

Table SI. Sequences of the primers used for reverse transcription-quantitative PCR.

Gene	Primer sequence (5'- 3')
<i>JUNB</i>	Fwd: ATACACAGCTACGGGATACGG Rev: GCTCGGTTTCAGGAGTTTGT
<i>JUN</i>	Fwd: CCAAAGGATAGTGCGATGTTT Rev: CTGTCCCTCTCCACTGCAAC
<i>FOS</i>	Fwd: CTACCACTCACCCGCAGACT Rev: AGGTCCGTGCAGAAGTCCT
<i>JUND</i>	Fwd: CAGCGAGGAGCAGGAGTT Rev: GAGCTGGTTCTGCTTGTGTAAAT
<i>ATF3</i>	Fwd: TCTCATCTTCTGGAGTCCTC Rev: TTGAGGAGCTCAAGAACG
<i>ATF7</i>	Fwd: AAGATCCAGATGAGCGAC Rev: TTCTTCTCTAGGGAGGACAC
<i>JDP2</i>	Fwd: TCAATCTGGGTCTTCAGC Rev: AGAACAAAGTCGCAGCAG
<i>CREB1</i>	Fwd: TTGACTTGTGGAGACTG Rev: TCATGCAACATCATCTGC
<i>CREB3L4</i>	Fwd: ACAACCTCATAGAGCATAGGAG Rev: TCAAGAGAGTGAGCCTGAAG
<i>PVRL4</i>	Fwd: CTCCAGGACCAAAGGATCAC Rev: TCAAGGCCCTCACAGAG
<i>HPRT1</i>	Fwd: TGACCTTGATTTATTTTGCATACC Rev: CGAGCAAGACGTTTCAGTCCT

Fwd, forward primer; Rev, reverse primer; ATF, activating transcription factor; CREB, cAMP responsive element binding protein; HPRT1, hypoxanthine phosphoribosyltransferase 1; JDP2, Jun dimerization protein 2.

Table SII. Target sequence of the siRNAs used in the present study.

Gene	Target sequence (5'- 3')
Control	UGGUUUACAUGUCGACUAA UGGUUUACAUGUUGUGUGA UGGUUUACAUGUUUUCUGA UGGUUUACAUGUUUCCUA
<i>JUN</i>	AUAUGGAAUUGCUUACCAAAGGATA
<i>JUNB</i>	GAAACACGCACUUAGUCUCUAAAGA
<i>JUND</i>	CGAGUCCACAUUCCUGUUUGUAATC
<i>FOS#1</i>	GGGGCAAGGUGGAACAGUUAUCUCC
<i>FOS#2</i>	CAGUGGAACCUGUCAAGAGCAUCAG
<i>FOS#3</i>	CUGAUUAGAAAUGACCAAUAUUATA
<i>ATF3</i>	GCAUUUGAUUAACAUGCUC AACCTT
<i>ATF7</i>	GCUAGCUCCUUUGAACAUGAAUUCA
<i>JDP2</i>	CGUGAAAAGUGAGCUAGAUGAGGAA
<i>CREB1</i>	GAGAGAGGUCCGUCUAAUG
<i>CREB3L4</i>	GCUAAUUGCUCAAACUCCAACAAA
<i>PVRL4#1</i>	CCAAACUCUUCUUAGUCUGAAAUCT
<i>PVRL4#2</i>	CAAGGGAUUCUCAGGUCACUGUGGA

ATF, activating transcription factor; CREB1, CAMP responsive element binding protein 1; CREM, cAMP responsive element modulator; FOSL2, FOS-like 2; HPRT1, hypoxanthine phosphoribosyltransferase 1; JDP2, Jun dimerization protein 2.

Table SIII. Sequences of the primers used for the cloning of reporter plasmids.

Region cloned into pGL4.23, bp	Primer sequence (5' - 3') ^a	Restriction enzyme
5'-flanking region #1, -3402/-2093	Fwd: <u>ccgctcgagcgg</u> TCTTCTCTGACCTGGGACTTC	<i>XhoI</i>
	Rev: <u>gaagatcttc</u> AGGAGAACTGAGGCACAG	<i>BglIII</i>
5'-flanking region #2, -4687/-3352	Fwd: <u>cgggtacc</u> ccgTTCGGTAGGACCCATACTTG	<i>KpnI</i>
	Rev: <u>cccaagcttggg</u> TGGTGAGTAATATCCTGTCTGC	<i>HindIII</i>
5'-flanking region #3, -6717/-5455	Fwd: <u>ccgctcgagcgg</u> TCTGCAGTGGTCTATAATGACC	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> TCAGTCAAAGGAGCTCTCAAG	<i>HindIII</i>
5'-flanking region #4, -7420/-6689	Fwd: <u>ccgctcgagcgg</u> TCCCAACTATAGCTTTCCCTG	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> TGACAAGGTCATTATAGACCACTG	<i>HindIII</i>
5'-flanking region #5, -8058/-7270	Fwd: <u>ccgctcgagcgg</u> ACTCCTACCCTTCCCTGATATC	<i>XhoI</i>
	Rev: <u>gaagatcttc</u> TAGCCAGTGAGAAAGAAGCC	<i>BglIII</i>
intron #6, +671/+910	Fwd: <u>ccgctcgagcgg</u> TGGTGACCCAGTGTAGATG	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> TCCTGGTGCTCTAGAACATC	<i>HindIII</i>
intron #7, +4277/+4498	Fwd: <u>ccgctcgagcgg</u> AATCTGGTGTGTCCTGAGAG	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> AGCTCCCAGCACAGATAAG	<i>HindIII</i>
intron #8, +5164/+5421	Fwd: <u>ccgctcgagcgg</u> TAGTGGTGAGAAGGTTGGC	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> TCTGTGGTATGTGGCTGTG	<i>HindIII</i>
intron #9, +5433/+6630	Fwd: <u>ccgctcgagcgg</u> ATAGCCTCACATGCGTTTG	<i>XhoI</i>
	Rev: <u>gaagatcttc</u> TATACTAGCCAACCACCTGC	<i>BglIII</i>
Intron #10, +14831/+15174	Fwd: <u>ccgctcgagcgg</u> TCCAATTACCTCTACACCCAG	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> AAGACAGGGAAGCTGAAGG	<i>HindIII</i>
intron #11, +15750/+15891	Fwd: <u>ccgctcgagcgg</u> CAAGTATGCACACGCACA	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> AAGGTGAGCCTATCCTGG	<i>HindIII</i>
intron #12, +16185/+16582	Fwd: <u>ccgctcgagcgg</u> TGTCTTTCCAGTGAGGAG	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> AACAGGTCCATTCTCTCCTG	<i>HindIII</i>
3'-flanking region #13, +55848/+56085	Fwd: <u>ccgctcgagcgg</u> TTAGCCAGGTGTGGTGAC	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> AGTGCATAATCATGGCTCAC	<i>HindIII</i>
3'-flanking region #14, +98811/+99037	Fwd: <u>ccgctcgagcgg</u> TTACAGGCACGTGCTACTAC	<i>XhoI</i>
	Rev: <u>cccaagcttggg</u> TTCAACCTGCTCTACTGTCC	<i>HindIII</i>

^aText in uppercase denotes the sequence in the genome; underlined text denotes the restriction enzyme site. Fwd, forward primer; Rev, reverse primer.

Table SIV. Primer sequences for the #10 region for the cloning of reporter plasmids.

Region	Primer sequence (5' - 3') ^a	Restriction enzyme
10-I	Fwd: ccgctc <u>gag</u> cggTCCAATTACCTCTACACCCAG	<i>XhoI</i>
	Rev: ccca <u>agctt</u> gggAACGTGTCCAGACAGCTC	<i>HindIII</i>
#10-II	Fwd: ccgctc <u>gag</u> cggTCCAATTACCTCTACACCCAG	<i>XhoI</i>
	Rev: ccca <u>agctt</u> gggAGAGGTCCTTTCTCCCTGC	<i>HindIII</i>
#10-III	Fwd: ccgctc <u>gag</u> cggGAGCAGGGAGAAAGGACCTC	<i>XhoI</i>
	Rev: ccca <u>agctt</u> gggAACGTGTCCAGACAGCTC	<i>HindIII</i>

^aText in uppercase denotes the sequence in the genome; underlined text denotes the restriction enzyme site. Fwd, forward primer; Rev, reverse primer.

Table SV. Sequences of the primers used for the cloning of mutant plasmids.

Mutant	Primer sequence (5' - 3')
#10-III FOS binding site Mut-1	Fwd: GCCTGCGGCTCTGGGCAACGTCATGGGGGTGG Rev: CCACCCCATGACGTTGCCAGAGCCGCAGGC
#10-III FOS binding site Mut-2	Fwd: CGGCTCTGGGTGACGCAATGGGGGTGGAGGGT Rev: ACCCTCCACCCCATTTGCGTCACCCAGAGCCG
#10-III FOS binding site Mut-1 + 2	Fwd: GCCTGCGGCTCTGGGCAACGCAATGGGGGTGG Rev: CCACCCCATTTGCGTTGCCAGAGCCGCAGGC

Fwd, forward primer; Rev, reverse primer.

Table SVI. Primer sequences for the pCMV-FOS plasmid.

Gene	Primer sequence (5' - 3') ^a	Restriction enzyme
<i>FOS</i>	Fwd: ccggaattcggATGATGTTCTCGGGCTTCAA	<i>EcoRI</i>
	Rev: ccgctcgagTCACAGGGCCAGCAGCGTGG	<i>XhoI</i>

^aText in uppercase denotes the sequence in the genome; underlined text denotes the restriction enzyme site. Fwd, forward primer; Rev, reverse primer.

Table SVII. Primer sequences for the Assay for Transposase-Accessible Chromatin Sequencing.

Region	Primer sequence (5' - 3')
#10	Fwd: TCCAATTACCTCTACACCCAG
	Rev: AAGACAGGGAAGCTGAAGG
#11	Fwd: CAAGTATGCACACGCACA
	Rev: AAGGTGAGCCTATCCTGG
#13	Fwd: TTAGCCAGGTGTGGTGAC
	Rev: AGTGCATAATCATGGCTCAC
#14	Fwd: TTACAGGCACGTGCTACTAC
	Rev: TTCAACCTGCTCTACTGTCC

Fwd, forward primer; Rev, reverse primer.

Table SVIII. Primer sequences for the chromatin conformation capture assay.

PCR	Region	Primer sequence (5' - 3')
1st PCR	Promoter	
	5'	AGAACTCTGCAGCTTCCTG
	3'	ATCCTCCTTGCTCTCAACC
	Enhancer	
	5'	AGAGAGTAGGAGTTGAGGTGAG
	3'	AAGTCCACACAACAGGACAC
Nested PCR	Promoter	
	5'	TCCTTATTCAAGTCTGCAGC
	3'	AATCCAGGACCACAGTATGG
	Enhancer	
	5'	TCTGGATGTTTCTCTGGGTTG
	3'	TGATGTTGCATGGCAGGATAC

Table SIX. *PVRL4* enhancer region and NC primer sequences for the chromatin immunoprecipitation sequencing.

Region	Primer sequence (5' - 3')
<i>PVRL4</i> enhancer	Fwd: GAGCAGGGAGAAAGGACCTC
	Rev: AACGTGTCCAGACAGCTC
NC (exon 1 of <i>GAPDH</i>)	Fwd: AGCTCAGGCCTCAAGACCTT
	Rev: AAGAAGATGCGGCTGACTGT

GAPDH, glyceraldehyde-3-phosphate dehydrogenase; NC, negative control; Fwd, forward primer; Rev, reverse primer.

Table SX. Changes in expression of genes related to “Cytokine Signaling in Immune System” upregulated following treatment with siPVRL4#1 and #2.

Gene	siPVRL4#1			siPVRL4#2		
	Relative expression, Log ₂ FC	P-value	q-value	Relative expression, Log ₂ FC	P-value	q-value
<i>IFIT1</i>	11.45	9.5x10 ⁻¹³	1.2x10 ⁻⁹	4.62	0.004	0.107
<i>IFI44</i>	10.90	7.4x10 ⁻⁹	2.6x10 ⁻⁶	4.23	0.029	0.301
<i>IFI44L</i>	9.88	3.6x10 ⁻⁷	7.5x10 ⁻⁵	3.51	0.084	0.490
<i>MX1</i>	9.82	5.5x10 ⁻¹⁴	1.0x10 ⁻¹⁰	3.30	0.012	0.190
<i>XAF1</i>	9.67	2.6x10 ⁻⁶	4.5x10 ⁻⁴	4.05	0.052	0.391
<i>OAS2</i>	9.22	1.5x10 ⁻¹³	2.6x10 ⁻¹⁰	2.95	0.020	0.254

IFIT1, interferon induced protein with tetratricopeptide repeats 1; IFI44, interferon induced protein 44; IFI44L, interferon induced protein 44-like; MX1, MX dynamin-like GTPase 1; XAF1, XIAP associated factor 1; OAS2, 2'-5'-Oligoadenylate Synthetase 2; si, small interfering; FC, fold change.