

# Canonical and noncanonical Wnt pathway: A comparison among normal ovary, benign ovarian tumor and ovarian cancer

LEVON BADIGLIAN FILHO<sup>1</sup>, CELINA TIZUKO FUJIYAMA OSHIMA<sup>2</sup>, FLÁVIO DE OLIVEIRA LIMA<sup>2</sup>,  
HENRIQUE DE OLIVEIRA COSTA<sup>2</sup>, ROBÉRIO DE SOUSA DAMIÃO<sup>1</sup>,  
THIAGO SIMÃO GOMES<sup>2</sup> and WAGNER JOSÉ GONÇALVES<sup>1</sup>

<sup>1</sup>Discipline of Gynecologic Oncology, Department of Gynecology, and

<sup>2</sup>Department of Pathology, Federal University of Sao Paulo, Sao Paulo, Brazil

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**Abstract.** The Wnt family is involved in tumorigenesis of several tissues. In ovarian cancer, the role played by Wnts and its pathways is not clearly defined. In order to analyze the canonical and noncanonical Wnt pathway in normal ovary, benign ovarian tumor and ovarian cancer, we evaluated the immunohistochemical expression of Wnt1, Frizzled-1 (FZD1), Wnt5a, Frizzled-5 (FZD5) and  $\beta$ -catenin. Ovarian specimens were obtained from surgeries performed between 1993 and 2004. The patients were divided in three groups: group A, epithelial ovarian cancer (n=38); group B, benign epithelial neoplasia (n=28); and group C, normal ovaries (n=26). Immunoreactivity for Wnt1, FZD1, Wnt5a, FZD5 and  $\beta$ -catenin was scored for each group. The proportion of Wnt1 positive women in group A (29.4%) was significantly higher than in group B (4.3%) and C (9.1%) ( $p=0.020$ ). The proportion of FZD1 positive patients in group C (54.5%) was significantly lower than in group A (97.1%) and B (90.0%) ( $p<0.001$ ). The proportion of Wnt5a positive women was significantly higher for group A (80.0%) compared to group B (25.0%) and C (27.3%) ( $p<0.001$ ). The proportion of  $\beta$ -catenin positive patients in group C (95.8%) was significantly higher than group B (52.4%) ( $p=0.004$ ). Comparison of the survival curves in group A according to Wnt5a expression showed a significant difference between positive and negative patients, whereas the Wnt5a positive women showed worse results ( $p=0.050$ ). Our findings suggest that the pathways related to Wnt5a have an important role in ovarian malignant neoplasia. Furthermore, Wnt5a was found to be a predictor of poor prognosis for ovarian cancer.

## Introduction

In recent years, several studies have shown the importance of Wnt family in tumorigenesis (1-5). Wnts are an evolutionarily highly conserved family of genes/proteins that act through four signaling pathways (6).

The canonical pathway: In the absence of Wnt signaling, a multiprotein complex that includes adenomatous polyposis coli (APC), glycogen synthase kinase 3 (GSK3) and Axin ensure the degradation of  $\beta$ -catenin, thereby limiting the free intracytoplasmic pool of  $\beta$ -catenin. The presence of Wnt signal through the Frizzled (FZD) receptor and low density lipoprotein receptor-related Protein 5 and 6 (LRP5/6) receptor complex inactivates GSK3 and causes its dissociation from Axin preventing the phosphorylation of  $\beta$ -catenin. The intracytoplasmic pool of  $\beta$ -catenin thus increases and it translocates to the nucleus where it complexes with members of the LEF/TCF family of transcription factors to mediate transcriptional induction of target genes such as c-myc, cyclin D, VEGF and others.

The noncanonical pathway: In noncanonical or planar cell polarity (PCP) signaling, Wnt signaling is transduced through Frizzled independent of LRP5/6. This pathway mediates cytoskeletal changes through activation of the small GTPases Rho and Rac (7).

Certain Wnts can activate both the canonical and the noncanonical pathway, such as Wnt3a; others appears to be specific to the noncanonical pathway, such as Wnt5a (6,7).

The WntCa<sup>2+</sup> pathway: Wnt signaling via Frizzled mediates activation of heterotrimeric G-proteins, which engage Dsh, phospholipase C calcium-calmodulin kinase 2 (CamK2) and protein kinase C (PKC). This pathway modulates cell adhesion and motility.

The protein kinase A pathway: Chen *et al* demonstrated that adenylyl cyclase signaling via protein kinase A (PKA) and its target transcription factor cAMP responsive element-binding protein (CREB) are required for Wnt-directed myogenic gene expression. They have shown that Wnt proteins can also stimulate CREB-mediated transcription (6,8).

In 1990, six members of the Wnt family were identified, including Wnt5a. It encodes a protein involved in cell-cell signaling during embryonic development (9-13). In 1997,

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**Correspondence to:** Dr Levon Badiglian Filho, Discipline of Gynecologic Oncology, Department of Gynecology, Al Lorena, 131, cj 51, Sao Paulo - SP, 01424-000, Brazil  
E-mail: dr.levon@terra.com.br

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Olson *et al* suggested that Wnt5a inactivation would lead to tumorigenesis (14). Thereby, some authors affirm that the noncanonical pathway would assume an antagonistic effect on the canonical pathway through different mechanisms and the Wnt5a would be a tumor suppressor (15-18). It can antagonize the canonical pathway at TCF/LEF level, in the nucleus (17). Furthermore, Wnt5a was able to increase the expression of the APC and Siah2 proteins which would lead to  $\beta$ -catenin degradation independent of CAMK2 (18).

However, many authors found Wnt5a amplified in several malignancies such as stomach, esophagus, prostate, melanoma, colon/rectum and breast. Ricken *et al* considered the possibility that the Wnt5a could be involved in ovarian carcinogenesis (19).

### Patients and methods

Ovarian specimens were obtained from surgeries performed between 1993 and 2004 at Federal University of Sao Paulo for patients who underwent exploratory laparotomy for adnexal tumor or prophylactic oophorectomy during surgery for benign disease. None of the patients had received any preoperative therapy. Borderline tumors were excluded from this study.

The patients were divided in three groups: group A, malignant epithelial neoplasia (n=38); group B, benign epithelial neoplasia (n=28); and group C, normal ovaries (n=26). The study was approved by the Institutional Ethics Committee.

**Immunohistochemistry.** Sections were deparaffinized in three changes of xylene and rehydrated in a graded series of ethanol finishing in destilated water. For antigen retrieval slides were placed in 0.01 M citrate-buffer pH 6.0 and heated in a steamer for 30 min. Endogenous peroxidases were quenched by incubating in 3% H<sub>2</sub>O<sub>2</sub>, 20 min at room temperature. Sections were incubated overnight at 4°C with antibody against Wnt1 (1:100), R&D Systems, Minneapolis, MN, USA; FZD1 (1:100) R&D Systems; Wnt5a (1:100) R&D Systems; FZD5 (1:100) R&D Systems; and  $\beta$ -catenin (1:100) Santa Cruz Biotechnology, Santa Cruz, CA, USA. Subsequently, sections were incubated with biotinylated secondary antibody (LSAB, Dakocytomation) for 30 min, washed in PBS, and incubated with streptavidin-peroxidase conjugate (LSAB, Dakocytomation) for 30 min. Finally, the reaction was revealed using 3,3'-Diaminobenzidine tetrahydrochloride (Sigma) for 5 min. Slides were briefly counterstained in hematoxylin and dehydrated, and cover slips added. Negative and positive controls were made to run simultaneously. Positive controls were represented by mammary tissue. Negative controls were made by eliminating the primary antibody.

The presence of tumor tissue was confirmed previously in each core. Immunostaining was scored by two trained independent observers without prior knowledge of the clinicopathological parameters. Discordant cases were reviewed and agreed upon before data were statistically analyzed.

Immunoreactivity of Wnt1, FZD1, Wnt5a, FZD5 and  $\beta$ -catenin was assigned a score based on the proportion of positive tumor cells over total tumor cells (percent positivity) ranging from 0 to 100%. The percentage of positive tumor cells was graded as follows: 0, none; 1, 1-25%; 2, 26-50%;

Table I. Malignant group.

Variables	(n=38)
Cytoreduction	n (%)
Optimal	22 (57.9%)
Suboptimal	16 (42.1%)
Stage	
I	10 (26.3%)
II	2 (5.3%)
III	16 (42.1%)
IV	10 (26.3%)
Hystology	
Serous	22 (57.9%)
Mucinous	7 (18.4%)
Endometrioid	4 (10.5%)
Clear cell	1 (2.6%)
Undifferentiated	4 (10.5%)

3, 51-75%; and 4, 76-100%. Staining intensity was evaluated as 0, negative; 1, weak; 2, moderate; and 3, strong. The score was calculated multiplying the positivity percent rating by staining intensity (20). From this score, values ranging from 0 to 2 (none or weak reaction) were considered negative and values ranging from 3 to 9 (moderate or strong reaction) were considered positive.

Owing to too few cells in certain samples, 61 out of 460 (group A, 190; group B, 140; group C, 130; total, 460) samples were uninterpretable. For these specimens, a score of NA was given.

**Statistical analyses.** The software used was the Statistical Package for the Social Sciences (SPSS, Chicago, IL). The groups were compared to quantitative variables by ANOVA when normal distribution of variables or, otherwise, by Kruskal-Wallis test.

The associations between expression of biomarkers and clinicopathological parameters were tested with contingency tables and Pearson's Chi-square test. Survival of patients was estimated by Kaplan-Meier analysis and the covariates were analyzed by the log-rank test for univariate analysis.  $p < 0.05$  was considered statistically significant.

### Results

The groups were homogeneous in terms of distribution of the following variables: age ( $p=0.145$ ), body mass index ( $p=0.454$ ), number of pregnancies ( $p=0.061$ ), age at menarche ( $p=0.236$ ), menopausal status ( $p=0.070$ ), age at menopause ( $p=0.603$ ), and cigarette smoking ( $p=0.443$ ). Data regarding group A are shown in Table I.

For the expression of Wnt1, there was a significant association among the groups. The proportion of positive women in group A (29.4%) was significantly higher than group B (4.3%) and C (9.1%) ( $p=0.020$ ). There was a significant association among the groups for the FZD1 expression ( $p < 0.001$ ). The proportion of positive scored patients in



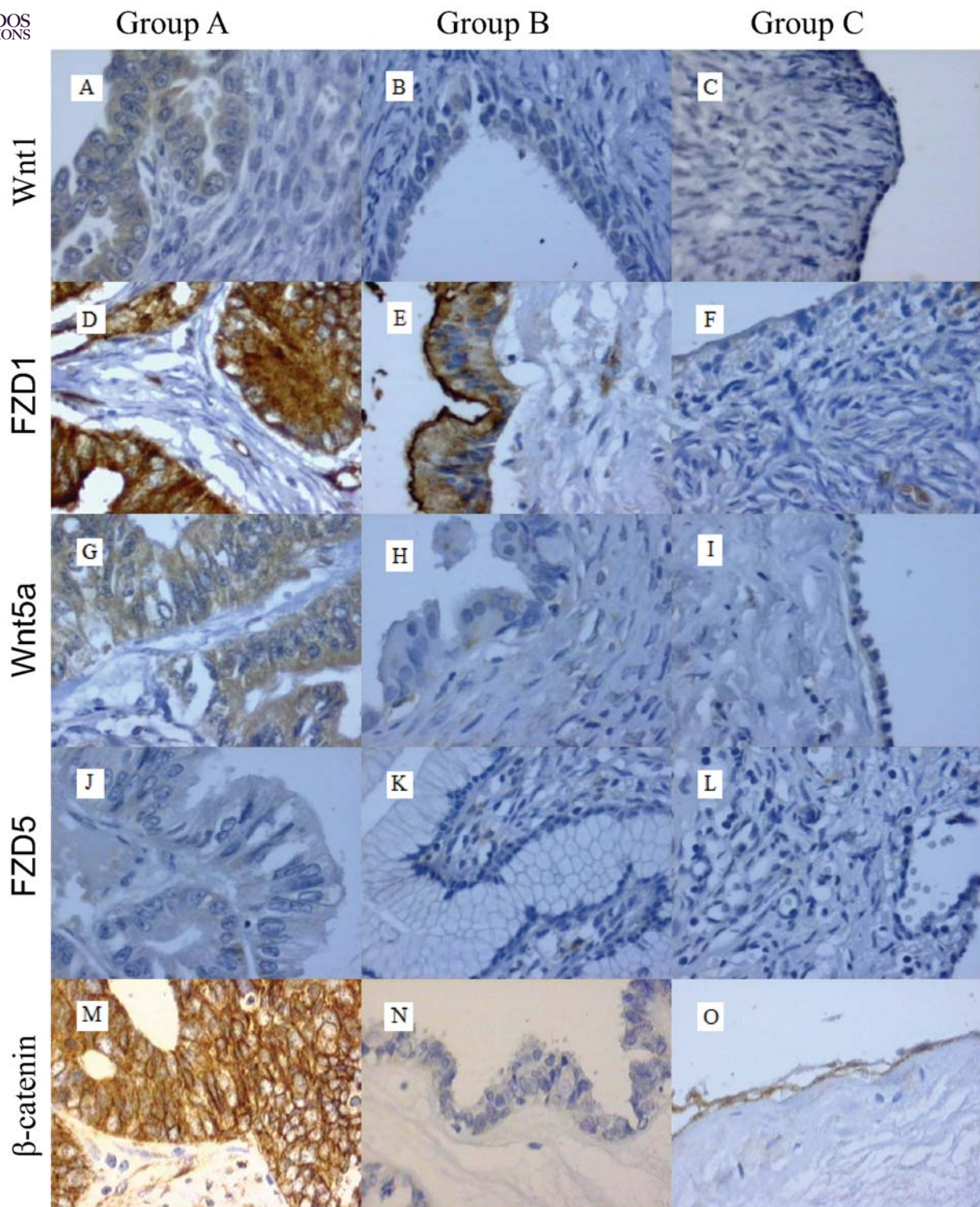


Figure 1. Immunostaining for Wnt1: strong staining for malignant tumor (A) and weak staining for benign tumor (B) and normal ovary (C). Immunostaining for FZD1: strong staining for malignant (D) and benign tumor (E) and weak staining for normal ovary (F). Immunostaining for Wnt5a: strong staining for malignant tumor (G) and weak staining for benign tumor (H) and normal ovary (I). Immunostaining for FZD5: weak staining for all groups (J-L). Immunostaining for β-catenin: strong staining for malignant tumor (M) and normal ovary (O), and weak staining for benign tumor (N) (x400).

group C (54.5%) was significantly lower than the other groups (97.1% for group A and 90.0% for group B).

A significant association was observed among the groups for the Wnt5a expression ( $p < 0.001$ ). The proportion of Wnt5a positive women was significantly higher for group A (80.0%) compared to group B (25.0%) and C (27.3%). No significant association was observed between FZD5 expression and the

patient group since the expression for groups A, B and C were 14.3, 4.0 and 8.7%, respectively ( $p = 0.380$ ).

The proportion of patients in group C with a positive score (95.8%) for β-catenin staining was significantly higher than group B (52.4%) ( $p = 0.004$ ). Group A (74.3% of positive cases) did not significantly differ from the other groups (Figs. 1 and 2).

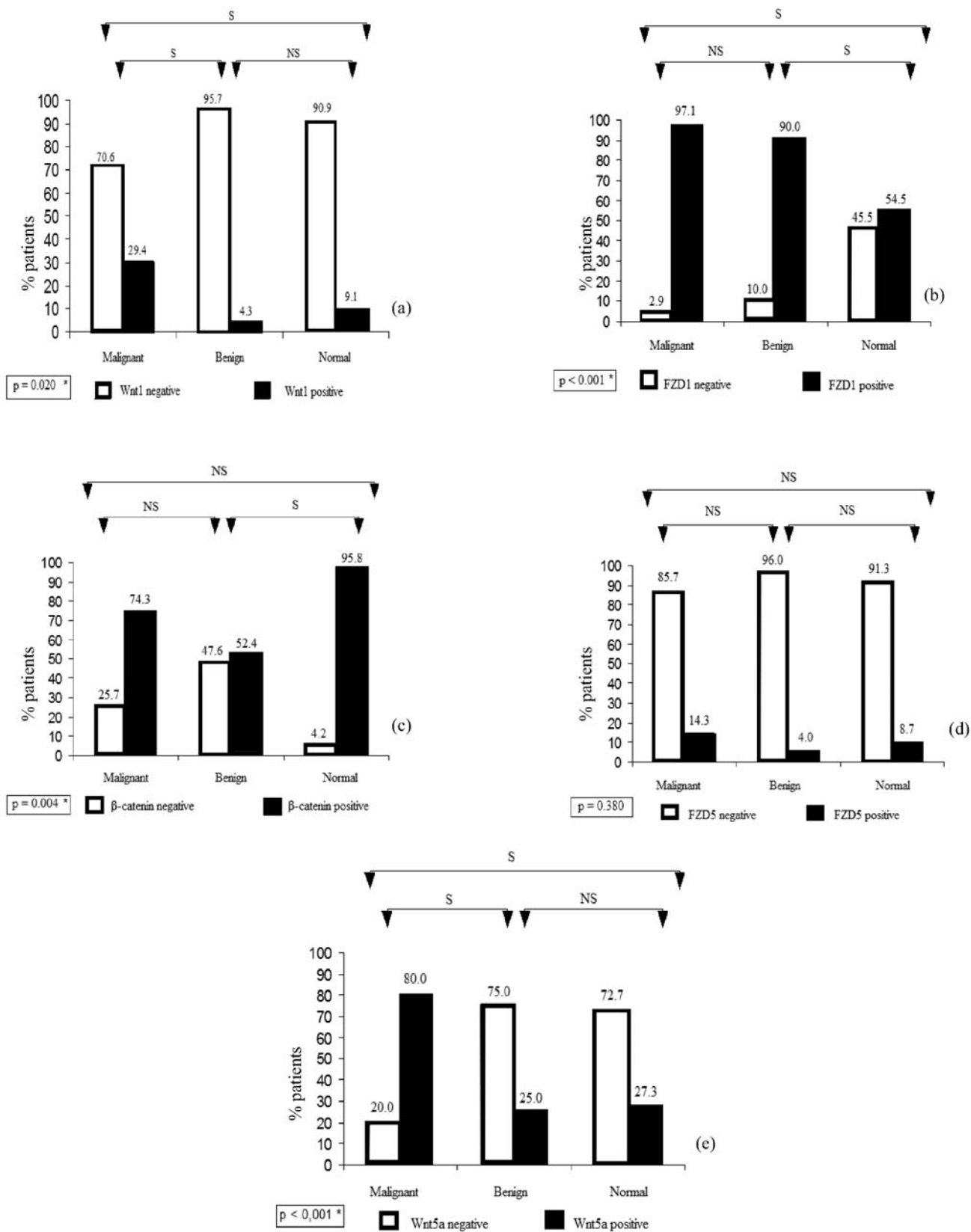


Figure 2. Expression of Wnt1 (a), FZD1 (b), β-catenin (c), FZD5 (d) and Wnt5a (e). A significant proportion of patients from malignant group stained for Wnt5a.

There was not significant association of the survival curves according to the β-catenin expression (p=0.062) in group A. Also in group A, no significant association of the survival curves was observed according to Wnt1 (p=0.497)

and FZD5 (p=0.550) expression. The survival curves according to the FZD1 expression could not be calculated because only one patient in group A was negative for this protein.

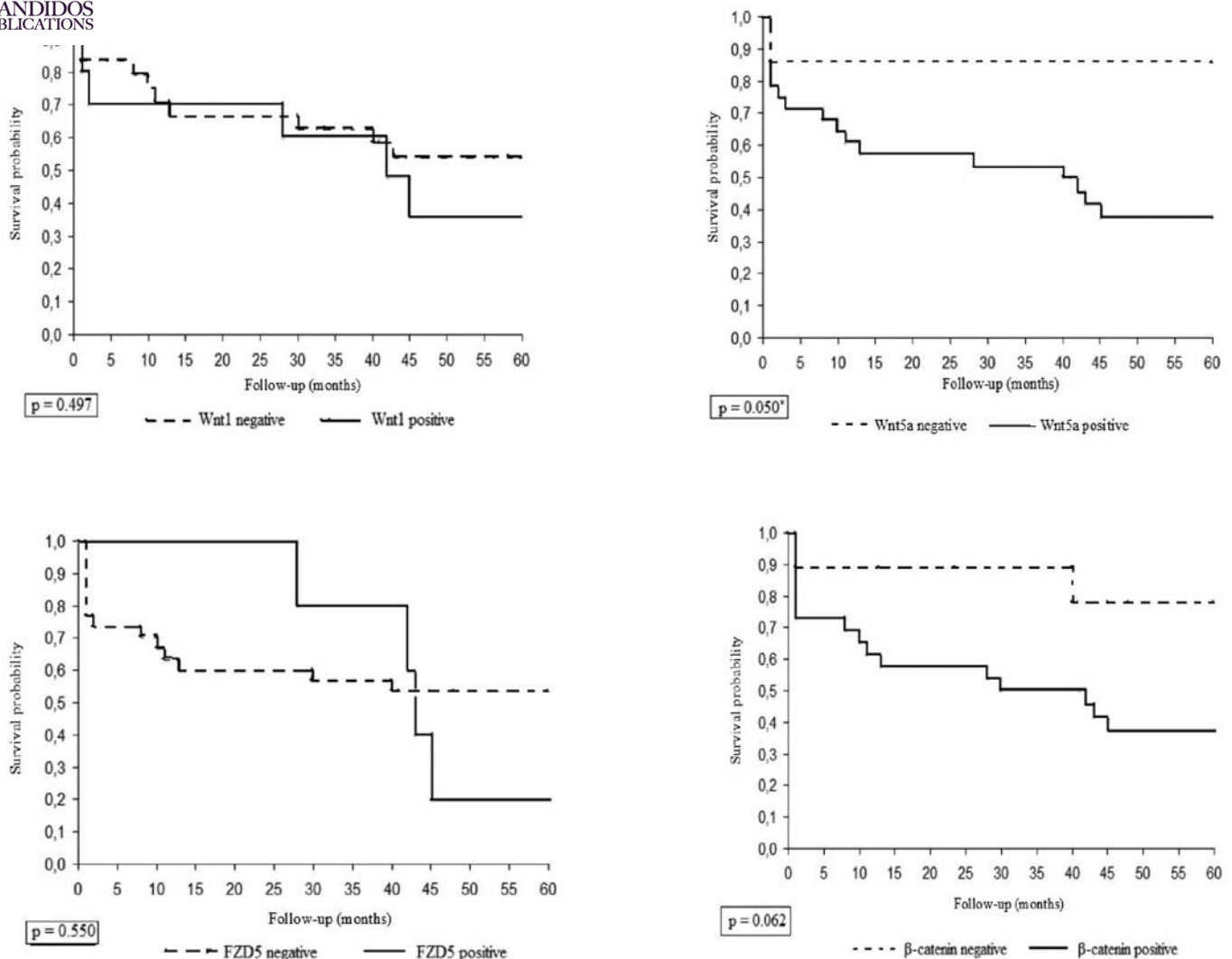


Figure 3. Kaplan-Meier overall survival curves for Wnt1, Wnt5a, FZD5 and  $\beta$ -catenin in group A. The curve for FZD1 could not be calculated because only one patient in group A was negative for this protein.

Comparison of the survival curves in group A according to Wnt5a expression showed a significant difference between Wnt5a positive and Wnt5a negative patients, whereas the Wnt5a positive women showed worse results ( $p=0.050$ ) (Fig. 3).

## Discussion

**Wnt1.** Wnt1 is overexpressed in several types of human cancer. In our series, Wnt1 was overexpressed in ovarian carcinoma, which would be expected to lead to overexpression of  $\beta$ -catenin, but it did not. It could be speculated that  $\beta$ -catenin was inhibited by noncanonical pathways through diverse mechanisms, such as activation of PKC and/or Siah2 (18). It is an intriguing issue to be solved, since the expression of FZD1 was higher in malignant group as well.

**Frizzled1.** Frizzled proteins are found exclusively at the plasma membrane. FZD1 was found to be up-regulated (21) in breast cancer as well as in poorly differentiated colon cancer (22). However, the complexity of Wnt pathway extends to its receptors. When the co-receptor LRP5/6 interaction to FZD1

is considered, the transmission of the canonical Wnt signaling occurs as expected; otherwise, considering the co-receptor LRP1 interaction to FZD1, canonical signaling (23) is repressed. An interaction between Wnt5a and FZD1 might exist, however, it is known that FZD1 is activated by Wnt3a, Wnt3, Wnt1 and to a lesser extent Wnt2, but not by Wnt4, Wnt5a, Wnt5b, Wnt6, Wnt7a or Wnt7b (24).

**Wnt5a.** We found a strong staining for group A compared to groups B and C and, furthermore, we demonstrated a prognostic role for Wnt5a. Staining for Wnt5a in malignant group correlated to worse prognosis. It is worth noting that only one Wnt5a negative patient died in group A. Several aspects could explain the positive Wnt5a expression and ovarian cancer.

Considering the three  $\beta$ -catenin independent pathways. PCP pathway: It was found that alendronate inhibits lysophosphatidic acid-induced migration of human ovarian cancer cells by attenuating the activation of Rho (binding domain of Rhoketin), we suppose that Wnt5a signaling activates Rho, leading to the progress of ovarian carcinoma. Moreover, the activation of Rac leads to activation of JNK;



phosphorylation of c-jun by JNK acts synergistically with TCF/LEF on the promoter of the canonical target gene c-jun. Alternatively, Wnt5a can bind to the receptor tyrosine kinase Ror2 which results in Rho GTPase-independent activation of JNK (25).

WntCa<sup>2+</sup>: Wnt5a signals through noncanonical pathways which involves stimulation of intracellular Ca<sup>2+</sup> release and activation of PKC and CAMK2 (18). PKC takes an important role in this pathway since several studies have demonstrated that it is overexpressed in ovarian carcinomas and it is related to worst prognosis (26,27). Furthermore, down-regulation of PKC enhances the sensitivity of human ovarian carcinoma to various types of platinum compounds and PKC inhibitors can decrease the invasiveness of ovarian cancer. D'Souza and colleagues showed that PKC phosphorylates claudin-4 (a transmembrane proteins essential to the formation and maintenance of tight junctions) which would lead to disruption of barrier function in ovarian cancer cells (28). In melanoma cells, there is a direct correlation with Wnt5a expression, PKC activation and increased melanoma cell invasion. Disruption of the Wnt5a/Frizzled-5 pathways results in an inhibition of PKC activation and reduced invasiveness of melanoma cells, apparently due to motility function related to PKC (29).

Interestingly, it have been shown that Wnt5a can activate PKC as well PKC can activate Wnt5a (30). Thus, independently of PKC, Wnt5a/Ca<sup>2+</sup> activates nuclear factor associated with T cells (NFAT), which is involved in tumorigenesis (25).

Protein kinase A pathway: Also D'Souza and colleagues suggested that PKA phosphorylates claudin3 leading to rupture of tight junctions and causing disruption of barrier function in ovarian cancer cells (31).

The overexpression of Wnt5a in some tumors is not a result of gene amplification or rearrangement, suggesting that the level of Wnt5a is being modulated by some further regulatory apparatus (29). Since Wnt5a was overexpressed in malignant neoplasms in our findings, it could be speculated that  $\beta$ -catenin was inhibited by Wnt5a (17,18). Mikels and Nusse demonstrated that Wnt5a activates or inhibits  $\beta$ -catenin pathway depending on the receptor context. They showed that Wnt5a can activate Wnt/ $\beta$ -catenin signaling in the presence of FZD4 and LRP5 and can inhibit it in the presence of the transmembrane receptor Ror2 (32-34). Also, Liu *et al* demonstrated that the mechanism of endogenous receptor functionally distinguishes prototype canonical and non-canonical Wnts, since they linked noncanonical Wnt5a with the C-terminal half of Dickkopf-2 (Dkk2C) creating a Wnt5a/Dkk2C chimera that was capable of activating canonical signaling (35). Moreover, the function of Wnt5a as either a suppressor or promoter of malignant progression is beyond intracellular signaling and seems to be modulated by intercellular interactions (36).

Contrary to us, Dejmek *et al* found that the expression of Wnt5a in primary Dukes' B colon cancer constitutes a good prognostic marker for a longer survival (37). On the other hand, Kurayoshi *et al* found that the expression of Wnt5a is correlated with aggressiveness of gastric cancer by stimulating cell migration and invasion (38).

*Frizzled5*. Although FZD5 is considered by some authors a key molecule functioning as Wnt5a receptor, no significant association was observed by us between FZD5 expression and the patient group. This fact reflects the diversity of receptors and coreceptors that interacts with Wnt5a, such as FZD4, FZD2, Ror2 and others (39).

$\beta$ -catenin.  $\beta$ -catenin is considered a central molecule of the Wnt signaling pathway and has a dual function in cell adhesion (mainly through controlling E-cadherin-mediated cell adhesion at the plasma membrane) and transcription. Both of these function are involved in human tumorigenesis. The role of  $\beta$ -catenin in ovarian tumorigenesis is not clear yet. Rask *et al* found significant increase of  $\beta$ -catenin in ovarian cancer compared to normal ovary (40). To explain the fact that  $\beta$ -catenin is not mutated (and consequently, it is not stabilized) in most ovarian cancers, it was suggested that other factors that could influence the expression of  $\beta$ -catenin, such as overexpression of frequently rearranged in advanced T-cell lymphomas-1 (FRAT1) (41), overexpression of GSK3 and reduction of APC (40).

It is known that  $\beta$ -catenin has an important function in colorectal tumorigenesis (42). However, it does not seem to be the same for ovarian cancer. Most of the authors have shown that  $\beta$ -catenin mutation is found practically only in endometrioid ovarian type carcinomas and are rare in serous, clear cell and mucinous carcinomas. It is noteworthy that  $\beta$ -catenin is used to differentiate primary ovarian mucinous carcinoma from colorectal adenocarcinoma metastatic to the ovary, considering that  $\beta$ -catenin is overexpressed in colorectal adenocarcinoma. Furthermore, only the nuclear sublocalization of  $\beta$ -catenin is regarded as an indicator of dysregulation (43,44). In our specimens, we had only four endometrioid ovarian carcinomas from 38 patients (10%), and none from the three groups studied stained a nuclear sublocalization of  $\beta$ -catenin.

Davies *et al* found that  $\beta$ -catenin is consistently expressed in normal ovarian surface epithelium and benign tumors, and reduced or absent in ovarian carcinomas (45), which is in agreement to our findings whereas the malignant group did not differ from the others in terms of  $\beta$ -catenin expression. Even more, the proportion of patients in group C (normal) with positive score for  $\beta$ -catenin was significantly higher than group B (benign). Unfortunately, most studies of the surface epithelium are limited by its fragility; it is usually denuded by allowing the surface to dry intraoperatively or by touching or rubbing it during removal or gross pathologic examination (46).

We found a tendency to worst prognosis for  $\beta$ -catenin positive patients in group A, however it did not reach the significance level. Kildal *et al* found that cytoplasmic or membranous  $\beta$ -catenin staining were not of prognostic importance. Interestingly, they found that nuclear  $\beta$ -catenin staining was associated with improved survival, and occurred preferentially in endometrioid carcinoma. Similar results were described by Gamallo *et al*, they found that nuclear  $\beta$ -catenin indicates good prognosis in contrast to patients who expressed only membranous  $\beta$ -catenin (47), mostly in endometrioid ovarian carcinomas. For breast cancer patients, Dolled-Filhart *et al* also found worse outcome with decreased expression of  $\beta$ -catenin (48).



SPANDIDOS are a promising research field for new therapeutic strategies in cancer. It is known that certain drugs act through Wnt pathways. Although, most studies on the relationship of ovarian cancer and Wnt pathway is focused on the canonical/ $\beta$ -catenin pathway. Our findings suggest that the pathways related to Wnt5a also have a very important role in ovarian tumorigenesis and warrant further investigation. Wnt5a was found to be a predictor of poor prognosis for ovarian cancer.

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## References

- Boerboom D, Paquet M, Hsieh M, Liu J, Jamin SP, Behringer RR, Sirois J, Taketo MM and Richards JS: Misregulated Wnt/ $\beta$ -catenin signaling leads to ovarian granulosa cell tumor development. *Cancer Res* 65: 9206-9215, 2005.
- Nusse R, Brown A, Papkoff J, Scambler P, Shackleford G, McMahon A, Moon R and Varmus H: A new nomenclature for int-1 and related genes: the Wnt gene family. *Cell* 64: 231-232, 1991.
- McMahon JA and McMahon AP: Nucleotide sequence, chromosomal localization and developmental expression of the mouse int-1-related gene. *Development* 107: 643-650, 1989.
- Nusse R and Varmus HE: Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome. *Cell* 31: 99-109, 1982.
- Van Ooyen A and Nusse R: Structure and nucleotide sequence of the putative mammary oncogene int-1; proviral insertions leave the protein-encoding domain intact. *Cell* 39: 233-240, 1984.
- Johnson ML and Rajamannan N: Diseases of Wnt signaling. *Rev Endocr Metab Disord* 7: 41-9, 2006. Erratum in: *Rev Endocr Metab Disord* 8: 183, 2007.
- Habas R and Dawid IB: Dishevelled and Wnt signaling: is the nucleus the final frontier? *J Biol* 4: 2, 2005.
- Chen AE, Ginty DD and Fan CM: Protein kinase A signalling via CREB controls myogenesis induced by Wnt proteins. *Nature* 433: 317-322, 2005.
- Sarkar L and Sharpe PT: Expression of Wnt signalling pathway genes during tooth development. *Mech Dev* 85: 197-200, 1999.
- Iozzo RV, Eichstetter I and Danielson KG: Aberrant expression of the growth factor Wnt-5A in human malignancy. *Cancer Res* 55: 3495-3499, 1995.
- Danielson KG, Pillarisetti J, Cohen IR, Sholehvar B, Huebner K, Ng LJ, Nicholls JM, Cheah KS and Iozzo RV: Characterization of the complete genomic structure of the human WNT-5A gene, functional analysis of its promoter, chromosomal mapping, and expression in early human embryogenesis. *J Biol Chem* 270: 31225-31234, 1995.
- Hollyday M, McMahon JA and McMahon AP: Wnt expression patterns in chick embryo nervous system. *Mech Dev* 52: 9-25, 1995.
- Yamaguchi TP, Bradley A, McMahon AP and Jones S: A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo. *Development* 126: 1211-1223, 1999.
- Olson DJ, Gibo DM, Siggers G, Debinski W and Kumar R: Reversion of uroepithelial cell tumorigenesis by the ectopic expression of human wnt-5a. *Cell Growth Differ* 8: 417-423, 1997.
- Kremenevskaja N, von Wasielewski R, Rao AS, Schöfl C, Andersson T and Brabant G: Wnt-5a has tumor suppressor activity in thyroid carcinoma. *Oncogene* 24: 2144-2154, 2005.
- Jonsson M, Dejmeck J, Bendahl PO and Andersson T: Loss of Wnt-5a protein is associated with early relapse in invasive ductal breast carcinomas. *Cancer Res* 62: 409-416, 2002.
- Ishitani T, Kishida S, Hyodo-Miura J, Ueno N, Yasuda J, Waterman M, Shibuya H, Moon RT, Ninomiya-Tsuji J and Matsumoto K: The TAK1-NLK mitogen-activated protein kinase cascade functions in the Wnt-5a/Ca(2+) pathway to antagonize Wnt/ $\beta$ -catenin signaling. *Mol Cell Biol* 23: 131-139, 2003.
- Topol L, Jiang X, Choi H, Garrett-Beal L, Carolan PJ and Yang Y: Wnt-5a inhibits the canonical Wnt pathway by promoting GSK-3-independent  $\beta$ -catenin degradation. *J Cell Biol* 162: 899-908, 2003.
- Ricken A, Lochhead P, Kontogiannou M and Farookhi R: Wnt signaling in the ovary: identification and compartmentalized expression of wnt-2, wnt-2b, and frizzled-4 mRNAs. *Endocrinology* 143: 2741-2749, 2002.
- Krajewska M, Krajewski S, Epstein JI, Shabaik A, Sauvageot J, Song K, Kitada S and Reed JC: Immunohistochemical analysis of bcl-2, bax, bcl-X, and mcl-1 expression in prostate cancers. *Am J Pathol* 148: 1567-1576, 1996.
- Milovanovic T, Planutis K, Nguyen A, Marsh JL, Lin F, Hope C and Holcombe RF: Expression of Wnt genes and frizzled 1 and 2 receptors in normal breast epithelium and infiltrating breast carcinoma. *Int J Oncol* 25: 1337-1342, 2004.
- Holcombe RF, Marsh JL, Waterman ML, Lin F, Milovanovic T and Truong T: Expression of Wnt ligands and Frizzled receptors in colonic mucosa and in colon carcinoma. *Mol Pathol* 55: 220-226, 2002.
- Zilberberg A, Yaniv A and Gazit A: The low density lipoprotein receptor-1, LRP1, interacts with the human frizzled-1 (HFz1) and down-regulates the canonical Wnt signaling pathway. *J Biol Chem* 279: 17535-17542, 2004.
- GeneCards (<http://genome-www.stanford.edu/cgi-bin/genecards/carddisp.pl?gene=FZD1&search=Frizzled&stuff=txt>).
- Pukrop T and Binder C: The complex pathways of Wnt 5a in cancer progression. *J Mol Med* 86: 259-266, 2008.
- Zhang L, Huang J, Yang N, Liang S, Barchetti A, Giannakakis A, Cadungog MG, O'Brien-Jenkins A, Massobrio M, Roby KF, Katsaros D, Gimotty P, Butzow R, Weber BL and Coukos G: Integrative genomic analysis of protein kinase C (PKC) family identifies PKC $\alpha$  as a biomarker and potential oncogene in ovarian carcinoma. *Cancer Res* 66: 4627-4635, 2006.
- Weichert W, Gekeler V, Denkert C, Dietel M and Hauptmann S: Protein kinase C isoform expression in ovarian carcinoma correlates with indicators of poor prognosis. *Int J Oncol* 23: 633-639, 2003.
- D'Souza T, Indig FE and Morin PJ: Phosphorylation of claudin-4 by PKC $\epsilon$  regulates tight junction barrier function in ovarian cancer cells. *Exp Cell Res* 313: 3364-3375, 2007.
- Weeraratna AT, Jiang Y, Hostetter G, Rosenblatt K, Duray P, Bittner M and Trent JM: Wnt5a signaling directly affects cell motility and invasion of metastatic melanoma. *Cancer Cell* 1: 279-288, 2002.
- Jönsson M, Smith K and Harris AL: Regulation of Wnt5a expression in human mammary cells by protein kinase C activity and the cytoskeleton. *Br J Cancer* 78: 430-438, 1998.
- D'Souza T, Agarwal R and Morin PJ: Phosphorylation of claudin-3 at threonine 192 by cAMP-dependent protein kinase regulates tight junction barrier function in ovarian cancer cells. *J Biol Chem* 280: 26233-26240, 2005.
- Mikels AJ and Nusse R: Wnts as ligands: processing, secretion and reception. *Oncogene* 25: 7461-7468, 2006.
- Mikels AJ and Nusse R: Purified Wnt5a protein activates or inhibits  $\beta$ -catenin-TCF signaling depending on receptor context. *PLoS Biol* 4: 115, 2006.
- Gordon MD and Nusse R: Wnt signaling: multiple pathways, multiple receptors, and multiple transcription factors. *J Biol Chem* 281: 22429-22433, 2006.
- Liu G, Bafico A and Aaronson SA: The mechanism of endogenous receptor activation functionally distinguishes canonical and noncanonical Wnts. *Mol Cell Biol* 25: 3475-3482, 2005.
- Pukrop T, Klemm F, Hagemann T, Gradl D, Schulz M, Siemes S, Trümper L and Binder C: Wnt 5a signaling is critical for macrophage-induced invasion of breast cancer cell lines. *Proc Natl Acad Sci USA* 103: 5454-5459, 2006.
- Dejmeck J, Dejmeck A, Sjöholm A, Sjölander A and Andersson T: Wnt-5a protein expression in primary Dukes' B colon cancers identifies a subgroup of patients with good prognosis. *Cancer Res* 65: 9142-9146, 2005.
- Kurayoshi M, Oue N, Yamamoto H, Kishida M, Inoue A, Asahara T, Yasui W and Kikuchi A: Expression of Wnt-5a is correlated with aggressiveness of gastric cancer by stimulating cell migration and invasion. *Cancer Res* 66: 10439-10448, 2006.

39. Yamamoto H, Yoo SK, Nishita M, Kikuchi A and Minami Y: Wnt5a modulates glycogen synthase kinase 3 to induce phosphorylation of receptor tyrosine kinase Ror2. *Genes Cells* 12: 1215-1223, 2007.
40. Rask K, Nilsson A, Brännström M, Carlsson P, Hellberg P, Janson PO, Hedin L and Sundfeldt K: Wnt-signalling pathway in ovarian epithelial tumours: increased expression of beta-catenin and GSK3beta. *Br J Cancer* 89: 1298-1304, 2003.
41. Wang Y, Hewitt SM, Liu S, Zhou X, Zhu H, Zhou C, Zhang G, Quan L, Bai J and Xu N: Tissue microarray analysis of human FRAT1 expression and its correlation with the subcellular localisation of beta-catenin in ovarian tumours. *Br J Cancer* 94: 686-691, 2006.
42. Behrens J: The role of the Wnt signalling pathway in colorectal tumorigenesis. *Biochem Soc Trans* 33: 672-675, 2005.
43. Chou YY, Jeng YM, Kao HL, Chen T, Mao TL and Lin MC: Differentiation of ovarian mucinous carcinoma and metastatic colorectal adenocarcinoma by immunostaining with beta-catenin. *Histopathology* 43: 151-156, 2003.
44. Brembeck FH, Rosário M and Birchmeier W: Balancing cell adhesion and Wnt signaling, the key role of beta-catenin. *Curr Opin Genet Dev* 16: 51-59, 2006.
45. Davies BR, Worsley SD and Ponder BA: Expression of E-cadherin, alpha-catenin and beta-catenin in normal ovarian surface epithelium and epithelial ovarian cancers. *Histopathology* 32: 69-80, 1998.
46. Bell DA: Origins and molecular pathology of ovarian cancer. *Mod Pathol* 18: S19-S32, 2005.
47. Gamallo C, Palacios J, Moreno G, Calvo de Mora J, Suárez A and Armas A: Beta-catenin expression pattern in stage I and II ovarian carcinomas: relationship with beta-catenin gene mutations, clinicopathological features, and clinical outcome. *Am J Pathol* 155: 527-536, 1999.
48. Dolled-Filhart M, McCabe A, Giltane J, Cregger M, Camp RL and Rimm DL: Quantitative in situ analysis of beta-catenin expression in breast cancer shows decreased expression is associated with poor outcome. *Cancer Res* 66: 5487-5494, 2006.