

Molecular alterations and clinical relevance in cervical carcinoma and precursors (Review)

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Received May 19, 2020; Accepted September 22, 2020

DOI: 10.3892/or.2020.7804

Abstract. Cervical cancer is one of the most common types of cancer and the fourth leading cause of cancer-related deaths in women. The occurrence and development of cervical cancer is a multifactorial and multilevel process, which usually occurs alongside a continuous high-risk human papillomavirus infection. With further developments in molecular biology and the advancement of sequencing technology, the role of biomarkers in cervical diseases has been gradually recognized. Therefore, it remains a priority to identify key molecular markers that can be used for the screening and triaging of the lesions. In recent years, numerous studies have been conducted in order to identify important markers for cervical diseases. The present review aimed to summarize the molecular alterations and clinical relevance of chromosomal alterations, DNA polymorphisms, the DNA methylation status, histone modifications, and alterations in microRNA and protein expression levels. Accumulating evidence suggests that molecular alterations may reflect the degree and the prognosis of the disease. Although significant progress has been made in the field of cervical cancer research, further samples and experiments are still required to identify crucial molecules.

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1. Introduction

Cervical cancer is one of the ten most common types of malignancies affecting women. According to the cancer statistics in 2018, there are ~570,000 new cases of cervical cancer and 311,000 deaths due to cervical cancer worldwide each year (1). In addition, Chinese cancer data have estimated ~98,500 cases and 30,500 deaths from cervical cancer, accounting for 17% of cases and 10% of deaths globally. Contrary to the decreasing trend of morbidity in developed countries, the incidence rates of cervical cancer in China have increased significantly since 2000 (2).

The major histological type of cervical cancer is squamous carcinoma of the cervix (SCC). SCC has been confirmed to be caused by high-risk human papillomavirus (HR-HPV) infection. Half of HPV infections are cleared within 6-12 months; however, 10% of HPV infections persist (3). Following HR-HPV infection, cervical cells may undergo the precursor steps of SCC, which are termed squamous intraepithelial lesions (SILs) (4). SILs are classified into low grade SILs (LSILs) and high grade SILs (HSILs), which correspond to the traditional histological classification, known as cervical intraepithelial neoplasia (CIN). CIN1 is the synonym for LSIL, and CIN2 and CIN3 are classified as HSILs (5). The histological diagnosis of CIN is the gold standard to guide subsequent treatment; however, the reproducibility of CIN diagnosis is poor, especially for CIN2, with a diagnostic consistency rate of <50% (6). Moreover, the prognosis of CIN is different. According to a survey, the 10-year transition probability from CIN1 to CIN2 was 4.37%, and from CIN2 to CIN3+ was 25.58% (7). In addition, a recent meta-analysis revealed that the regression rate of CIN2 after a 24-month follow-up was 50% (11 studies, 819/1,470 women) and 60% (4 studies, 638/1,069 women, age <30 years), respectively. Patients with CIN2 who have a plan for future pregnancies can attend screening tests and there is no requirement for immediate treatment (8).

It is imperative to identify molecular markers for the screening and triage of cervical cancer and precancerous lesions. For patients with cervical precancerous lesions,

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Key words: cervical cancer, cervical squamous intraepithelial lesions, precursors, biomarkers, molecules

the identified biomarkers may predict the development of the disease and help to guide subsequent treatments and avoid overtreatment. For patients with cervical cancer, the biomarkers could help to predict the prognosis and potentially be used as therapeutic targets.

For both cervical cancer and precursors lesions, the majority of previous research has investigated the molecular alterations and clinical relevance using PCR, fluorescence *in situ* hybridization, microarrays, ELISAs, western blotting and immunohistochemistry. The present review aimed to provide a summary of the progression of cervical cancer and precursor lesions due to the alteration of chromosomes, DNA polymorphisms, the DNA methylation status, histone modifications, and alterations in the expression levels of microRNAs (miRNAs) and proteins.

2. Chromosomal alterations

The presence of chromosomal aberrations has been confirmed in SCC and its precursors. For example, Policht *et al* (9) reported the gain or loss in copies of 8q24, Xp22, 20q13, 3p14, 3q26 and CEP15 in the cervical tissue of CIN and cancer lesions, and it was further reported that 8q24 and 3q26 were the most useful molecules for detecting HSILs and SCCs. Rodolakis *et al* (10) analyzed the gain of 3q26 in 40 patients and discovered that none of the 3q26(-) progressed to HSILs/CIN2⁺ after 17.5 months, while 38% of the 3q26(+) patients progressed. The study also revealed that the gain of 3q26 could predict the progression with a negative predictive value (NPV) of 100%. Another meta-analysis indicated a potential association between the gain of 3q26 and disease prognosis (8 studies, 407 patients), with positive predictive values ranging from 50 to 93% and a NPV ranging from 75 to 100% (11). In addition to 3q26, gains in 5p15 were also identified in cervical lesions of increasing severity (12). Based on the above findings, 3q26 was hypothesized to have an important role in cervical cancer and it should be further studied. The details of these molecules are presented in Table I.

3. DNA polymorphisms

DNA polymorphisms are a type of genetic variation that do not change the gene expression levels. Cezar-Dos-Santos *et al* (13) reported that the forkhead box P3 (*FOXP3*) rs3761548 homozygous genotype may be associated with the resistance to HPV infection, while the rs2232365 homozygous genotype (G/G) was a risk factor for HPV infection [odds ratio (OR)=2.10 (95% confidence interval (CI): 1.06-4.15)]. In addition, the Arg72Arg genotype and Arg72 alleles of tumor protein p53 (*TP53*) were also suggested to be related to the susceptibility of HPV infection [OR=1.85 (95% CI: 1.03-3.32) and 1.94 (95% CI: 1.20-3.15), respectively] (14).

The relationship between genetic polymorphisms and cervical cancer susceptibility has also been studied. Chen *et al* (15) suggested that polymerase II polypeptide E (*POLR2E*) may be associated with the susceptibility of cervical cancer and breast cancer. For Uyghur women, the apolipoprotein B mRNA editing enzyme-catalytic polypeptide-like 3G (*APOBEC3G*) and interleukin-1 β (*IL1B*) polymorphisms were discovered to be associated with the

susceptibility of cervical cancer (16,17). For Han Chinese women, the NAD(P)H: Quinone oxidoreductase 1 (*NQO1*) rs1800566 TT genotype presented with an increased risk of cervical cancer development compared with the CT and CC genotypes (18). Notably, there seems to be ethnic differences in the presence of DNA polymorphisms; for instance, according to a meta-analysis, the cytotoxic T-lymphocyte associated antigen-4 (*CTLA4*) gene rs5742909 polymorphism was related to the susceptibility of cervical cancer in Asians, but it had little association with cervical cancer in Caucasians (19). Similarly, genetic polymorphisms, such as in the deoxyuridine triphosphatase (*DUT*) gene, were also discovered to be associated with HSIL susceptibility (20). Thus, DNA polymorphisms have been suggested to serve a predictive role for the susceptibility of cervical lesions. Predicting the early occurrence of cervical cancer can help prevent its occurrence, thus it is worthy of research. The details of these findings are presented in Table II.

4. DNA methylation

Sakane *et al* (21) investigated the methylation of distal-less homeobox 4 (*DLX4*) and SIM bHLH transcription factor 1 (*SIMI*) in LSILs; significant differences were identified in the methylation frequency of *DLX4* and *SIMI* between LSILs that persisted for >1 year and LSILs that progressed to HSILs within a year (P=0.044 and P=0.005, respectively). LSIL cases with *SIMI* methylation were identified to progress to HSILs faster compared with DNA methylation-negative cases (P=0.033). According to a meta-analysis of 1,055 patients in 7 studies, paired box gene 1 (*PAX1*) methylation was also discovered to be a protective factor for CIN1 to CIN2/3 progression and CIN2/3 to cervical cancer progression, demonstrating an OR of 0.09 and 0.16, respectively (22). Through studying plasma samples, the methylation of maternally expressed 3 (*MEG3*) in CIN3 and cervical cancer was identified to be significantly increased compared with that in healthy controls, exhibiting an OR of 13.033 and 17.100, respectively. In addition, the methylation status of *MEG3* was increased in cervical cancer tissues compared with normal tissues, which indicated that the methylation status of *MEG3* may have a diagnostic value in plasma and tissues (23).

In another study, the methylation patterns of 15 genes in the normal cervix and CIN1-3 cervixes were analyzed using quantitative methylation-specific PCR. The methylation of hsa-miR-124-2, SRY-box transcription factor 1 (*SOX1*), telomerase reverse transcriptase (*TERT*) and LIM homeobox transcription factor 1- α (*LMX1A*) genes were discovered to be independent predictors associated with the diagnosis of high-grade cervical lesions, exhibiting ORs of 5.1, 2.8, 2.2, 2.0, respectively (24). Verlaet *et al* (25) discovered that the methylation of growth hormone secretagogue receptor (*GHSR*), somatostatin (*SST*) and Zic family member 1 (*ZIC1*) were also associated with gain in 3q and an increased severity of cervical lesions (P<0.005). Finally, De Strooper *et al* (26) followed 1,040 HPV-positive women for 14 years and discovered that women with negative family with sequence similarity 19 (chemokine (C-C)-motif)-like-member A4 (*FAM19A4*)/miR-124-2 methylation had a lower risk of cervical cancer. The findings described above are presented in Table III.

Table I. Chromosomal alterations in cervical disease.

Name	Population (Refs.)	Sample	Cases	Methods	Potential role	Alteration
8q24						↑
3q26	USA (9)	NC, CIN, SCC	136	FISH	Diagnosis	↑
3q26	Slovakia (12)	NC, LSIL, HSIL, SCC/AC	131	FISH	Diagnosis	↑
5p15						↑
3q26	Greece (10)	ASCUS/LSIL	40	FISH	Prognosis	-
3q26	Norway (11)	CIN2/3	19	FISH	Prognosis	-

NC, normal cervix; CIN, cervical intraepithelial neoplasia; SCC, squamous carcinoma of cervix; ASCUS, atypical squamous cells of undetermined significance; LSIL, low grade squamous intraepithelial lesion; HSIL, high grade squamous intraepithelial lesion; AC, adenocarcinoma; ASC, adenosquamous carcinoma; FISH, fluorescence *in situ* hybridization. ↑, indicates that the molecule is upregulated in cervical diseases. -, indicates that the amount of molecules was not compared in different tissues.

Table II. DNA polymorphisms in cervical disease.

Name	Variation	Population (Refs.)	Cases	Methods
Association with HPV infection				
<i>FOXP3</i>	rs3761548, rs2232365	Brazil (13)	426	PCR
<i>TP53</i>	rs1042522	Kyrgyz (14)	205	PCR
Association with cancer susceptibility				
<i>POLR2E</i>	rs3787016	China (15)	884	PCR
<i>APOBEC3G</i>	rs5757465	Uygur (16)	529	First-generation
<i>IL1B</i>	rs1143627	Uygur (17)	569	PCR
<i>NQO1</i>	rs1800566	China (18)	1,018	PCR
<i>CTLA4</i>	rs5742909	Asian (19)	8,507	Meta ^a
<i>DUT</i>	rs3784619, rs11637235	China (20)	2,000	PCR
Association with CIN3 susceptibility				
<i>FOXP3</i>	rs3761548	Brazil (13)	426	PCR
<i>DUT</i>	rs3784619, rs11637235	China (20)	2,000	PCR

FOXP3, forkhead box P3; *TP53*, tumor protein p53; *POLR2E*, polymerase II polypeptide E; *APOBEC3G*, apolipoprotein B mRNA editing enzyme-catalytic polypeptide-like 3G; *IL1B*, interleukin-1β; *NQO1*, NAD(P)H: Quinone oxidoreductase 1; *CTLA4*, cytotoxic T-lymphocyte associated antigen-4; *DUT*, deoxyuridine triphosphatase; PCR, polymerase chain reaction. ^aMeta-analysis study.

5. Histone modifications

Histone modifications involve processes in which histones undergo acetylation, methylation or other modifications under the action of related enzymes. Upon analyzing the expression levels of histone H3 acetyl K9 (H3K9ac) and histone H3 tri methyl K4 (H3K4me3) in cervical cancer, Beyer *et al* (27) discovered that both histones were related to the clinicopathological variables of patients. In addition, the staining intensity of H3K9ac was also identified to be associated with the 10-year survival rate. These findings revealed the important role of histone acetylation and methylation in cervical cancer. Zhang *et al* (28) also discovered that HPV 18 E6/E7 enhanced the transcriptional activity of enhancer of zeste homolog 2 (*EZH2*), thereby enhancing the expression levels of histone 3 tri methyl K27 (H3K27me3) and exerting a positive effect on the development of cervical cancer. Polycomb repressive complex 2 (PRC2) can also catalyze the methylation

of histones, thereby inhibiting gene expression. Shi *et al* (29) identified C10ORF12 as an interactor of PRC2, which was found to positively regulate H3K27me3 modifications. At present, inhibitors for enzymes controlling histone modifications have been developed and are being used in clinical cancer treatment (30). However, to the best of our knowledge, related studies in cervical cancer are rare. Therefore, further research into histone modifications in cervical cancer is required.

6. miRNA alterations

Zeng *et al* (31) compared the expression levels of nine miRNAs in normal cervical, LSIL, HSIL and cervical cancer tissues; the results revealed that in cervical cancer, miR-218 expression levels were downregulated by 0.175-fold ($P=0.002$), while miR-21 expression levels were upregulated by 5.677-fold ($P=0.001$) compared with the normal tissues. Zhu *et al* (32) discovered that the upregulation of miR-21-5p

Table III. DNA methylation in cervical disease.

Name	Population (Refs.)	Sample	Cases	Methods	Alteration
<i>DLX4</i>					↑
<i>SIM1</i>	Japan (21)	NC, CIN, SCC	113	PCR, IHC	↑
<i>PAX1</i>	Meta ^a (22)	NC, CIN, SCC/AC	1,055	Meta ^a	↓
<i>MEG3</i>	China (23)	NC, CIN, SCC, AC	168	MSP	↑
<i>hsa-miR-124-2</i>	Brazil (24)	NC, CIN	447	PCR	↑
<i>SOX1</i>					↑
<i>TERT</i>					↑
<i>LMX1A</i>					↑
<i>GHSR</i>	The Netherlands (25)	NC, CIN, SCC	233	NGS	↑
<i>SST</i>					↑
<i>ZIC1</i>					↑
<i>FAM19A4/miR-124-2</i>	The Netherlands (26)	LSIL	1,040	PCR	-

DLX4, distal-less homeobox 4; *SIM1*, SIM bHLH transcription factor 1; *PAX1*, paired box gene 1; *MEG3*, maternally expressed 3; *SOX1*, SRY-box transcription factor 1; *TERT*, telomerase reverse transcriptase; *LMX1A*, LIM homeobox transcription factor 1- α ; *GHSR*, growth hormone secretagogue receptor; *SST*, somatostatin; *ZIC1*, Zic family member 1; *FAM19A4*, family with sequence similarity 19 (chemokine (C-C)-motif)-like)-member A4; NC, normal cervix; CIN, cervical intraepithelial neoplasia; SCC, squamous carcinoma of cervix; AC, adenocarcinoma; PCR, polymerase chain reaction; IHC, immunohistochemistry; NGS, 'next-generation' sequencing technology; MSP, methylation-specific polymerase chain reaction. ^aMeta-analysis study. ↑, indicates that the molecule is upregulated in cervical diseases; ↓, indicates that the molecule is downregulated in cervical diseases; -, indicates that the amount of molecules was not compared in different tissues.

expression levels and the downregulation of miR-34a expression levels were associated with the severity of cervical lesions ($P < 0.05$). In addition, miR-409-3p was negatively associated with E6 mRNA, and subsequent cell experiments revealed that it exerted an inhibitory effect on cervical cancer cells (33).

Recently, numerous studies have focused on the expression levels of miRNAs and their target genes in cervical cancer. Jin *et al* (34) compared the expression levels of miR-612 in normal and cancerous cervical tissues and cells, and discovered that they were downregulated in cancer tissues and cells, and that the target of miR-612 was nin one binding protein (*NOB1*). Zhao *et al* (35) reported that miR-15a-5p expression levels were upregulated in cervical cancer, and TP53 regulated inhibitor of apoptosis 1 (*TP53INP1*) was identified as the target gene. In fact, numerous miRNAs have been discovered to be downregulated in cervical cancer, including miR-889-3p (36), miR-299-3p (37), miR-140-3p (38), miR-505-5p (39), miR-877 (40), miR-636 (41), miR-144-3p (42), miR-139-5p (43), miR-126 (44), miR-138 (45), miR-526b (46), miR-432 (47), miR-543 (48) and miR-503 (49). Conversely, miRNAs that have been identified to be upregulated in cervical cancer include miR-93-5p (50) and miR-150-5p (51).

In addition, the detection of miRNA in the blood has also been suggested as a feasible method to diagnose cervical diseases. For example, the expression levels of miR-3142 in the serum of patients with cervical cancer were reported to be significantly upregulated compared with these levels in healthy individuals, and the high expression levels of miR-3142 were associated with a poor prognosis (52). In addition, Zheng *et al* (53) performed miRNA sequencing of plasma samples and screened out two significant miRNAs, let-7d-3p and miR-30d-5p; these two miRNAs were discovered to be

able to distinguish between CIN1⁻ and CIN2⁺ lesions [area under the curve (AUC)=0.828].

The details of these studies described above are presented in Table IV. It is worth mentioning that previous studies investigating the therapeutic ability of miRNAs in treating cancer have been performed, such as for the treatment of liver and breast cancer (54). However, there still remains a long way to go for the clinical application of miRNAs for the treatment of cervical cancer.

7. Protein alterations

The effect of the p16INK4a (p16), Ki-67 and cytokeratin 7 (CK7) proteins have been studied in cervical cancer and precancerous lesions. p16 is a tumor-suppressor protein that serves an important role in cell cycle regulation by decelerating the progression of cells from the G₁ phase to S phase. Ki-67 is a protein that is present during the active phase of the cell cycle and is involved in the proliferative activity of tumors. High p16 expression levels and >50% of Ki-67 expression in CIN2 lesions was discovered to have a higher probability of progressing to CIN3 and cancerous lesions ($P < 0.001$), with a hazard ratio of 2.58 and 2.84, respectively (55). Another study demonstrated that all of the HSIL/CIN2 patients with p16-negative expression had either regressed to normal or CIN1 tissue during the 12 months of follow-up, while both persistent and progressive CIN2 lesions were p16-positive (56). Therefore, these findings suggested that p16 and Ki-67 may be used to predict the outcome of CIN2.

While it is controversial to predict the outcome of CIN1, a follow-up study of an average of 28 months revealed that p16 staining had limited value in predicting the progression of LSILs to higher-grade lesions (57). In addition, HPV16/18 was discovered

Table IV. miRNAs in cervical disease.

Name	Population (Refs.)	Sample	Cases	Methods	Target gene	Alteration
miR-612	China (34)	NC, CC	52	PCR	<i>NOB1</i>	↓
miR-15a-5p	China (35)	NC, CC	30	PCR	<i>TP53INP1</i>	↑
miR-889-3p	China (36)	NC, CC	49	PCR	<i>FGFR2</i>	↓
miR-299-3p	China (37)	Cell lines	0	PCR	<i>TCF4</i>	↓
miR-140-3p	China (38)	NC, CC	44	PCR	<i>RRM2</i>	↓
miR-505-5p	China (39)	NC, CC	60	PCR	<i>CDK5</i>	↓
miR-877	China (40)	NC, CC	57	PCR	<i>MACC1</i>	↓
miR-636	China (41)	NC, CC	40	PCR	<i>BCL2, CDK6</i>	↓
miR-144-3p	China (42)	NC, CC	23	PCR	<i>MAPK6</i>	↓
miR-139-5p	China (43)	NC, CC	40	PCR	<i>TCF4</i>	↓
miR-126	China (44)	NC, CC	30	PCR	<i>ZEB1</i>	↓
miR-138	China (45)	Cell lines	0	PCR	<i>H2AX</i>	↓
miR-526b	China (46)	NC, SCC, AC	85	PCR	<i>PBX3</i>	↓
miR-432	China (47)	NC, CC	47	PCR	<i>FN1</i>	↓
miR-543	China (48)	NC, SCC, AC	69	PCR	<i>TRPM7</i>	↓
miR-503	China (49)	NC, CC	52	PCR	<i>AKT2</i>	↓
miR-93-5p	China (50)	NC, CIN, CC	328	PCR	<i>BTG3</i>	↑
miR-150-5p	China (51)	Cell lines	0	PCR	<i>SRCIN1</i>	↑

NC, normal cervix; CIN, cervical intraepithelial neoplasia; SCC, squamous carcinoma of cervix; CC, cervical cancer; AC, adenocarcinoma; PCR, polymerase chain reaction; FISH, fluorescence *in situ* hybridization. ↑, indicates that the molecule is upregulated in cervical diseases. ↓, indicates that the molecule is downregulated in cervical diseases.

to be more capable of predicting LSIL progression compared with other HR-HPVs; however, there was no association identified between p16/Ki-67 staining and prognosis (58). Therefore, further research is required for p16 and Ki67. In addition, other previous p16-related research has been conducted (59-61).

Cytokeratin 7 (CK7) is a squamocolumnar junction-related immunomarker. Paquette *et al* (62) identified that CK7-positive LSILs progressed with more ease to HSILs compared with negative CK7 LSILs (32.0 vs. 11.1%; $P=0.05$). Mills *et al* (63) proved that high levels of CK7 staining were associated with the progression of CIN1 to CIN2 (OR=2.8; $P=0.021$) and to CIN3 (OR=5.7; $P=0.018$). Cao *et al* (64) also reported the role of CK7 in CIN.

Wu *et al* (65) determined that the expression levels of cancerous inhibitor of PP2A (CIP2A) increased alongside the development of cervical lesions. CIP2A could bind to the onco-gene H-Ras and activate the MEK/ERK signaling pathway, which subsequently promoted epithelial-mesenchymal transition (EMT) in cervical cancer progression. Human discs large tumor suppressor (DLG1) is a component of the Scribble polarity complex; through a 2-year follow-up study, Cavatorta *et al* (66) identified that the cases progressing from LSILs to HSILs had diffuse DLG1 expression, and that LSILs with a DLG1 staining pattern similar to normal tissue were more likely to regress. Myosin IB (MYO1B) is a member of class I myosin, which was discovered to participate in the cell migration of zebrafish embryonic cells. In addition, MYO1B expression levels were upregulated in squamous cervical cancer and cervical cancer cell lines, where it served a role in cancer cell proliferation, migration and invasion (67).

A previous study investigating the expression levels of multiple proteins in exfoliated cervical cells indicated that the expression levels of Sialyl-Lewis A in cervical cancer were significantly downregulated compared with normal and CIN lesions ($P<0.01$). In addition, the expression of HPV L1 and p53 in cervical cancer were increased compared with normal and CIN lesions ($P<0.05$) (68). Compared with normal cervical tissue, the expression of T lymphoma invasion and metastasis 1 (Tiam1) was significantly increased in CIN and cervical cancer ($P<0.05$ and $P<0.01$, respectively), and the upregulated expression levels of Tiam1 were discovered to be associated with a poor prognosis in patients with cervical cancer. In addition, Tiam1 promoted the proliferation and migration of cancer cells by activating EMT (69). Mizushima *et al* discovered that following the development of normal cervical tissue to CIN, as the severity of the lesions increased, the expression of atypical protein kinase C λ/ι (aPKC λ/ι) also increased. In fact, aPKC λ/ι overexpression and nuclear localization were identified as independent factors for CIN1 progression, with hazard ratios of 4.26 ($P=0.007$) and 3.59 ($P=0.019$), respectively (70). Hester *et al* also discovered that prostaglandin E2-receptor 3 (EP3) expression was decreased with increasing grades of cervical lesions (from normal to CIN1-3; $P<0.05$). Notably, the proportion of EP3-positive cells in progressed CIN2 was decreased compared with in regressed CIN2 ($P=0.04$) (71).

In recent years, numerous studies have been conducted to determine the relationship between protein levels in the blood and cervical diseases. Sawada *et al* (72) found that patients with cervical cancer with high levels of vascular endothelial

Table V. Protein alterations in cervical disease.

Name	Population (Refs.)	Sample	Cases	Methods	Alteration
Potential diagnosis markers					
p16	Italy (59)	CIN1/3	66	IHC	↑
p16	China (61)	NC, CIN, SCC	254	WB	↑
CK7	USA (62)	NC, CIN, SCC	326	IHC	↑
CK7					↓
HPV L1	China (64)	LSIL, HSIL	100	IHC	↑
CIP2A	China (65)	NC, CIN, SCC	105	PCR, IHC, WB	↑
MYO1B	China (67)	NC, CIN, SCC	335	IHC	↓
SLeA	Korea (68)	NC, CIN, CC	146	ELISA, WB, IP	↑
HPV L1					↑
p53					↑
Tiam1	China (69)	NC, CIN, CC	298	IHC	↑
aPKC λ/ι	Japan (70)	NC, CIN	205	IHC	↑
EP3	Germany (71)	NC, CIN	124	IHC	↓
RAP1	Brazil (74)	NC, CIN	183	IHC	↑
COX-2					↑
EGFR	Brazil (75)	NC, CIN, SCC	412	IHC	↑
NCL	China (76)	NC, CIN, SCC	175	IHC	↑
HBXIP	China (77)	NC, CIN, SCC	243	IHC	↑
ERK1/2	China (78)	NC, CIN, SCC	176	PCR, IHC	↑
A3G	Japan (79)	NC, CIN, SCC	34	PCR, IHC	↑
HPV 16					↑
hnRNP K	China (80)	NC, CIN	204	FH, WB	↑
MFN2	Korea (81)	NC, CIN, SCC	191	IHC	↑
ADAR1	China (82)	NC, CIN, SCC	303	IHC	↑
Geminin	China (83)	NC, CIN	95	IHC	↑
SIRT1	USA (84)	CIN, SCC	101	IHC	↑
Gankyrin	China (85)	NC, CIN, SCC	76	IHC	↑
Potential prognosis markers for CIN1					
HPV16/18	Spain (58)	LSIL	200	IHC	-
HPV L1	Italy (59)	CIN1/3	66	IHC	↓
p16					
HPV L1	Japan (60)	CIN	199	PCR, IHC	-
CK7	USA (62)	NC, CIN, SCC	326	IHC	↑
CK7	USA (63)	CIN1	517	IHC	-
CK7					↑
HPV L1	China (64)	LSIL, HSIL	100	IHC	↓
DLG1	Argentina (66)	LSIL	30	IHC	-
aPKC λ/ι	Japan (70)	NC, CIN	205	IHC	↑
RAP1	Brazil (74)	NC, CIN	183	IHC	↑
Potential prognosis markers for CIN2					
p16/Ki-67	Japan (55)	CIN2	122	IHC	-
p16	Spain (56)	HSIL/CIN2	96	IHC	-
p16					
HPV L1	Japan (60)	CIN	199	PCR, IHC	-
EP3	Germany (71)	NC, CIN	124	IHC	↓

NC, normal cervix; CIN, cervical intraepithelial neoplasia; SCC, squamous carcinoma of cervix; CC, cervical cancer; LSIL, low grade squamous intraepithelial lesion; HSIL, high grade squamous intraepithelial lesion; IHC, immunohistochemistry; PCR, polymerase chain reaction; WB, western blot analysis; ELISA, enzyme linked immunosorbent assay; IP, immunoprecipitation; FH, flow-through hybridization; ↑, indicates that the molecule is upregulated in cervical diseases; ↓, indicates that the molecule is downregulated in cervical diseases; -, indicates that the amount of molecules was not compared in different tissues.

growth factor A (VEGF-A) and vascular endothelial growth factor receptor 2 (VEGFR-2) in the serum had a poor prognosis. Maestri *et al* (73) discovered that the serum levels of MBL-associated serine proteases (MASP)-2, MASP-1 and MAP-19 in patients with cervical cancer were significantly upregulated compared with in CIN and normal tissues ($P<0.0001$, $P=0.012$, $P=0.025$, respectively). These findings indicated that detecting the levels of specific proteins in the blood may help diagnose and predict the prognosis of cervical diseases.

Other proteins discovered to be involved in cervical cancer and precursors include RAS proximate 1 (RAP1) (74), cyclooxygenase-2 (Cox-2), epidermal growth factor receptor (EGFR), human epidermal growth factor receptor-2 (ERBB-2) (75), nucleolin (NCL) (76), hepatitis B virus X-interacting protein (HBXIP) (77), extracellular signal-regulated kinases 1/2 (ERK1/2) (78), APOBEC3G (79), heterogeneous nuclear ribonucleoproteins K (hnRNP K) (80), mitofusin-2 (MFN2) (81), RNA-dependent adenosine deaminase (ADAR1) (82), geminin (83), sirtuin 1 (SIRT1) (84) and gankyrin (85), among others. The details of these molecules are listed in Table V.

8. Conclusion

A significant amount of research has accumulated regarding the possible development of biomarkers for the early diagnosis of cervical lesions and the risk assessment of precursors. The development of cervical cancer is a multifactorial process; the transition from normal cervix tissue to precursors/cervical cancer is associated with chromosomal alterations, DNA polymorphisms, the DNA methylation status, histone modifications, and alterations to miRNA and protein expression levels. The majority of the experimental studies are conducted using cervical tissues and cells, while a small number of specimens are studied in the blood of patients. Since liquid biopsies represent a detection method with demonstrated diagnostic and monitoring value for cancer, which exert little harm to the body due to the non-invasive nature, they warrant further research in the future. Although there has been significant progress in the field of cervical cancer research, the identification of important molecules that could help predict the progression and prognosis of cervical cancer are still required. However, future studies require more samples and improved experimental designs.

Acknowledgements

Not applicable.

Funding

The present study was funded by teacher grants from Central South University (Changsha, Hunan, China).

Availability of data and materials

Not applicable.

Author's contributions

JS and YX wrote the manuscript and constructed the tables. LS and QH designed and revised the manuscript. JS, YX, LS and

QH were responsible for the submission of the manuscript and the final approval of the version to be published. All authors were involved in the literature search and review.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 68: 394-424, 2018.
2. Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ and He J: Cancer statistics in China, 2015. *CA Cancer J Clin* 66: 115-132, 2016.
3. Schiffman M, Wentzensen N, Wacholder S, Kinney W, Gage JC and Castle PE: Human papillomavirus testing in the prevention of cervical cancer. *J Natl Cancer Inst* 103: 368-383, 2011.
4. zur Hausen H: Papillomaviruses and cancer: From basic studies to clinical application. *Nat Rev Cancer* 2: 342-350, 2002.
5. Lu Z and Chen J: Introduction of WHO classification of tumours of female reproductive organs, fourth edition. *Zhonghua Bing Li Xue Za Zhi* 43: 649-650, 2014 (In Chinese).
6. Dalla Palma P, Giorgi Rossi P, Collina G, Buccoliero AM, Ghiringhello B, Gilioli E, Onnis GL, Aldovini D, Galanti G, Casadei G, *et al*: The reproducibility of CIN diagnoses among different pathologists: Data from histology reviews from a multicenter randomized study. *Am J Clin Pathol* 132: 125-132, 2009.
7. Zhang SK, Kang LN, Chang JJ, Zhao FH, Hu SY, Chen W, Shi JF, Zhang X, Pan QJ, Li SM and Qiao YL: The natural history of cervical cancer in Chinese women: Results from an 11-year follow-up study in China using a multistate model. *Cancer Epidemiol Biomarkers Prev* 23: 1298-1305, 2014.
8. Tainio K, Athanasiou A, Tikkinen KAO, Aaltonen R, Cardenas J, Hernandez, Glazer-Livson S, Jakobsson M, Joronen K, Kiviharju M, *et al*: Clinical course of untreated cervical intraepithelial neoplasia grade 2 under active surveillance: Systematic review and meta-analysis. *BMJ* 360: k499, 2018.
9. Policht FA, Song M, Sitailo S, O'Hare A, Ashfaq R, Muller CY, Morrison LE, King W and Sokolova IA: Analysis of genetic copy number changes in cervical disease progression. *BMC Cancer* 10: 432, 2010.
10. Rodolakis A, Biliatis I, Symiakaki H, Kershner E, Kilpatrick MW, Haidopoulos D, Thomakos N and Antsaklis A: Role of chromosome 3q26 gain in predicting progression of cervical dysplasia. *Int J Gynecol Cancer* 22: 742-747, 2012.
11. Koenen MM, Ovestad IT, Janssen EAM, Ummelen M, Kruitwagen RFP, Hopman AH and Kruse AJ: Gain of chromosomal region 3q26 as a prognostic biomarker for high-grade cervical intraepithelial neoplasia: Literature overview and pilot study. *Pathol Oncol Res* 25: 549-557, 2019.
12. Kudela E, Visnovsky J, Balharek T, Farkasova A, Zubor P, Plank L and Danko J: Different amplification patterns of 3q26 and 5p15 regions in cervical intraepithelial neoplasia and cervical cancer. *Ann Diagn Pathol* 35: 16-20, 2018.
13. Cezar-Dos-Santos F, Ferreira RS, Okuyama NCM, Trugilo KP, Sena MM, Pereira ER, Pereira APL, Watanabe MAE and de Oliveira KB: FOXP3 immunoregulatory gene variants are independent predictors of human papillomavirus infection and cervical cancer precursor lesions. *J Cancer Res Clin Oncol* 145: 2013-2025, 2019.
14. Isakova J, Vinnikov D, Bukuev N, Talaibekova E and Aldasheva N: TP53 Codon 72 polymorphism and human papilloma virus-associated cervical cancer in Kyrgyz women. *Asian Pac J Cancer Prev* 20: 1057-1062, 2019.

15. Chen B, Jiao Y, Yaolong F, Li T, Liu Y, Wang M, Xiuli G and Feng X: The POLR2E rs3787016 polymorphism is strongly associated with the risk of female breast and cervical cancer. *Pathol Res Pract* 215: 1061-1065, 2019.
16. Sui S, Chen H, Han L, Wang L, Niyazi M and Zhu K: Correlation of APOBEC3G polymorphism with human papillomavirus (HPV) persistent infection and progression of cervical lesions. *Med Sci Monit* 25: 6990-6997, 2019.
17. Wang L, Zhao W, Hong J, Niu F, Li J, Zhang S and Jin T: Association between IL1B gene and cervical cancer susceptibility in Chinese Uygur population: A case-control study. *Mol Genet Genomic Med* 7: e779, 2019.
18. Yang S, Zhao J and Li L: NAD(P)H: Quinone oxidoreductase 1 gene rs1800566 polymorphism increases the risk of cervical cancer in a Chinese Han sample: A STROBE-complaint case-control study. *Medicine (Baltimore)* 99: e19941, 2020.
19. Hu S, Pu D, Xia X, Guo B and Zhang C: CTLA-4 rs5742909 polymorphism and cervical cancer risk: A meta-analysis. *Medicine (Baltimore)* 99: e19433, 2020.
20. Ye F, Wang H, Liu J, Cheng Q, Chen X and Chen H: Genetic variants of the dUTPase-encoding gene DUT increase HR-HPV infection rate and cervical squamous cell carcinoma risk. *Sci Rep* 9: 513, 2019.
21. Sakane J, Taniyama K, Miyamoto K, Saito A, Kuraoka K, Nishimura T, Sentani K, Oue N and Yasui W: Aberrant DNA methylation of DLX4 and SIM1 is a predictive marker for disease progression of uterine cervical low-grade squamous intraepithelial lesion. *Diagn Cytopathol* 43: 462-470, 2015.
22. Luan T, Hua Q, Liu X, Xu P, Gu Y, Qian H, Yan L, Xu X, Geng R, Zeng X and Li P: PAX1 methylation as a potential biomarker to predict the progression of cervical intraepithelial neoplasia: A Meta-analysis of related studies. *Int J Gynecol Cancer* 27: 1480-1488, 2017.
23. Zhang J, Yao T, Lin Z and Gao Y: Aberrant Methylation of MEG3 functions as a potential plasma-based biomarker for cervical cancer. *Sci Rep* 7: 6271, 2017.
24. Rogeri CD, Silveira HCS, Causin RL, Villa LL, Stein MD, de Carvalho AC, Arantes LM, Scapulatempo-Neto C, Possati-Resende JC, Antoniazzi M, *et al*: Methylation of the hsa-miR-124, SOX1, TERT, and LMX1A genes as biomarkers for precursor lesions in cervical cancer. *Gynecol Oncol* 150: 545-551, 2018.
25. Verlaet W, Snijders PJ, Novianti PW, Wilting SM, De Strooper LM, Trooskens G, Vandersmissen J, Van Criekinge W, Wisman GB, Meijer CJ, *et al*: Genome-wide DNA methylation profiling reveals methylation markers associated with 3q gain for detection of cervical precancer and cancer. *Clin Cancer Res* 23: 3813-3822, 2017.
26. De Strooper LM, Berkhof J, Steenbergen RD, Lissenberg-Witte BI, Snijders PJ, Meijer CJ and Heideman DA: Cervical cancer risk in HPV-positive women after a negative FAM19A4/mir124-2 methylation test: A post hoc analysis in the POBASCAM trial with 14 year follow-up. *Int J Cancer* 143: 1541-1548, 2018.
27. Beyer S, Zhu J, Mayr D, Kuhn C, Schulze S, Hofmann S, Dannecker C, Jeschke U and Kost BP: Histone H3 Acetyl K9 and Histone H3 Tri Methyl K4 as prognostic markers for patients with cervical cancer. *Int J Mol Sci* 18: 477, 2017.
28. Zhang L, Tian S, Pei M, Zhao M, Wang L, Jiang Y, Yang T, Zhao J, Song L and Yang X: Crosstalk between histone modification and DNA methylation orchestrates the epigenetic regulation of the costimulatory factors, Tim3 and galectin9, in cervical cancer. *Oncol Rep* 42: 2655-2669, 2019.
29. Shi Y, Ma HL, Zhuang YW, Wang XX, Jiang Y and Xu HE: C10ORF12 modulates PRC2 histone methyltransferase activity and H3K27me3 levels. *Acta Pharmacol Sin* 40: 1457-1465, 2019.
30. Egger G, Liang G, Aparicio A and Jones PA: Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 429: 457-463, 2004.
31. Zeng K, Zheng W, Mo X, Liu F, Li M, Liu Z, Zhang W and Hu X: Dysregulated microRNAs involved in the progression of cervical neoplasm. *Arch Gynecol Obstet* 292: 905-913, 2015.
32. Zhu Y, Han Y, Tian T, Su P, Jin G, Chen J and Cao Y: MiR-21-5p, miR-34a, and human telomerase RNA component as surrogate markers for cervical cancer progression. *Pathol Res Pract* 214: 374-379, 2018.
33. Sommerova L, Anton M, Bouchalova P, Jasickova H, Rak V, Jandakova E, Selingerova I, Bartosik M, Vojtesek B and Hrstka R: The role of miR-409-3p in regulation of HPV16/18-E6 mRNA in human cervical high-grade squamous intraepithelial lesions. *Antiviral Res* 163: 185-192, 2019.
34. Jin Y, Zhou X, Yao X, Zhang Z, Cui M and Lin Y: MicroRNA-612 inhibits cervical cancer progression by targeting NOB1. *J Cell Mol Med* 24: 3149-3156, 2020.
35. Zhao XQ, Tang H, Yang J, Gu XY, Wang SM and Ding Y: MicroRNA-15a-5p down-regulation inhibits cervical cancer by targeting TP53INP1 in vitro. *Eur Rev Med Pharmacol Sci* 23: 8219-8229, 2019.
36. Sun Y, Cheng Y, Zhang Y and Han K: MicroRNA-889-3p targets FGFR2 to inhibit cervical cancer cell viability and invasion. *Exp Ther Med* 18: 1440-1448, 2019.
37. Yu Y, Zhao JD and Yang H: MiR-299-3p inhibits proliferation and invasion of cervical cancer cell via targeting TCF4. *Eur Rev Med Pharmacol Sci* 23: 5621-5627, 2019.
38. Ma J, Zhang F and Sun P: miR-140-3p impedes the proliferation of human cervical cancer cells by targeting RRM2 to induce cell-cycle arrest and early apoptosis. *Bioorg Med Chem* 28: 115283, 2020.
39. Kapora E, Feng S, Liu W, Sakhautdinova I, Gao B and Tan W: MicroRNA-505-5p functions as a tumor suppressor by targeting cyclin-dependent kinase 5 in cervical cancer. *Biosci Rep* 39: BSR20191221, 2019.
40. Meng F, Ou J, Liu J, Li X, Meng Y, Yan L, Deng P and Sun B: MicroRNA-877 is downregulated in cervical cancer and directly targets MACC1 to inhibit cell proliferation and invasion. *Exp Ther Med* 18: 3650-3658, 2019.
41. Hu QL, Xu ZP, Lan YF and Li B: miR-636 represses cell survival by targeting CDK6/Bcl-2 in cervical cancer. *Kaohsiung J Med Sci* 36: 328-335, 2020.
42. Wu J, Zhao Y, Li F and Qiao B: MiR-144-3p: A novel tumor suppressor targeting MAPK6 in cervical cancer. *J Physiol Biochem* 75: 143-152, 2019.
43. Ji X, Guo H, Yin S and Du H: miR-139-5p functions as a tumor suppressor in cervical cancer by targeting TCF4 and inhibiting Wnt/ β -catenin signaling. *Oncotargets Ther* 12: 7739-7748, 2019.
44. Xu J, Wang H, Wang H, Chen Q, Zhang L, Song C, Zhou Q and Hong Y: The inhibition of miR-126 in cell migration and invasion of cervical cancer through regulating ZEB1. *Hereditas* 156: 11, 2019.
45. Yuan M, Zhao S, Chen R, Wang G, Bie Y, Wu Q and Cheng J: MicroRNA-138 inhibits tumor growth and enhances chemosensitivity in human cervical cancer by targeting H2AX. *Exp Ther Med* 19: 630-638, 2020.
46. Li H, Wang J, Xu F, Wang L, Sun G, Wang J and Yang Y: By downregulating PBX3, miR-526b suppresses the epithelial-mesenchymal transition process in cervical cancer cells. *Future Oncol* 15: 1577-1591, 2019.
47. Wang S, Gao B, Yang H, Liu X, Wu X and Wang W: MicroRNA-432 is downregulated in cervical cancer and directly targets FN1 to inhibit cell proliferation and invasion. *Oncol Lett* 18: 1475-1482, 2019.
48. Liu X, Gan L and Zhang J: miR-543 inhibits cervical cancer growth and metastasis by targeting TRPM7. *Chem Biol Interact* 302: 83-92, 2019.
49. Fu Y, Meng Y, Gu X, Tian S, Hou X and Ji M: miR-503 expression is downregulated in cervical cancer and suppresses tumor growth by targeting AKT2. *J Cell Biochem*: Jan 29, 2019 (Epub ahead of print). doi: 10.1002/jcb.28099.
50. Li J, Chu ZP, Han H, Zhang Y, Tian F, Zhang JQ and Huang XH: Suppression of miR-93-5p inhibits high-risk HPV-positive cervical cancer progression via targeting of BTG3. *Hum Cell* 32: 160-171, 2019.
51. Zhu J and Han S: miR-150-5p promotes the proliferation and epithelial-mesenchymal transition of cervical carcinoma cells via targeting SRCIN1. *Pathol Res Pract* 215: 738-747, 2019.
52. Luo Q, Wang H and Li J: Serum miR-3142 could be used as a potential biomarker to screen cervical cancer patients from healthy controls. *Clin Lab* 65, 2019.
53. Zheng M, Hou L, Ma Y, Zhou L, Wang F, Cheng B, Wang W, Lu B, Liu P, Lu W and Lu Y: Exosomal let-7d-3p and miR-30d-5p as diagnostic biomarkers for non-invasive screening of cervical cancer and its precursors. *Mol Cancer* 18: 76, 2019.
54. Hayes J, Peruzzi PP and Lawler S: MicroRNAs in cancer: Biomarkers, functions and therapy. *Trends Mol Med* 20: 460-469, 2014.
55. Miyamoto S, Hasegawa J, Morioka M, Hirota Y, Kushima M and Sekizawa A: The association between p16 and Ki-67 immunohistostaining and the progression of cervical intraepithelial neoplasia grade 2. *Int J Gynaecol Obstet* 134: 45-48, 2016.
56. Miralpeix E, Genoves J, Maria Sole-Sedeno J, Mancebo G, Lloveras B, Bellosillo B, Alameda F and Carreras R: Usefulness of p16(INK4a) staining for managing histological high-grade squamous intraepithelial cervical lesions. *Mod Pathol* 30: 304-310, 2017.

57. Sagasta A, Castillo P, Saco A, Torne A, Esteve R, Marimon L, Ordi J and Del Pino M: p16 staining has limited value in predicting the outcome of histological low-grade squamous intraepithelial lesions of the cervix. *Mod Pathol* 29: 51-59, 2016.
58. Rodriguez-Trujillo A, Marti C, Angeles MA, Sierra A, Esteve R, Saco A, Barnadas E, Marimon L, Nicolas I, Torne A, *et al*: Value of HPV 16/18 genotyping and p16/Ki-67 dual staining to predict progression to HSIL/CIN2⁺ in negative cytologies from a colposcopy referral population. *Am J Clin Pathol* 150: 432-440, 2018.
59. Negri G, Bellisano G, Zannoni GF, Rivasi F, Kasal A, Vittadello F, Antoniazzi S, Faa G, Ambu R and Egarter-Vigl E: p16 ink4a and HPV L1 immunohistochemistry is helpful for estimating the behavior of low-grade dysplastic lesions of the cervix uteri. *Am J Surg Pathol* 32: 1715-1720, 2008.
60. Hoshikawa S, Sano T, Yoshida T, Ito H, Oyama T and Fukuda T: Immunohistological analysis of HPV L1 capsid protein and p16 protein in low-grade dysplastic lesions of the uterine cervix. *Pathol Res Pract* 206: 816-820, 2010.
61. Jia WL, Ding L, Ren ZY, Wu TT, Zhao WM, Fan SL and Wang JT: Effects of both folic acid, p16 protein expression and their interaction on progression of cervical cancerization. *Zhonghua Liu Xing Bing Xue Za Zhi* 37: 1647-1652, 2016 (In Chinese).
62. Paquette C, Mills AM and Stoler MH: Predictive value of cytokeratin 7 immunohistochemistry in cervical low-grade squamous intraepithelial lesion as a marker for risk of progression to a high-grade lesion. *Am J Surg Pathol* 40: 236-243, 2016.
63. Mills AM, Paquette C, Terzic T, Castle PE and Stoler MH: CK7 immunohistochemistry as a predictor of CIN1 progression: A retrospective study of patients from the quadrivalent HPV vaccine trials. *Am J Surg Pathol* 41: 143-152, 2017.
64. Cao L, Sun PL, Yao M, Chen S and Gao H: Clinical significance of CK7, HPV-L1, and koilocytosis for patients with cervical low-grade squamous intraepithelial lesions: A retrospective analysis. *Hum Pathol* 65: 194-200, 2017.
65. Wu Y, Gu TT and Zheng PS: CIP2A cooperates with H-Ras to promote epithelial-mesenchymal transition in cervical-cancer progression. *Cancer Lett* 356: 646-655, 2015.
66. Cavatorta AL, Di Gregorio A, Bugnon Valdano M, Marziali F, Cabral M, Bottai H, Cittadini J, Nocito AL and Gardiol D: DLG1 polarity protein expression associates with the disease progress of low-grade cervical intraepithelial lesions. *Exp Mol Pathol* 102: 65-69, 2017.
67. Zhang HR, Lai SY, Huang LJ, Zhang ZF, Liu J, Zheng SR, Ding K, Bai X and Zhou JY: Myosin 1b promotes cell proliferation, migration, and invasion in cervical cancer. *Gynecol Oncol* 149: 188-197, 2018.
68. Jin Y, Kim SC, Kim HJ, Ju W, Kim YH and Kim HJ: Use of protein-based biomarkers of exfoliated cervical cells for primary screening of cervical cancer. *Arch Pharm Res* 41: 438-449, 2018.
69. Yang Y, Wu Q, Li N, Che S, Jin T, Nan Y, Lin Z and Chen L: Upregulation of Tiam1 contributes to cervical cancer disease progression and indicates poor survival outcome. *Hum Pathol* 75: 179-188, 2018.
70. Mizushima T, Asai-Sato M, Akimoto K, Nagashima Y, Taguri M, Sasaki K, Nakaya MA, Asano R, Tokinaga A, Kiyono T, *et al*: Aberrant expression of the cell polarity regulator aPKC ζ is associated with disease progression in cervical intraepithelial neoplasia (CIN): A possible marker for predicting CIN prognosis. *Int J Gynecol Pathol* 35: 106-117, 2016.
71. Hester A, Ritzer M, Kuhn C, Schmoeckel E, Mayr D, Kolben T, Dannecker C, Mahner S, Jeschke U and Kolben TM: The role of EP3-receptor expression in cervical dysplasia. *J Cancer Res Clin Oncol* 145: 313-319, 2019.
72. Sawada M, Oishi T, Komatsu H, Sato S, Chikumi J, Nonaka M, Kudoh A, Osaku D and Harada T: Serum vascular endothelial growth factor A and vascular endothelial growth factor receptor 2 as prognostic biomarkers for uterine cervical cancer. *Int J Clin Oncol* 24: 1612-1619, 2019.
73. Maestri CA, Nishihara R, Mendes HW, Jensenius J, Thiel S, Messias-Reason I and de Carvalho NS: MASP-1 and MASP-2 serum levels are associated with worse prognostic in cervical cancer progression. *Front Immunol* 9: 2742, 2018.
74. Pascoal-Xavier MA, Figueiredo AC, Gomes LI, Peruhype-Magalhaes V, Calzavara-Silva CE, Costa MA, Reis IA, Bonjardim CA, Kroon EG, Oliveira JG and Ferreira PC: RAP1 GTPase overexpression is associated with cervical intraepithelial neoplasia. *PLoS One* 10: e0123531, 2015.
75. Fukazawa EM, Baiocchi G, Soares FA, Kumagai LY, Faloppa CC, Badiglian-Filho L, Coelho FR, Goncalves WJ, Costa RL and Goes JC: Cox-2, EGFR, and ERBB-2 expression in cervical intraepithelial neoplasia and cervical cancer using an automated imaging system. *Int J Gynecol Pathol* 33: 225-234, 2014.
76. Meng GZ, Zi Y, Li HQ, Huang M and Gao T: Nucleolin expression is correlated with carcinogenesis and progression of cervical squamous cell carcinoma. *Nan Fang Yi Ke Da Xue Xue Bao* 35: 1511-1514, 2015 (In Chinese).
77. Li N, Wang Y, Che S, Yang Y, Piao J, Liu S and Lin Z: HBXIP over expression as an independent biomarker for cervical cancer. *Exp Mol Pathol* 102: 133-137, 2017.
78. Fan SL, Ding L, Ren ZY, Chen X, Sun XS, Li CC, Liu CL, Jia WL, Li QL and Wang JT: Effect of extracellular signal-regulated kinases 1/2 expression and HPV16 infection and their interaction in progression of cervical cancerization. *Zhonghua Liu Xing Bing Xue Za Zhi* 38: 96-101, 2017 (In Chinese).
79. Iizuka T, Wakae K, Nakamura M, Kitamura K, Ono M, Fujiwara H and Muramatsu M: APOBEC3G is increasingly expressed on the human uterine cervical intraepithelial neoplasia along with disease progression. *Am J Reprod Immunol* 78, 2017.
80. Ding L, Feng MJ, Liu CL, Wang L, Song ZC, Yang Q, Li XX, Song L, Gao W and Wang JT: Effect of hnRNP K and its interaction with HPV16 on cervical intraepithelial neoplasia. *Zhonghua Liu Xing Bing Xue Za Zhi* 39: 1630-1635, 2018 (In Chinese).
81. Ahn SY, Li C, Zhang X and Hyun YM: Mitofusin-2 expression is implicated in cervical cancer pathogenesis. *Anticancer Res* 38: 3419-3426, 2018.
82. Chen Y, Wang H, Lin W and Shuai P: ADAR1 overexpression is associated with cervical cancer progression and angiogenesis. *Diagn Pathol* 12: 12, 2017.
83. Xing Y, Wang C and Wu J: Expression of geminin, p16, and Ki67 in cervical intraepithelial neoplasm and normal tissues. *Medicine (Baltimore)* 96: e7302, 2017.
84. Velez-Perez A, Wang XI, Li M and Zhang S: SIRT1 overexpression in cervical squamous intraepithelial lesions and invasive squamous cell carcinoma. *Hum Pathol* 59: 102-107, 2017.
85. Liu Y, Zhang J, Qian W, Dong Y, Yang Y, Liu Z, Feng Y, Ma D, Zhang Z and Wu S: Gankyrin is frequently overexpressed in cervical high grade disease and is associated with cervical carcinogenesis and metastasis. *PLoS One* 9: e95043, 2014.