

Short interfering RNA directed against the E2F-1 gene suppressing gastric cancer progression *in vitro*

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Abstract. Gastric cancer is the third most common cancer in China. The sustained overexpression of E2F-1 is a characteristic feature of gastric cancer. RNA interference (RNAi), which has been proven to be a powerful tool for suppressing gene expression, may provide a promising way forward in gastric cancer therapy. In this study, we constructed the recombinant Psilencer 4.1- E2F-1 siRNA plasmids and transfected them into gastric cancer MGC-803 cells *in vitro*. Our data demonstrated that E2F-1 siRNA led to inhibition of endogenous E2F-1 mRNA and protein expression as determined by real-time quantitative RT-PCR and Western blotting. Furthermore, simultaneous silencing of E2F-1 resulted in a reduction of tumor cell proliferation activity and a higher percentage of apoptotic cells. The inhibition of migration and invasion potential of tumor cells was investigated *in vitro*. In summary, siRNA targeting of E2F-1 can effectively inhibit gastric cancer progression and may be used as a potent therapy.

Introduction

Gastric cancer is one of the most frequent cancers and one of the most frequent causes of cancer-related mortality in China, with an incidence of 0.4 million new cases and 0.3 million deaths annually, and it ranks the third most common cancer in China (1). Although diagnostic and surgical techniques for treatment of gastric cancer have advanced in recent years, post-surgery survival rates have not improved in the last decade. The overall 5-year survival rate generally remains less than 30%, even after three-field lymph node dissection or combined chemotherapy and radiotherapy.

Tumor cell proliferation, migration and invasion which are processes controlled by a very complex cell cycle machinery are main causes of death in patients with gastric cancer.

Multiple components such as cyclins, cyclin dependent kinases (Cdks), Cdk inhibitors, Rb, and E2Fs are implicated in the transition from one cell cycle phase to another. For example, D-type cyclins bind and activate Cdk4/Cdk6, which phosphorylate the Rb gene product pRb, whereas Cdk inhibitors such as p21 and p16 can inhibit the function of Cdks. As a result of phosphorylation of pRb, E2F is released from the pRb-E2F complex. The E2F is an active transcription factor that promotes the transcription of the genes required for DNA synthesis and drives cells from the G1 phase into the S phase (2-4).

Dysregulation of the normal cell cycle-regulatory machinery is integral to the neoplastic process, and there is now compelling evidence implicating loss of cell cycle control in the development and progression of most human cancers (5). The transcription factor E2F-1 is the best-known ultimate transcription factor activated in the cyclin-Cdk-Rb pathway. E2F-1 can regulate progression through checkpoints in the cell cycle and has a dual role in cancer development with the capacity to act as both a tumor suppressor gene and an oncogene (6). E2F-1 enhances proliferation in gastrointestinal carcinomas while conversely stimulating apoptosis in bladder cancers independent of pRb expression (7-9). Overexpression of E2F-1 in head and neck squamous carcinoma cell (HNSCC) lines has the capacity to stimulate cell cycle reentry but is also associated with increased invasiveness, suggesting a role in metastasis (10).

The RNA interference (RNAi) means the use of double-stranded RNA (dsRNA) to target specific mRNAs for degradation, thereby silencing their expression. RNAi is one manifestation of a broad class of RNA silencing phenomena that are found in plants, animals and fungi. The discovery of RNAi has changed our understanding of how cells guard their genomes, led to the development of new strategies for blocking gene function, and may yet yield RNA-based drugs to treat human disease (11). Biochemical and molecular evidence suggests that short interfering RNAs (siRNAs) are generated in cells by endonucleolytic cleavage of exogenous dsRNA. The siRNAs are then thought to incorporate into a protein-RNA complex, the RNA-induced silencing complex (RISC), where they serve to guide a nuclease to the target mRNA (12).

To study whether E2F-1 plays a role in tumor progression, we transfected gastric cancer MGC-803 cells with E2F-1 siRNA to inhibit E2F-1 expression and investigated the

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Table I. The base sequence of primers and probes for real-time quantitative RT-PCR.

Gene	Primer	Base sequence	PCR product (bp)
E2F-1	Forward	5'-ACCCTGCAGAGCAGATGGTT-3'	100
	Reverse	5'-TTTGCTCTTAAGGGAGATCTGAA-3'	
β -actin	Forward	5'-AACTCCATCATGAAGTGTGA-3'	247
	Reverse	5'-ACTCCTGCTTGCTGATCCAC-3'	

ability of proliferation, migration and invasion suppressed by E2F-1 siRNA in gastric cancer cells.

Materials and methods

Cell culture. The human gastric carcinoma MGC-803 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) (Invitrogen, Gaithersburg, MD, USA). All media were supplemented with 10% FBS, penicillin (100 U/ml), and streptomycin (100 μ g/ml). The cells were cultured in an incubator at 5% CO₂ and 37°C, with medium changes every 3 days.

Construction and transfection of the siRNA plasmid expression vector. Double chains of oligonucleotide with complementary sequences that can code siRNA were obtained from Genesil Biotechnology Company (China). There were two reversed repeated sequences with 19 inserted sequences (CTGACTCTGCCACCATAGT, ACTATGGTGGCAGAGTCAG) in this complementary sequence, with *Bam*HI and *Hind*III in both sides for the ligation to Psilencer 4.1 vector. The siRNA targeting site of the transcribed product was nucleotides 124-1437 of E2F-1 mRNA (GeneBank No. NM-005225). The negative control was the siRNA sequence from the transcribed product of double-chain oligonucleotide with no homology to any human gene sequence.

After the ligation, the plasmid was transformed into *Escherichia coli* TOP10 cells, and then planted on solid LB medium. Ampicillin-resistant colonies were cultured at 37°C overnight in a rocking bed. The recombinant E2F-1 siRNA plasmid was prepared, and the sequences were verified by electrophoresis of the digested product. The inserted siRNA was 55 bp. MGC-803 cells (1 \times 10⁵) were inoculated into a 6-well plate and transfected with Psilencer 4.1- E2F-1(+) or Psilencer 4.1- E2F-1(-) recombination plasmids when the confluence was 90%. Forty-eight hours after transfection, MGC-803 cells were diluted to 1:10 for passage, and cultured for at least 2 weeks in medium with G418. The cells were divided into 3 groups: MGC-803/Silence(+) cells, MGC-803/Silence(-) cells and MGC-803 cells.

Real-time quantitative RT-PCR. Total RNAs were extracted from positive cell clone using TRIzol Reagent (Invitrogen). Neo gene segments were amplified and verified by real-time RT-PCR. Complementary deoxyribonucleic acids (cDNAs) were reverse-transcribed from 2 μ g of total RNA. Table I shows the PCR primer sequences used in this study. The products of PCR were checked by agarose gel electro-

phoresis, and the abundance of each mRNA was detected and normalized to that of glyceraldehyde-3-phosphate dehydrogenase (β -actin) mRNA.

Western blot analysis. Cell lysates were prepared in a buffer containing 0.1 M NaCl, 0.01 M Tris-Cl (pH 7.6), 0.001 M EDTA (pH 8.0), 1 μ g/ml aprotinin, 100 μ g/ml PMSF, and 1%(v/v) NP40. After protein quantitation using the Lowery protein assay, equal amounts of proteins were separated by SDS-PAGE and blotted onto nitrocellulose membranes by the semi-dry blotting method using a three buffer system. The membrane was blocked with 5%(w/v) BSA in PBST (PBS, pH 7.5, containing 0.1% Tween-20) and incubated with a 1:500 dilution of primary antibody (anti-E2F-1) (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA) overnight at 4°C. The membrane was then washed with PBST and incubated with a peroxidase-conjugated secondary antibody (1:1000) (Santa Cruz Biotechnology) for 1 h. Specific antibody binding was detected using a chemiluminescence detection system (Pierce, USA), according to the manufacturer's recommendations. Western blot film was scanned, and the net intensities of the bands were quantified using Image-Quant software (Molecular Dynamics, Sunnyvale, CA, USA). After development, the membrane was stripped and reprobed with antibody against β -actin (1:1000) (Santa Cruz Biotechnology) to confirm equal sample loading.

Cell proliferation and survival assay. The viability and proliferation of MGC-803 cells were determined by MTS assay using a CellTiter 96 AQueous assay system (Promega, Madison, WI, USA), according to the manufacturer's instructions. The assay measures the dehydrogenase enzyme activity in metabolically active tumor cells, as reflected by the conversion of MTS to formazan, which is soluble in tissue culture medium and was detected by absorbance at 490 nm. The production of formazan is proportional to the number of living cells, with the intensity of the produced color serving as an indicator of the cell viability. Briefly, the MGC-803 cells were plated at 1 \times 10⁵ cells per well in 96-well plates and cultured for 72 h. MTS mixed with medium without serum was added to the cell cultures after removing the old medium. The plates were incubated at 37°C for 4 h, and the absorbance (A) at 490 nm was determined using a 96-well Opsys MR™ microplate reader (ThermoLab systems, Chantilly, VA, USA) and Revelation™ QuickLink software. The blank control wells with medium only were set as zero absorbance. The percentage of cell survival was calculated using the background-corrected absorbance: % cell viability



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A of experimental well/A of untreated control well.
periments were performed at least three times, and
representative data are presented in Fig. 3.

Colony formation assay. Cell suspensions from each group were diluted in DEME with 10% FBS, and immediately replated in 6-well plates at a density of 20 cells/cm². The plates were incubated until cells in control wells formed sufficiently large colonies. After that, the colonies were fixed with 6% glutaraldehyde and stained with 0.5% crystal violet. The plates were photographed and their digital images were analyzed manually to known the colony number.

Cell cycle analysis by flow cytometry. For cell cycle analysis, MGC-803 cells (1x10⁶) were washed twice with ice-cold PBS, treated with trypsin, and then fixed in 70% cold ethanol at 4°C for 30 min. The cell pellet was incubated in a solution containing 50 ng/ml propidium iodide, 0.2 mg/ml RNase, and 0.1% Triton X-100 at room temperature for 30 min, and then analyzed by flow cytometry using a FACscan (Becton-Dickinson, Mountain View, CA, USA). The data were analyzed with the MultiCycle for Windows (Phoenix Flow Systems, San Diego, USA).

Apoptosis assay by flow cytometry. Apoptotic cells were determined using Annexin V-FITC Apoptosis Detection Kit (Jingmei Biotech Co., Shenzhen, China) and an EPICS XL-MCL flow cytometry (Becton-Dickinson) according to the manufacturer's instructions. Briefly, 1x10⁶ cells were stained with Annexin V/FITC for 30 min at 4°C in the dark and then propidium iodide for 10 min before flow cytometric analyses.

Wound healing assay. The cells were cultured to confluence on 6-well plates. A central linear wound was made with a 200 μ l sterile pipet tip. Media were changed gently to remove any floating cells. Phase micrographs of the wound cultures were taken at 0, 24, 36 and 48 h. The photographs were analyzed by measuring the distance from the wound edge of the cell sheet to the original wound site. Migration activity was calculated as the mean distance between edges of three points in 12 fields per well. Relative motility = (mean original distance - mean distance at a time point)/mean original distance x 100%. Each test group was assayed in triplicate.

Cell invasion assay. Cell invasion was assessed using Transwell chambers (6.5 mm; Corning, New York, USA) with sera-free DMEM 50 μ l containing 1 μ g/ μ l Matrigel (Department of Biology, Beijing University, China) in upper chamber. The lower chamber was filled with DMEM 50 μ l containing 0.1 μ g/ μ g fibronectin (Beijing University). Cells (1x10⁵) were suspended with 100 μ l DMEM with 1% fetal calf serum and plated into the upper chamber. PBS (5%) 500 μ l was added in lower chamber. After 24 h incubation at 5% CO₂ and 37°C, the number of cells with Giemsa staining on the undersurface of the polycarbonate membranes was scored visually in five random fields at x400 magnification using a light microscope.

Statistical analysis. Data are expressed as mean \pm standard error of the mean (SEM) analyzed by SPSS 13.0 (SPSS Inc.,

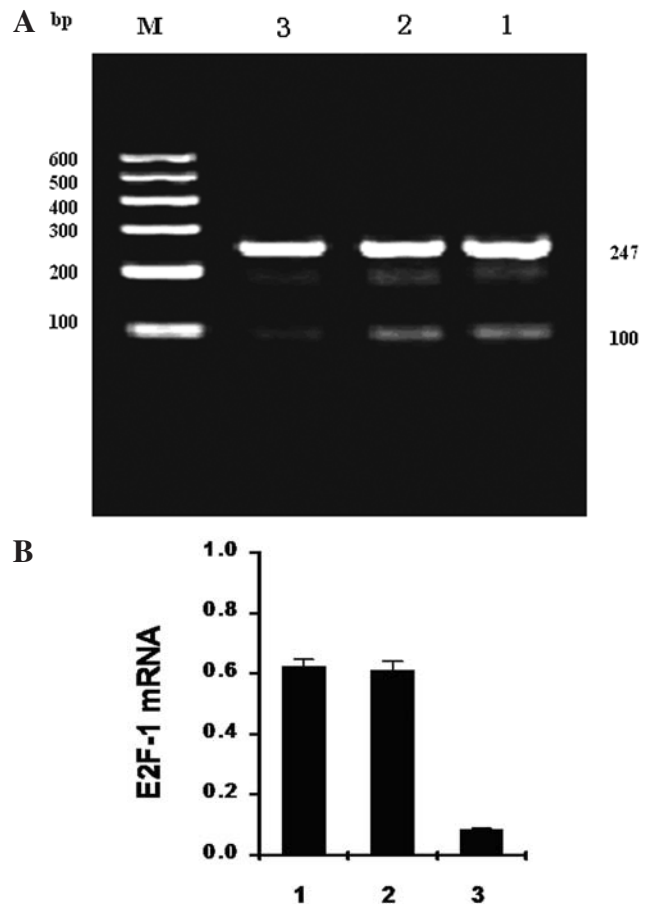


Figure 1. (A) The band of E2F-1 mRNA checked by real-time quantitative RT-PCR. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells. M, 600 bp marker. (B) The expression of E2F-1 mRNA in MGC-803/Silence(+) cells was strongly suppressed.

Chicago, IL, USA) and Origin 7.5 software programs (OriginLab Co., Northampton, MA, USA). Student's t-test was used to measure statistical significance between two treatment groups. Multiple comparisons were performed with a one-way analysis of variance (ANOVA). Data were considered significant at $P < 0.05$.

Results

Construction and verification of E2F-1 siRNA plasmid. Recombinant Psilencer 4.1- E2F-1(+) and Psilencer 4.1- E2F-1(-) sequences were verified by electrophoresis. The inserted siRNA was 55 bp with digestion sites *Bam*HI and *Hind*III on both sides, consistent with the designed series. This confirmed that the construction of the E2F-1 siRNA expression plasmid was successful.

Inhibitory effect of the expression of E2F-1 mRNA and protein. The transfection of Psilencer 4.1- E2F-1(+) plasmid into MGC-803 cells led to remarkable inhibition of E2F-1 mRNA and protein expression. Densitometric analysis showed that E2F-1 mRNA and protein in MGC-803/Silence(+) cells were about 6- and 5-fold lower than those of two control groups, respectively ($P < 0.05$), while no difference was found between MGC-803/Silence(-) cells and MGC-803 cells ($P > 0.05$) (Figs. 1 and 2).

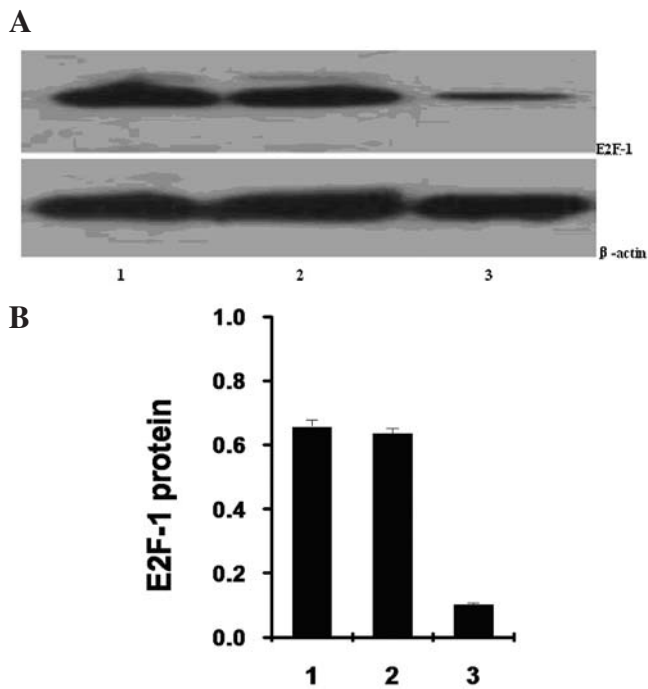


Figure 2. (A) The expression of E2F-1 protein in cells by Western blot analysis. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells. β -actin: internal control protein. (B) Statistical evaluation showed that E2F-1 siRNA significantly reduced E2F-1 protein concentrations in cells.

Psilencer 4.1- E2F-1(+) inhibits cell growth and proliferation in gastric cancer MGC-803 cells. Next, we determined the *in vitro* survival rates of gastric tumor cells stably transfected with *Psilencer 4.1- E2F-1(+)* plasmids, using gastric carcinoma cell line MGC-803 as a model for gastric cancer. As shown in Fig. 3, *Psilencer 4.1- E2F-1(+)* significantly reduced cell survival ($P < 0.05$), as assessed by MTS assay. MGC-803/Silence(-) cells and MGC-803 cells exhibited 3.3- and 3.2-fold higher mean proliferation rates, respectively than MGC-803/Silence(+) cells. Additionally, we observed that MGC-803/Silence(+) cells obviously grew slower than MGC-803/Silence(-) cells and MGC-803 cells, which was consistent with the decreased levels of E2F-1 in MGC-803/Silence(+) cells and indicated a suppressive effect of *Psilencer 4.1- E2F-1(+)* on MGC-803 cell growth and survival.

To confirm the inhibitory effect of *Psilencer 4.1- E2F-1 (+)* on the growth of MGC-803 cells, we performed colony formation assays for measuring the capability of the cell lines to grow in an anchorage-independent environment by culturing the cells in soft agarose. As shown in Fig. 4, three cell lines were able to form colonies in soft agarose but the number of colony formation in MGC-803/Silence(+) cells after three weeks was 46.1 ± 2.5 , with a 64.2 and 62.3% decrease, as compared with two control groups, respectively ($P < 0.05$). Together, these data suggest that E2F-1 siRNA inhibits cell growth and proliferation in the gastric cancer cell system.

Effect of Psilencer 4.1- E2F-1(+) on cell cycle control in gastric cancer MGC-803 cells. We used flow cytometry to determine whether the inhibitory effect of E2F-1 siRNA on

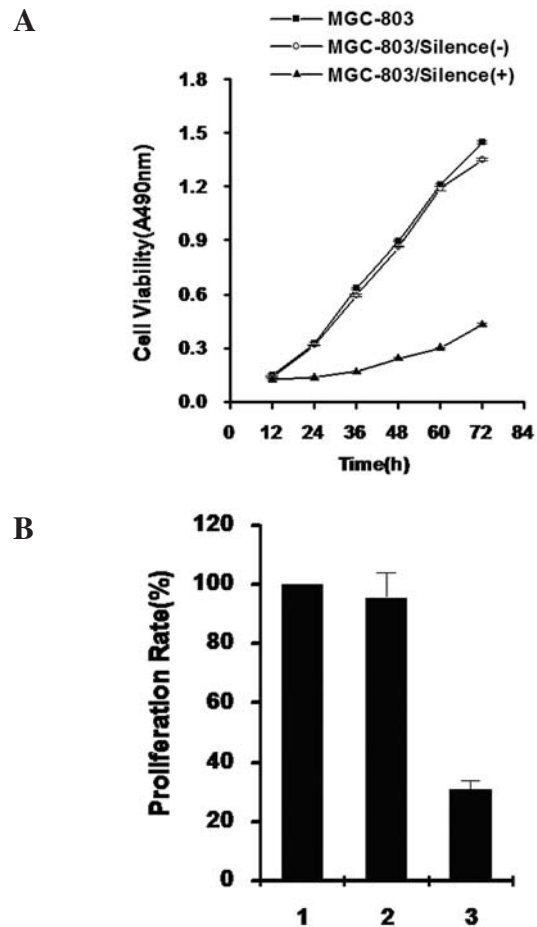


Figure 3. Cell survival assessed by MTS assay. (A) The inhibition of proliferation was observed after MTS treatment. (B) The mean cell proliferation rate of MGC-803/Silence(+) cells was significantly lower. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells.

Table II. Cell cycle analysis by flow cytometry in gastric cancer cells.

Group	Cell cycle phase		
	G ₀ /G ₁ (%)	S (%)	G ₂ /M (%)
MGC-803 cells	31.9	35.1	32.9
MGC-803/Silence(-) cells	36.8	29.7	33.4
MGC-803/Silence(+) cells	19.4 ^a	21.3 ^a	59.3 ^a

^a $P < 0.05$, MGC-803/Silence(+) cells vs. MGC-803 cells and MGC-803/Silence(-) cells.

MGC-803 cell proliferation was mediated, at least in part, through affecting cell cycle progression. We found that MGC-803/Silence (+) cells were 19.4% at G₀/G₁ phase and 59.3% at G₂/M phase, with 80.2 and 77.5% increase in G₂/M phase cell population and 39.2 and 47.3% decrease in G₀/G₁ phase cell population, as compared to MGC-803 cells and MGC-803/Silence(-) cells ($P < 0.05$) (Table II and Fig. 5). These data indicate that cell growth inhibition by *Psilencer 4.1-*

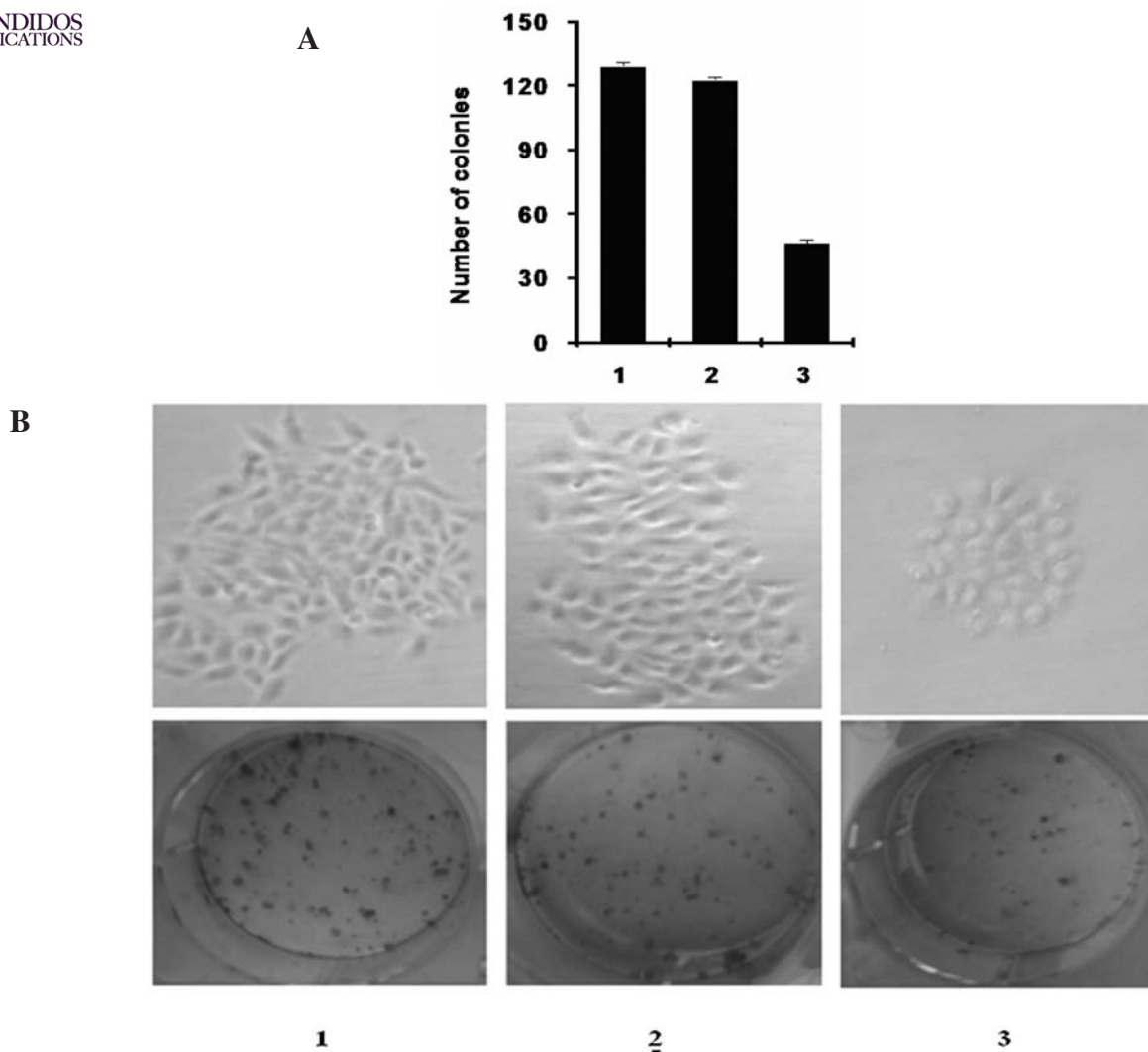


Figure 4. Results of colony formation assay. (A) MGC-803/Silence(+) cells exhibited fewer colonies than MGC-803/Silence(-) cells or MGC-803 cells. (B) A representative result of the colony formation assay. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells.

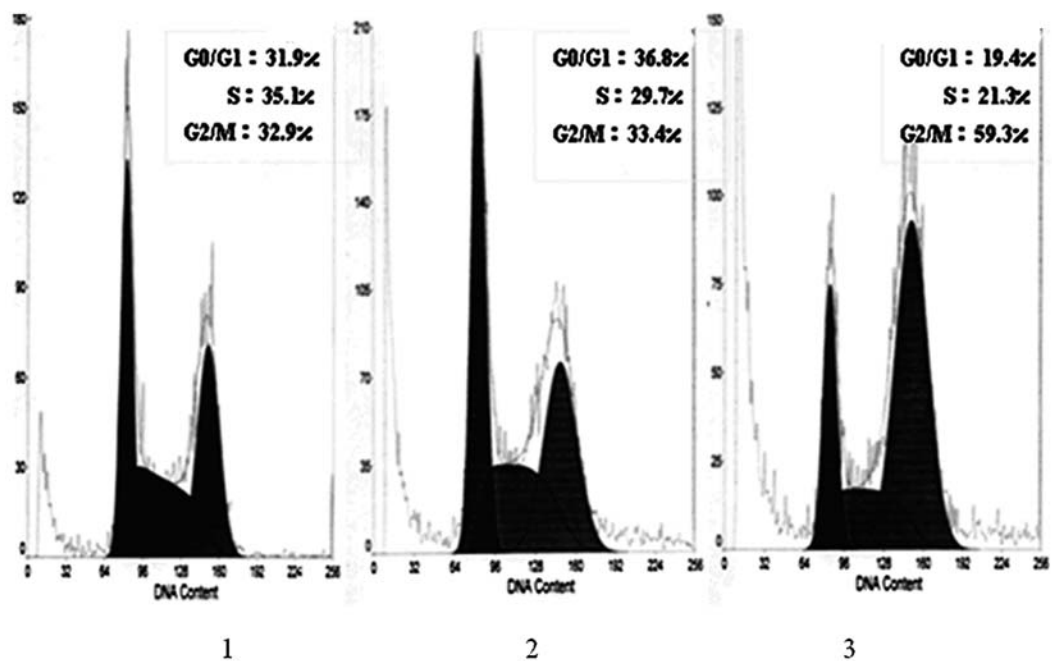


Figure 5. Cell cycle analysis by flow cytometry. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells.

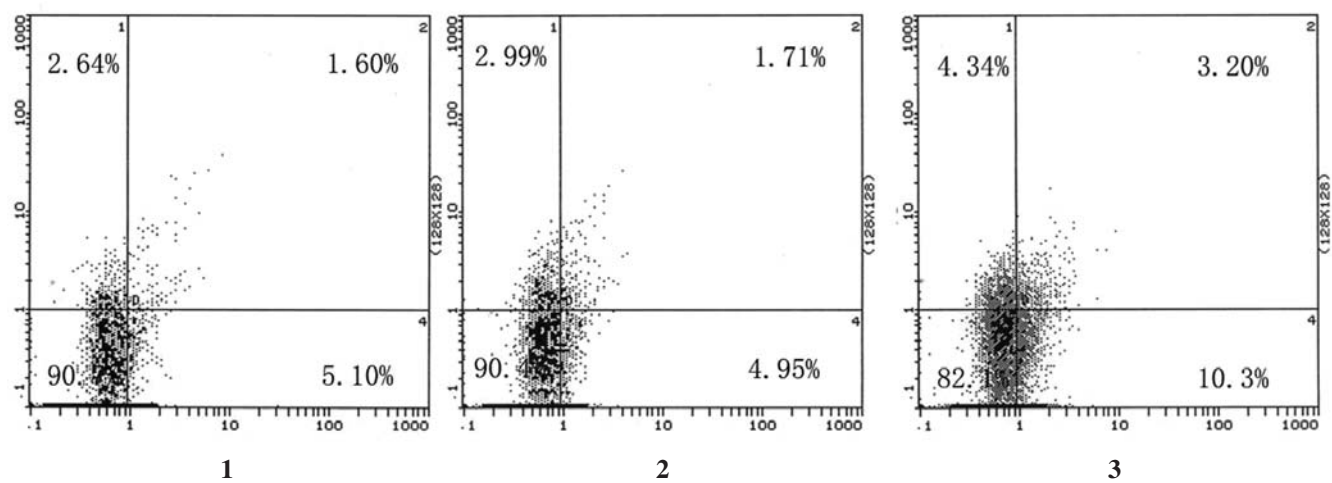
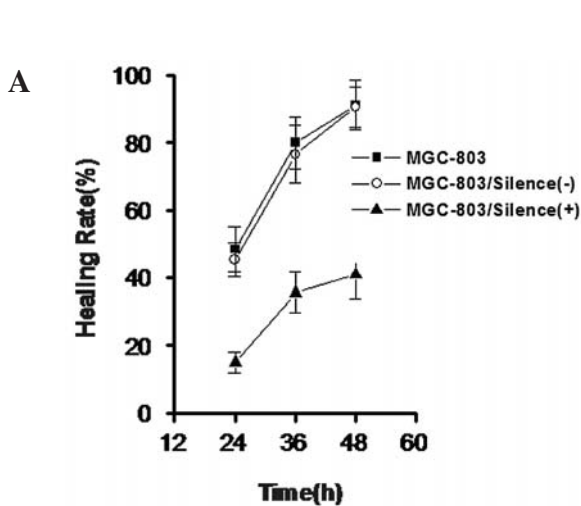


Figure 6. Percentages of apoptotic cells analyzed by flow cytometry. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells.



E2F-1(+) is associated with significant cell cycle arrest in G₂/M phase and suggest that siRNA directed against the E2F-1 gene suppresses cell proliferation by controlling the G₂/M checkpoint and inducing a specific block in cell cycle progression.

Psilencer 4.1- E2F-1(+) transfection induces cellular apoptosis. To further study the effect of E2F-1 on MGC-803 cell apoptosis, cells were stained with Annexin V-FITC and PI, and then subsequently analyzed by flow cytometry. This assay was based on the translocation of phosphatidylserine from the inner leaflet of the plasma membrane to the cell surface in the early apoptotic cells. The dual parameter

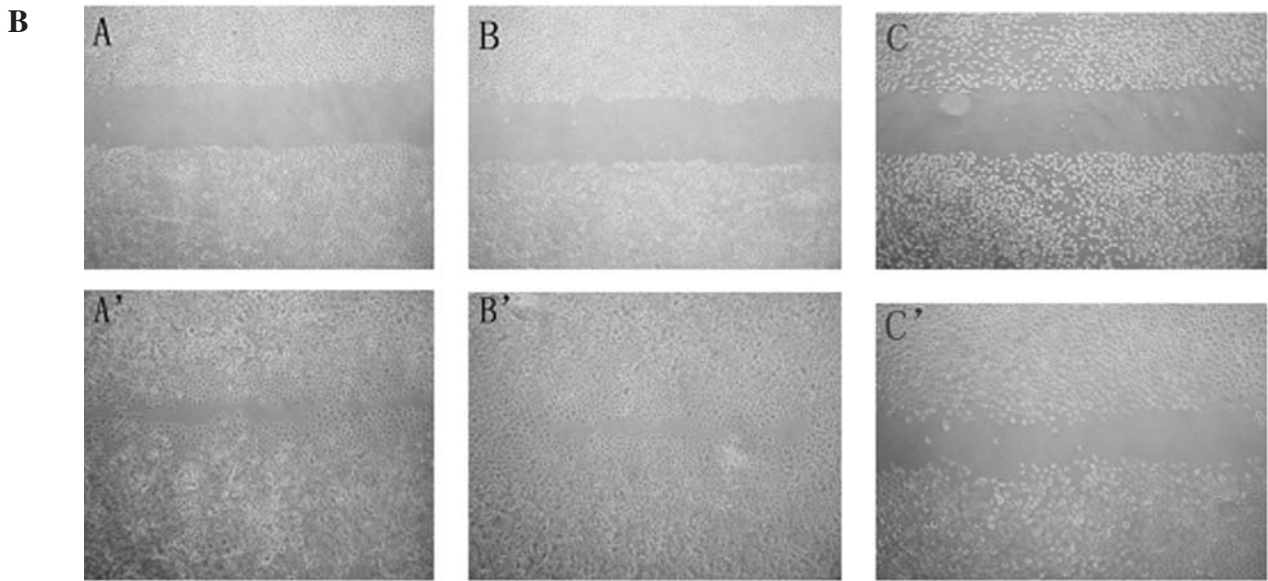


Figure 7. (A) E2F-1 siRNA decreased migration of MGC-803 cells in wound healing assay. (B) A representative result of cell migration. (A) MGC-803 cells at 0 h; (A') MGC-803 cells at 48 h; (B) MGC-803/Silence(-) cells at 0 h; (B') MGC-803/Silence(-) cells at 48 h; (C) MGC-803/Silence(+) cells at 0 h; (C') MGC-803/Silence(+) cells at 48 h.

SPANDIDOS PUBLICATIONS dot plots showed that the viable cells were in the $Annexin V^-/PI^-$ quadrant (Annexin V-/PI-), the cells at the early apoptosis were in the lower right quadrant (Annexin V+/PI-), and the ones at the late apoptosis or necrosis were in the upper right quadrant (Annexin V+/PI+). As indicated in Fig. 6, the percentage of MGC-803/Silence(+) cells with Annexin V+/PI- was significantly higher than that of MGC-803/Silence(-) or MGC-803 cells ($P<0.05$). It implies that inhibition of E2F-1 was able to induce apoptosis in gastric cancer MGC-803 cells.

Psilencer 4.1- E2F-1(+) transfection decreases migration and invasion of gastric cancer cells. We measured the migration ability of three cell groups using the wound healing assay by scratching the single-layer cells. The distance between the wound edges was determined at 0, 24, 36 and 48 h and the healing rate was calculated in the three groups. MGC-803/Silence(+) cells showed a lower migration ability at the time-points evaluated than MGC-803/Silence(-) cells ($P<0.05$) and MGC-803 cells ($P<0.05$). The healing rate of MGC-803/Silence(+) cells after 48 h was $40.9\pm7.2\%$, with 54.8 and 55.1% decrease, as compared with MGC-803/Silence(-) cells and MGC-803 cells (Fig. 7).

Because siRNA targeting of E2F-1 inhibited the expression of E2F-1 gene in gastric cancer cells, we assessed its ability to inhibit cell invasion. After incubation for 24 h in the invasion assay, the numbers of MGC-803/Silence(-) and MGC-803 cells invaded through the membrane of Matrigel chamber were 2.7- and 3.0-fold greater than that

of MGC-803/Silence(+) cells, respectively ($P<0.05$) (Fig. 8). The results indicate that E2F-1 siRNA reduces migration and invasion ability of gastric cancer MGC-803 cells.

Discussion

Gastric cancer is a worldwide problem. Besides the undetermined etiological factors, there are also limitations in surgery, chemotherapy and radiotherapy, the major therapies for gastric cancer to date (13). Many patients lose the chance of surgery because of their systemic condition, while many cannot tolerate the side effects of chemotherapy or radiotherapy. It is important to find a new way to effectively inhibit cancer cell growth and avoid the side effects of drugs or radioactive rays. Gene target therapies have proved to be a promising way to achieve this goal.

The recent discovery of RNAi, is a strong tool for silencing the function of specific genes, compared with the conventional gene-suppression technologies such as chemical inhibitors, antisense technology, ribozymes and deoxyribozymes, which often evoke non-specific side effects and/or offer only transient and partial suppression of the gene of interest. RNAi is a more potent inhibitor of gene expression with less toxicity (14). More importantly, given the potency and effectiveness of RNAi as a gene silencing tool, RNAi has significant therapeutic potential for human applications. Thus, RNAi-mediated gene silencing has opened new avenues in cancer therapy. The feasibility of RNAi-mediated gene silencing as a novel tool to arrest tumor growth and kill cancer cells is being tested in several laboratories and their initial results are promising (15).

In the present study, we constructed recombinant Psilencer 4.1-E2F-1 siRNA plasmids and successfully transfected these plasmids into gastric cancer MGC-803 cells. Our results clearly showed that the transfection of Psilencer 4.1-E2F-1(+) plasmid led to remarkable inhibition of E2F-1 mRNA and protein expression in MGC-803 cells, and it inhibited cell growth and made cell cycle arrest in G_2/M phase. The results in our study also showed that E2F-1 siRNA induced cell apoptosis. Furthermore, RNAi-directed targeting

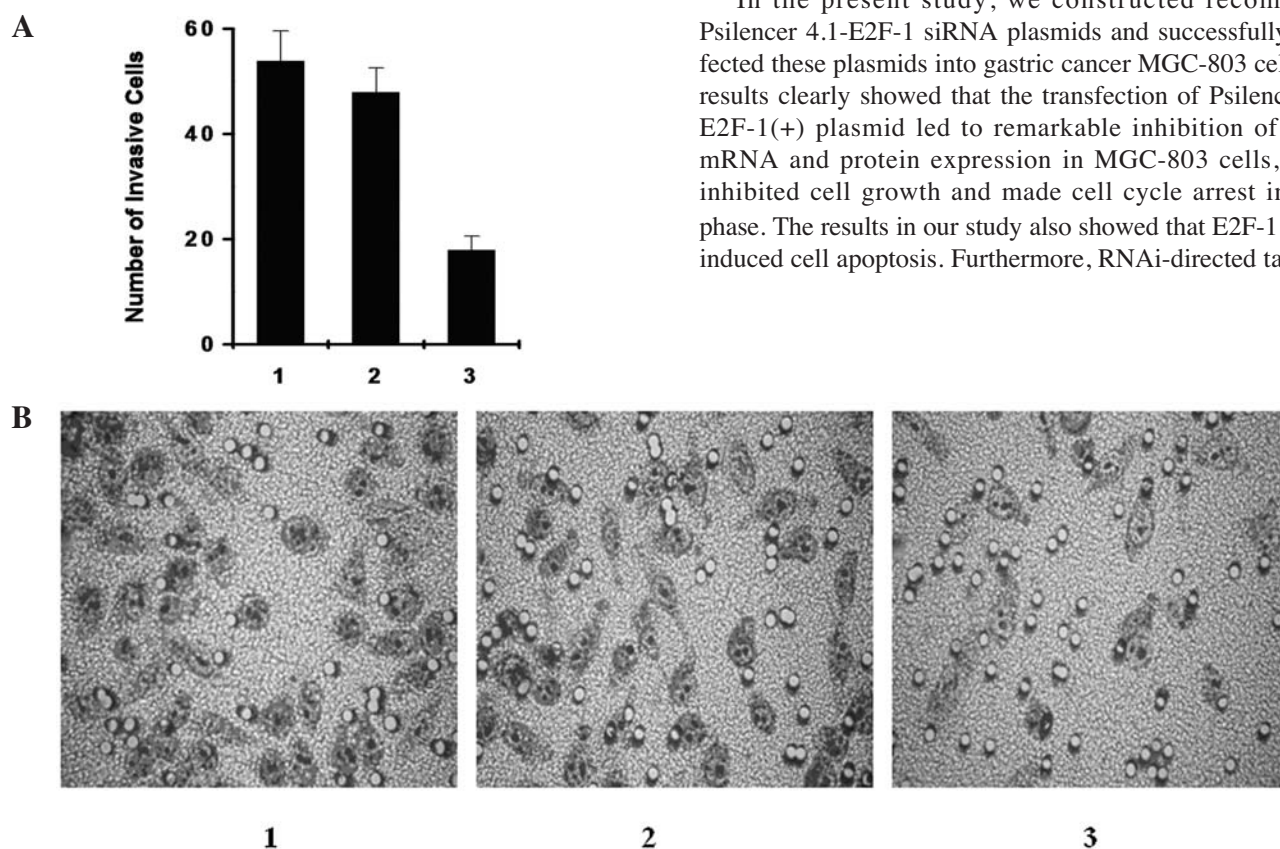


Figure 8. (A) E2F-1 siRNA decreased invasion of MGC-803 cells. (B) A representative result of cell invasiveness obtained from three independent experiments. Lane 1, MGC-803 cells; lane 2, MGC-803/Silence(-) cells; lane 3, MGC-803/Silence(+) cells.

of E2F-1 in MGC-803 cells could reduce the capability of cell motion, invasion and colony formation. The above evidence indicates that siRNA targeting E2F-1 plays an important role in the inhibition of gastric cancer progression *in vitro*. These inhibitory effects were due to post-transcriptional silence mechanisms, in which the synthetic, double-stranded siRNA effectively silenced E2F-1 in gastric cancer MGC-803 cells. This is consistent with many studies, which showed that the introduction of a 21 nt dsRNA into cancer cells strongly suppressed the expression of specific mRNAs (16,17). The sequence of this dsRNA was highly homologous and unique to a region of the targeted mRNA (18). The expression of E2F-1 protein was therefore inhibited, and the percentage of apoptotic cells was higher than that of control groups and the tumor growth was obviously suppressed.

The E2F transcription factor can regulate expression of numerous cellular genes controlling proliferation, including proto-oncogenes and genes regulating cell cycle progression. Therefore, the genes comprising the E2F gene family could potentially contribute to carcinogenesis. E2F-1 was the first cloned and is the better characterized member of this gene family. E2F-1 is the best-known transcription factor regulated by the cyclin-Cdk-Rb pathway. E2F-1 controls the expression of genes encoding various DNA replication proteins and cell cycle regulators, and it seems logical that E2F-1-dependent expression of these genes would be needed to sustain cell proliferation (19,20).

Previous observations indicated that inhibition of E2F-1 activity led to a perturbation of cell cycle progression. Thus, an E2F-1 dominant-negative mutant lacking the DP-binding site inhibited both the transcriptional activity of E2F-1 and cell proliferation (21). Likewise, a dominant-negative DP-1 mutant that forms inactive heterodimers with E2F proteins has been reported to induce a cell cycle arrest in G₁, and RNA aptamers that specifically blocked E2F DNA binding also inhibited cell proliferation after microinjection into primary fibroblasts (22,23). Our work confirms and extends these observations. In the present study, we reported the inhibitory effects of specific siRNA on E2F-1 expressed in human gastric cancer cells and the results showed that E2F-1 siRNA was able to significantly reduce cell growth and induced a cell cycle arrest in G₂/M.

Apart from its essential function in cell cycle progression, a role for E2F-1 in regulating apoptosis is well documented. The physiological relevance of E2F-1 control in promoting cell death has been clearly demonstrated by the development of spontaneous tumors in E2F-1 null mice, indicating that E2F-1 can act as a tumor suppressor gene (24). This function of E2F-1 is in part because of its ability to activate proapoptotic genes, such as p73 and Apaf-1 (25,26). The observations made in the present study are in apparent contrast to this proapoptotic function of E2F-1, but is in line with several other reports in the literature. A short DP-1-derived peptide linked to penetratin to enable cellular translocation inhibited E2F-DP heterodimerization and caused apoptosis of tumor cell lines (27). It has been described that E2F activity is essential for the survival of Myc-overexpressing tumor cells regardless of their ARF and p53 status (28). It would thus appear that E2F-1 can exert opposing functions with respect

to the regulation of apoptosis, and that the precise outcome of E2F-1 inhibition may depend on the genetic status of the cell and the loss-of-function approach.

One characteristic feature of any malignant tumor is the ability of tumor cells to invade and migrate into surrounding and/or distal tissue. Cell invasion is due to degradation of the ECM and the basement membrane surrounding the primary tumor by proteases. These proteases are involved in tumor growth and invasion at both primary and metastatic sites, particularly at the invading foci. Although several extracellular protease systems have been implicated in the tissue degradation and remodeling that often accompanies cancer invasion, several studies demonstrated that the E2F-1 system is important to these processes (29). In the present study, we found inhibition of migration and invasion potential of MGC-803 cells *in vitro* because of the transfection of Psilencer 4.1- E2F-1(+) plasmid.

Our previous study showed that E2F-1 overexpression in gastric cancer MKN-45 cells produced similar results to E2F-1 siRNA (30). However, the inhibition by E2F-1 overexpression was associated with cell cycle arrest in G₁/S phase but not G₂/M phase as E2F-1 siRNA. It suggests that appropriate activity and expression level of E2F-1 are necessary for normal cell cycle, even in promoting tumor proliferation and regression. Upregulation or downregulation of E2F-1 can suppress gastric cancer progression *in vitro* but with different mechanisms.

In conclusion, the downregulation of E2F-1 using RNAi successfully reduced the proliferation, migration and invasion of MGC-803 cells *in vitro*. These findings suggest that RNAi-directed targeting of E2F-1 can be used as a potent and specific therapeutic tool for the treatment of gastric cancer, especially in inhibiting and/or preventing cancer cell progression.

Acknowledgements

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