

# A LOW number of tumor-infiltrating FOXP3-positive cells during primary systemic chemotherapy correlates with favorable anti-tumor response in patients with breast cancer

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**Abstract.** Cancer cells induce proliferation and local accumulation of immunosuppressive cells, such as FOXP3-positive cells known as regulatory T cells (Tregs), leading to tumor-induced immune tolerance. Although cancer chemotherapy is usually considered immunosuppressive, some chemotherapeutic agents activate an anticancer immune response. Therefore, we postulated that the number of tumor-infiltrating FOXP3-positive cells during primary systemic chemotherapy (PSC) correlates with therapeutic outcomes in patients with breast cancer. Between September 2000 and January 2005, we examined 93 patients with breast cancer diagnosed by core-needle biopsy and treated with PSC. Core-needle biopsy (CNB) and surgical resected specimens were stained with a FOXP3 mouse monoclonal antibody to compare the numbers of FOXP3-positive cells in the tumors before and after PSC. A median cut-off value of >16.3/high power field (HPF) and >6.6/HPF defined high numbers of Tregs in CNB and in surgical specimens, respectively. We then assigned the patients into 4 groups (HH, high number of FOXP3-positive cells in both CNB and surgical specimen; LL, low number in both specimens; HL, high in CNB and low in the surgical specimen; LH, low in CNB and high in surgical specimen). Lymph vessel invasion-positive, clinically non-responder and

ER-negative tumors contained significantly more FOXP3-positive cells after PSC ( $p=0.04$ ,  $p=0.03$  and  $p=0.04$ , respectively). Prognosis was better among patients with low numbers than high numbers of FOXP3-positive cells both in CNB and in surgically resected specimens. In multivariate analysis, LL group demonstrated significantly better recurrence-free survival with risk ratio of 5.81 (95%CI, 1.09-107.5;  $p=0.04$ ) rather than that of non-LL group (LH, HL and HH). These findings suggest that the number of FOXP3-positive cells identified during PSC represents a promising predictive factor that might also be an important therapeutic target for breast cancer.

## Introduction

Primary systemic treatment (PST) before surgery has become a standard therapeutic strategy for operable breast cancer. Primary systemic chemotherapy (PSC) is as effective as adjuvant systemic chemotherapy, and it also has several theoretical advantages. Firstly, full-dose chemotherapy can be administered before any potential debilitation from surgery that might increase the likelihood of non-compliance with chemotherapy at a time when micrometastases are likely to be at their smallest. Second, it permits radiological and pathological assessment of the effect of a regimen that might guide the selection of postoperative chemotherapy. Thirdly, it can reduce tumor burden and increase the likelihood of breast-conserving surgery with negative margins.

Systemic chemotherapy is usually considered to be immunosuppressive, considering its toxicity towards hematopoietic cells. However, some chemotherapeutic agents can activate an anticancer immune response by depleting immunosuppressive cells such as CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells and myeloid-derived suppressor cells (1-3). The X chromosome-encoded forkhead transcription factor FOXP3 has been established as a key transcription factor for CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells (Tregs) (4) that play a vital role in preventing autoimmunity and pathology inflicted by uncontrolled immune

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responses to infection. However, increasing evidence indicates that Tregs also play an important role in immune evasion mechanisms used by cancers (5-13). Tumors actively recruit and induce Tregs to block innate and adaptive immune priming, effector function and memory response, which can result in a favorable environment for cancer progression.

Therefore, we postulated that a low number of tumor-infiltrating FOXP3-positive cells during PSC constitutes an unfavorable environment for breast cancer and that it correlates with the antitumor response in patients with breast cancer. The present study examines correlations between the number of tumor-infiltrating FOXP3-positive cells during PSC and therapeutic effects (tumor response, RFS and OS) in patients with breast cancer.

## Materials and methods

**Patients and tissue sampling.** Ninety-three patients who underwent primary systemic chemotherapy (PSC) followed by definitive surgery during September 2000 and January 2005 at Tokyo Metropolitan Cancer and Infectious Diseases Center Komagome Hospital, Tokyo, Japan were consecutively enrolled in this study. Informed consent was obtained from all patients before PSC and institutional review board of Komagome hospital approved.

All patients were diagnosed with invasive breast carcinoma by needle biopsy; 3 were excluded because excisional biopsy was performed before PSC and 3 others in pathological complete remission were also excluded because of difficulties associated with identifying 'tumor-infiltrating' Tregs. Thus, 87 of the originally enrolled patients were evaluated.

**Clinical and pathological response evaluation.** Clinical responses to PSC were evaluated by ultrasonography based on the Response Evaluation Criteria in Solid Tumors (RECIST).

We applied the Japanese pathological response criteria defined as follows: grade 0, no chemotherapeutic change in remnant cancer cells; grade 1a, 0-1/3 of remnant cancer cells in degeneration or necrosis; grade 1b, 1/3-2/3; grade 2, >2/3; grade 3, no viable cancer cells in duct and stroma (14,15).

**Immunohistochemical staining and quantification of FOXP3 positive cell.** Thick paraffin sections (4  $\mu$ m) mounted on silane-coated glass slides were dewaxed in xylene, rehydrated through a graded ethanol series and then incubated in 1 mM EDTA at 98°C for 40 min, followed by 0.3% hydrogen peroxide in methanol for 15 min to inhibit endogenous peroxidase activity, primary FOXP3 mouse monoclonal antibody (clone: mAbcam22509) diluted 1:50 in PBS for 30 min at room temperature and the reagent provided with the Elite ABC kit (Vectastain, Vector Laboratories, Burlingame, CA, USA). Cells were visualized using the chromogen diaminobenzidine and counterstained with Meyer's hematoxylin.

Average numbers of FOXP3-positive cells within the neoplastic epithelium and immediately adjacent stroma were determined from scores in 3 and 5 random high-power fields (HPF; x40 objective and x10 eyepiece, CFWN 10x) in CNB and surgical specimens, respectively. A median cut-off value

Table I. Patient background (n=87).

Age (years)	23-69 (median, 51)
Follow-up period	5.3-89.1 months (median, 46.3)
Tumor size before PSC	0.9-13.5 cm (median, 4.7)
PSC regimen	Anthracycline only 17 (FEC, 500-100-500/m <sup>2</sup> or EC, 90-600/m <sup>2</sup> q3w) Taxane containing 70 (Docetaxel 75/m <sup>2</sup> q3w or Paclitaxel 80/m <sup>2</sup> qw)
Clinical response <sup>a</sup>	
CR	3
PR	57
SD	22
PD	3
Unknown	2
Pathological response <sup>b</sup>	
Grade 0	15
Grade 1a	37
Grade 1b	17
Grade 2	18

<sup>a</sup>Evaluated by US based on RECIST criteria; CR, complete remission; PR, partial response; SD, stable disease; PD, progressive disease.

<sup>b</sup>Pathological response defined as: grade 0, no chemotherapeutic change in remnant cancer cells; grade 1a, 0-1/3 of remnant cancer cells in degeneration or necrosis; grade 1b, 1/3-2/3; grade 2, >2/3; grade 3, no viable cancer cells in duct and stroma.

of >16.3 and >6.6/HPF defined patients with high numbers of Tregs in CNB and surgical specimens, respectively.

**ER, PgR and HER2 status.** For estrogen receptor (ER), progesterone receptor (PgR), and human epidermal growth factor receptor-2 (HER2) evaluation, immunohistochemical staining were performed using anti-ER mouse monoclonal antibody (clone 1D5; Dako), anti-PgR mouse monoclonal antibody (clone PgR636; Dako) and a HercepTest kit (Dako), respectively. Hormone receptor status was evaluated as the percentage of positive nuclear staining among cancer cells, and the cut-off value was set to 10%. HER2 scoring was carried out according to the standard Hercep Test Guidelines.

**Statistical analysis.** The association between number of Tregs and clinicopathological/biological features was examined using the  $\chi^2$  test and Fisher's exact probability test. Correlations between the numbers of FOXP3 and RFS and OS were determined using the log-rank and Wilcoxon tests. Associations between changes in numbers of FOXP3-positive cells during PSC and prognosis were assessed using the Cox proportional hazard model.

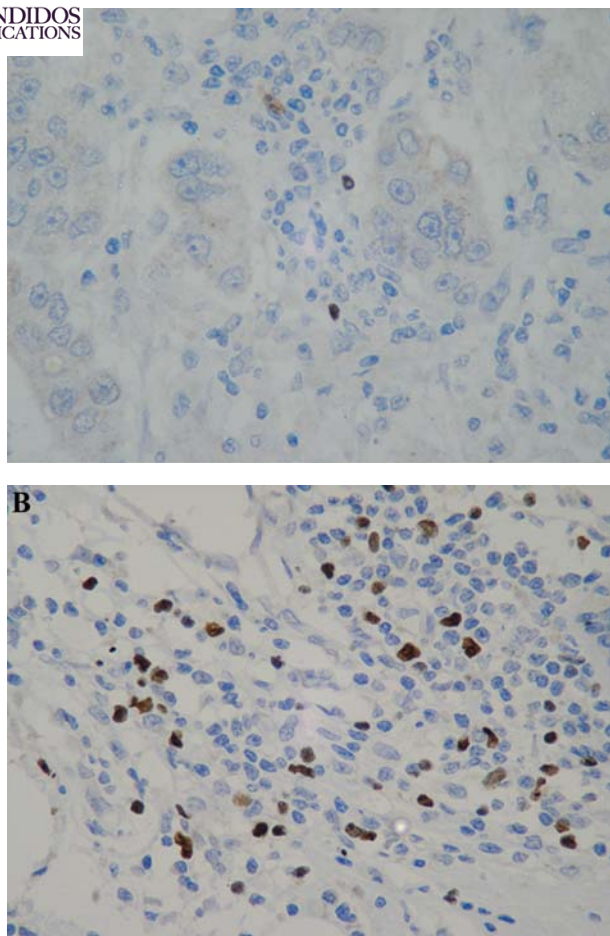


Figure 1. Representative immunohistochemical staining with FOXP3 antibody. (A) Low number FOXP3-positive cells; (B) high number FOXP3-positive cells.

The data were analyzed using the JMP(r) version 7 software package (SAS institute Inc, USA). A p-value <0.05 was considered statistically significant.

## Results

**Patient background.** The median age of the enrolled patients was 51 years (range, 23-69 years), median tumor size before PSC was 4.7 cm (range, 0.9-13.5 cm) and median follow-up period was 46.3 months (range, 5.3-89.1 months). All patients received a regimen containing anthracycline (FEC or EC) and 70 patients (80.5%) received a sequential regimen containing taxane. None of the patients were administered with trastuzumab or hormonal therapy before surgery. Sentinel lymph node biopsy was conducted on 32 patients before starting PSC and 8 patients did not undergo axillary lymph node dissection after PSC. Fifty-seven patients achieved partial response (PR) and 22 remained stable disease (SD). Three patients each were in clinically complete remission (CR) and progressive disease (PD), respectively.

Fifteen patients were diagnosed with a grade 0 pathological response, and 37, 17 and 18 patients were diagnosed with grades 1a, 1b and 2, respectively. Three patients achieved complete pathological remission and were excluded from analysis (Table I).

Table II. Correlation between number of tumor-infiltrating Tregs and clinicopathological features of 87 CNB specimens before PSC.

Variable	FOXP3 <16.3 Low (n=43)	FOXP3 ≥16.3 High (n=44)	P-value
Age (years)			
<50	20	19	0.75
≥50	23	25	
ER status			
Negative	4	18	0.007
Positive	39	26	
PgR status			
Negative	12	29	0.004
Positive	31	15	
HER2 status			
0, 1, 2	39	37	0.45 <sup>a</sup>
3	4	6	
Unknown	0	1	
Clinical response			
CR + PR	30	30	0.87
SD + PD	12	13	
Unknown	1	1	
Pathological response			
Grade 0, 1	33	36	0.56
Grade 2	10	8	
Recurrence-free survival			
		Worse	0.285 <sup>b</sup>
			0.188 <sup>c</sup>
Overall survival			
		Worse	0.036 <sup>b</sup>
			0.092 <sup>c</sup>

n, total number of patients; ER, estrogen receptor; PgR, progesterone receptor; HER2, human epidermal growth factor receptor-2; CR, complete remission; PR, partial response; SD, stable disease; PD, progressive disease. <sup>a</sup>Fisher's exact probability test; <sup>b</sup>log-rank test; <sup>c</sup>Wilcoxon test.

**FOXP3 expression in CNB specimens and surgically resected specimens.** The number of FOXP3-positive cells ranged from 0 to 112/HPF in CNB specimens (median 16.3) and from 0 to 66.8/HPF in surgically resected specimens (median 6.6). Patients were classified according to the median number of FOXP3-positive cells (Fig. 1). Negative ER was associated with significantly more FOXP3 infiltrates before and after PSC (p=0.007 and p=0.04, respectively) (Tables II and III). The findings were the same among PgR-negative patients before, but not after PSC (p=0.004 and p=0.60, respectively).

Lymph vessel invasion was prominent in the group with a high number of FOXP3 infiltrates (p=0.04) (Table III).



Table III. Correlation between number of tumor-infiltrating Tregs and clinicopathological features of 87 specimens resected after PSC.

Variable	FOXP3 <6.6 Low (n=43)	FOXP3 ≥6.6 High (n=44)	P-value
Age (years)			
<50	20	19	0.75
≥50	23	25	
PSC regimen			
Anthracycline only	6	12	0.13
Taxane included	37	32	
Nodal status			
Negative	16	21	0.70
Positive	20	22	
Ax not performed	7	1	
Remnant tumor size (cm)			
≤2	27	23	0.32
>2	16	21	
Lymph vessel invasion			
Negative	26	17	0.04
Positive	17	27	
ER status			
Negative	9	18	0.04
Positive	34	26	
PgR status			
Negative	27	30	0.60
Positive	16	14	
HER2 status			
0, 1, 2	29	32	0.45 <sup>a</sup>
3	1	5	
Unknown	13	7	
Clinical response			
CR + PR	33	27	0.03
SD + PD	9	16	
Unknown	1	1	
Pathological response			
Grade 0, 1	32	37	0.27
Grade 2	11	7	
Recurrence-free survival		Worse	0.031 <sup>b</sup> 0.028 <sup>c</sup>
Overall survival		Worse	0.060 <sup>b</sup> 0.142 <sup>c</sup>

n, total number of patients; PSC, primary systemic chemotherapy; ER, estrogen receptor; PgR, progesterone receptor; HER2, human epidermal growth factor receptor-2; CR, complete remission; PR, partial response; SD, stable disease; PD, progressive disease.  
<sup>a</sup>Fisher's exact probability test; <sup>b</sup>log-rank test; <sup>c</sup>Wilcoxon test.

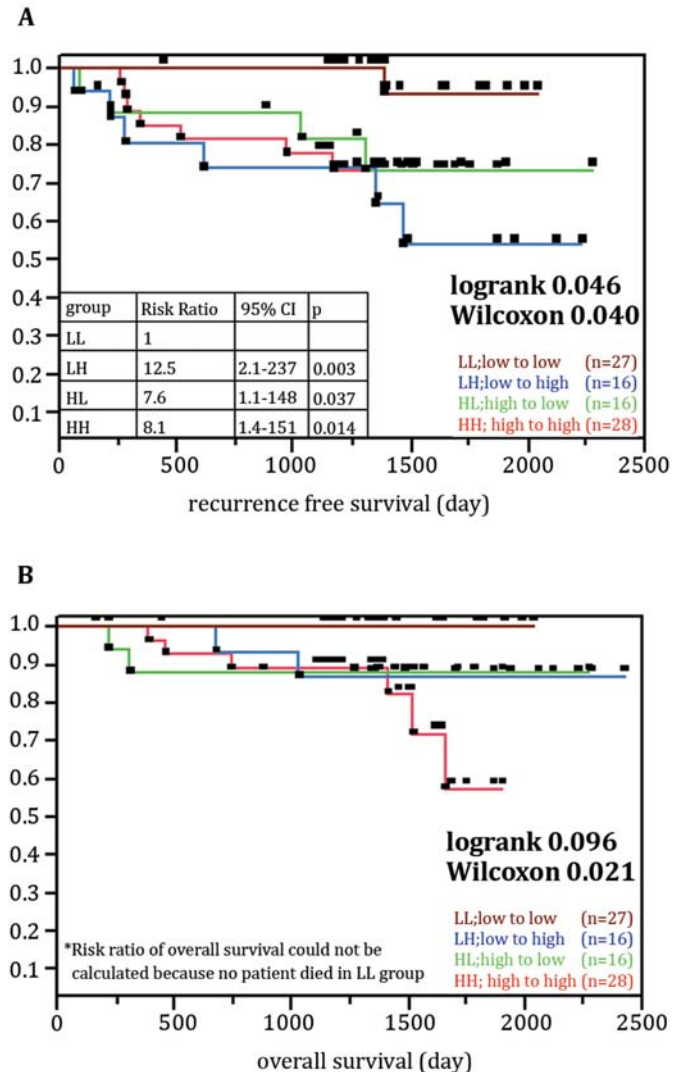


Figure 2. Changes in numbers of FOXP3-positive cells before and after chemotherapy. (A) Recurrence-free survival; (B) overall survival; HH, high number of FOXP3-positive cells in both CNB and surgical specimens; LL, low numbers in both specimens; HL, high number in CNB and low number in surgical specimen; LH, low number in CNB and high number in surgical specimen.

However, nodal status and the number of FOXP3-positive cells ( $p=0.70$ ) did not significantly differ. Importantly, a low number of Tregs in tumors after PSC correlated with clinical response (CR+PR) ( $p=0.03$ ), although numbers of Tregs did not correlate with pathological response ( $p=0.27$ ). Numbers of Tregs in tumors before PSC did not have any predictive impact on clinical and pathological response (Tables II and III). The trend of recurrence-free and overall survival in low-FOXP3 patients evaluated either by CNB or surgical specimen was clearly favorable (Tables II and III).

We also assigned all patients into 4 groups according to the numbers of FOXP3-positive cells in both CNB and surgical specimen: high numbers in both (HH;  $n=28$ ), low numbers in both (LL;  $n=27$ ), high in CNB and low in surgical specimen (HL;  $n=16$ ), and low in CNB and high in surgical specimen (LH;  $n=16$ ). Prognosis was significantly better in the LL group (Fig. 2). In multivariate analysis, LL group



Variable	RFS			OS		
	Risk ratio	95% CI	P-value	Risk ratio	95% CI	P-value
ER	3.10	1.09-8.61	0.03	36.3	5.26-776.3	<0.0001
FOXP3						
(LL vs. others)	5.81	1.09-107.5	0.04	<sup>a</sup> 1		0.32
Nodal status	1.48	0.51-4.71	0.48	12.1	1.40-299.2	0.02
Age	1.43	0.54-89	0.47	7.4	1.28-52.0	0.02

<sup>a</sup>1, risk ratio of overall survival could not be calculated because no patient died in LL group.

demonstrated significantly better recurrence-free survival with risk ratio of 5.81 (95%CI, 1.09 to 107.5;  $p=0.04$ ) rather than that of non-LL group, however, no significant correlation was found in overall survival (Table IV).

## Discussion

FOXP3 is a specific marker for regulatory T cells (Tregs), which comprise a distinct group of T lymphocytes with immunosuppressive properties that normally serve to prevent harmful autoimmune responses. However, Tregs can also interfere with beneficial immune responses in humans, such as anti-tumor immunity. FOXP3-positive Tregs are associated with tumor genesis and progression in lung (16), colon (17), melanoma (18), gastric (19), hepatic (20), and pancreatic (21) carcinoma, lymphoma (22) and sarcoma (23). High levels of FOXP3 mRNA expression or high numbers of FOXP3-positive cells are associated with unfavorable tumor features and a poor prognosis among patients with breast cancer (24-28).

An appropriate cut-off value must be established before the numbers of FOXP3-positive cells in tumor cells can be clinically applied. Our study showed that high numbers, defined as median cut-off values of 16.3 and 6.6 cells/HPF in CNB and in surgical resected specimens, respectively, are associated with aggressive tumor features and poor prognosis. Bates *et al* (28) reported that a high number of FOXP3 cells is associated with unfavorable nodal status, tumor grade, ER status, HER2 status and relapse-free survival with a median cut-off of 15 cells/mm diameter invasive tumor cores (approximately 3 cells/HPF). Bohling *et al* (27) reported that the average number of FOXP3-positive cells/HPF in a group with good clinical feature ranges from 15 to 20/HPF. Although optimization of the cut-off value for clinical use is necessary, these data indicate that the appropriate cut-off to differentiate high and low numbers of FOXP3-positive cells in tumors might be in the range 3-20. Interestingly, we found that the median cut-off value for surgical specimens after PSC was decreased to 6.6/HPF. This might have a detrimental effect on FOXP3-positive Tregs in patients undergoing chemotherapy and could be another important factor for optimizing the cut-off value of FOXP3-positive cell number using tumor specimens after PSC.

Although one possible prognostic factor is pCR after PSC (29), few prognostic factors are available for patients with operable breast cancer after PSC. We found here that fewer FOXP3-positive cells before and after PSC indicates a better prognosis and in the multivariate analysis, preserving relatively low number of FOXP3 during PSC (LL group) is significantly associated with a better RFS; therefore, the number of FOXP3-positive cells in the tumors during PSC could become another prognostic factor in the treatment for breast cancer patients with PSC.

Further analysis discovered changes in the number of FOXP3-positive cells in tumors after PSC (Fig. 2). Although we cannot simply show that numbers of FOXP3-positive cells changed after PSC in the LH and HL groups, the prognosis of the LH group was significantly worse than that of the LL group. Furthermore, the risk ratio for overall survival of HL to HH was 0.57, and HL seems to indicate a better overall survival than HH, although the p-value did not reach significance (data not shown). Given these findings and the previous experimental reports that control of Tregs augments antitumor immune responses which provide clinical benefits and some chemotherapeutic agents such as cyclophosphamide specifically diminish Tregs (30,31), we speculate that some chemotherapeutic agents could clinically control FOXP3-positive cells and this might be one of the mechanism of chemotherapeutic agents besides its cytotoxic effect for patients with breast cancer. Further studies are required to prove our hypothesis.

In conclusion, numbers of FOXP3-positive Tregs could constitute an important prognostic factor for patients with breast cancer treated with PSC, and FOXP3-positive cells in tumors could be a novel therapeutic target that could improve outcomes for such patients.

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