

1 paclitaxel encapsulated in cationic liposomes: A new option for neovascular targeting for the treatment of prostate cancer

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Abstract. Neovascular targeting is an established approach for the therapy of prostate cancer (PCa). Cationic liposomes have been shown to be absorbed by immature vascular endothelial cells due to negative electric charge of their outer cell membrane. We aimed to evaluate the antitumoural efficacy of paclitaxel encapsulated in cationic liposomes for the treatment of PCa. Tumours were generated by subcutaneous injection of 10⁶ MatLu tumour cells into the right hind leg of 21 male Copenhagen rats. After tumour growth, the animals were treated by an i.v. infusion with either 5% glucose (Gl), paclitaxel (Pax), cationic liposomes (CL) or paclitaxel encapsulated in cationic liposomes (EndoTAG-1) on days 12, 14, 16 and 19. Treatment was initiated on day 12 after tumour inoculation at mean tumour volumes of 0.31±0.13 mm³. On the last day of treatment, animals treated with EndoTAG-1 had the significantly lowest tumour volumes with 2.49±0.84 cm³ vs. Pax (5.59±0.45 cm³) vs. CL (3.87±1.25 cm³) vs. GL (5.17±1.70 cm³). The quantification of MVD showed the lowest count for EndoTAG-1-treated tumours (11.78±2.68 vessels/mm²) followed by Gl (15.64±6.68 vessels/mm³), Pax (18.22± 9.50 vessels/mm³) and CL (40.9±32.8 vessels/mm³). The data confirm that neovascular targeting with EndoTAG-1 is a promising new method for the treatment of PCa by reducing the primary tumour mass and demonstrating benefits in the suppression of angiogenesis in comparison with the conventional treatment.

Introduction

Prostate cancer (PCa) is the most frequently diagnosed cancer in men in North America and Western Europe (1). The therapeutic concept of PCa conforms to the tumour stage, age and general condition of the patient. As a local PCa is curable by an operation or radiotherapy, the advanced or disseminated form of PCa is treated with androgen deprivation therapy in a palliative intention. However, most patients with an anti-hormonal therapy have a disease progression after a median of 13-20 month (2). After failure of an anti-hormonal therapy, the options for reducing the tumour mass are limited by a lack of effective therapy options.

Angiogenesis is necessary for tumour growth and metastasis (3). Tumour endothelial cells display higher proliferation rates than the endothelium in non-malignant tissue which make them more vulnerable for cytotoxic agents (4). 'Neovascular targeting' is an established approach in cancer therapy by destructing the tumours microvasculature (5). This concept was already demonstrated to be successful in preclinical studies for PCa with different antiangiogenic molecules (6,7).

Paclitaxel is a microtubule-stabilizing antineoplastic cytotoxic drug which has antiangiogenic activity (8). It is approved for the treatment of different advanced solid cancers like metastatic ovarian and breast cancer and non-small cellular lung carcinoma. Currently, an application in PCa therapy exists only within study protocols. Because of the lipophilic character of paclitaxel, it is used with the solvent Cremophor EL, which provokes serious side-effects like neurotoxicity and hypersensitivity reactions.

Liposomes were first discovered as a reproducible and efficient method to introduce plasmid DNA into cells (9). Since then, liposomes represent an established drug delivery system for lipophilic substances by improving the therapeutic index and increasing the application dose of antitumoural agents (10). However, the clinical usage is impaired by the rapid clearance of liposomes by the reticuloendothelial system and their low bioavailability (11). In contrast to these conventional liposomes, positively charged cationic liposomes were originally developed for non-viral gene transfer into mammalian cells *in vitro*. On the background of

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negatively charged capillary endothelium in tumours (12), cationic liposome preparations were evaluated to be preferentially bound to and internalized by angiogenic negatively charged endothelial cells in tumours and chronic inflammation (13). Recently, cationic liposomes loaded with chemotherapeutic drugs have been evaluated as delivery vehicle for *in vivo* targeting of drugs to the tumour vasculature (14,15).

In this study, we investigated the concept of neovascular targeting with paclitaxel encapsulated in cationic liposomes in an animal model as a new approach for advanced PCa. Therefore, we compared the macro- and microscopic aspects of tumour progression of the novel and a conventional therapeutic concept.

Materials and methods

Animal model. The trial was performed in accordance with the approved institutional protocol and the guidelines of the Institutional Animal Care and Use Committee. The Dunning rat PCa model is established for the evaluation of new treatment options against PCa (16). Male Copenhagen rats (Harlan-Winkelmann, Borken, Germany) were used in this study. After four weeks of acclimatisation, the animals were kept in separate cages after implantation of a central venous catheter.

Cell line. The Dunning R3327H rat prostatic adenocarcinoma subline MatLu was obtained from the ECACC (European Collection of Cell Cultures, catalogue number 94102735, Salisbury, UK). This subline displays a high metastatic ability with no androgen sensitivity and spreads almost exclusively via blood vessels into the lung. The tumour doubling time is estimated to be 2.7 ± 0.3 days *in vitro* and 3 ± 0.2 days *in vivo* (17).

The cells were cultivated at 37°C, 5% CO₂ and 100% humidity in 10 ml RPMI-1640 medium (Invitrogen, Paisley, Scotland, UK) supplemented with 1 ml 10% fetal bovine serum (PAN Biotech, Aidenbach, Germany), 0.2 ml 2 mM glutamine and 2.8 µl 250 nM dexamethasone (both from Sigma, Taufkirchen, Germany).

Tumour implantation. Cell suspensions of 5×10^6 cells/ml were generated and 1×10^6 cells were injected subcutaneous into the right flank region of the rat with a 25G cannula (Becton-Dickinson, Heidelberg, Germany) under general anaesthesia with isoflurane (Forene®, Abott, Wiesbaden, Germany).

Drug preparation. EndoTAG-1 (Munich Biotech, Munich, Germany) and Paclitaxel (Bristol-Myers Squibb, Munich, Germany) were applied at a dose of 5.0 mg/kg b.w. Empty cationic liposomes (without paclitaxel) were provided by Munich Biotech and administered at the same lipid concentration as EndoTAG-1. All preparations were given volume adapted.

The treatment protocol contained an i.v. injection volume of 12 µl/g b.w. per animal administered over a 3 min period under general anaesthesia with isoflurane. EndoTAG-1 and empty liposomes, which were stored as lyophilisate at 4°C,

were prepared for injection by addition of 13.8 ml aqua ad injectabilia. Paclitaxel was solved in 5% glucose (B. Braun, Melsungen, Germany).

Treatment protocol. After tumour implantation on day 0 and tumour growth for 10 days, 24 h prior to the beginning of treatment, the animals received a central venous catheter (PAE-microtube 0.58x0.96 mm, NeoLab, Heidelberg, Germany) in the left V. femoralis under general anaesthesia with ketamine (100 mg/kg/b.w. i.p.; Hostaket®, Intervet, Wiesbaden, Germany) and Xylazine (3 mg/kg/b.w. i.p.; Rompun 2%®, Bayer, Leverkusen, Germany). The animals were randomized into four groups (n=5 in Pax, Cl and Gl group; n=6 in EndoTAG-1 group) and were treated with either 5% glucose (Gl), Pax, cationic liposomes (CL), or Pax encapsulated in cationic liposomes (EndoTAG-1) on day 12, 14, 16 and 19.

Besides the evaluation of the general animal condition and nutritional status including body weight, the tumour was measured in three levels by calliper and volume which was calculated according to the formula $V = a \times b \times c \times 0.5$.

Immediately after euthanasia of the animals on day 20, gross necropsy was performed with special emphasis on tumour, lungs, liver, kidneys and spleen. The tumour tissue was snap-frozen and stored at -80°C, the removed organs were fixed in 4% unbuffered formaldehyde (Merck, Darmstadt, Germany) and embedded in paraffin by a tissue embedding system (Leica TP 1020®, Wetzlar, Germany).

Microvessel density. The cryo-preserved tumour tissue was sectioned at 6 µm thickness and transferred to Super-Frostplus® slides (Langenbrinck, Emmendingen, Germany). After drying overnight, the slides were fixed for 10 min in acetone and subsequently air-dried.

The microvasculature of the tumour was evaluated by immunohistochemical staining of von Willebrand factor: To block endogenous peroxidase activity the sections were treated with 0.3% hydrogen peroxide in methanol for 30 min and rinsed in PBS (pH=7.0). The tumour tissue was incubated with avidine and after PBS washing with biotine (Avidine-Biotine-Blocking Kit®, Linaris, Wertheim-Bettinger, Germany) for 15 min respectively. Followed by washing with PBS, the slides were incubated with goat serum (ABC-kit Elite PK-6101®, Vector Laboratories, Burlingame, CA, USA) for 30 min. Polyclonal rabbit anti-human von Willebrand factor (PAK010M®, Linaris) was used as the primary antibody in a dilution of 1:500 for 1 h. Before incubating with the biotinylated goat anti-rabbit as the secondary antibody (ABC-kit Elite PK-6101) for 30 min, the sections were rinsed in PBS. The tissue was coated with avidine-biotine-complex for 30 min. The sections were finally stained with diaminobenzidine (DAB-Peroxidase kit SK-4100®, Vector; CA, USA) and washed in water. Last, the sections were counterstained with haematoxylin, dehydrated, and mounted.

The evaluation of microvessel density (MVD) was determined in the area of highest vascularisation. These vascular 'hot spots' were selected by scanning the tumour section at a low magnification (x40). Vessels were counted within the three highest vascularised areas by one investigator at a

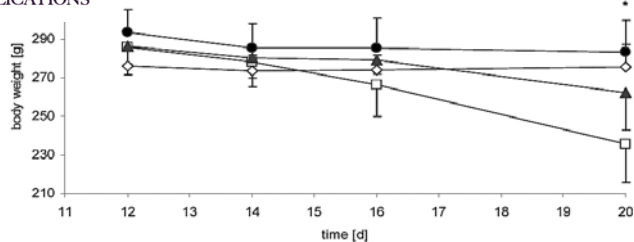


Figure 1. Development of mean body weight between the four groups GL (○), EndoTAG-1 (□), Pax (▲) and CL (●) during treatment. No difference was observed between the four groups on day 12, 14 and 16. On day 20 animals in EndoTAG-1 and Pax group revealed a significant decrease in body weight (* $p < 0.002$ EndoTAG-1 and Pax vs. CL and GL; variance analysis).

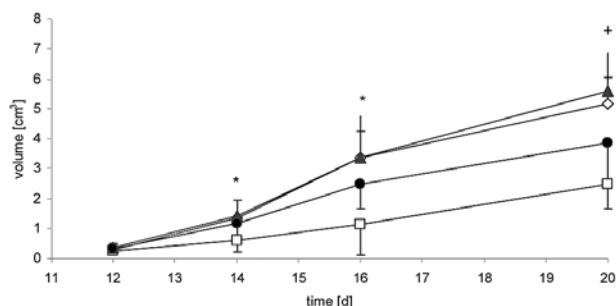


Figure 2. Development of mean tumour volume between the four groups GL (○), EndoTAG-1 (□), Pax (▲) and CL (●) during treatment. Significant difference in tumour volume between the groups were observed from the second day of therapy (day 14 $p < 0.138$; day 16 $p < 0.006$; day 20 $p < 0.003$; variance analysis). EndoTAG-1 group developed the significant smallest tumours vs. all other treatment arms on day 14 (* $p < 0.018$) and day 16 (* $p < 0.002$); EndoTAG-1 vs. GL and Pax on day 20 (* $p < 0.003$; all F-test).

magnification of $\times 200$. The average microvessel density was measured in vessels per mm^2 by a standardised counting grid. This protocol constitutes an established method for the quantification of MVD (18).

Histopathology. Organ sections of $3 \mu\text{m}$ were deparaffinated and rehydrated according to standard procedures and soaked in haematoxylin and water for 10 min. The sections were stained in 0.1% eosin for 2 min, dehydrated and mounted.

Statistics. The statistical analysis was performed using SAS (Release 8.02; SAS Institute Inc., Cary, NC, USA). Results are presented as mean \pm SEM. $P < 5\%$ was considered to be significant.

An ANOVA was used to show differences between the four treatment groups for the body weight of animals and for the tumour volume. Scheffe's test compared the body weight of the EndoTAG-1- and Pax group. Dunnett's test, F-test and t-test investigated differences in tumour volume between EndoTAG-1 group and the other groups. Kruskal-Wallis-Test and Mann-Whitney U-test were used to compare the MVD between the therapy groups.

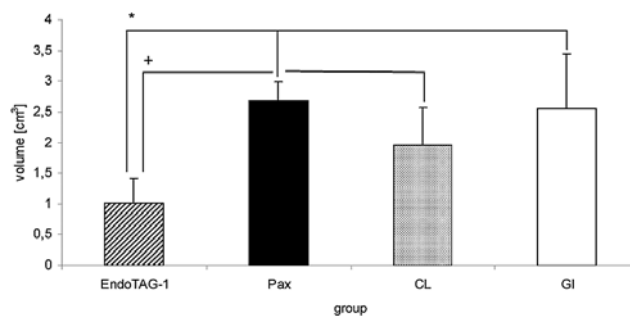


Figure 3. Group-specific mean tumour volume over the whole treatment period. EndoTAG-1 treated tumours possessed the smallest volume (*GL vs. EndoTAG-1 $p = 0.01$; *Pax vs. EndoTAG-1 $p = 0.01$; *Liposome vs. EndoTAG-1 $p = 0.05$; Dunnett's test).

Results

Every animal developed a palpable subcutaneous tumour in the right flank region between day 7 and 10 after cell implantation. All animals finished the trial except for one animal in the EndoTAG-1 group. Due to the distinct deterioration of general condition and nutritional status, it was euthanized on day 17.

At the beginning of the i.v. treatment protocol on day 12, there were no differences in general condition and body weight between the four groups (285 ± 12.8 g; $p = 0.20$, variance analysis).

Animals which were treated with glucose (274.4 ± 11.8 g) and pure cationic liposomes (283.2 ± 16.1 g) finished treatment without a significant change of the body weight. Body weight of animals in the chemotherapeutic groups was reduced significantly during treatment (day 20: EndoTAG-1 235.6 ± 19.9 g, $p = 0.0002$; Pax 262.2 ± 19.5 g, $p = 0.0237$, variance analysis) without a significant difference between the EndoTAG-1 and the Pax group ($p > 0.05$, Scheffe's test) (Fig. 1).

Histopathology. On histopathological examination of organs, the majority of animals showed, regardless of group affiliation, mural thrombus consisting of fibrin, inflammatory cells, detritus and tumour cells.

The status of the lymphatic system in animals of the EndoTAG-1 and Pax group differed from the control groups. It displayed an absence of lymphocytes in lymph nodes and lymphatic tissues whereas the animals from GL and CL groups showed signs of an activated immune response.

Tumour progression. At the beginning of the treatment (day 12), there was no interrelation between body weight and tumour volume ($p = 0.38$, $r = 0.21$, variance analysis) and no difference in tumour volume between all groups (0.31 ± 0.12 cm^3 , $p = 0.68$ variance analysis). Significant differences of the tumour volume between the groups were observed from the second day of therapy (day 14 $p < 0.138$; day 16 $p < 0.006$; day 20 $p < 0.003$; all variance analysis). At the end of the trial (day 20), lowest tumour volume was found in the EndoTAG-1 group (2.49 ± 0.8 cm^3) followed by CL (3.87 ± 1.3 cm^3), GL (5.17 ± 2.1 cm^3) and Pax (5.59 ± 0.5 cm^3). EndoTAG-1 treated tumours possessed the significantly

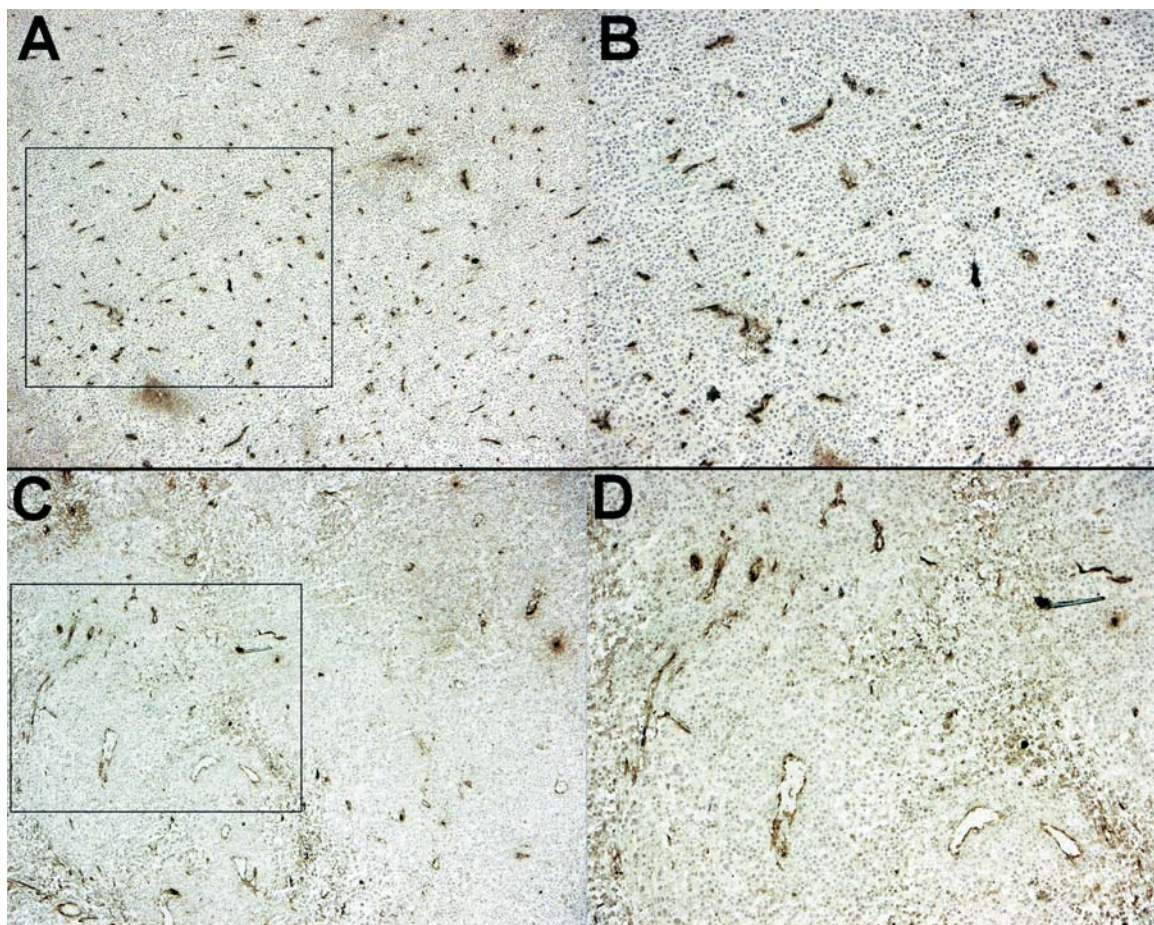


Figure 4. Immunohistology of tumour tissue. Endothelial cells were stained with vWF antibodies to evaluate MVD of the tumour. The vascular 'hot spots' were shown in low magnification (x40) on the left (A and C) and in high magnification (x200) on the right (B and D). Highest MVD was found in the CL group (A and B), EndoTAG-1 treated tumours revealed the lowest MVD (C and D).

smallest tumour volumes compared with Gl (day 14 $p < 0.008$; day 16 $p < 0.001$; day 20 $p < 0.002$) and Pax (day 14 $p < 0.004$; day 16 $p < 0.001$; day 20 $p < 0.001$). In contrast to CL group, the neoplasms in the EndoTAG-1 group were significantly smaller on day 14 ($p < 0.033$) and day 16 ($p < 0.011$), but not on day 20 ($p < 0.077$; all t-test) (Fig. 2). The group-specific mean tumour volume over the whole treatment differed significantly (EndoTAG-1 $1.01 \pm 0.4 \text{ cm}^3$, CL $1.96 \pm 0.6 \text{ cm}^3$, Gl $2.55 \pm 0.8 \text{ cm}^3$, Pax $2.69 \pm 0.3 \text{ cm}^3$, $p = 0.0008$, variance analysis). The mean tumour volume in the EndoTAG-1 group was significantly smaller than in all other groups (Gl $p = 0.01$; Pax $p = 0.01$; CL $p = 0.05$; Dunnett's test) (Fig. 3).

The MVD was heterogeneous within the groups, so the results showed only a trend between the four treatment arms ($p = 0.0521$, Kruskal-Wallis-Test). Lowest MVD was found in the EndoTAG-1 group ($11.78 \pm 2.68 \text{ vessels/mm}^2$) followed by Gl ($15.64 \pm 6.68 \text{ vessels/mm}^2$), Pax ($18.22 \pm 9.50 \text{ vessels/mm}^2$) and CL ($40.9 \pm 32.8 \text{ vessels/mm}^2$), thereby EndoTAG-1 and CL significantly differed in their MVD to the other two groups ($p = 0.032$, Mann-Whitney U-test) (Fig. 4).

The formation of metastases in the tumour model was confirmed in this study. With exception of one case of a solid metastasis in perirenal fatty tissue (Gl group), metastatic cancer cells were exclusively found within the lung of the animals. All animals developed lung metastases except for

one animal in the Pax group. The distribution of solid metastases and diffuse tumour cells was homogeneous within and between the four treatment arms.

Discussion

Therapeutic options for advanced prostate cancer after androgen deprivation therapy are still limited. Taxane based chemotherapy has demonstrated to extend survival especially in combination with novel agents like anti-angiogenic drugs (19).

A recent strategy in the treatment of neoplasms is to attack the tumour microvasculature as suggested by Denekamp *et al* (5). The approach of specific inhibition of the tumour supplying vessels is supported by the genetically stability of endothelial cells in comparison to neoplastic cells. Without blood vessels, tumours are not able to grow beyond a critical size or metastasize to other organs. Furthermore, the destruction of >100 tumour cells by deletion of one endothelial cell is an efficient way of targeted anticancer therapy (20).

The inhibition of endothelial cell proliferation can be managed with the taxane paclitaxel, a chemotherapeutic drug with an antiangiogenic activity (8). The treatment with paclitaxel has already been evaluated to prolong mean



time in advanced PCa (21,22). However, the therapy by its low therapeutic index and the serious side-effects of the drug itself and the conventional solvent Cremophor EL.

Cationic liposomes were first utilised in gene therapy for application of DNA *in vivo*, later their accumulation in activated endothelial cells was observed (23). Thurston *et al* have shown that angiogenic endothelial cells exhibit a preferential uptake of cationic liposomes by endocytosis (13). The negatively charged glycocalyx of proliferating endothelial cells is suggested to promote the selective uptake of cationic particles. Further investigations have revealed a selective accumulation and prolonged retention in tumour tissue and vasculature for positive charged liposomes whereas neutral and anionic liposomes did not show these attributes (24).

In this study, we evaluated the effect of paclitaxel encapsulated in cationic liposomes in hormone refractory and disseminated PCa in comparison to the commercial available formulation form of paclitaxel. During treatment, the animals in the chemotherapeutic groups (Pax and EndoTAG-1) suffered from reduced general condition which was reflected in a loss of body weight. Because of the normal nutritional condition in the control groups and the predescribed mean survival time of 35 ± 1 days after tumour implantation (17), the partly serious reduction of the general condition towards the end of treatment is supposed to be substantiated by chemotherapy without an emphasis on the form of drug delivery. Toxicity studies with intravenous application of comparable dosages of paclitaxel on rats revealed a loss of body weight and a reduced food intake (25). Furthermore, the described loss of infection defence, myelosuppression and hypoplasia of lymphatic organs agrees with the histopathological findings in this study.

Our experiment showed the strongest inhibition of tumour growth in the EndoTAG-1 treated animals compared with the standard paclitaxel treatment and the control groups. Surprisingly, together with GI group, Pax treated animals displayed the highest tumour volume of all groups despite the fact that paclitaxel was applied in equal amounts in both chemotherapeutic arms. A previous study that investigated the efficacy of EndoTAG-1 and soluble paclitaxel under a similar treatment protocol in a Syrian Golden hamster melanoma model revealed controversial results (26). In our study, we did not evaluate the effect of the formulations of paclitaxel in MatLu-cells *in vitro*, however, an equal antiproliferative effect was proven for a melanoma mouse model *in vitro* (27).

The effect of EndoTAG-1 on metastases was missing in contrast to the already described deceleration of incidence of metastases. Filiae of MatLu-tumours are valid to response to chemotherapy, but only a quantitative reduction was described (28). So, the cause for the lack of differential results between the therapy arms could exist in the exclusive qualitative evaluation of the metastatic status.

The previously observed antitumoural efficacy of empty cationic liposomes was confirmed by our findings (29). On day 20, the tumour volume in the CL group was even smaller than in the Pax group. But the significant differences in MVD for EndoTAG-1 and CL groups do not prove an

antiangiogenic mechanism of action responsible for the therapeutic effect of empty cationic liposomes. In our study due to the observed strong variation of MVD within the control groups a statistically significant antivascular action of EndoTAG-1 superior to the paclitaxel or glucose control could not be demonstrated. Nevertheless EndoTAG-1 showed the strongest antiangiogenic activity which reached significance against the CL group. Previous studies with a comparable experimental setup in other tumour entities were able to show a statistically significant EndoTAG-1 related inhibition of angiogenesis compared to other treatment arms (26,27).

In conclusion, our data suggest that paclitaxel encapsulated in cationic liposomes is able to reduce tumour growth significantly in comparison to the standard application formula. The reduction of side-effects and a suppression of metastasis by the treatment with EndoTAG-1 were not confirmed in this study. The microvessel density was the least in EndoTAG-1 group with a weak significance compared to conventional Paclitaxel suggesting an antiangiogenic action of EndoTAG-1 in this rat tumour model.

Our study demonstrates that cationic liposomes can increase the therapeutic index of paclitaxel by reducing the primary tumour mass in an animal model. The therapeutic effect may be accomplished by reduced angiogenesis.

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