodies (1:1,000; cat. no. 4874) were obtained from Cell Signaling Technology, Inc. (Danvers, MA, USA).

Cell culture. The HCC MHCC97-H cell line with wild-type p53 gene was purchased from Shanghai Institute of Biotechnology and Cell Biology of the Chinese Academy of Sciences (Shanghai, China). Cells were cultured in RPMI-1640 complete culture medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA), supplemented with 10% FBS (HyClone; GE Healthcare Life Sciences, Logan, UT, USA), 100 μ g/ml streptomycin and 100 U/ml penicillin in a humidified atmosphere of 5% CO₂ at 37°C.

Cell viability assay. MHCC-97H cells (5×10^3) were seeded into 96-well plates and treated with different concentrations of resveratrol (0–200 μ M) for 24 or 48 h. The purple formazan was dissolved in dimethyl sulfoxide. Cell viability was measured by MTT assay at 450 nm, according to the manufacturer's protocol. All experiments were performed in triplicate and repeated at least twice.

Cell invasion assay. The Transwell invasion chambers coated with Matrigel (1 mg/ml) (BD Biosciences, Franklin Lakes, NJ, USA) were used to perform an invasion assay of MHCC-97H cells. Briefly, 100 μ l serum-free Dulbecco's modified Eagle's medium (DMEM; Gibco; Thermo Fisher Scientific, Inc.) containing 1×10^5 cells/well was added into the upper chamber,

while the lower chamber contained 600 μ l DMEM supplemented with FBS (Gibco; Thermo Fisher Scientific, Inc.) containing 10% FBS. After 24 h of incubation, the cells in the upper chamber were removed and the cells that had invaded through the Matrigel matrix were fixed in 95% ethanol overnight at 4°C and stained with hematoxylin at 4°C for 2 h. The cell numbers were counted and images were captured under an inverted microscope (magnification, x400; Olympus Corporation, Tokyo, Japan) on 5 randomly selected fields in each well.

Wound healing assay. Cell migration ability was determined using a wound healing assay. Cells (1.5×10^6 cells/well) were seeded into 6-well plates and cultured overnight at 37°C until the cells reached 90% confluence. Next, a sterile pipette tip was used to create a straight scratch. The destroyed cells were gently rinsed off with PBS 3 times and cultured in MCDB153 medium (Sigma-Aldrich; Merck KGaA) for another 24 h. Cell migration was observed and images were captured under a confocal microscope (magnification, x200; NIKON A1; Nikon Corporation, Tokyo, Japan) at 0 and 24 h.

Western blot analysis. MHCC-97H cells in different groups were lysed and the protein was extracted using radioimmunoprecipitation assay buffer (10X; Cell Signaling Technology, Inc.), containing 1% phenylmethylsulfonyl fluoride. The extracted protein was determined using a bicinchoninic acid protein assay kit (Beijing Solarbio Science & Technology Co., Ltd., Beijing, China). Equal amounts of proteins (20 μ g) from each sample were separated by 5% SDS-PAGE, prior to being transferred onto polyvinylidene difluoride membranes (EMD Millipore, Billerica, MA, USA). Following blocking with 5% skimmed milk at room temperature for 2 h, the membranes were incubated with the aforementioned primary antibodies [diluted in 0.1 mmol/l PBS (PH 7.4) + 1% BSA + 0.1% Na₂N₃] at 4°C overnight and the corresponding HRP-conjugated secondary antibodies at room temperature for 1 h. The signal was detected using an enhanced chemiluminescence commercial kit (GE Healthcare, Chicago, IL, USA) according to the manufacturer's protocol. Analysis was performed using ImageJ software (version 1.48; National Institutes of Health, Bethesda, MD, USA).

Analysis of autophagy. MHCC-97H cells were transfected with green fluorescent protein (GFP)-LC3 plasmid (Invitrogen; Thermo Fisher Scientific, Inc.) using Lipofectamine 3000™ (Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. Following transfection for 48 h, cells were treated with different concentrations of resveratrol, the intensity of GFP-LC3 fluorescence in cells was determined under a fluorescence microscope (Olympus Corporation). Cells with >5 puncta were considered to have accumulated autophagosomes.

Cell proliferation assay. Cell Counting Kit-8 (CCK-8) assay was used to perform a cell proliferation assay. Cells were seeded into a 96-well plate with 2×10^3 cells/well in triplicate. CCK-8 reagent was added to each well (5 mg/ml) at indicated time points and then incubated in the dark at 37°C for 2 h. Absorbance was determined at a wavelength of 450 nm. All the experiments were repeated at least 3 times.

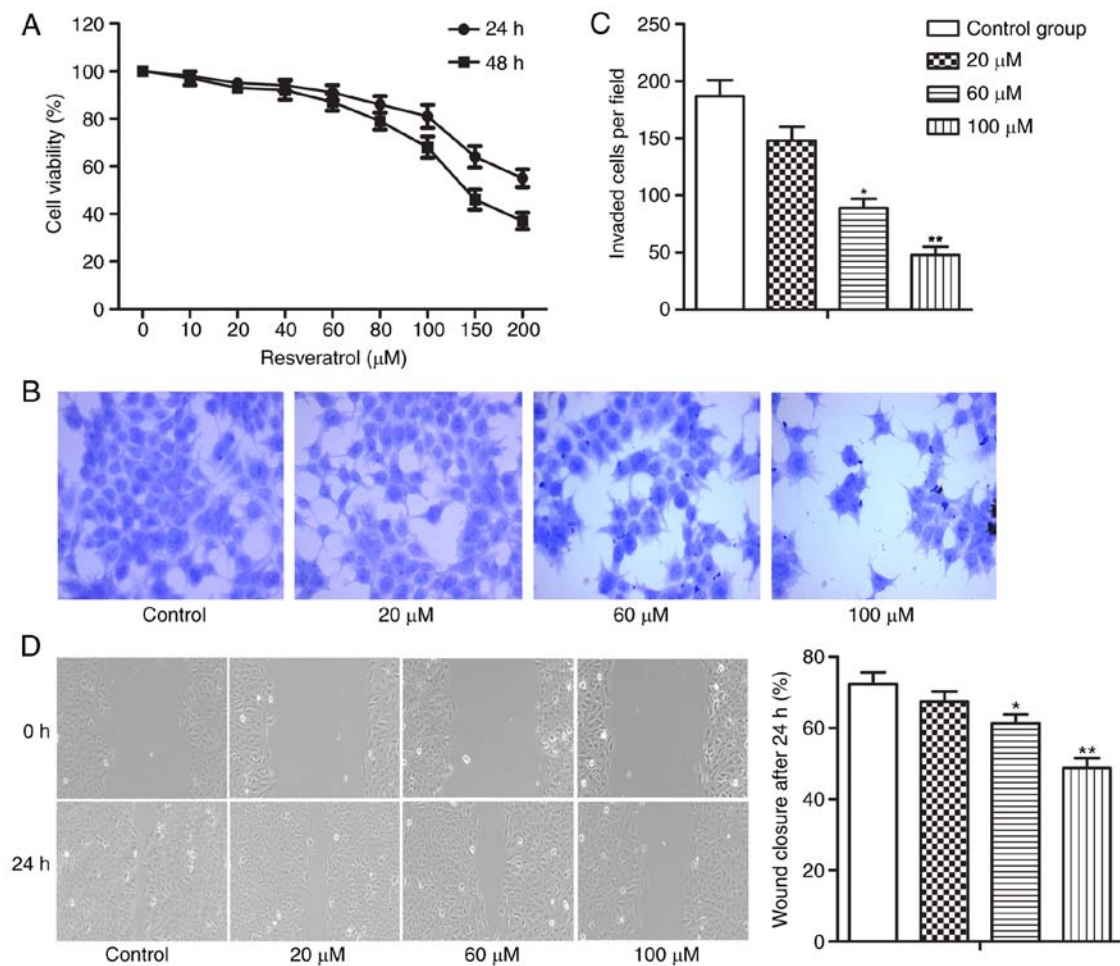


Figure 1. Resveratrol inhibits the viability, invasion and migration of hepatocellular carcinoma cells. MHCC-97H cells were treated with different concentrations of resveratrol (0, 20, 60 or 100 μ M). (A) Cell viability was detected using an MTT assay. (B) Images and (C) quantitative analysis of cell invasive ability was detected using Transwell invasion assay. (D) Cell migration ability was detected using a wound healing assay. All data are presented as the mean \pm standard deviation of 3 independent experiments. * $P < 0.05$ and ** $P < 0.01$, compared with the control group.

Statistical analysis. All data are presented as the mean \pm standard deviation. SPSS 19.0 software (IBM Corp., Armonk, NY, USA) was used for analysis. Paired Student's t-tests were used to compare means of two groups and one-way analysis of variance, followed by Bonferroni's post hoc test, was used to compare means of multiple samples. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Resveratrol inhibits the viability, invasion and migration of HCC cells. To begin with, the present study investigated the potential therapeutic effect of resveratrol in the treatment of HCC. It was revealed that resveratrol treatment suppressed the viability of MHCC-97H cells in a dose- and time-dependent manner (Fig. 1A). Next, the invasion and migration abilities of MHCC-97H cells treated with different concentrations of resveratrol (0, 20, 60 or 100 μ M) were compared in the present study. The results demonstrated that resveratrol dose-dependently inhibited the invasion and migration abilities of MHCC-97H cells (Fig. 1B-D; * $P < 0.05$ and ** $P < 0.01$). These results indicated that resveratrol exerted antitumor effects in HCC through suppressing the viability, invasion and migration of HCC cells.

Resveratrol induces autophagy in HCC cells. As resveratrol can act as an autophagy inducer during tumor progression, the present study investigated whether resveratrol induced autophagy in HCC cells. The relative expression of Beclin1 and LC3 II/I, two well-validated biomarkers of autophagy, was increased while the expression of p62, which can clear autophagosomes, was significantly decreased by resveratrol in a dose-dependent manner (Fig. 2A-D; * $P < 0.05$ and ** $P < 0.01$). Furthermore, LC3⁺ puncta formation, which represented autophagosome formation, was markedly dose-dependently upregulated by resveratrol treatment (Fig. 2E and F; * $P < 0.05$ and ** $P < 0.01$). The aforementioned results suggested that resveratrol can induce autophagy in HCC cells.

Autophagy inhibitor 3-MA counteracts the effect of resveratrol. The present study further investigated the effect of autophagy on the antitumor effect of resveratrol in HCC cells. 3-MA (10 mmol/l) was used to inhibit resveratrol-induced autophagy in MHCC-97H cells. The results demonstrated that suppressing resveratrol-induced autophagy by 3-MA significantly promoted the proliferation, invasion and migration abilities of MHCC-97H cells compared with the 100 μ M resveratrol group (Fig. 3A-E; * $P < 0.05$ and ** $P < 0.01$). The results elucidated that suppressing autophagy counteracted the

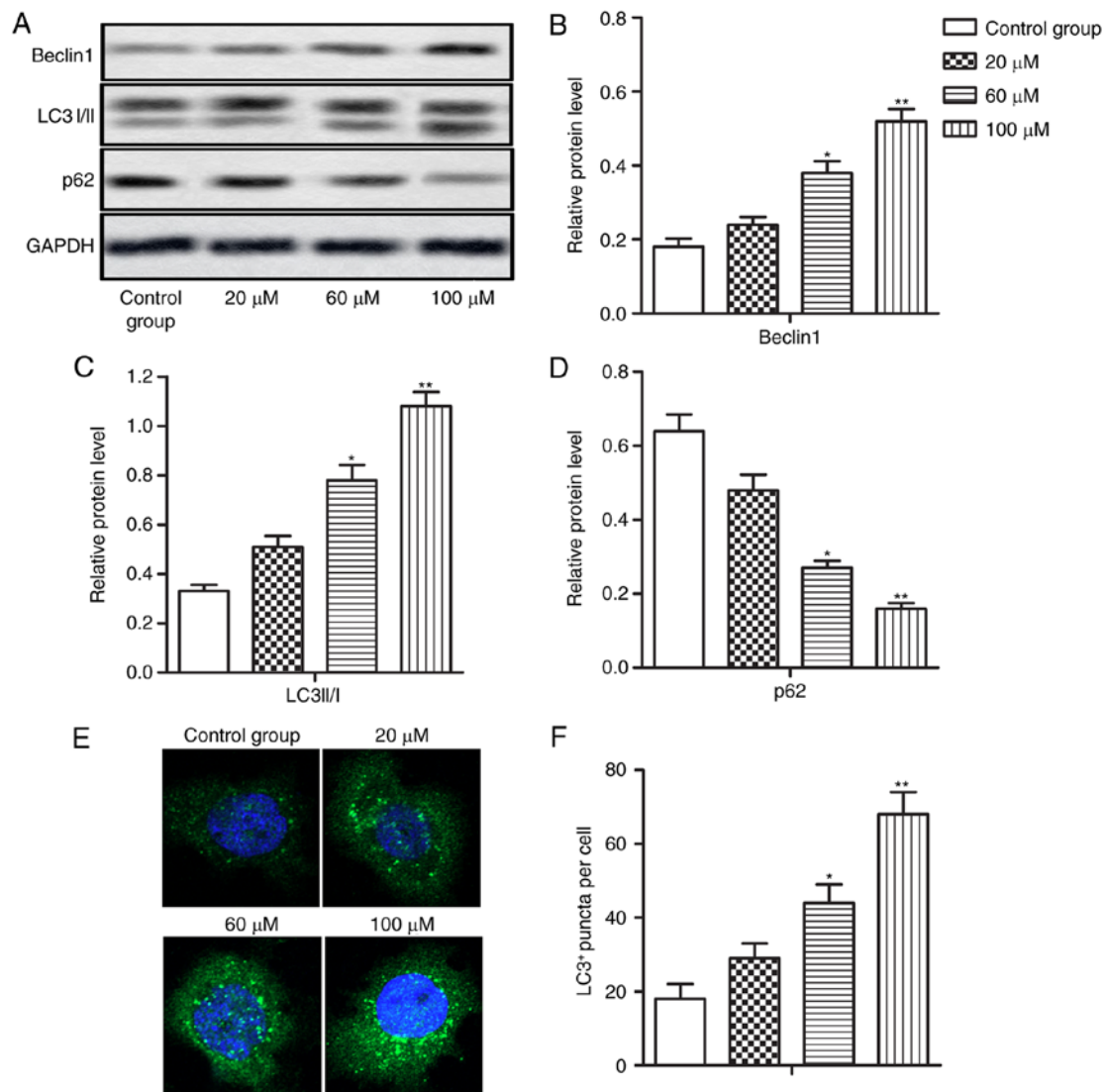


Figure 2. Resveratrol induces autophagy in hepatocellular carcinoma cells. MHCC-97H cells were treated with different concentrations of resveratrol (0, 20, 60 or 100 μ M). (A) Relative protein level of Beclin1, LC3 I/II and p62 was detected through western blot analysis. (B) Relative expression level of Beclin1. (C) Relative expression level of LC3 I/II. (D) Relative expression level of p62. (E) Immunofluorescence of MHCC-97H cells was quantified by the number of GFP-LC3 dots, which represent autophagosomes. (F) The number of GFP-LC3 dots were analyzed by bar graph. All data are presented as the mean \pm standard deviation of 3 independent experiments. * $P < 0.05$ and ** $P < 0.01$, compared with the control group.

inhibitory effect of resveratrol on the proliferation, invasion and migration abilities of HCC cells.

Resveratrol induces autophagy through regulating the p53 and PI3K/Akt pathways. Resveratrol was reported to exert its antitumor effects through the p53 or PI3K/Akt pathways (17,18). To investigate whether these signaling pathways were also involved in resveratrol-induced autophagy in HCC cells, western blot analysis was performed to evaluate the expression of p53, p-Akt and Akt. It was revealed that resveratrol upregulated the expression of p53 while decreasing the ratio of p-Akt/Akt in HCC cells dose-dependently, indicating that resveratrol activates p53 while suppressing the PI3K/Akt pathway in HCC cells (Fig. 4A and B; * $P < 0.05$ and ** $P < 0.01$). P53 inhibitor pifithrin- α (30 μ M) and Akt activator IGF-1 (50 ng/ml) were used to treat MHCC-97H cells. The results demonstrated that pifithrin- α and IGF-1 significantly decreased the expression of Beclin1, suggesting

that inhibiting p53 or activating PI3K/Akt suppressed resveratrol-induced autophagy in HCC cells (Fig. 4C-F; * $P < 0.05$ and ** $P < 0.01$). In summary, the results of the present study indicated that resveratrol induced autophagy through activating p53 while suppressing the PI3K/Akt pathway in HCC cells.

Pifithrin- α and IGF-1 counteract the inhibitory effect of resveratrol on proliferation, invasion and migration of HCC cells. The present study then investigated the effects of pifithrin- α and IGF-1 on the motility and proliferation of HCC cells. Pifithrin- α and IGF-1 treatment inhibited the invasion, migration and proliferation abilities of MHCC-97H cells compared with the 100 μ M resveratrol group (Fig. 5A-F; ** $P < 0.01$ and * $P < 0.05$). The results of the present study suggested that pifithrin- α and IGF-1 counteracted the inhibitory effect of resveratrol on the proliferation, invasion and migration of HCC cells.

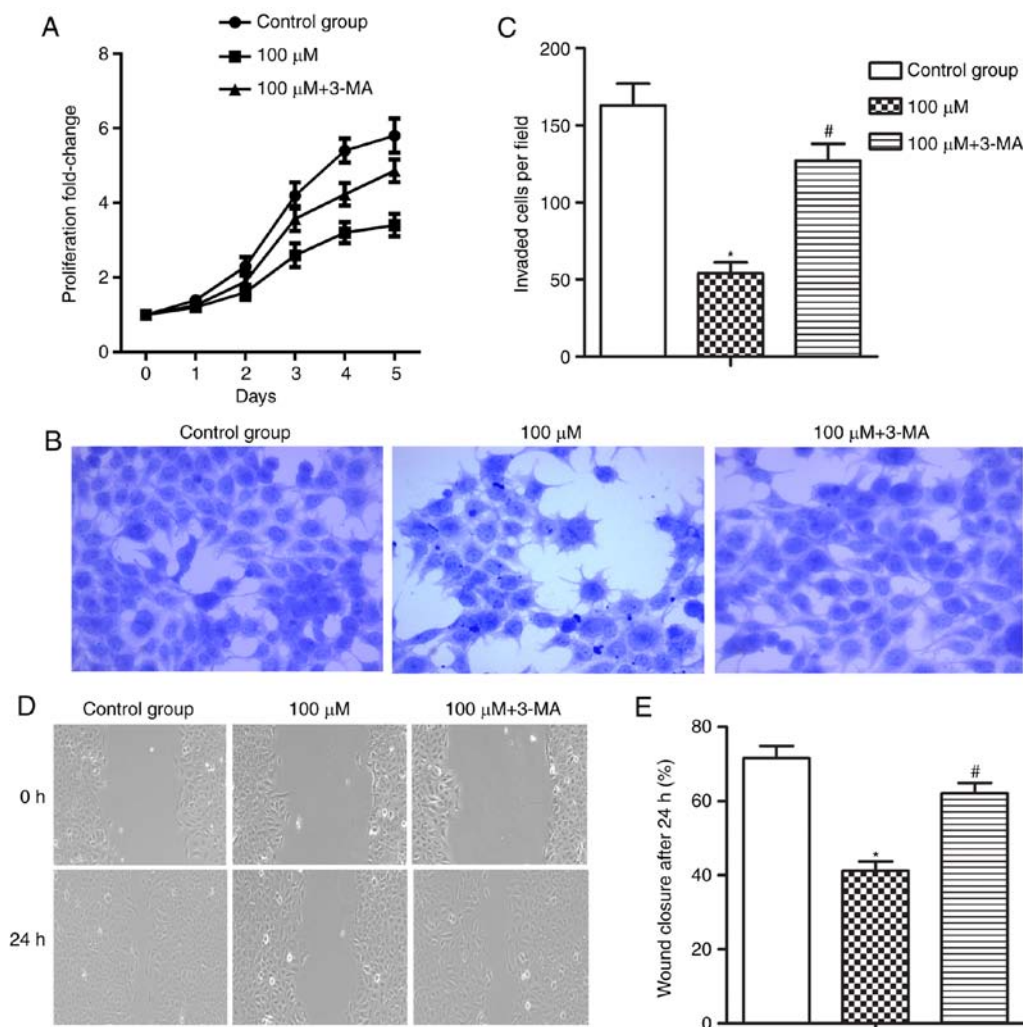


Figure 3. Autophagy inhibitor 3-MA counteracts the effect of resveratrol. MHCC-97H cells were treated with 100 μ M resveratrol with or without 3-MA treatment. (A) Cell proliferation was detected using a CCK-8 assay. (B) The cell invasion ability was detected using a Transwell invasion assay. (C) Invaded cells per field. (D) The cell migration ability was detected using a wound healing assay. (E) Wound closing rate. All data are presented as the mean \pm standard deviation of 3 independent experiments. * P <0.05, compared with the control group; # P <0.05, compared with the 100 μ M resveratrol group.

Discussion

Resveratrol, as a natural product compound, has been demonstrated to have multiple potential chemoprotective activities in various types of cells and animal models (19,20). In addition to antioxidation and chemopreventive effects, resveratrol has also been demonstrated to exert antitumor effects through suppressing proliferation, and inducing apoptosis and autophagy in a variety of cancer cells (21). However, the mechanism of the antitumor effect of resveratrol in HCC remained unclear.

The present study was designed to investigate the antitumor effect of resveratrol in HCC and its potential mechanism. The results demonstrated that resveratrol treatment inhibited cell viability, proliferation, invasion and migration through inducing autophagy in HCC cells via activating p53 while suppressing PI3K/Akt pathway in a time- and dose-dependent manner.

Numerous studies have reported that resveratrol inhibited cell proliferation and suppressed cell mobility in various types of cancer cells. For instance, Chai *et al* (18) reported that

resveratrol induced mitochondrial apoptosis and decreased cell migration, invasion and epithelial-mesenchymal transition (EMT)-inducing transcription factor in oral squamous cell carcinoma cells. Harikumar *et al* (19) indicated that resveratrol suppressed invasion, colony forming capacity and cell proliferation in colorectal cancer cells by modulation of focal adhesion molecules. The inhibitory effects of resveratrol on the adhesion, migration and invasion had also been observed in human bladder cancer cells (22). In agreement with the results of these studies, it was observed that resveratrol inhibited cell proliferation, invasion and migration in HCC cells in a time- and dose-dependent manner, suggesting that resveratrol exerted an antitumor effect in HCC.

Autophagy is a major degradation system that promotes the lysosomal digestion of organelles and cytoplasmic components (23). The role of autophagy in cancer cells is diverse: On the one hand, autophagy can promote cancer progression through driving cell metabolism (24); on the other hand, autophagy itself can induce cell death, which is known as autophagic cell death and induction of autophagy is also reported to facilitate the activation of apoptosis (25,26).

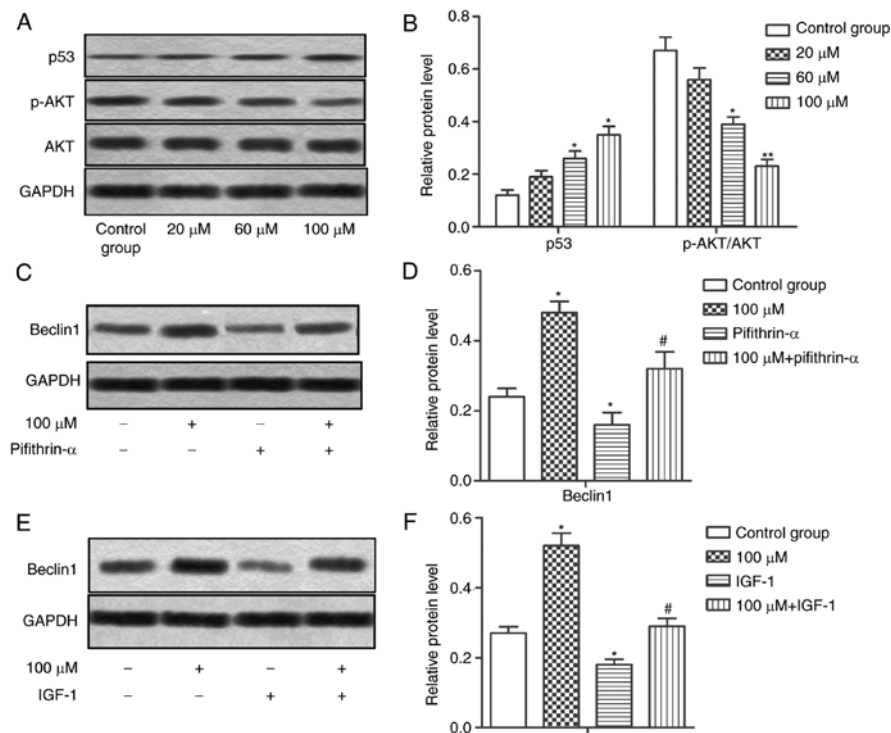


Figure 4. Resveratrol induces autophagy through regulating the p53 and phosphoinositide 3-kinase/Akt pathways. MHCC-97H cells were treated with different concentrations of resveratrol (0, 20, 60 or 100 μ M) with or without p53 inhibitor pifithrin- α or Akt activator IGF-1. (A) Relative protein expression level of p53, p-Akt and Akt was detected through western blot analysis. (B) Densitometric analysis of the relative expression level of p53, p-Akt and Akt. (C) Relative protein expression of Beclin1 treated with resveratrol (100 μ M) with or without p53 inhibitor pifithrin- α was detected through western blot analysis. (D) Densitometric analysis of the relative protein expression of Beclin1. (E) Relative protein expression of Beclin1 treated with resveratrol (100 μ M) with or without Akt activator IGF-1 was measured by western blot. (F) Densitometric analysis of the relative protein expression of Beclin1. All data are presented as the mean \pm standard deviation from 3 independent experiments. * P <0.05 and ** P <0.01, compared with the control group; # P <0.05, compared with the 100 μ M group. Akt, protein kinase B; IGF-1, insulin-like growth factor-1.

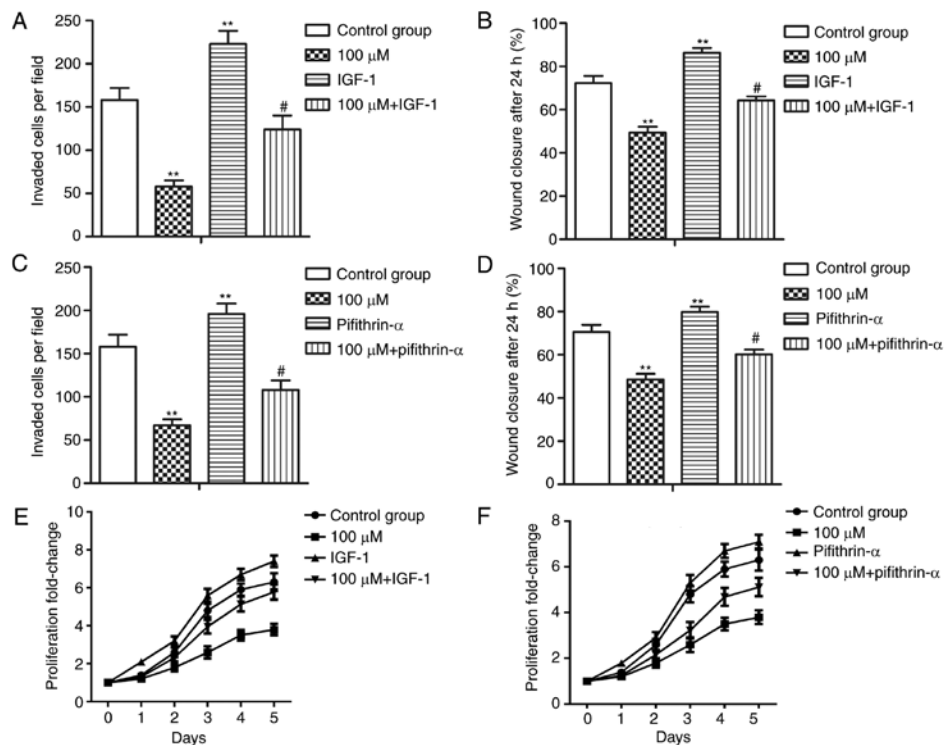


Figure 5. Pifithrin- α and IGF-1 counteract the inhibitory effect of resveratrol on the proliferation, invasion and migration of HCC cells. MHCC-97H cells were treated with 100 μ M resveratrol with or without pifithrin- α or IGF-1. (A) The invasion ability of MHCC-97H cells treated with 100 μ M resveratrol with or without pifithrin- α was detected using Transwell invasion assay. (B) Cell migration ability was detected using a wound healing assay. (C) The invasion ability of MHCC-97H cells treated with 100 μ M resveratrol with or without IGF-1 was detected using a Transwell invasion assay. (D) Cell migration ability was measured using a wound healing assay. (E) The proliferation of MHCC-97H cells treated with 100 μ M resveratrol with or without pifithrin- α was detected using a CCK-8 assay. (F) The proliferation of MHCC-97H cells treated with 100 μ M resveratrol with or without IGF-1 was detected using a CCK-8 assay. All data are presented as the mean \pm standard deviation of 3 independent experiments. ** P <0.01, compared with the control group; # P <0.05, compared with the 100 μ M group.

Although the relevance of autophagy with therapeutic efficacies of antitumor agents has been discussed, the conclusions are contradictory. For instance, Lamy *et al* (25) indicated that autophagy promoted cancer cell growth, which may counteract the effect of resveratrol in Ishikawa endometrial cancer cells. However, Mariño *et al* (26) determined that inhibition of autophagy attenuated resveratrol-induced ovarian cancer cell death. Autophagy-related proteins were detected through western blot analysis and autophagosomes were quantified in the present study. Beclin1 is a specific marker of autophagy and LC3 II, generated by enzymatic cleavage, indicates the initiation of autophagy while p62 is reported to clear the autophagosome (27,28). In agreement with the results of a study undertaken by Zhu *et al* (29), the present study observed dose-dependently enhanced autophagic activity in the resveratrol treatment group. Furthermore, inhibiting autophagy with 3-MA counteracted the inhibitory effects of resveratrol on proliferation, invasion and migration of HCC cells. The results indicated that resveratrol inhibited HCC progression through inducing autophagy, and enhancing autophagy can augment the antitumor effect of resveratrol in HCC.

As resveratrol can induce apoptosis and autophagy in cancer cells, the present study investigated the potential signaling pathway involved in these processes. Nuclear translocation of p53 is important for apoptotic induction. It was reported that p53 was crucial for resveratrol-induced apoptotic cell death in human ovarian cancer cells (30). The PI3K/Akt pathway is an important signaling pathway associated with autophagy (31,32). He *et al* (31) suggested that resveratrol suppressed renal cell carcinoma viability and migration while promoting apoptosis via the p53/AMPK/mTOR-induced autophagy signaling pathway. Deng *et al* (32) also reported that resveratrol-induced autophagy was regulated by the PI3K/Akt/mTOR pathway. Similarly, the present study demonstrated that resveratrol treatment activated p53 while inhibiting the PI3K/Akt pathway in HCC cells. In addition, p53 inhibitor pifithrin- α (30 μ M) and Akt activator IGF-1 treatment decreased the expression of Beclin1 and significantly inhibited the proliferation, invasion and migration of HCC cells. The aforementioned results demonstrated that resveratrol suppressed the proliferation and mobility of HCC cells through inducing autophagy via activating p53 while suppressing the PI3K/Akt signaling pathways. As mutant p53 cancer cell lines were more resistant to imiquimod-induced apoptosis than wild-type p53 cancer cell lines (33), our future studies will investigate the effect of resveratrol in HCC cells containing p53 mutant genes. It is worth investigating the role of resveratrol in converting mutant p53 protein to transcriptionally active wild-type p53.

Taken together, the results of the present study revealed that resveratrol inhibited the viability and mortality of HCC cells through inducing autophagy via regulating the p53 and PI3K/Akt pathways. Enhancing autophagy can augment the antitumor effects of resveratrol in HCC. Therefore, combined treatment with resveratrol and an autophagy inducer may be a viable option for treating HCC. To the best of our knowledge, the present study was the first to elucidated that autophagy was involved in the effect of resveratrol on HCC cells. Specifically, resveratrol could inhibit the progression of HCC through inducing autophagy via regulating the p53 and PI3K/Akt pathways.

Acknowledgements

The authors would like to thank the members of The Second Hospital of Dalian Medical University (Dalian, China), for providing technical support concerning the present study.

Funding

Funding information is not applicable.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

BZ analyzed and interpreted the data regarding autophagy. XY was responsible for conducting western blot and statistical analysis. SS was responsible for the design and drafting of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ and He J: Cancer statistics in China, 2015. *CA Cancer J Clin* 66: 115-132, 2016.
- Dugo EB, Yedjou CG, Stevens JJ and Tchounwou PB: Therapeutic potential of arsenic trioxide (ATO) in treatment of hepatocellular carcinoma: Role of oxidative stress in ATO-induced apoptosis. *Ann Clin Pathol* 5: 1101, 2017.
- Oh IS, Sinn DH, Kang TW, Lee MW, Kang W, Gwak GY, Paik YH, Choi MS, Lee JH, Koh KC, *et al*: Liver function assessment using albumin-bilirubin grade for patients with very early-stage hepatocellular carcinoma treated with radiofrequency ablation. *Dig Dis Sci* 62: 3235-3242, 2017.
- Kanda T, Ogasawara S, Chiba T, Haga Y, Omata M and Yokosuka O: Current management of patients with hepatocellular carcinoma. *World J Hepatol* 7: 1913-1920, 2015.
- Gomha SM, Edrees MM, Muhammad ZA and El-Reedy AA: 5-(Thiophen-2-yl)-1,3,4-thiadiazole derivatives: Synthesis, molecular docking and in vitro cytotoxicity evaluation as potential anticancer agents. *Drug Des Dev Ther* 12: 1511-1523, 2018.
- Rawat D, Shrivastava S, Naik RA, Chhonker SK, Mehrotra A and Koiri RK: An overview of natural plant products in the treatment of hepatocellular carcinoma. *Anticancer Agents Med Chem*: Jun 3, 2018 (Epub ahead of print). doi: 10.2174/1871520618666180604085612.
- Piotrowska H, Kucinska M and Murias M: Biological activity of piceatannol: Leaving the shadow of resveratrol. *Mutat Res* 750: 60-82, 2012.
- Johnson WD, Morrissey RL, Usborne AL, Kapetanovic I, Crowell JA, Muzzio M and McCormick DL: Subchronic oral toxicity and cardiovascular safety pharmacology studies of resveratrol, a naturally occurring polyphenol with cancer preventive activity. *Food Chem Toxicol* 49: 3319-3327, 2011.

9. Szekeres T, Saiko P, Fritzer-Szekeres M, Djavan B and Jager W: Chemopreventive effects of resveratrol and resveratrol derivatives. *Ann NY Acad Sci* 1215: 89-95, 2011.
10. Fabre KM, Saito K, DeGraff W, Sowers AL, Thetford A, Cook JA, Krishna MC and Mitchell JB: The effects of resveratrol and selected metabolites on the radiation and antioxidant response. *Cancer Biol Ther* 12: 915-923, 2011.
11. Kunimasa K, Ohta T, Tani H, Kato E, Eguchi R, Kaji K, Ikeda K, Mori H, Mori M, Tatefuji T, *et al*: Resveratrol derivative-rich melinjo (*Gnetum gnemon* L.) seed extract suppresses multiple angiogenesis-related endothelial cell functions and tumor angiogenesis. *Mol Nutri Food Res* 55: 1730-1734, 2011.
12. Li Q, Yue Y, Chen L, Xu C, Wang Y, Du L, Xue X, Liu Q, Wang Y and Fan F: Resveratrol sensitizes carfilzomib-induced apoptosis via promoting oxidative stress in multiple myeloma cells. *Front Pharmacol* 9: 334, 2018.
13. Fan Y, Chiu JF, Liu J, Deng Y, Xu C, Zhang J and Li G: Resveratrol induces autophagy-dependent apoptosis in HL-60 cells. *BMC Cancer* 18: 581, 2018.
14. Chang CH, Lee CY, Lu CC, Tsai FJ, Hsu YM, Tsao JW, Juan YN, Chiu HY, Yang JS and Wang CC: Resveratrol-induced autophagy and apoptosis in cisplatin-resistant human oral cancer CAR cells: A key role of AMPK and Akt/mTOR signaling. *Int J Oncol* 50: 873-882, 2017.
15. Mizushima N: Autophagy: Process and function. *Genes Dev* 21: 2861-2873, 2007.
16. Fan TF, Bu LL, Wang WM, Ma SR, Liu JF, Deng WW, Mao L, Yu GT, Huang CF, Liu B, *et al*: Tumor growth suppression by inhibiting both autophagy and STAT3 signaling in HNSCC. *Oncotarget* 6: 43581-43593, 2015.
17. Singh SK, Banerjee S, Acosta EP, Lillard JW and Singh R: Resveratrol induces cell cycle arrest and apoptosis with docetaxel in prostate cancer cells via a p53/p21^{WAF1/CIP1} and p27^{KIP1} pathway. *Oncotarget* 8: 17216-17228, 2017.
18. Chai R, Fu H, Zheng Z, Liu T, Ji S and Li G: Resveratrol inhibits proliferation and migration through SIRT1 mediated posttranslational modification of PI3K/AKT signaling in hepatocellular carcinoma cells. *Mol Med Rep* 16: 8037-8044, 2017.
19. Harikumar KB, Kunnumakkara AB, Sethi G, Diagaradjane P, Anand P, Pandey MK, Gelovani J, Krishnan S, Guha S and Aggarwal BB: Resveratrol, a multitargeted agent, can enhance antitumor activity of gemcitabine in vitro and in orthotopic mouse model of human pancreatic cancer. *Int J Cancer* 127: 257-268, 2010.
20. Bai Y, Mao QQ, Qin J, Zheng XY, Wang YB, Yang K, Shen HF and Xie LP: Resveratrol induces apoptosis and cell cycle arrest of human T24 bladder cancer cells in vitro and inhibits tumor growth in vivo. *Cancer Sci* 101: 488-493, 2010.
21. Ko JH, Sethi G, Um JY, Shanmugam MK, Arfuso F, Kumar AP, Bishayee A and Ahn KS: The role of resveratrol in cancer therapy. *Int J Mol Sci* 18: E2589, 2017.
22. Bai Y, Yang H, Zhang G, Hu L, Lei Y, Qin Y, Yang Y, Wang Q, Li R and Mao Q: Inhibitory effects of resveratrol on the adhesion, migration and invasion of human bladder cancer cells. *Mol Med Rep* 15: 885-889, 2017.
23. Mizushima N and Komatsu M: Autophagy: Renovation of cells and tissues. *Cell* 147: 728-741, 2011.
24. Guo JY, Xia B and White E: Autophagy-mediated tumor promotion. *Cell* 155: 1216-1219, 2013.
25. Lamy L, Ngo VN, Emre NC, Shaffer AL III, Yang Y, Tian E, Nair V, Kruhlak MJ, Zingone A, Landgren O, *et al*: Control of autophagic cell death by caspase-10 in multiple myeloma. *Cancer Cell* 23: 435-449, 2013.
26. Mariño G, Niso-Santano M, Baehrecke EH and Kroemer G: Self-consumption: The interplay of autophagy and apoptosis. *Nat Rev Mol Cell Biol* 15: 81-94, 2014.
27. Kadowaki M and Karim MR: Cytosolic LC3 ratio as a quantitative index of macroautophagy. *Methods Enzymol* 452: 199-213, 2009.
28. Wang S, Wang C, Yan F, Wang T, He Y, Li H, Xia Z and Zhang Z: N-acetylcysteine attenuates diabetic myocardial ischemia reperfusion injury through inhibiting excessive autophagy. *Mediators Inflamm* 2017: 9257291, 2017.
29. Zhu H, Ding J, Wu J, Liu T, Liang J, Tang Q and Jiao M: Resveratrol attenuates bone cancer pain through regulating the expression levels of ASIC3 and activating cell autophagy. *Acta Biochim Biophys Sin* 49: 1008-1014, 2017.
30. Lang F, Qin Z, Li F, Zhang H, Fang Z and Hao E: Apoptotic cell death induced by resveratrol is partially mediated by the autophagy pathway in human ovarian cancer cells. *PLoS One* 10: e0129196, 2015.
31. He W and Cheng Y: Inhibition of miR-20 promotes proliferation and autophagy in articular chondrocytes by PI3K/AKT/mTOR signaling pathway. *Biomed Pharmacother* 97: 607-615, 2018.
32. Deng F, Ma YX, Liang L, Zhang P and Feng J: The pro-apoptosis effect of sinomenine in renal carcinoma via inducing autophagy through inactivating PI3K/AKT/mTOR pathway. *Biomed Pharmacother* 97: 1269-1274, 2018.
33. Huang SW, Chang SH, Mu SW, Jiang HY, Wang ST, Kao JK, Huang JL, Wu CY, Chen YJ and Shieh JJ: Imiquimod activates p53-dependent apoptosis in a human basal cell carcinoma cell line. *J Dermatol Sci* 81: 182-191, 2016.