

Comparative effects of EGCG, green tea and a nutrient mixture on the patterns of MMP-2 and MMP-9 expression in cancer cell lines

M.W. ROOMI, J.C. MONTERREY, T. KALINOVSKY, M. RATH and A. NIEDZWIECKI

Dr Rath Research Institute, 1260 Memorex Drive, Santa Clara, CA 95050, USA

Received May 18, 2010; Accepted June 21, 2010

DOI: 10.3892/or_00000917

Abstract. Type IV collagenase matrix metalloproteinases (MMPs), especially MMP-2 and MMP-9, have been found to promote invasion and metastasis of malignant tumors. Extracellular matrix (ECM) degradation by MMPs and increased expression of MMPs in cancer cells and tumor microvascular endothelial cells make MMPs an attractive target for cancer. Focused on a common pathomechanism of cancer growth and invasion, the disintegration of connective tissue, we used natural approaches to increase the integrity and strength of connective tissues. Utilizing the principle of nutrition synergy, we developed a novel micronutrient mixture (NM) containing lysine, proline, ascorbic acid and green tea extract. This study evaluates the potency of the components EGCG and green tea extract independently compared to that of NM on modulation of patterns of MMP-2 and MMP-9 expression in four cancer cell lines expressing MMP-2, MMP-9 or both. Human fibrosarcoma (HT-1080), hepatocellular carcinoma (SK-Hep-1), glioblastoma (T-98G), uterine leiomyosarcoma (SK-UT-1) cell lines were obtained from ATCC and grown in minimum essential medium (MEM) supplemented with 10% FBS, penicillin (100 U/ml) and streptomycin (100 mg/ml) in 24-well tissue culture plates. At near confluence, the cells were treated with agents dissolved in media and tested at concentrations indicated in triplicate at each dose. Cells were also treated with PMA 100 ng/ml to study enhanced expression of MMP-9. MMP expression was assessed by gelatinase zymography. Fibrosarcoma and hepatocellular carcinoma cells expressed both MMP-2 and MMP-9. Glioblastoma cells expressed MMP-2 and PMA treatment induced MMP-9 expression. Uterine leiomyosarcoma cells expressed no MMPs but PMA induced MMP-9. NM was the most potent dose-dependent inhibitor of MMPs, followed by green tea extract and EGCG. In conclusion, these results suggest the enhanced efficacy of nutrients working in synergy to modulate complex pathways such as MMP expression.

Introduction

Metastasis is responsible for a majority of cancer-related deaths. Tumor invasion of surrounding tissue and subsequent metastasis results from a multi-step process that includes proteolytic degradation of the surrounding extracellular matrix (ECM), allowing malignant cells to move into and through the ECM and basement membrane (1-5). Type IV collagenase matrix metalloproteinases (MMPs), in particular, MMP-2 and MMP-9, gelatinase A and B, respectively, have been found to promote invasion and metastasis of malignant tumors (4,5). Elevated levels of MMP-2 and MMP-9 are often correlated with malignancies of liver (6,7), brain (8), breast (9), prostate (10), cervical (11), ovarian (12) and other cancers. Degradation of the ECM by MMPs and the increased expression of MMPs associated with tumor growth, angiogenesis and metastasis point to MMPs as an attractive target for cancer treatment.

We focused on a common pathomechanism of cancer growth and invasion, disintegration of connective tissue, and have developed strategies to inhibit cancer development by inhibiting MMPs, especially MMP-2 and MMP-9 using natural products. Focusing on natural approaches to increase the integrity and strength of connective tissues and utilizing the principle of synergy, we combined lysine, proline, ascorbic acid and green tea extract among other micronutrients to develop a novel micronutrient mixture (NM).

The micronutrient mixture has been shown to inhibit MMP expression in a number of cancer cell lines (13,14). It also exhibited a broad spectrum of anti-tumor activity against a number of cancer cell lines, including inhibition of cell proliferation, cell invasion, anti-metastatic potential, inhibition of angiogenesis and pro-apoptotic effects against a number of human cancer cell lines *in vitro* and *in vivo* (15-17).

The NM was formulated by selecting nutrients that act on critical physiological targets in cancer progression and metastasis. Adequate supplies of ascorbic acid and the amino acids lysine and proline are essential for optimal ECM formation and structure, as these nutrients ensure proper synthesis and hydroxylation of collagen fibers. Lysine also contributes to ECM stability as a natural inhibitor of plasmin-induced proteolysis (18,19). Manganese and copper are also essential for collagen formation. Green tea extract has been shown to modulate cancer cell growth, metastasis, angiogenesis and other aspects of cancer progression (20-26). N-acetyl cysteine

Correspondence to: Dr Aleksandra Niedzwiecki, Dr Rath Research Institute, 1260 Memorex Drive, Santa Clara, CA 95050, USA
E-mail: author@drath.com

Key words: EGCG, green tea, nutrient mixture, MMP-2, MMP-9

Table I. NM equivalents in test solutions.

Treatment/Stock solution no.	0	1	2	3	4	5
Nutrient mixture (μg)	0	50	100	250	500	1000
EGCG equivalent (μg)	0.0000	3.8308	7.6617	19.154	38.309	76.617
GTE equivalent (μg)	0.0000	8.7562	17.512	43.781	87.562	175.12

and selenium have been observed to inhibit MMP-9 and invasive activities of tumor cells, as well as migration of endothelial cells through ECM (27-31). Ascorbic acid has been demonstrated to inhibit tumor growth via anti-angiogenic activity (32) and to inhibit cell division and growth *in vitro* through production of hydrogen peroxide (33). Arginine is a precursor of nitric oxide (NO); any deficiency of arginine can limit the production of NO, which has been shown to predominantly act as an inducer of apoptosis, as in breast cancer cells (34).

In this study we examined the relative inhibitory effects on MMP-2 and MMP-9 of epigallocatechin gallate (EGCG), the main polyphenol in green tea extract (GTE) to that of green tea extract and the nutrient mixture as a whole on several cancer cell lines to elucidate whether there is an additive or synergistic effect from combination of these agents.

Materials and methods

Cancer cell line and culture. Human fibrosarcoma (HT-1080), hepatocellular carcinoma (SK-Hep-1), glioblastoma (T-98G) and uterine leiomyosarcoma (SK-UT-1) cell lines and their recommended media, minimum essential medium (MEM), were obtained from ATCC (American Type Culture Collection, Rockville, MD). Cells were maintained in MEM supplemented with 10% fetal bovine serum, 100 U/ml penicillin and 100 $\mu\text{g}/\text{ml}$ streptomycin in 24-well tissue culture plates (Costar, Cambridge, MA). Cells were incubated with 1 ml of media at 37°C in a tissue culture incubator equilibrated with 95% air and 5% CO₂. At near confluence, the cells were treated with the nutrient mixture, green tea extract, epigallocatechin gallate or other agents dissolved in media and tested at the concentrations indicated in triplicate at each dose. The media and sera used were obtained from ATCC, and antibiotics (penicillin and streptomycin) were from Gibco BRL, Long Island, NY. EGCG and other chemicals were obtained from Sigma.

Composition of the nutrient mixture. The nutrient mixture was composed of the following in the ratio indicated: Vitamin C (as ascorbic acid and as Mg, Ca and palmitate ascorbate) 700 mg, L-lysine 1000 mg, L-proline 750 mg, L-arginine 500 mg, N-acetyl cysteine 200 mg, standardized green tea extract [derived from green tea leaves, was obtained from US Pharma Lab; the certificate of analysis indicated the following characteristics: total polyphenol 80%, catechins 60%, epigallocatechin gallate (EGCG) 35% and caffeine 1.0%] 1000 mg, selenium 30 μg , copper 2 mg, manganese 1 mg. Stock solutions of GTE and EGCG were prepared

as nutrient mixture equivalents in the composition of NM. The respective amounts for each equivalent of NM are indicated in Table I.

In addition, the following components of NM were tested at one dose only - the equivalent of their concentration in 1000 $\mu\text{g}/\text{ml}$ NM. A combination of 400 μM -lysine, 120 μM -proline, 100 μM -AA (LPA); N-acetyl cysteine (NAC) at 43.8 μg ; and L-selenomethionine at 1.3 μg .

Gelatinase zymography. MMP expression in conditioned media was determined by gelatinase zymography. Gelatinase zymography was performed in 10% Novex Pre-Cast SDS Polyacrylamide Gel (Invitrogen Corp.) in the presence of 0.1% gelatin under non-reducing conditions. Culture media (20 μl) were mixed with sample buffer and loaded for SDS-PAGE with tris glycine SDS buffer as suggested by the manufacturer (Novex). Samples were not boiled before electrophoresis. Following electrophoresis the gels were washed twice in 2.5% Triton X-100 for 30 min at room temperature to remove SDS. The gels were then incubated at 37°C overnight in substrate buffer containing 50 mM Tris-HCl and 10 mM CaCl₂ at pH 8.0 and stained with 0.5% Coomassie Brilliant Blue R250 in 50% methanol and 10% glacial acetic acid for 30 min and destained. Upon renaturation of the enzyme, the gelatinases digest the gelatin in the gel and give clear bands against an intensely stained background. Protein standards were run concurrently and approximate molecular weights were determined by plotting the relative mobilities of known proteins.

Gelatinase zymograms were scanned using CanoScan 9950F Canon Scanner at 300 dpi. The intensity of the bands was evaluated using the pixel-based densitometer program Un-Scan-It, Version 5.1, 32-bit, by Silk Scientific Corp. (Orem, UT, USA), at a resolution of 1 Scanner Unit (1/100 of an inch for an image that was scanned at 100 dpi). The pixel densitometer calculates the optical density of each pixel (values 0-255) using the darkly stained background of the gel as a pixel value of 0. A logarithmic optical density scale was used since the optical density of films and gels is logarithmically proportional to the concentration. The pixel densitometer sums the optical density of each pixel to give a band's density. In all graphs, band densities were reported as percentages of the sums of all pixels in a given lane (treatment) of a gel.

The effect of EGCG, GTE and NM on MMP expression was assessed by treating cells to increasing concentrations of EGCG, GTE and NM and expressed as relative band densities (as % of control value). The cumulative effect of EGCG, GTE or NM, was determined by summing all the band densities

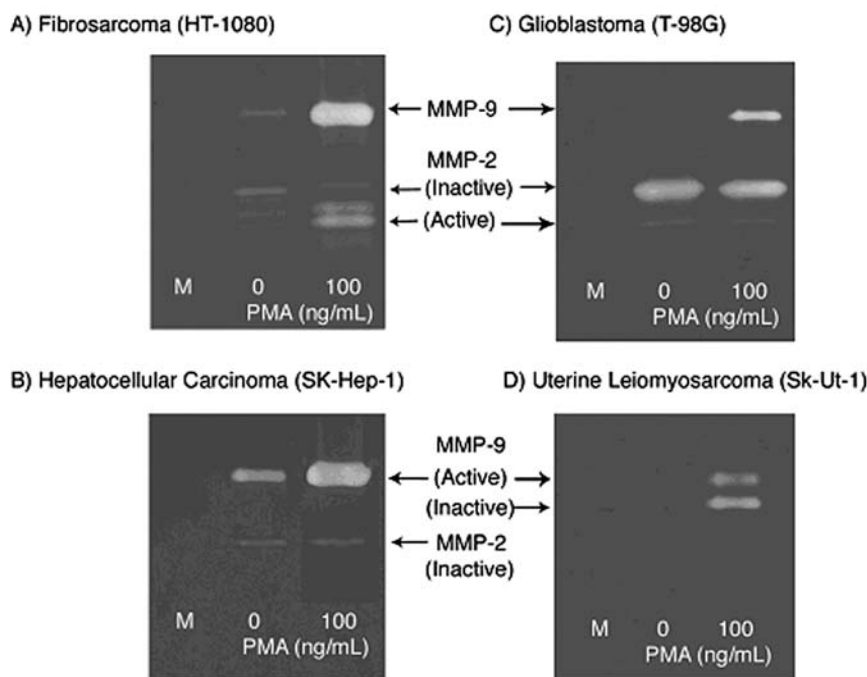


Figure 1. Zymograms of MMP-2 and MMP-9 expression by fibrosarcoma HT-1080 (A), hepatocellular carcinoma SK-Hep-1 (B), glioblastoma T-98G (C) and uterine leiomyosarcoma SK-UT-1 (D).

within a treatment group, and comparing these densities between treatment groups. The extent of total MMP band expression in a treatment group was reported as 'total band pixels'.

Results and Discussion

MMP-2 and MMP-9 expression in normal and PMA-treated fibrosarcoma, hepatocellular carcinoma, glioblastoma and uterine leiomyosarcoma cell lines. Hepatocellular carcinoma SK-Hep-1 exhibited a major band at 92 kDa for MMP-9 and a faint band at 72 kDa for MMP-2, whereas human glioblastoma cell line T-98G had only one band, with molecular weight corresponding to MMP-2. Uterine leiomyosarcoma SK-UT-1 did not express MMP-2 or MMP-9 in untreated conditioned media. PMA treatment of the cell lines induced MMP-9. Fibrosarcoma HT-1080 showed MMP-9 (92 kDa) as a major band and MMP-2 (72 kDa) as a minor band with PMA treatment. With PMA treatment, MMP-9 expression was increased in SK-Hep-1, and in T-98G, PMA led to the expression of both MMP-2 and MMP-9 in equal intensities. Interestingly, SK-UT-1, on PMA treatment, exhibited both active and inactive MMP-9 in equal proportions. The MMP profiles of these cancer cell lines are depicted in Fig. 1.

Effect of lysine, proline, ascorbic acid, NAC and selenium on MMP expression. Initially we investigated the effect of a combination of NM components 400 μ M-lysine + 120 μ M-proline + 100 μ M-AA (LPA), the equivalent concentration of these agents in NM 1000 μ g/ml, on MMP expression in conditioned and PMA-stimulated media. LPA had no effect on expression of either MMP-2 or MMP-9 in all four cell lines, regardless of PMA treatment at the dose tested. We

also studied the effect of NM components N-acetyl cysteine (NAC) and selenium on MMP expression in both uninduced and induced media. As shown in Figs. 2 and 3, neither NAC nor selenium had any effect on MMP-2 and MMP-9 expression at the dose tested. Figs. 2 and 3 present the zymograms and densitometric analyses for results from treatment of SK-Hep-1 and human glioblastoma cell lines with LPA, NAC and Se.

Effect of NM, GTE and EGCG on MMP-2 and MMP-9 secretion. Our next step was to explore the comparative effects of NM to that of green tea extract (GTE), an important constituent of the nutrition mixture and EGCG, because EGCG is the main constituent of green tea. The green tea extract and EGCG solutions were prepared to match the equivalent amount found in 50, 100, 250, 500 and 1000 μ g/ml concentrations of nutrition mixture.

Effect of NM, GTE and EGCG on MMP-2 and -9 secretion by fibrosarcoma cells. Fibrosarcoma (HT-1080) cells expressed both MMP-2 and MMP-9, and the nutrient mixture was the most potent dose-dependent inhibitor of both MMPs, followed by green tea extract and then EGCG, as shown in Fig. 4 and Table II. Although EGCG and green tea extract greatly reduced MMP-2 expression at 100 μ g/ml equivalent of NM, they did not completely block MMP-2 expression. However, the nutrient mixture at 100 μ g/ml concentration completely abolished MMP-2 expression. MMP-9 expression decreased in a dose-dependent fashion with EGCG, GTE and NM treatment, but only GTE and NM were able to completely abolish MMP-9 expression at 500 μ g/ml. Treating the fibrosarcoma cell line with PMA induced further MMP-9 expression, but the inhibitory effects of EGCG, GTE and the NM on the expression of MMP-2 and MMP-9 was generally the

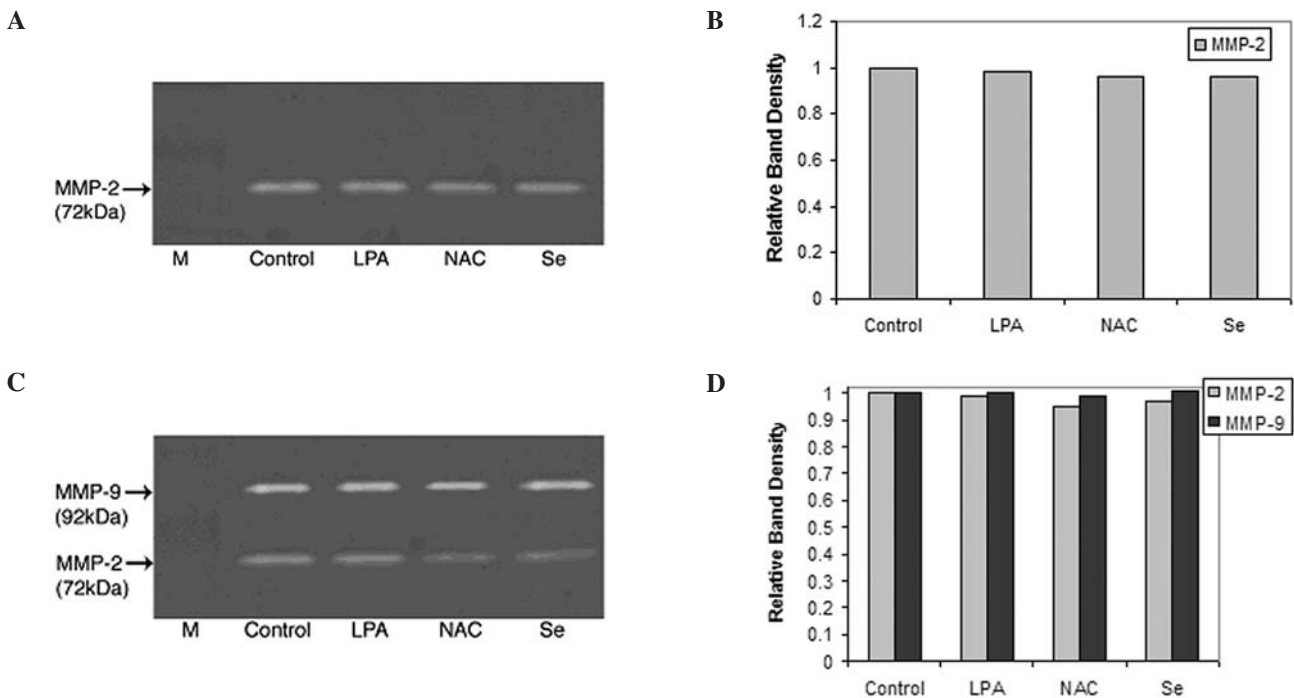


Figure 2. Effect of LPA, NAC and Se on MMP-2 and MMP-9 expression in PMA-treated and non-treated human glioblastoma cells. (A) Gelatinase zymogram of T-98G cells showing no effect of any of the tested agents on MMP-2 expression. (B) Quantitative densitometry showing no effect of tested agents on T-98G cell MMP-2 expression. (C) Gelatinase zymogram of PMA (100 ng/ml)-treated T-98G cells showing MMP-9 induction with PMA treatment and no effect of any of the agents tested on MMP-2 or MMP-9 expression. (D) Quantitative densitometry showing no effect of tested agents on PMA-treated T-98G cell MMP-2 or MMP-9 expression.

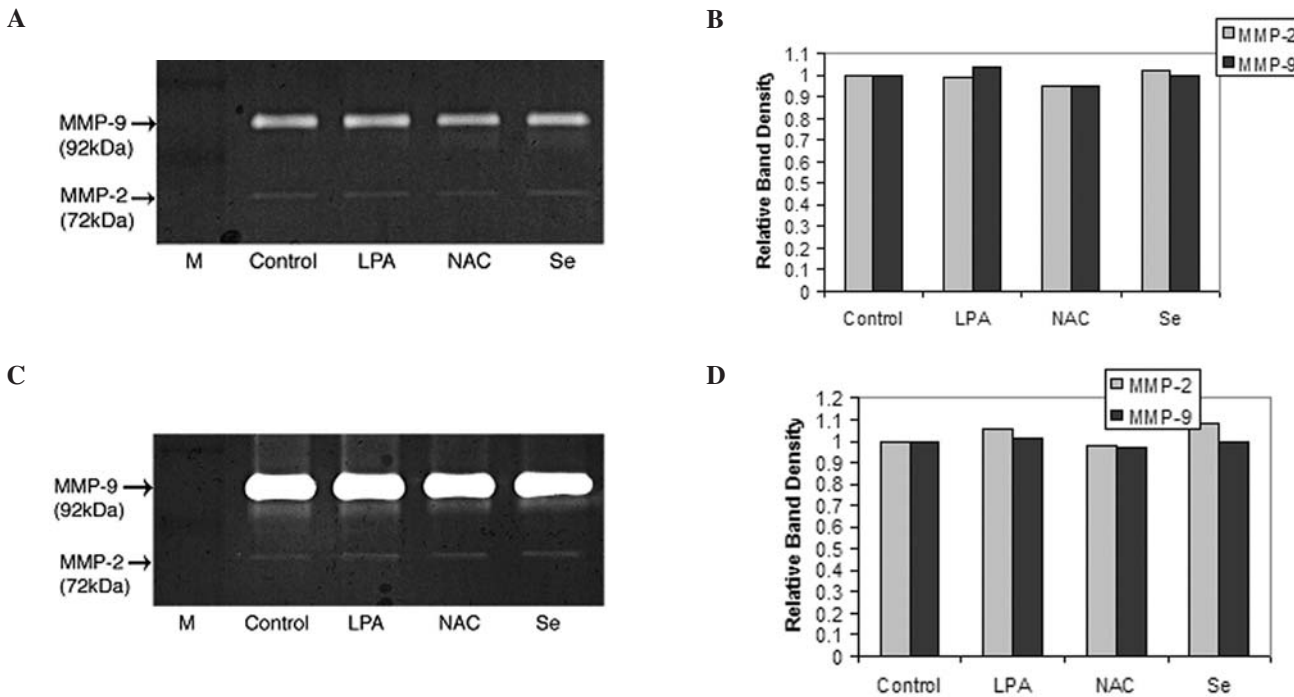


Figure 3. Effect of LPA, NAC and Se on MMP-2 and MMP-9 expression in PMA-treated and non-treated human hepatocellular carcinoma cells. (A) Gelatinase zymogram of SK-Hep-1 cells showing no effect of any of the tested agents on MMP-2 or MMP-9 expression. (B) Quantitative densitometry showing no effect of tested agents on SK-Hep-1 cell MMP-2 and MMP-9 expression. (C) Gelatinase zymogram of PMA (100 ng/ml)-treated SK-Hep-1 cells showing increased MMP-9 expression with PMA treatment and no effect of any of the agents tested on MMP-2 or MMP-9 expression. (D) Quantitative densitometry showing no effect of tested agents on PMA-treated SK-Hep-1 cell MMP-2 or MMP-9 expression.

same, as shown in Fig. 5 and Table II. The nutrient mixture was the most potent dose-dependent inhibitor of MMP expression, followed by green tea extract and then EGCG.

MMP-2 appeared to be more sensitive to MMP inhibition by inhibitor treatment than MMP-9 in PMA treated cells (MMP-2 expression was abolished by 50 μ g/ml with the nutrient

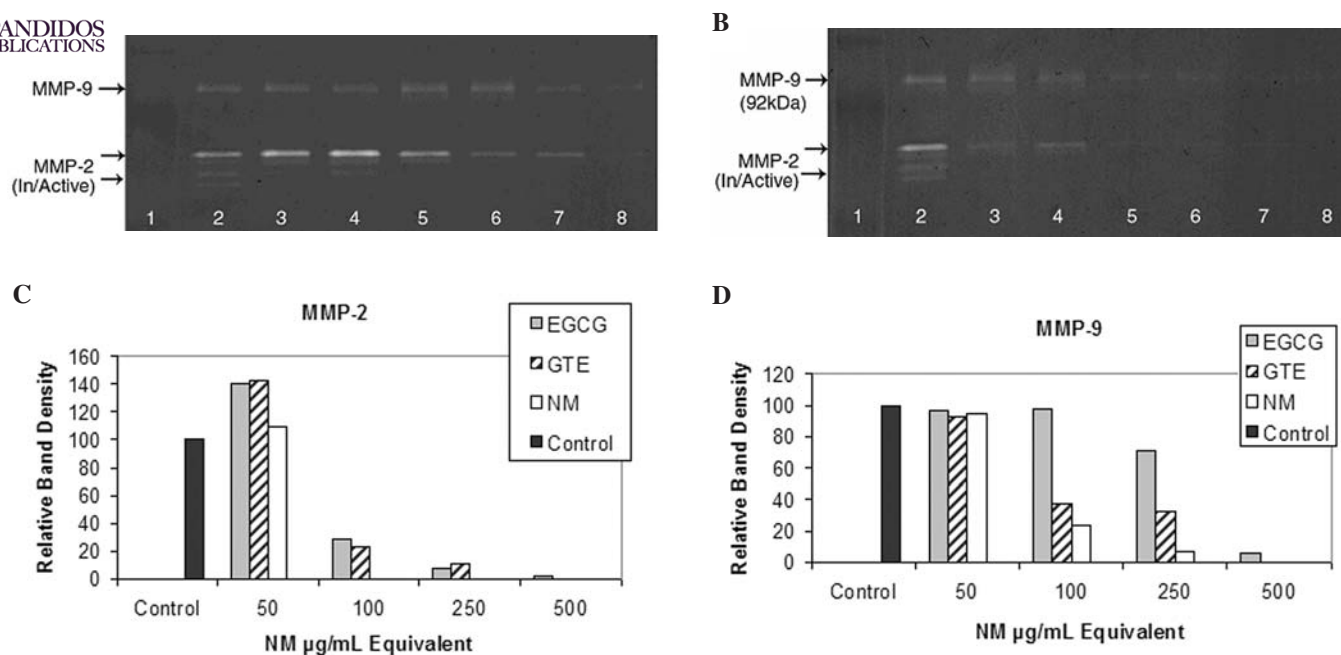
SPANDIDOS
PUBLICATIONS

Figure 4. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of normal human fibrosarcoma HT-1080 cells. (A, B) Gelatinase zymogram of HT-1080 cells showing dose-dependent inhibition of MMP-2 and MMP-9 expression; NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 µg/ml NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 µg/ml NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 µg/ml NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 µg/ml NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on HT-1080 cell (C) MMP-2 and (D) MMP-9 expression.

Table II. Effect of EGCG, GTE and NM on MMP-2 and -9 secretion by fibrosarcoma (HT-1080).

	Without PMA		With PMA (100 ng/ml)	
	MMP-2 (%)	MMP-9 (%)	MMP-2 (%)	MMP-9 (%)
EGCG (µg/ml)				
0	100	100	100	100
50	140	96.5	71	120
100	29	98	0	120
250	8	70.6	0	74.7
500	2	6.2	0	19.8
GTE (µg/ml)				
0	100	100	100	100
50	143	93	70	104
100	23	37	0	58.6
250	11	33	0	53.6
500	0	6.2	0	6.24
Nutrient mixture (µg/ml)				
0	100	100	100	100
50	109	95	0	96.7
100	0	23.7	0	26.2
250	0	7.4	0	19.5
500	0	0	0	0

Table III. Effect of EGCG, GTE and NM on MMP-2 and -9 secretion by hepatocellular carcinoma (SK-Hep-1).

	Without PMA		With PMA (100 ng/ml)	
	MMP-2 (%)	MMP-9 (%)	MMP-2 (%)	MMP-9 (%)
EGCG (µg/ml)				
0	100	100	100	100
50	197	114	45	69.7
100	0	25.1	12.8	59
250	0	0.6	0	65.8
500	0	1.4	0	56
GTE (µg/ml)				
0	100	100	100	100
50	179	99.6	41	79.6
100	0	1.9	0	55.2
250	0	0.5	0	58.1
500	0	1.0	0	46
Nutrient mixture (µg/ml)				
0	100	100	100	100
50	0	11.7	21.5	62.1
100	0	1.9	0	50
250	0	0.9	0	32.2
500	0	0	0	0

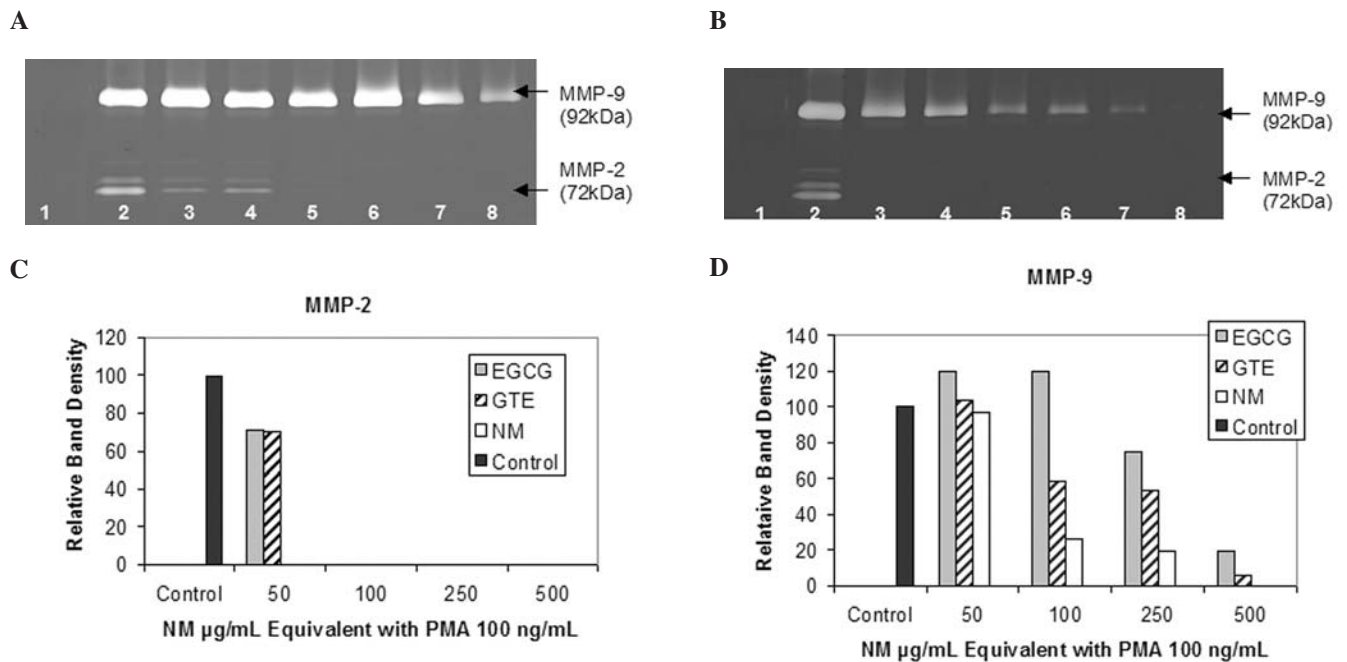


Figure 5. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of PMA (100 ng/ml)-treated human fibrosarcoma HT-1080 cells. (A, B) Gelatinase zymogram of HT-1080 cells showing increased MMP-9 expression with PMA treatment and dose-dependent inhibition of MMP-2 and MMP-9 expression with NM, GTE and EGCG. NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on PMA-treated HT-1080 cell (C) MMP-2 and (D) MMP-9 expression.

Table IV. Effect of EGCG, GTE and NM on MMP-2 and -9 secretion by glioblastoma T-98G.

	Without PMA		With PMA (100 ng/ml)	
	MMP-2 (%)	MMP-9 (%)	MMP-2 (%)	MMP-9 (%)
EGCG ($\mu\text{g/mL}$)				
0	100	100	100	100
50	140	96.5	71	120
100	29	98	0	120
250	8	70.6	0	74.7
500	2	6.2	0	19.8
GTE ($\mu\text{g/mL}$)				
0	100	100	100	100
50	143	93	70	104
100	23	37	0	58.6
250	11	33	0	53.6
500	0	6.2	0	6.24
Nutrient mixture ($\mu\text{g/mL}$)				
0	100	100	100	100
50	109	95	0	96.7
100	0	23.7	0	26.2
250	0	7.4	0	19.5
500	0	0	0	0

Table V. Effect of EGCG, GTE and NM on MMP-2 and -9 secretion by uterine leiomyosarcoma (SK-UT-1).

	With PMA	
	MMP-9 (Inactive)	MMP-9 (Active)
EGCG ($\mu\text{g/mL}$)		
0	100	100
50	207	138
100	82.2	79.2
250	28.8	29.5
500	3	8.3
GTE ($\mu\text{g/mL}$)		
0	100	100
50	192	148
100	79.4	62.5
250	39.5	46.6
500	0.0	1.0
Nutrient mixture ($\mu\text{g/mL}$)		
0	100	100
50	130	133
100	57.7	64.8
250	25.6	29.0
500	0.0	0.0

mixture and 100 $\mu\text{g/mL}$ with EGCG and GTE, while MMP-9 expression was only abolished by the NM, at 500 $\mu\text{g/mL}$).

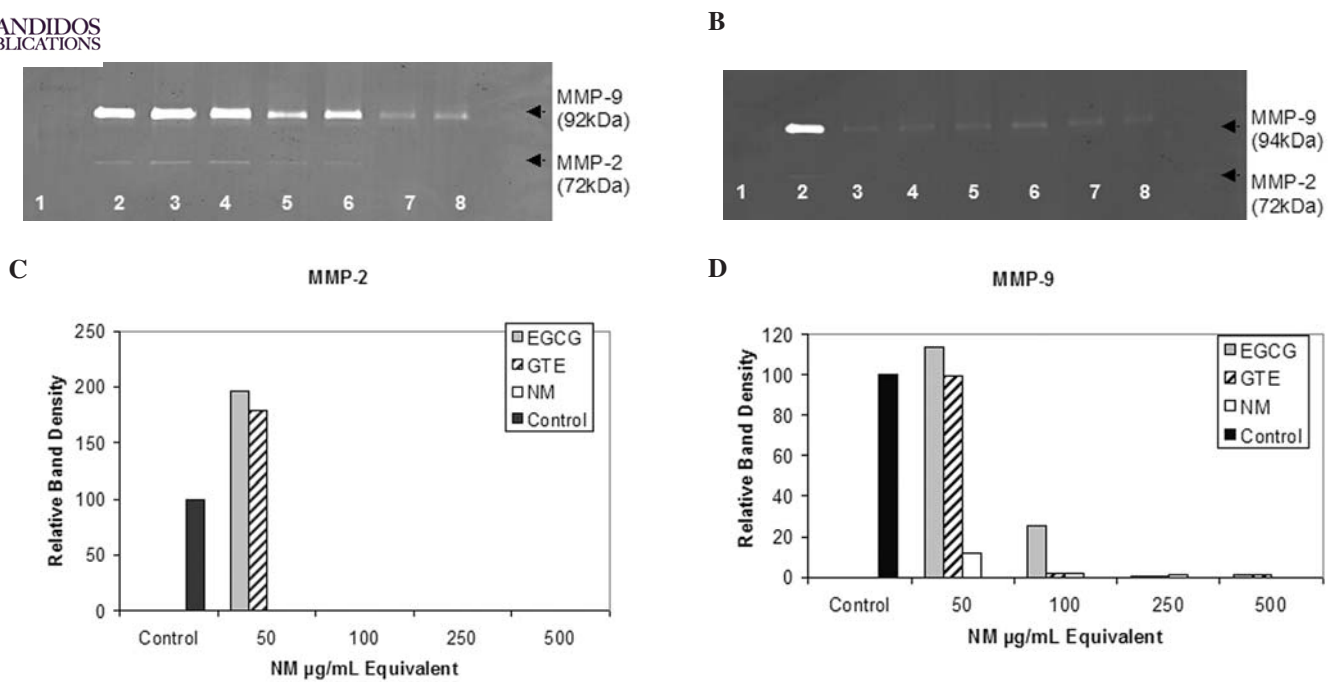


Figure 6. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of normal human hepatocellular carcinoma SK-Hep-1 cells. (A, B) Gelatinase zymogram of normal SK-Hep-1 cells showing dose-dependent inhibition of MMP-2 and MMP-9 expression. NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on SK-Hep-1 cell (C) MMP-2 and (D) MMP-9 expression.

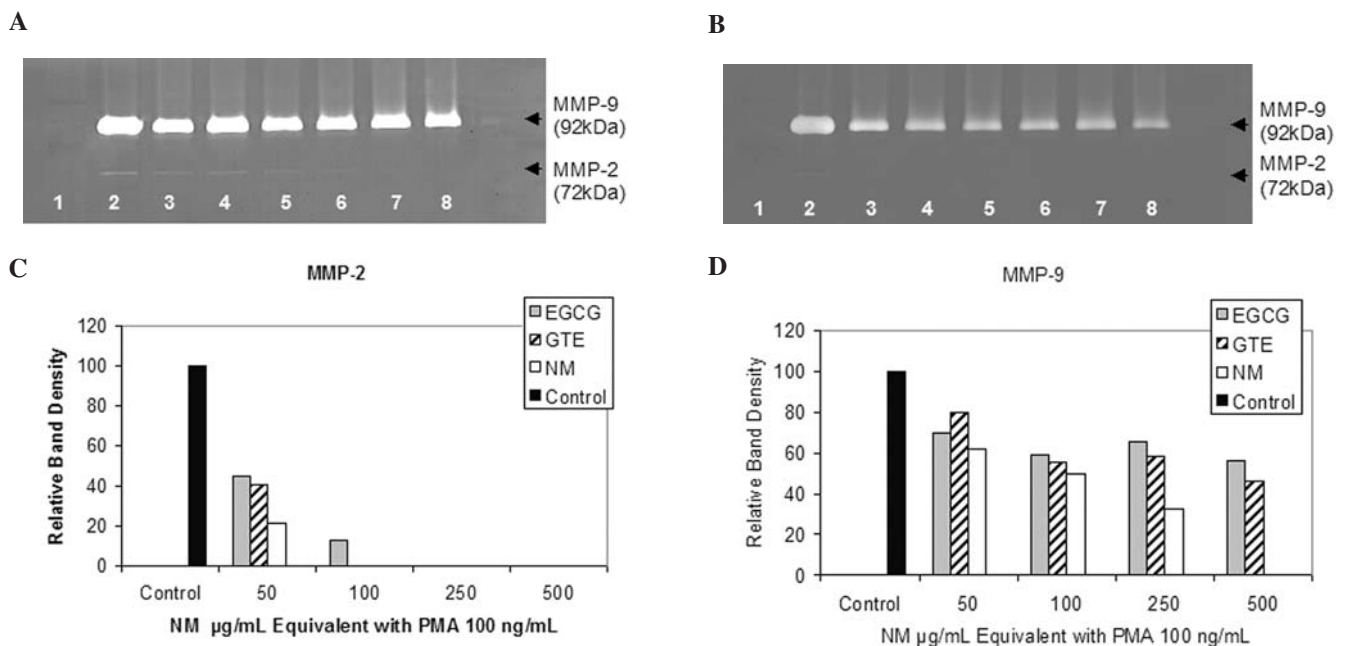


Figure 7. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of PMA (100 ng/ml)-treated human hepatocellular carcinoma SK-Hep-1 cells. (A, B) Gelatinase zymogram of SK-Hep-1 cells showing increased MMP-9 expression with PMA treatment and dose-dependent inhibition of MMP-2 and MMP-9 expression. NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on PMA-treated SK-Hep-1 cell (C) MMP-2 and (D) MMP-9 expression.

This is understandable because PMA greatly increased MMP-9 expression in fibrosarcoma cells, but not MMP-2.

The cumulative effect of EGCG, GTE, and NM on MMP expression was assessed by treating cells to increasing con-

centrations of EGCG, GTE and NM, summing all the band densities within a treatment group, and comparing these densities between treatments. As shown in Table VI, cumulative MMP-2 expression of uninduced fibrosarcoma cells treated

Table VI. Comparative cumulative expression (total band pixels in thousands) of MMP-2 and -9 in cell lines treated with EGCG, GTE and NM.

	EGCG	GTE	NM	EGCG with PMA	GTE with PMA	NM with PMA
HT-1080						
MMP-2	7.88	7.47	3.29	0.21	0.20	0.00
MMP-9	5.74	3.02	1.58	209.06	139.84	93.54
SK-Hep-1						
MMP-2	1.21	1.10	0.00	0.77	0.55	0.29
MMP-9	256.51	187.28	26.59	611.90	593.80	508.28
T-98G						
MMP-2	109.97	86.63	65.84	178.16	140.09	53.20
MMP-9	0.17	0.17	0.10	92.69	82.67	52.50
SK-UT-1						
MMP-9 (inactive)	0.00	0.00	0.00	51.36	49.97	34.30
MMP-9 (active)	0.00	0.00	0.00	87.42	87.30	77.95

with EGCG, GTE and NM resulted in total band pixels (in thousands) of 7.9, 7.5 and 3.3, respectively, showing the significantly more modulating effect of NM. Similarly, cumulative MMP-9 expression of uninduced fibrosarcoma cells with EGCG, GTE and NM resulted in total band pixels (in thousands) of 5.7, 3.0 and 1.6, respectively. Cumulative MMP-2 and MMP-9 expression followed the same pattern with potent inhibition by NM followed by GTE and EGCG.

Effect of NM, GTE and EGCG on MMP-2 and -9 secretion by hepatocellular carcinoma cells. Hepatocellular carcinoma (SK-Hep-1) cells expressed both MMP-2 and MMP-9, but MMP-9 to a much greater extent, as shown in Fig. 6 and Table III. Our results demonstrated the nutrient mixture to be the most potent inhibitor of both MMP-2 and MMP-9, followed by green tea extract and then EGCG. The low expression of MMP-2 rendered the nutrient mixture able to fully inhibit MMP-2 expression by 50 $\mu\text{g/ml}$, whereas green tea extract and EGCG completely blocked MMP-2 expression at 100 $\mu\text{g/ml}$. MMP-9 expression decreased in a dose-dependent fashion but was not completely abolished. However, minimal MMP-9 expression was detected at 100 $\mu\text{g/ml}$ GTE and NM and at 250 $\mu\text{g/ml}$ EGCG treatment. PMA treatment of SK-Hep-1 further induced MMP-9 expression and left the slight expression of MMP-2 by hepatocellular carcinoma cells unchanged. The effect of the inhibitors remained the same, with the nutrient mixture being the most effective, followed by green tea extract, and then EGCG, as shown in Fig. 7 and Table III. MMP-2 expression was again abolished by 100 $\mu\text{g/ml}$ by the nutrient mixture, and by 250 $\mu\text{g/ml}$ by EGCG. MMP-9 expression was decreased in a dose-dependent fashion by all treatments, with the nutrient mixture being the most effective, although MMP-9 expression was not fully abolished (understandably due to the concurrent treatment by PMA).

As shown in Table VI, cumulative MMP-2 expression of uninduced hepatocellular carcinoma cells treated with EGCG,

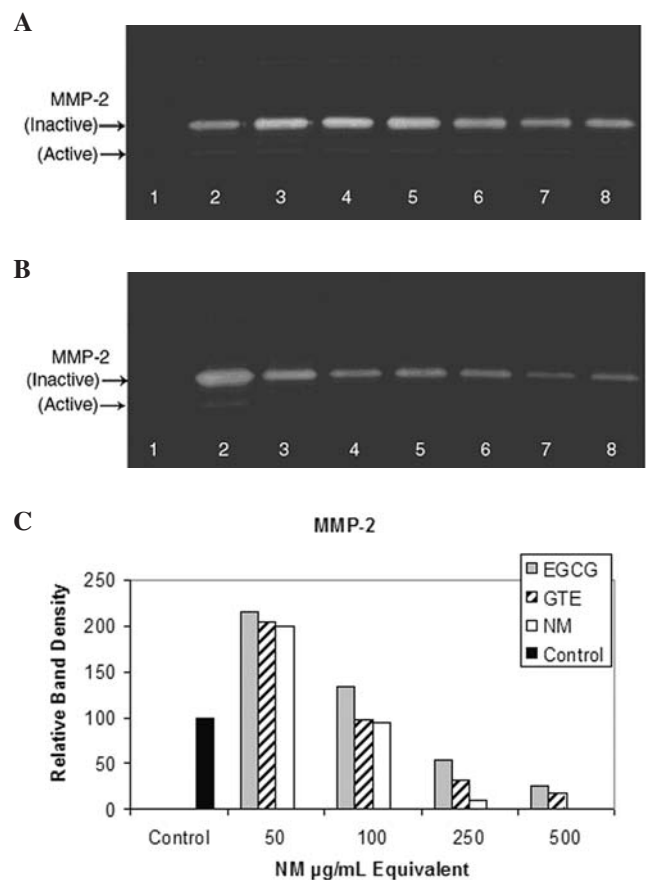


Figure 8. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of normal human glioblastoma T-98G cells. (A, B) Gelatinase zymogram of normal T-98G cells showing secretion of only MMP-2, which was inhibited in a dose-dependent manner by all three test agents, most potently by NM. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/ml}$ NM EQ of EGCG, GTE and NM, respectively. (C) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on normal T-98G cell MMP-2 expression.

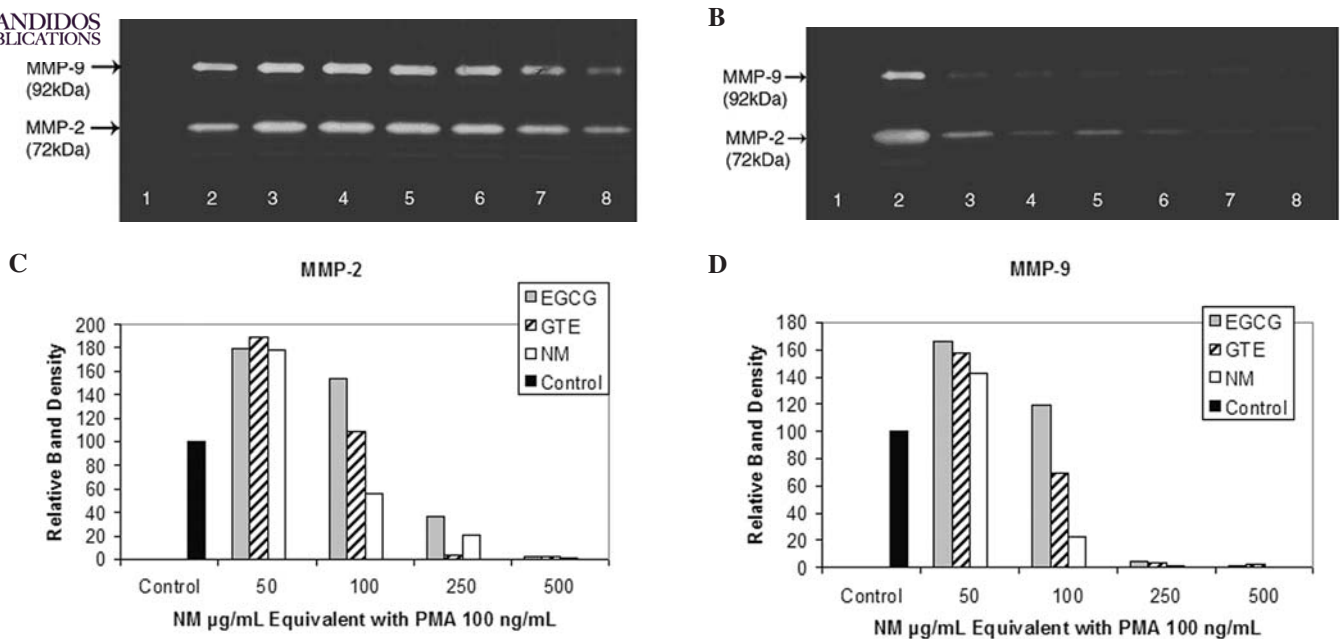
SPANDIDOS
PUBLICATIONS

Figure 9. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of PMA (100 ng/ml)-treated human glioblastoma T-98G cells. (A, B) Gelatinase zymogram of SK-Hep-1 cells showing induced MMP-9 expression with PMA treatment and dose-dependent inhibition of MMP-2 and MMP-9 expression. NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on PMA-treated T-98G cell (C) MMP-2 and (D) MMP-9 expression.

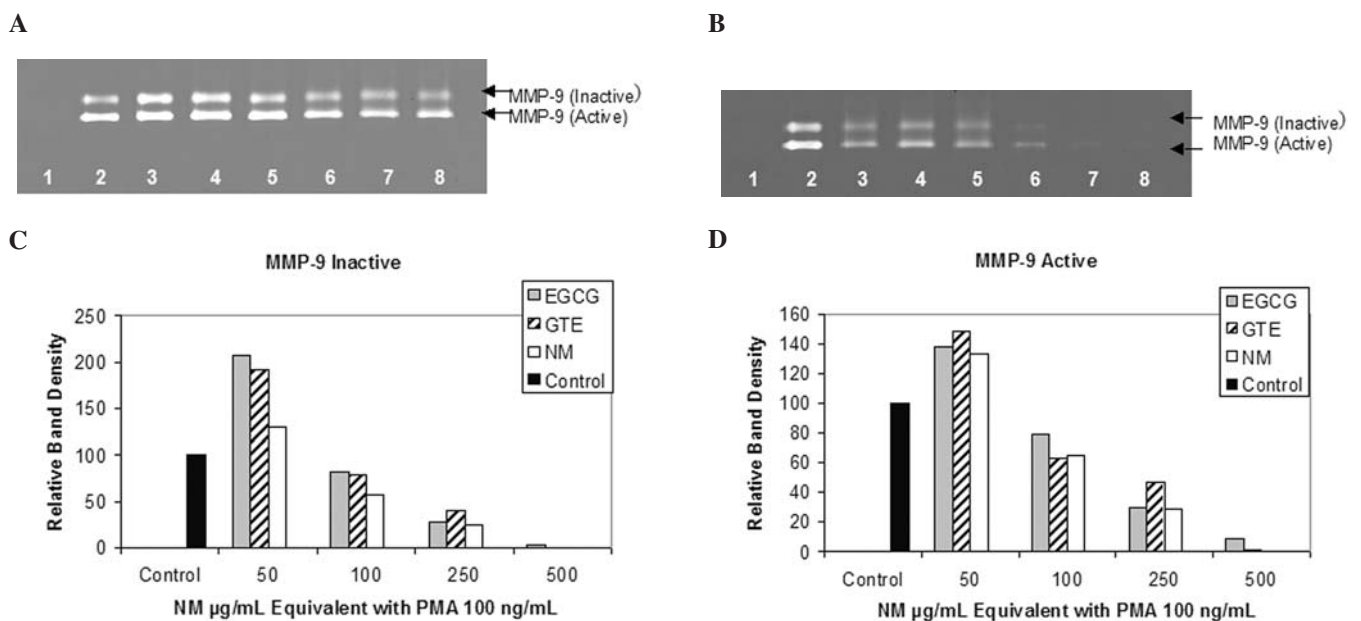


Figure 10. Comparative effects of EGCG, GTE and NM on MMP-2 and MMP-9 expression of PMA (100 ng/ml)-treated human leiomyosarcoma SK-UT-1 cells. (A, B) Gelatinase zymogram of SK-UT-1 cells showing induced secretion of MMP-9 (inactive and active), which was inhibited in a dose-dependent manner by all three test agents; NM was the most potent followed by GTE and EGCG. (A) Legend: 1-Markers, 2-control, 3-5, 50 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 100 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (B) Legend: 1-Markers, 2-control, 3-5, 250 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively; 6-8, 500 $\mu\text{g/mL}$ NM EQ of EGCG, GTE and NM, respectively. (C, D) Quantitative densitometry showing comparative effects of NM, GTE and EGCG on PMA-treated SK-UT-1 cell (C) MMP-9 (inactive) and (D) MMP-9 (active) expression.

GTE and NM resulted in total band pixels (in thousands) of 1.21, 1.10 and 0, respectively, showing the significantly more modulating effect of NM. Similarly, cumulative MMP-9 expression of uninduced fibrosarcoma cells with EGCG, GTE

and NM resulted in total band pixels (in thousands) of 256.5, 187.3 and 26.6, respectively. Cumulative MMP-2 and MMP-9 expression followed the same pattern with potent inhibition by NM followed by GTE and EGCG.

Effect of NM, GTE and EGCG on MMP-2 and -9 secretion by glioblastoma cells. Human glioblastoma (T-98G) cells expressed only MMP-2, which decreased in a dose-dependent fashion with treatment with all three test agents, but with the nutrient mixture being the most effective, as shown in Fig. 8 and Table IV. Treating T-98G with PMA induced MMP-9 expression to an equal extent to which MMP-2 was expressed. Both MMP-2 and MMP-9 expression decreased with increasing concentration of treatment, with the nutrient mixture being most effective, followed by green tea extract and then EGCG, as shown in Fig. 9 and Table IV.

As shown in Table VI, cumulative MMP-2 expression of uninduced glioblastoma cells treated with EGCG, GTE and NM resulted in total band pixels (in thousands) of 110, 86.6 and 65.8, respectively, showing the significantly more modulating effect of NM. Cumulative MMP-2 and MMP-9 expression followed the same pattern with potent inhibition by NM followed by GTE and EGCG.

Effect of NM, GTE and EGCG on MMP-2 and -9 secretion by uterine leiomyosarcoma cells. Uterine leiomyosarcoma (SK-UT-1) cells expressed no MMPs, and only MMP-9 after PMA treatment. PMA-treated SK-UT-1 cells showed a dose-dependent decrease of MMP-9 expression with increasing concentration of inhibitor treatment. The most effective inhibition of MMP-9 expression was seen with the nutrient mixture, abolishing inactive and active MMP-9 expression by 500 µg/ml, as shown in Fig. 10 and Table V.

As shown in Table VI, cumulative inactive MMP-9 expression of PMA-treated leiomyosarcoma cells treated with EGCG, GTE and NM resulted in total band pixels (in thousands) of 51.4, 50.0 and 34.3, respectively, showing the significantly more modulating effect of NM. Cumulative active MMP-9 expression of PMA-treated leiomyosarcoma cells with EGCG, GTE and NM resulted in total band pixels (in thousands) of 87.4, 87.3 and 77.9, respectively.


In conclusion, investigation of the comparative effects of NM, GTE and EGCG on uninduced and PMA-treated cancer cell lines (fibrosarcoma, hepatocellular carcinoma, glioblastoma and leiomyosarcoma) with different patterns of MMP-2 and MMP-9 expression, demonstrated the superior inhibitory action of NM over GTE and EGCG on modulation of MMPs. The superior potency of NM over GTE and EGCG can be understood from the more comprehensive treatment offered by the combination of nutrients in NM over individual components of NM since MMP-2 and MMP-9 are mediated by differential pathways. LPA, NAC and selenium had no effect on MMP-2 and MMP-9 expression by these cancer cell lines. However, green tea and its component EGCG inhibited MMP expression but to a smaller degree than NM, which is composed of a combination of LPA, NAC, selenium and GTE. Thus NM, the mixture of all these components, demonstrated superior inhibitory activity on MMP expression from a synergistic, rather than from an additive effect. This study suggests the importance of nutrients working in cooperation to more effectively treat complex biochemical pathways.

Acknowledgements

This study was funded by the Dr Rath Health Foundation (Plantation, FL), a non-profit organization.

References

1. Fidler IJ: Molecular biology of cancer: invasion and metastasis. In: Cancer: Principles and Practice of Oncology. 5th edition. De Vita VT, Hellman S and Rosenberg SA (eds). Lippincott-Raven, Philadelphia, PA, pp135-152, 1997.
2. Folkman J: Role of angiogenesis in tumor growth and metastasis. *Semin Oncol* 29: 15-18, 2002.
3. Egeblad M and Werb Z: New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2: 161-174, 2002.
4. Chambers AF and Matrisian LM: Changing views on the role of matrix metalloproteinases in metastasis. *J Natl Cancer Inst* 89: 1260-1270, 1997.
5. Kleiner DL and Stetler-Stevenson WG: Matrix metalloproteinases and metastasis. *Cancer Chemother Pharmacol* 43 (Suppl.): S42-S51, 1999.
6. Sakamoto Y, Mafune K, Mori M, Shiraishi T, Imamura H, Mori M, Takayama T and Makuuchi M: Overexpression of MMP-9 correlates with growth of small hepatocellular carcinoma. *Int J Oncol* 17: 237-243, 2000.
7. Kashiwagi A: Matrix metalloproteinases in hepatocellular carcinoma. A study in resected specimens from 25 patients. *Tokyo Jikeikai Med J* 118: 33-48, 2001.
8. Ozen O, Krebs B, Hemmerlein B, Pekrun A, Kretschmar H and Herms J: Expression of matrix metalloproteinases and their inhibitors in medulloblastoma and their prognostic relevance. *Clin Cancer Res* 10: 4746-4753, 2004.
9. Loppä S, Saarto T, Vehmanen L, Blomqvist C and Elomaa I: A high serum matrix metalloproteinase-2 level is associated with an adverse prognosis in node-positive breast carcinoma. *Clin Cancer Res* 10: 1057-1063, 2004.
10. Stearns M and Stearns ME: Evidence for increased activated metalloproteinase-2 (MMP-2a) expression associated with human prostate cancer progression. *Oncol Res* 8: 69-75, 1996.
11. Sato T, Sakai T, Noguchi Y, Takta M, Hirakawa S and Ito A: Tumor-stromal cell contact promotes invasion of human uterine cervical carcinoma cells by augmenting the expression and activation of stromal matrix metalloproteinases. *Gynecol Oncol* 92: 47-56, 2004.
12. Di Nezza LA, Misajon A, Zhang J, Jobling T, Quinn MA, Ostör AG, Nie G, Lopata A and Salamonsen LA: Presence of active gelatinases in endometrial carcinoma and correlation of matrix metalloproteinase expression with increasing tumor grade and invasion. *Cancer* 94: 1466-1475, 2002.
13. Roomi MW, Monterrey JC, Kalinovsky T, Rath M and Niedzwiecki A: *In vitro* modulation of MMP-2 and MMP-9 in human cervical and ovarian cancer cell lines by cytokines, inducers and inhibitors. *Oncol Rep* 23: 605-614, 2010.
14. Roomi MW, Monterrey JC, Kalinovsky T, Rath M and Niedzwiecki A: Modulation of MMP-2 and MMP-9 by cytokines, mitogens and inhibitors in lung cancer and malignant mesothelioma cell lines. *Oncol Rep* 22: 1283-1291, 2009.
15. Roomi MW, Roomi N, Ivanov V, Kalinovsky T, Niedzwiecki A and Rath M: Inhibitory effect of a mixture containing ascorbic acid, lysine, proline and green tea extract on critical parameters in angiogenesis. *Oncol Rep* 14: 807-815, 2005.
16. Roomi MW, Ivanov V, Kalinovsky T, Niedzwiecki A and Rath M: Inhibition of pulmonary metastasis of melanoma B16FO cells in C57BL/6 mice by a nutrient mixture consisting of ascorbic acid, lysine, proline, arginine and green tea extract. *Exp Lung Res* 32: 517-530, 2006.
17. Roomi MW, Ivanov V, Kalinovsky T, Niedzwiecki A and Rath M: In vivo and in vitro antitumor effect of ascorbic acid, lysine, proline, arginine and green tea extract on human fibrosarcoma cells HT-1080. *Med Oncol* 23: 105-112, 2006.
18. Rath M and Pauling L: Plasmin-induced proteolysis and the role of apoprotein(a), lysine and synthetic analogs. *Orthomolecular Med* 7: 17-23, 1992.
19. Sun Z, Chen YH, Wang P, Zhang J, Gurewich V, Zhang P and Liu JN: The blockage of high-affinity lysine binding sites of plasminogen by EACA significantly inhibits prourokinase-induced plasminogen activation. *Biochem Biophys Acta* 1596: 182-192, 2002.
20. Valcic S, Timmermann BN, Alberts DS, Wachter GA, Krutzsch M, Wymer J and Guillen JM: Inhibitory effect of six green tea catechins and caffeine on the growth of four selected human tumor cell lines. *Anticancer Drugs* 7: 461-468, 1996.

 SPANDIDOS PUBLICATIONS: M, Michaud-Levesque J, Annabi B, Gingras D, Jodoin J, Lamy S, Bertrand Y and Béliveau R: Green tea catechins as novel antitumor and antiangiogenic compounds. *Curr Med Chem Anticancer Agents* 2: 441-463, 2002.

22. Hara Y: *Green Tea: Health Benefits and Applications*. Marcel Dekker, Basel, New York, 2001.
23. Mukhtar H and Ahmed N: Tea polyphenols: prevention of cancer and optimizing health. *AM J Clin Nutr* 71: S1698-S1702, 2000.
24. Yang GY, Liao J, Kim K, Yurtow EJ and Yang CS: Inhibition of growth and induction of apoptosis in human cancer cell lines by tea polyphenols. *Carcinogenesis* 19: 611-616, 1998.
25. Taniguchi S, Fujiki H, Kobayashi H, Go H, Miyado K, Sadano H and Shimikawa R: Effect of (-) epigallocatechin gallate, the main constituent of green tea, on lung metastasis with mouse B16 melanoma cell lines. *Cancer Lett* 65: 51-54, 1992.
26. Wang X, Hao MW, Dong K, Lin F, Ren JH and Zhang HZ: Apoptosis induction effects of EGCG in laryngeal squamous cell carcinoma cells through telomerase repression. *Arch Pharm Res* 32: 1263-1269, 2009.
27. Kawakami S, Kageyama Y, Fujii Y, Kihara K and Oshima H: Inhibitory effects of N-acetyl cysteine on invasion and MMP 9 production of T24 human bladder cancer cells. *Anticancer Res* 21: 213-219, 2001.
28. Morini M, Cai T, Aluigi MG, *et al*: The role of the thiol N-acetyl cysteine in the prevention of tumor invasion and angiogenesis. *Int J Biol Markers* 14: 268-271, 1999.
29. Yoon SO, Kim MM and Chung AS: Inhibitory effects of selenite on invasion of HT 1080 tumor cells. *J Biol Chem* 276: 20085-20092, 2001.
30. Rayman MP: Selenium in cancer prevention: a review of the evidence and mechanism of action. *Proc Nutr Soc* 64: 527-542, 2005.
31. Schrauzer GN: Anticarcinogenic effects of selenium. *Cell Mol Life Sci* 57: 1864-1873, 2000.
32. Yeom CH, Lee G, Park JH, Yu J, Park S, Yi SY, Lee HR, Hong YS, Yang J and Lee S: High-dose concentration administration of ascorbic acid inhibits tumor growth in BALB/C mice implanted with sarcoma 180 cancer cells via restriction of angiogenesis. *J Transl Med* 7: 70, 2009.
33. Maramba C, Menon M, Balaji KC, Reddy PG and Laxmanan S: Effect of vitamin C on prostate cancer cells in vitro: effect on cell number, viability and DNA synthesis. *Prostate* 32: 188-195, 1997.
34. Cooke JP and Dzau VJ: Nitric oxide synthase: role in the genesis of vascular disease. *Annu Rev Med* 48: 489-509, 1997.