

# Resveratrol-induced *REG III* expression enhances chemo- and radiosensitivity in head and neck cancer in xenograft mice

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**Abstract.** Identifying the key molecules that enhance chemo- and radiosensitivity in head and neck squamous cell carcinoma (HNSCC) as well as reliable biomarkers for predicting recurrence and metastasis would be desirable to improve the prognosis of HNSCC. Previously, we have reported that *Regenerating gene III* (*REG III*) expression was associated with an improved survival rate for patients with HNSCC. In addition, resveratrol (3,4',5-trihydroxystilbene) significantly increased *REG III* expression in HNSCC cells, and significantly inhibited cell growth, enhanced chemo- and radiosensitivity, and blocked the cancer invasion of HNSCC cells *in vitro*. In the present study, the effect of resveratrol on cancer progression in HNSCC was investigated *in vivo* using a xenograft nude mouse model. The results revealed that resveratrol increased the mRNA level of *REG III* *in vivo*, which was in agreement with our previous *in vitro* findings. Furthermore, *REG III* increased the antitumor effect of radiation or cisplatin *in vivo*, and resveratrol sensitized HNSCC to irradiation and cisplatin *in vivo*. These results indicated that resveratrol could increase the efficacy of cisplatin and irradiation through the *REG III* expression pathway, resulting in the inhibition of HNSCC progression *in vivo*.

## Introduction

Head and neck squamous cell carcinoma (HNSCC) is the sixth most common cancer worldwide (1,2). Cancer in the head and neck region can arise from several functional disorders associated chewing, speech, swallowing and respiration.

Therefore, therapy for HNSCC is required to not only cure the disease, but to also preserve the function of the affected area in order to maintain the quality of life of patients with HNSCC. Despite the recent advancements in surgery, chemotherapy and radiotherapy, limited improvements in treating metastatic HNSCC have been achieved (3,4). Two-thirds of all patients present advanced stage III or IV tumors with low locoregional control rates and the long-term survival of patients with HNSCC has remained insufficient (5,6). Therefore, novel agents for potential alternative HNSCC treatments with greater efficacy are urgently required. The present study focused on the human regenerating gene (*REG*) as a reliable biomarker for predicting HNSCC progression. *REG* was first identified in regenerating pancreatic islets in studies on diabetology in 1988 (7). *REG* family proteins are classified into 4 subfamilies: Types I, II, III and IV. The human *REG* family consists of 5 members: *REG Ia*, *REG Ib*, *REG III*, *hepatocarcinoma-intestine-pancreas/pancreatitis-associated-protein* and *REG IV* (8-13). The *REG* family of proteins has been revealed to serve roles in normal tissue regeneration (14,15) and also the progression of various types of cancers, including esophageal, gastric, lung, liver, colorectal and prostate cancer (16-25). Recently, we reported that *REG III* expression was associated with improved survival rates for HNSCC, and that *REG III* enhanced chemo- and radiosensitivity *in vitro* (26). In addition, as *REG III* contributes to the improvement of the prognosis of HNSCC, in our previous study we searched for the substance that induced the expression of *REG III* and finally revealed that resveratrol (3,4',5-trihydroxy-*trans*-stilbene) significantly increased *REG III* expression in HNSCC cells, and also significantly inhibited cell growth, enhanced chemo- and radiosensitivity, and blocked HNSCC cell invasion *in vitro* (27). The aim of the present study was to investigate the effect of resveratrol on cancer progression in HNSCC *in vivo* for clinical application.

## Materials and methods

**Cell culture and reagent.** Human hypopharyngeal cell carcinoma FaDu cells were obtained from American Type Culture Collection (ATCC; Manassas, VA, USA) and grown

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and maintained in Dulbecco's modified Eagle's medium (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) containing 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.) and supplemented with 100 U/ml penicillin G and 100  $\mu$ g/ml streptomycin and 250 ng/ml amphotericin B (Antibiotic-Antimycotic; Gibco; Thermo Fisher Scientific, Inc.). The cells were incubated in 5% CO<sub>2</sub>/95% air with a humidified atmosphere at 37°C. 3,4',5-Trihydroxy-*trans*-stilbene (resveratrol) was obtained from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). Resveratrol was dissolved in dimethyl sulfoxide (DMSO; Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) and then diluted with normal saline to achieve the correct dose in 300  $\mu$ l of 2% DMSO.

**Reverse transcription-quantitative polymerase chain reaction (RT-qPCR).** Total RNA was isolated using a RNeasy Protect<sup>®</sup> Cell Mini kit (Qiagen GmbH, Hilden, Germany) from FaDu cells. cDNA was reverse-transcribed from 0.5–2  $\mu$ g samples of total RNA using a High Capacity cDNA Reverse Transcription kit (Applied Biosystems; Thermo Fisher Scientific, Inc.) as previously described (26–33). cDNA was subjected to RT-PCR with the following primers, which were synthesized and prepared by NGRL (Sendai, Japan):  $\beta$ -actin (NM\_001101) sense, 5'-GCGAGAAGATGACCCAGA-3' and antisense, 5'-CAG AGGCGTACAGGGATA-3'; and *REG III* (AB161037) sense 5'-GAATATTCTCCCCAACTG-3' and antisense, 5'-GAG AAAAGCCTGAAATGAAG-3'.

RT-qPCR was performed using the KAPA SYBR FAST qPCR Master Mix (Kapa Biosystems; Roche Diagnostics, Indianapolis, IN, USA) and Thermal Cycler Dice Real-Time System (Takara Bio, Inc., Otsu, Japan) as previously described (25–33). qPCR was performed with the following thermocycling conditions: An initial step of 3 min at 95°C followed by 40 cycles of 3 sec at 95°C and 20 sec at 60°C. The level of the target mRNA was normalized to the mRNA level of  $\beta$ -actin as an internal standard.

**Animals.** BALB/c nude male mice were purchased from CLEA Japan, Inc. (Tokyo, Japan). All protocols were approved by the Animal Care and Use Committee of Nara Medical University (Nara, Japan). Each cage housed 3 mice with food and water available *ad libitum* in a pathogen-free environment with a 12-h light/dark cycle.

**In vivo model for resveratrol-induced *REG III* expression in HNSCC cells.** A total of 6 BALB-c nude male mice were used in each experiment. FaDu cells (1 $\times$ 10<sup>6</sup> cells/100  $\mu$ l saline) were implanted subcutaneously into the right flanks of BALB/c nude male mice (4–5 weeks old). When the tumors reached ~100 mm<sup>3</sup> volume, the mice were randomly assigned into 2 groups (day 0): Vehicle control (normal saline and 2% DMSO) and resveratrol groups. Mice were treated intraperitoneally with resveratrol (100 mg/kg/day) or vehicle until the completion of the experiment. At 30 days following treatment, mice were sacrificed with intraperitoneal administration of pentobarbital (100 mg/kg) and the tumor tissues were harvested. The mRNA levels of intrinsic *REG III* were then examined in tumor tissues via RT-qPCR.

**In vivo model of *REG III*-induced chemo- and radiosensitivity.** A total of 12 BALB-c nude male mice (4–5 weeks old) were used to establish a xenograft model in each experiment. FaDu cells transfected with the *REG III* expression plasmid (FaDu-*REG III* cells) or the neomycin-resistance gene alone (FaDu-mock cells) were used as previously described (26). FaDu-*REG III* or FaDu-mock cells (1 $\times$ 10<sup>6</sup> cells in 100  $\mu$ l volume) were implanted subcutaneously in the right flanks of BALB/c nude male mice (4–5 weeks old). When the tumor sizes reached ~100 mm<sup>3</sup>, the mice were randomly assigned to the treatment groups (n=3/group; day 0). For chemosensitivity experiments, the treatment groups consisted of the vehicle control (normal saline) and cisplatin. On day 0, cisplatin (Nihon Kayaku Co., Tokyo, Japan; 4 mg/kg/week) or normal saline was administered intraperitoneally; a total of 4 injections of cisplatin were administered. For the radiosensitivity experiments, the treatment groups consisted of vehicle control (normal saline) and 6 Gy irradiation. Mice were exposed to radiation using a MBR-1520R system (Hitachi, Ltd., Tokyo, Japan) operated at 150 kV and 20 mA as previously described (27), which delivered the dose at 0.8 Gy/min. For both the chemo- and radiosensitivity experiments, the tumor size was assessed every 3 days. The tumor volume was calculated using the following formula: Volume=[Lx(W)<sup>2</sup>]/2, where L is the length and W is the width. At 24 days post-treatment, the mice were sacrificed with intraperitoneal administration of pentobarbital (100 mg/kg) and the tumor tissues were harvested.

**In vivo model for resveratrol-induced chemo- and radiosensitivity.** A total of 12 BALB-c nude male mice were used in each study. FaDu cells (1 $\times$ 10<sup>6</sup> cells/100  $\mu$ l saline) were implanted subcutaneously in the right flanks of BALB/c nude male mice (4–5 weeks old). When the tumors reached ~100 mm<sup>3</sup> volume, the mice were randomly assigned into the following treatment groups (n=3/group; day 0). For the chemosensitivity experiments, the treatment groups consisted of the vehicle control (normal saline and 2% DMSO), vehicle and cisplatin, resveratrol alone and cisplatin + resveratrol. On day 0, the mice were treated intraperitoneally with resveratrol or vehicle until the completion of the experiment; resveratrol was administered at doses of 100 mg/kg for 30 consecutive days. Cisplatin (3 mg/kg/week) or normal saline was administered intraperitoneally on day 0; a total of 4 cycles were administered. For the radiosensitivity experiments, the treatment groups consisted of the vehicle control (normal saline and 2% DMSO), vehicle and 6 Gy irradiation, resveratrol (100 mg/kg/day) alone and 6 Gy irradiation + resveratrol (100 mg/kg/day). For both the chemo- and radiosensitivity experiments, the tumor size was measured every 3 days. Following 30 days of treatment, the mice were sacrificed and the tumor tissues were harvested.

**Statistical analysis.** Data were expressed as the mean  $\pm$  standard error. Statistically significant differences between groups were determined by Student's t-test using StatMate IV (Abacus Concepts, Piscataway, NJ, USA). P<0.05 was considered to indicate a statistically significant difference.

## Results

**Induction of *REG III* mRNA by resveratrol in the tumor tissues of a xenograft mouse model.** To elucidate the

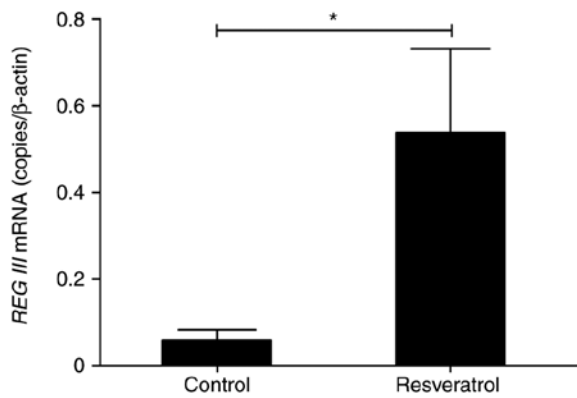


Figure 1. mRNA levels of *REG III* in the tumor tissues of xenograft nude mice treated with vehicle control or resveratrol. Nude male mice were implanted subcutaneously with FaDu cells. Mice were treated intraperitoneally with or without resveratrol. Following 30 days of treatment, tumor tissues were harvested and the mRNA levels of intrinsic *REG III* in tumor tissues were examined by reverse transcription-quantitative polymerase chain reaction. The ratios of the levels of target mRNA normalized to  $\beta$ -actin mRNA are displayed and data are presented as the mean  $\pm$  standard error of each group ( $n=3$ ). \* $P<0.05$ , as indicated. REG, regenerating gene.

induction of *REG III* in HNSCC cells by resveratrol *in vivo*, the present study investigated the levels of *REG III* mRNA in xenografted FaDu cells with or without resveratrol. The mRNA levels of *REG III* in the resveratrol-treated tumor tissues were significantly increased by resveratrol when compared with the untreated tumor tissues, which were used as the control (Fig. 1). This result was in agreement with the *in vitro* results we previously reported (27). Therefore, resveratrol induced *REG III* mRNA expression *in vivo* as well as *in vitro* in HNSCC.

*REG III* enhances the efficacy of radiation or cisplatin therapy in a HNSCC xenograft mouse model. To evaluate the effect of *REG III* on chemo- and radiosensitivity in HNSCC *in vivo*, the present study established a HNSCC xenograft model of BALB/c nude mice. An animal xenograft model was generated via injections in nude mice with FaDu cells stably transfected with the *REG III* expression plasmid (FaDu-REG III) or FaDu cells transfected the neomycin-resistance gene alone (FaDu-mock) as the control into the right flank. When tumors reached  $\sim 100 \text{ mm}^3$  following 2 weeks, the mice were randomly assigned into 2 groups: Non-treatment, and irradiation or cisplatin therapy groups (day 0), each specific treatment was then applied as aforementioned (Fig. 2A). The efficacy of the radiation or cisplatin therapy was evaluated by monitoring the tumor volume. Regarding the effect of radiation, there were no significant differences in tumor volume between the FaDu-REG III and FaDu-mock in the non-treatment groups. However, in the radiation groups the tumor volume of the FaDu-REG III-treated mice was significantly inhibited when compared with FaDu-mock from day 21 (Fig. 2B-D). Regarding the effect of cisplatin therapy, significant inhibition of tumor progression was observed in the FaDu-REG III group when compared with the FaDu-mock-treated groups from day 15, which corresponded with the results in the radiation groups (Fig. 2B-D). These results indicated that *REG III* enhanced chemo- and radiosensitivity in HNSCC *in vivo* as well as *in vitro*, as previously described (26).

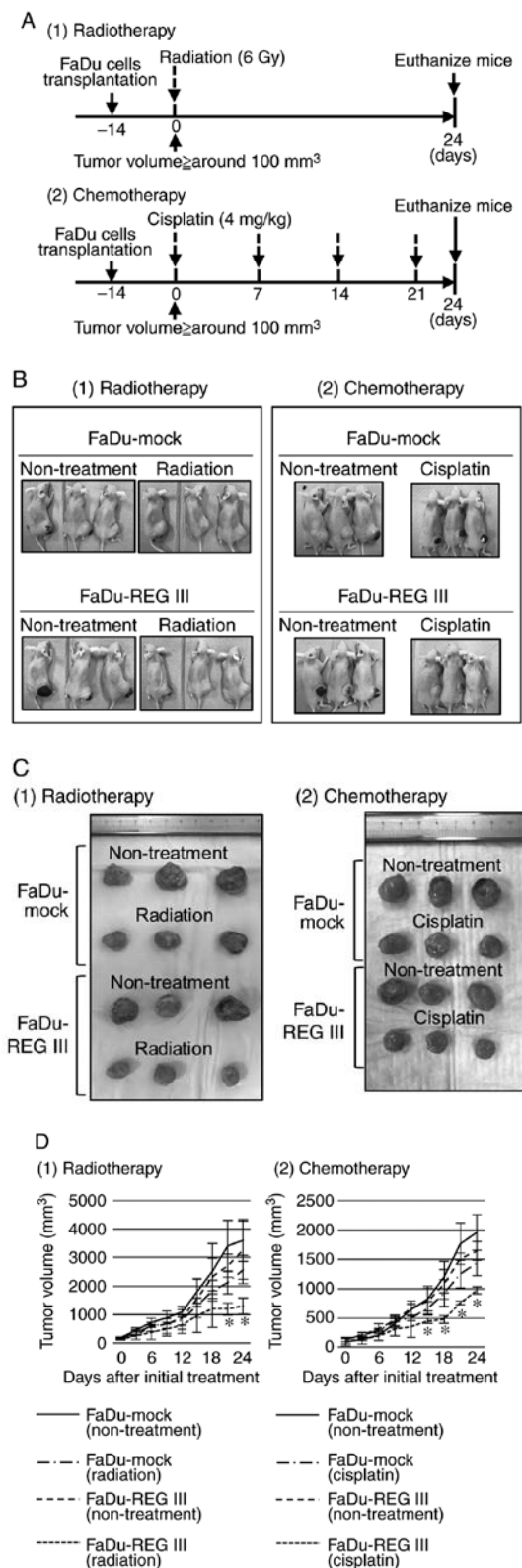


Figure 2. Enhancement of chemo- and radiosensitivity by *REG III* expression in xenograft tumors. A xenograft nude mouse model was generated via the injection of FaDu cells transfected with the *REG III* expression plasmid (FaDu-REG III) or FaDu cells transfected the neomycin-resistance gene alone (FaDu-mock). The mice were randomly assigned into 2 groups: Non-treatment and irradiation therapy (6 Gy), or cisplatin therapy (4 mg/kg/week). (A) The animals were treated as indicated and euthanized on day 24 of treatment. (B) Images of tumor-bearing mice in each group ( $n=3$ /group). (C) Images of the excised tumors in each group. (D) The tumor volumes in mice were measured during the course of the experiment (every 3 days) and were calculated using the formula:  $V=[L \times W^2]/2$ . Statistical significance was calculated by Student's t-test. \* $P<0.05$ . REG, regenerating gene.

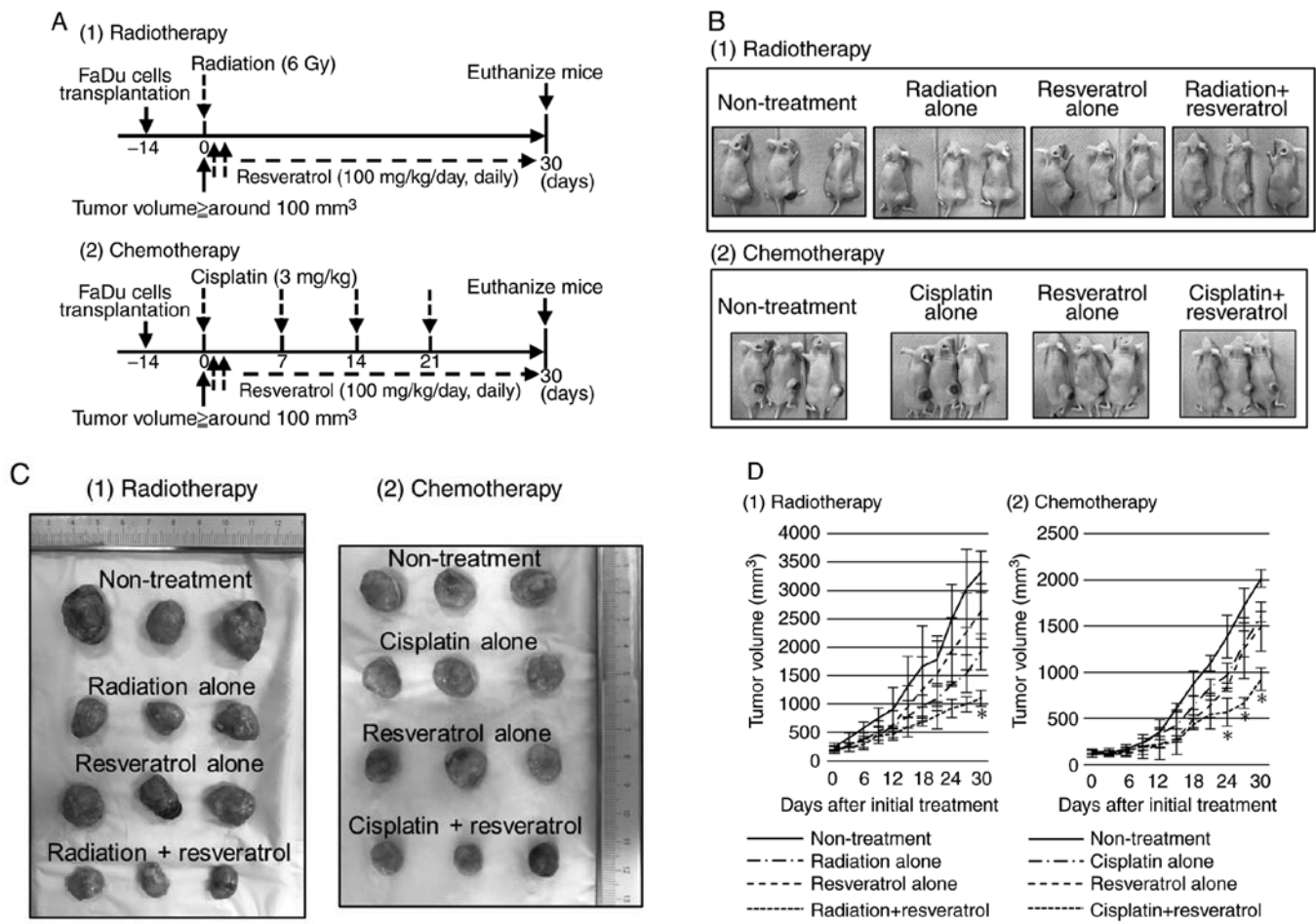


Figure 3. Resveratrol potentiates the effect of irradiation or cisplatin to inhibit the growth of human head and neck squamous cell carcinoma *in vivo*. Xenograft tumors were established by subcutaneous injection of FaDu cells into BALB/c mice. Mice were randomly divided into 4 groups ( $n=3$ /group). For radiotherapy experiments, the non-treatment group, radiation (6 Gy) alone group, resveratrol (100 mg/kg/day) alone group, and radiation (6 Gy) with resveratrol (100 mg/kg/day) group were applied. For chemotherapy, the non-treatment group, cisplatin (3 mg/kg/week) alone group, resveratrol (100 mg/kg/day) alone group, and cisplatin (3 mg/kg/week) with resveratrol (100 mg/kg/day) group were used. (A) A schematic representation of the experimental protocol. The animals were euthanized on day 30 of treatment. Images of (B) tumor-bearing mice and (C) excised tumors. (D) The tumor volumes in mice measured during the course of the experiment (every 3 days). Statistical significance was calculated by Student's *t*-test. \* $P<0.05$ .

*Irradiation or cisplatin therapy with resveratrol synergistically inhibits HNSCC xenograft tumor growth in vivo.* To assess the *in vivo* therapeutic potential of resveratrol, the present study examined tumor progression using a HNSCC xenograft model of BALB/c nude mice. Tumors in the right flanks of the mice were established for 2 weeks prior to the initiation of the treatments, then the mice were randomly assigned into 4 groups for experiments evaluating cisplatin therapy or irradiation, respectively (day 0; Fig. 3A). The potential of the treatment was evaluated by assessing the tumor volume. Regarding the potential of radiation therapy, mice in the resveratrol alone group had smaller tumor volumes than mice in the control group; however, the difference was not statistically significant. Mice in the radiation with resveratrol group had significantly smaller tumor volumes than mice in the control group. In addition, the radiation with resveratrol group exhibited significant antitumor effects when compared with the resveratrol alone group, or the radiation alone group on day 30 (Fig. 3B-D). Regarding the potential of cisplatin therapy, mice in the cisplatin alone and cisplatin with resveratrol groups had significantly smaller tumor volumes than mice in the control group. In addition, the cisplatin with resveratrol

group exhibited significant antitumor effects when compared with the resveratrol alone or cisplatin alone groups from day 24 (Fig. 3B-D). These results indicated that resveratrol enhanced chemo- and radiosensitivity in HNSCC *in vivo*, which was in agreement with the effects observed with *REG III* *in vivo*.

## Discussion

Despite the progression in the current cancer treatments available, such as surgery, radiation and chemotherapy, these have not been effective in improving the survival rate of HNSCC, particularly hypopharyngeal carcinoma (1-6). Chemo- and radioresistance can cause local recurrence and distant metastasis, which are associated with poor prognosis. Therefore, identification of reliable biomarkers that enhance sensitivity for the chemo- and radiotherapy of HNSCC is highly desirable to improve prognosis. As a biomarker of HNSCC, we have focused on *REG*, whose family of proteins have been associated with diseases such as chronic inflammation and malignant tumors (16-26).

We have previously reported that *REG III* expression was associated with an improved survival rate for patients with

HNSCC (26). In addition, resveratrol significantly increased *REG III* expression, could enhance the chemo- and radiosensitivity, and inhibit cancer progression through the *REG III* expression pathway in HNSCC cells *in vitro* (27). In the present study, the effect of resveratrol on cancer progression in HNSCC through the *REG III* expression pathway *in vivo* was investigated. The results of the present study demonstrated that resveratrol increased the mRNA level of *REG III* *in vivo*, which corresponded with our *in vitro* results previously reported (27). This result highlights the potential of resveratrol in inducing the *REG III* expression pathway *in vivo*.

Resveratrol (3,4',5-trihydroxystilbene) is a polyphenolic compound found in grapes and other food products that provides a number of anti-aging health benefits against metabolism, cardiovascular disease and carcinogenesis (34-37). Various previous studies indicated that resveratrol enhanced the sensitivity of chemo- or radiotherapy (38-42). Furthermore, some studies have demonstrated that resveratrol significantly decreased tumor progression (39,40,43). Such *in vitro* studies on the associations between resveratrol and cancer have been reported (34-37,44-50), while, to the best of our knowledge, there are fewer *in vivo* studies.

In the present study, a HNSCC xenograft nude mouse model was established to evaluate the effect of *REG III* on cancer progression in HNSCC *in vivo*. It was demonstrated that *REG III* increased the antitumor effect of radiation or cisplatin *in vivo*. In addition, the *in vivo* therapeutic potential of resveratrol was evaluated, and the results revealed that it significantly sensitized HNSCC to irradiation and cisplatin *in vivo*, although resveratrol is not likely to be a primary treatment for HNSCC. These results indicate that resveratrol has potential for use as an adjuvant anticancer therapy in HNSCC. In our previous studies, similar results were obtained in HSC-4 cells as well as FaDu cells (27). In terms of its effect for HNSCC *in vivo*, further studies whether HSC-4 cells have similar results based on the results of this study are required in the future. Moreover, it is necessary to consider which combination of chemotherapy, radiotherapy and resveratrol is the most effective treatment. The present study performed *in vivo* experiments using a xenograft mouse model. For clinical applications associated with the administration of resveratrol, experiments on an orthotopic transplant mouse model may be required in order to take into consideration the micro-environment in the future. Concerning the bioavailability of resveratrol, recent studies have indicated that resveratrol induces apoptosis or autophagy in several human cancer cell lines and in an animal model of carcinogenesis (49,50). It has been reported that resveratrol induces cell apoptosis and cell cycle arrest via the caspase/cyclin-cyclin dependent kinase (CDK) signaling pathway (49). The present study investigated the anti-carcinogenic potential of resveratrol by analyzing the *REG III* expression pathway. Preliminary experiments were performed to investigate the anti-carcinogenic mechanism of resveratrol, by analyzing the expression of apoptosis-related proteins. These preliminary results revealed that both resveratrol and *REG III* decreased the expression of cyclin D1, B-cell lymphoma-xL and activated caspase-3 compared with the control (data not shown). This indicated that resveratrol may inhibit cancer progression through the *REG III* pathway via the caspase/cyclin-CDK signaling pathway. However, the

underlying mechanism of how resveratrol enhances *REG III*, and how *REG III* enhances the chemo- and radiosensitivity of HNSCC remains unknown. Therefore, further studies are required in the future.

In conclusion, the results of the present study indicate that resveratrol increases the efficacy of cisplatin and irradiation through the *REG III* expression pathway, resulting in the inhibition of tumor growth, in treating HNSCC *in vivo*. The present study provides support for clinical trials using resveratrol as an adjuvant anticancer therapy and may help improve human HNSCC prognosis. However, additional studies will be required to fully define the therapeutic potential of resveratrol for HNSCC.

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## Availability of data and materials

The datasets used during the present study are available from the corresponding author upon reasonable request.

## Authors' contributions

IO, SM and ST conceived and designed the research. SM, TM and TU conducted the experiments. SM, HO, TKim and TU were involved in the analysis and interpretation of data. SM wrote the manuscript. IO and SM revised the manuscript for important intellectual content. TKit and IO were involved in editing the manuscript. All authors read and approved the final manuscript.

## Ethics approval and consent to participate

All protocols were approved by the Animal Care and Use Committee of Nara Medical University (Nara, Japan). All procedures performed in the studies were in accordance with the 1964 Declaration of Helsinki and its later amendments.

## Patient consent for publication

Not applicable.

## Competing interests

The authors state that they have no competing interests.

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