

Table SI. Effect of synthetic drugs against metabolic reprogramming in different cancer models.

A, Glucose metabolism					
First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Liu, 2012	GLUT1	WZB115	<i>In vitro</i> analysis	Preclinical	(1)
Liu <i>et al</i> , 2012		WZB117	Lung cancer	Preclinical	(2)
Gunnink <i>et al</i> , 2016		Curcumin	Fibroblast cells	Preclinical	(3)
Wood <i>et al</i> , 2008		Fesentin	Prostate cancer cells, leukemia cells	Preclinical	(4)
Gunnink <i>et al</i> , 2016		Cytochalasin B	Enzymatic assay	Preclinical	(3)
Chan <i>et al</i> , 2011		STF-31	Renal cell carcinoma	Preclinical	(5)
Wu <i>et al</i> , 2018, Lin <i>et al</i> , 2016	GLUT2	Phloretin	Breast cancer, colorectal cancer	Preclinical	(6,7)
Flaig <i>et al</i> , 2007	GLUT4	Silybin	Prostate cancer	Phase I	(8)
Dalva-Aydemir <i>et al</i> , 2015		Ritonavir	Myeloma cells	Preclinical	(9)
Yang <i>et al</i> , 2023	GLUT5	2,5-AM	Lung cancer cell	Preclinical	(10)
B, Glycolysis					
First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
De Marinis <i>et al</i> , 1999; Berruti <i>et al</i> , 2002	HK1	Lonidamine	Lung cancer, breast cancer	Phase III	(11,12)
Dai <i>et al</i> , 2015; Boocock <i>et al</i> , 2007	HK2	Resveratrol	Liver cell	Phase I	(13,14)
Mohanti <i>et al</i> , 1996; Stein <i>et al</i> , 2010		2-Deoxyglucose	Human cerebral gliomas, prostate cancer	Phase II	(15,16)
Ko <i>et al</i> , 2012; El Sayed <i>et al</i> , 2014		3-Bromopyruvate	Melanoma cell	Preclinical	(17,18)

Xu <i>et al</i> , 2017		Chrysin	Hepatocellular carcinoma cells	Preclinical	(19)
Li <i>et al</i> , 2017		Benserazide	<i>In vitro</i> analysis	Preclinical	(20)
Li <i>et al</i> , 2017		Astragalin	Hepatocellular carcinoma cells	Preclinical	(21)
Tao <i>et al</i> , 2017		Genistein-27	Breast cancer cells	Preclinical	(22)
Huang <i>et al</i> , 2012	PKM2	Orlistat	Ovarian cancer	Preclinical	(23)
Zhang <i>et al</i> , 2021		Shikonin	Esophageal squamous cell carcinomas	Preclinical	(24)
Chesney <i>et al</i> , 2015	PFKFB4	5MPN	Lung cancer, breast cancer, prostatic cancer and adenocarcinoma cell lines	Preclinical	(25)
De Oliveira <i>et al</i> , 2021	PFKFB3	KAN0438757	Colorectal cancer cells	Preclinical	(26)
Wang <i>et al</i> , 2020		Compounds 26	Enzymatic assay	Preclinical	(27)
Lea <i>et al</i> , 2016		PQP	Colon cancer cells, bladder cancer cells	Preclinical	(28)
Jiang <i>et al</i> , 2022		3PO/PFK158	Ovarian cancer	Phase I	(29)
Mateo <i>et al</i> , 2013	PDK-1	AR-12	Advanced solid tumors	Phase I	(30)
Powell <i>et al</i> , 2022	Pan-PDK	DCA	Head and neck squamous cell carcinoma	Phase II	(31)
Dhar <i>et al</i> , 2009		Mitaplatin (cisplatin and DCA fusion)	Advanced solid tumor	Phase I	(32)
Liberti <i>et al</i> , 2017; Liberti <i>et al</i> , 2018	GAPDH	Koningic acid	Enzymatic assay	Preclinical	(33,34)
Ganapathy-Kanniappan <i>et al</i> , 2018		Iodoacetate	Enzymatic assay	Preclinical	(35)
Ganapathy-Kanniappan <i>et al</i> , 2010		3-bromopyruvate	Enzymatic assay	Preclinical	(36)

---

### C, Tricarboxylic acid cycle

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Megias-Vericat <i>et al</i> , 2019	Mutant IDH1	IDH305	Leukemia cell lines	Phase IV	(37)
Norsworthy <i>et al</i> , 2019		Olutasidenib (FT-2102)	Brain tumor	Phase II	(38)
Heuser <i>et al</i> , 2020		Ivosidenib (AG-120)	Acute myeloid leukemia	Phase IV	(39)
Konteatis <i>et al</i> , 2020		BAY1436032	Leukemia cell lines	Phase I	(40)
Philip <i>et al</i> , 2019	Mutant IDH2	Enasidenib (AG-221)	Acute myeloid leukemia	Phase IV	(41)
Nyce, 2017	Mutant IDH1/2	Vorasidenib (AG-881)	Brain tumor	Phase III	(42)
Fang <i>et al</i> , 2016	PDH	CPI-613	Metastatic adenocarcinoma	Phase III	(43)

### D, Oxidative phosphorylation

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Arrieta <i>et al</i> , 2019	Complex I	Metformin	Lung adenocarcinoma	Clinical and preclinical	(44)
Dalton <i>et al</i> , 2021		Phenformin	Neuroblastoma cells	Clinical and preclinical	(45)
Guo <i>et al</i> , 2016	Complex II	Lonidamine	Human melanoma cells, colon cancer cells	Phase III	(46)
Fiorillo <i>et al</i> , 2016	Complex III	Atovaquone	Cancer stem cells	Early phase I	(47)
Sun <i>et al</i> , 2011	Complex IV	Arsenic trioxide	Breast cancer cells	Phase III	(48)
Moncada, 2015		Nitric oxide	Colon cancer cells	Clinical and preclinical	(49)

### E, Pentose phosphate pathway

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Nyce, 2017	G6PD	Polydatin	Skin cancer	Preclinical	(42)

Fang <i>et al</i> , 2016		Dehydroepiandrosterone	Cervical cancer cells	Phase I	(43)
Oronsky <i>et al</i> , 2016		RRx-001	Hepatocellular carcinoma, colorectal cancer	Phase II	(50)
Daneshmandi <i>et al</i> , 2021	6PGD	6-aminonicotinamide	T-cell metabolism	Preclinical	(51)

---

F, Lactate metabolism

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Le <i>et al</i> , 2010	LDHA	Oxamate	Pancreatic cancer cells	Preclinical	(52)
Zhou <i>et al</i> , 2010		FX11	Breast cancer cells	Preclinical	(53)
Colen <i>et al</i> , 2011		Galloflavin	Brain tumor	Preclinical	(54)
Guan <i>et al</i> , 2020	MCTs	$\alpha$ -cyano-4-hydroxycinnamic acid	Breast tumor	Preclinical	(55)
Guan <i>et al</i> , 2019	MCTs-1	AR-C155858	Breast tumor cancer cells	Preclinical	(56)
Noble <i>et al</i> , 2017		AZD3965	B-cell lymphoma	Phase I	(57)

---

G, Lipid metabolism

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Alwarawrah <i>et al</i> , 2016	FAS	Fasnall	Breast cancer	Preclinical	(58)
Sadowski <i>et al</i> , 2014		Triclosan	Prostate cancer	Preclinical	(59)
Zhou <i>et al</i> , 2003		C75	Breast cancer cells	Preclinical	(60)
Zhou <i>et al</i> , 2003; Flavin <i>et al</i> , 2010		C93	Breast cancer cells, prostate cancer	Preclinical	(60,61)
Hardwicke <i>et al</i> , 2014		GSK2194069	Gastric cell lines, non-small cell lung cancer cell lines	Preclinical	(62)
Vázquez <i>et al</i> , 2008		GSK837149A	Enzymatic assay	Preclinical	(63)
Kridel <i>et al</i> , 2004		Orlistat	Adenocarcinoma cell lines, prostate carcinoma cell lines	Preclinical	(64)
Shiragami <i>et al</i> , 2013		Cerulenin	Human colon cancer cells	Preclinical	(65)

Loomba <i>et al</i> , 2021		TVB-2640	Non-alcoholic steatohepatitis	Phase II	(66)
Lally <i>et al</i> , 2019	ACC	ND-654	Hepatocellular carcinoma	Preclinical	(67)
Li <i>et al</i> , 2019		ND-646	Non-small cell lung cancer	Preclinical	(68)
Beckers <i>et al</i> , 2007		Soraphen-A	Prostate cancer cells	Preclinical	(69)
Wang <i>et al</i> , 2009		5-(tetradecyloxy)-2-furoic acid	Breast cancer, prostate cancer	Preclinical	(70)
Bauer <i>et al</i> , 2005; Hatzivassiliou <i>et al</i> , 2005	ACLY	SB0204990	Hematopoietic cells	Preclinical	(71,72)
Shrypek <i>et al</i> , 2021	SCD	A939572	Pancreatic tumor	Preclinical	(73)
Fritz <i>et al</i> , 2010		BZ36	Prostate cancer	Preclinical	(74)
Li <i>et al</i> , 2014	SREBP	Fatostatin	Prostate cancer	Preclinical	(75)
Król <i>et al</i> , 2015		Betulin	Ovarian carcinoma, cervical carcinoma, glioblastoma multiforme	Preclinical	(76)

---

H, Amino acid metabolism

First author/s, year	Target	Drugs	Cancer model	Clinical phase	(Refs.)
Ju <i>et al</i> , 2019	MTHFD2	LY345899	Colorectal cancer	Preclinical	(77)
Fu <i>et al</i> , 2017	MTHFD1/2	Carolacton	Colon cancer cells	Preclinical	(78)
Okimoto <i>et al</i> , 2020	TS, DHFR, GARFT	Pemetrexed/MTA/LY231514	Lung cancer cells	Phase IV	(79)
Rana <i>et al</i> , 2020; Neradil <i>et al</i> , 2015	TS, DHFR	Amethopterin/MTX/methotrexate	Medulloblastoma and osteosarcoma cells	Phase IV	(80,81)
Sharma <i>et al</i> , 2008; Siddiqui <i>et al</i> , 2019	TS	Capecitabine/xeloda	Colorectal cancer, pancreatic cancer	Phase IV	(82,83)
Kim <i>et al</i> , 2022		5-Fluorouracil/Adrucil	Colon cancer	Phase IV	(84)
Atkinson <i>et al</i> , 1965;	PPAT	6-mercaptopurine	Ehrlich ascites tumor cells, malignant neoplasm	Phase III	(85,86)

Kodama <i>et al</i> , 2020					
Munshi <i>et al</i> , 2014		6-thioguanine	Colon cancer, gastric cancer, pancreatic cancer, head and neck cancer, renal cell cancer, breast cancer	Phase III	(87)
Elgogary <i>et al</i> , 2016	GLS	BPTES	Pancreatic cancer	Preclinical	(88)
Tannir <i>et al</i> , 2018		CB-839	Metastatic renal cell carcinoma	Phase II	(89)
Yamashita <i>et al</i> , 2021		JHU-083	Malignant glioma cells	Preclinical	(90)
Wang <i>et al</i> , 2010		969	Mesothelioma, pancreatic cancer, sarcoma, glioblastoma, gastrointestinal tumors	Preclinical	(91)

---

6PGD, 6-phosphogluconate dehydrogenase; ACC, acetyl CoA carboxylase; ACLY, ATP citrate lyase; DCA, dichloroacetate; DHFR, dihydrofolate reductase; FAS, fatty acid synthase; G6PD, glucose-6-phosphate dehydrogenase; GARFT, glycinamide ribonucleotide formyl transferase; GLS, glutaminase; GLUT, glucose transporter; HK, hexokinase; IDH, isocitrate dehydrogenase; LDHA, lactate dehydrogenase; MCT, monocarboxylate transporter; MTHFD, methylene tetrahydrofolate dehydrogenase; PDH, pyruvate dehydrogenase; PDK, pyruvate dehydrogenase kinase; PFKFB, fructose-2,6-bisphosphatase; PK, pyruvate kinase; PPAT, phosphoribosyl pyrophosphate aminotransferase; SCD, stearoyl CoA desaturase; SREBP, sterol regulatory element binding protein; TS, thymidylate synthase.

Table SII. Effect of flavonoids against metabolic reprogramming in different cancer models.

A, Glucose metabolism					
First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Melstrom <i>et al.</i> , 2008; Gonzalez-Menendez <i>et al.</i> , 2014	GLUTs	Apigenin	↓GLUT1 expression	Prostate cancer	(92,93)
Farhan, 2022		Catechin	↓GLUT1 expression	Prostate cancer	(94)
Li <i>et al.</i> , 2017		Genistein	↓GLUT1 expression	Hepatocellular carcinoma	(95)
Melstrom <i>et al.</i> , 2008		Phloretin	↓GLUT4 expression	Prostate cancer	(92)
Yang <i>et al.</i> , 2013		Hesperitin	↓GLUT1 level; translocation	↓GLUT4 Breast cancer	(96)
Azevedo <i>et al.</i> , 2015		Kaempferol	↓GLUT1 level	Breast cancer	(97)
Moreira <i>et al.</i> , 2013		Quercetin	↓GLUT1 activity	Breast cancer	(98)
Moreira <i>et al.</i> , 2013; Wei <i>et al.</i> , 2018		EGCG	↓GLUT1 expression	Breast cancer	(98,99)
Zhan <i>et al.</i> , 2011		Silibinin	↓GLUT4 activity	Non-tumor CHO	(100)
B, Glycolysis					
First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Liu <i>et al.</i> , 2019	HK2	Xanthohumol	↓HK2 expression	Colon cancer	(101)
Wu <i>et al.</i> , 2019		Quercetin	↓HK2 level	Hepatocellular cancer	(102)
Wei <i>et al.</i> , 2018		EGCG	↓HK2 activity	Breast cancer	(99)
Deng <i>et al.</i> , 2019		Chalcones (10v)	↓HK2 level	Colon cancer	(103)
Guo <i>et al.</i> , 2020		Wogonin (G1-v9)	↓HK2 expression	Breast cancer	(104)
Zhou <i>et al.</i> , 2016		Wogonin (Fv-429)	↓HK2 activity	Breast cancer	(105)

Tao <i>et al.</i> , 2017		Genistein 27	↓HK2 expression	Breast cancer	(22)
Li <i>et al.</i> , 2017		Astralangin	↓HK2 expression	Hepatocellular cancer	(21)
Li <i>et al.</i> , 2017		Hesperitin	↓HK2 expression	Hepatocellular cancer	(95)
Shan <i>et al.</i> , 2017	PKM2	Apigenin	↓PKM2 expression	Colon cancer	(106)
Feng <i>et al.</i> , 2019		Proanthocyanidin B2	↓PKM2 expression	Hepatocellular cancer	(107)
Wei <i>et al.</i> , 2018		EGCG	↓PKM2 expression	Breast cancer	(99)
Aslan <i>et al.</i> , 2016		Taxifolin	↓PKM2 activity	Breast cancer	(108)
		Neeriocitrin	↓PKM2 activity	Breast cancer	
		Fisetin	↓PKM2 activity	Breast cancer	
		Catechin gallate	↓PKM2 activity	Breast cancer	
		Epicatechin	↓PKM2 activity	Breast cancer	
Jia <i>et al.</i> , 2018		Quercetin	↓PKM2 activity	Breast cancer	(109)
Chen <i>et al.</i> , 2011		Shikonin	↓PKM2 activity	Breast cancer, lung cancer, melanoma	(110)
Mazlaghaninia <i>et al.</i> , 2019	LDH	Morin	↓LDH activity	<i>In vitro</i> study	(111)
Wu <i>et al.</i> , 2021		Kaempferol	↓LDH activity	Colon cancer	(112)
Jia <i>et al.</i> , 2018		Quercetin	↓LDH activity	Breast cancer	(109)
Bader <i>et al.</i> , 2015		Luteolin-7-O-β-D-glucoside	↓LDH enzymatic activity	<i>In vitro</i> study	(113)
Wei <i>et al.</i> , 2018		EGCG	↓LDH activity	Breast cancer	(99)
Li <i>et al.</i> , 2016	PFK	EGCG	↓PFK activity	Breast cancer	(114)
Gomez <i>et al.</i> , 2013		Resveratrol	↓PFK activity	Breast cancer	(115)
Dihal <i>et al.</i> , 2008	Aldolase	Quercetin	↓ Aldolase level	Colon mucosa	(116)



Dihal <i>et al</i> , 2008	Enolase	Quercetin	↓ Enolase level	Colon mucosa	(116)
Dihal <i>et al</i> , 2008	GAPDH	Quercetin	↓GAPDH level	Colon mucosa	(116)
Sellam <i>et al</i> , 2020	PD-L1	Silibinin	↓PD-L1 level	Prostate cancer	(117)

#### C, Tricarboxylic acid cycle

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Coricovac <i>et al</i> , 2021	PDH	Betulinic acid	↓PDH level	<i>In vitro</i>	(118)
Peeters <i>et al</i> , 2019	IDH	EGCG	↓ IDH activity	Colorectal cancer	(119)
Icard <i>et al</i> , 2021		Curcumin		In vitro study	(120)
Wei <i>et al</i> , 2022		Silibinin		Ovarian cancer	(121)
Frattaruolo <i>et al</i> , 2020		Resveratrol		In vitro assay	(122)
Shuvalov <i>et al</i> , 2023		Kaempferol		Enzymatic assay	(123)

#### D, OXPHOS

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Bianchi <i>et al</i> , 2018	ATP synthase	Curcumin	⊥ATP synthase	Breast cancer	(124)
Zheng <i>et al</i> , 2022		Kaempferol		Melanoma metastasis	(125)
Brecht <i>et al</i> , 2017	Mitochondrial chain complexes II and IV	Arctigenin	⊥Mitochondrial chain complexes II and IV and selectively killed only the OXPHOS-dependent cancer cells	Pancreatic cancer	(126)
Abotaleb <i>et al</i> , 2018	Mitochondrial membrane potential	Hesperetin	↓ Mitochondrial membrane potential ( $\Delta\Psi_M$ )	Gastric cancer	(127)
Patel <i>et al</i> , 2018		Naringenin	↓ Mitochondrial membrane potential	<i>In vitro</i>	(128)
Kicinska and	Complex I	Luteolin	Complex I, lower $H_2O_2$	Rat heart mitochondria	(129)

Jarmuszkiewicz, 2020		Myricetin	production		
		Fisetin			
		Rhamnetin			
		Baicalein			
Kincinskaand Jarmuszkiewicz, 2020	Complex III	Hispidulin	↓Complex III, lower H <sub>2</sub> O <sub>2</sub> production	Rat heart mitochondria	(129)

---

E, Pentose phosphate pathway

---

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Adem <i>et al</i> , 2014	G6PD	Caffeic acid	↓Regulation of G6PD	<i>In vitro</i> , cultured rainbow trout gill cells	(130)
Patrício, 2022		Polydatin	↓ Regulation of G6PD	<i>In vitro</i>	(131)
Adem <i>et al</i> , 2014		Ellagic acid	↓Regulation of G6PD	<i>In vitro</i> , enzymatic assay	(130)
Gomez <i>et al</i> , 2013	TK	Resveratrol	↓ Regulation of TK	Colon cancer	(115)
Adem <i>et al</i> , 2014	6PGD	Caffeic acid	↓ Regulation of 6PGD	<i>In vitro</i>	(130)
		Ellagic acid		<i>In vitro</i>	
Khan <i>et al</i> , 2021		Genistein		<i>In vitro</i>	(132)
Lin <i>et al</i> , 2015		Physcion		Lung cancer, leukemia cells, breast cancer	(133)

---

G, Lactate metabolism

---

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Wei <i>et al</i> , 2018	LDHA	EGCG	↓LDHA activity	<i>In vitro</i> study	(99)
Jia <i>et al</i> , 2018		Quercetin	↓LDHA activity	<i>In vitro</i> study	(109)

---

## H, Lipid metabolism

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Li <i>et al</i> , 2014	FAS	$\alpha$ -mangostin	↓Regulation of FAS	Breast cancer	(134)
Funabashi <i>et al</i> , 1989		Cerulenin	↓Regulation of FAS	<i>In vitro</i>	(135)
Lee <i>et al</i> , 2017		Emodin	↓Regulation of FAS	Colon cancer	(136)
Wang and Tian, 2001; Huang <i>et al</i> , 2009		EGCG	↓Regulation of FAS	Hepatocellular carcinoma	(137,138)
Li and Tian, 2004		Kaempferol	↓ Regulation of FAS	<i>In vitro</i>	(139)
Brusselmans <i>et al</i> , 2005		Luteolin	↓ Regulation of FAS	Breast cancer, prostate cancer	(140)
Li and Tian, 2004		Morin	↓ Regulation of FAS	<i>In vitro</i>	(139)
Lee and Sung, 2016		Platyphylloside	↓ Regulation of FAS	Preadipocytes	(141)
Brusselmans <i>et al</i> , 2005		Quercetin	↓ Regulation of FAS	Breast cancer, prostate cancer	(140)
Khan <i>et al</i> , 2014		Resveratrol	↓ Regulation of FAS	Breast cancer	(142)
Gao <i>et al</i> , 2014	ACLY	Cucurbitacins	⊥Xenograft growth in an ACLY-dependent manner	Prostate cancer	(143)
Wang and Tian, 2001; Huang <i>et al</i> , 2009	ACC	EGCG	⊥ACC	Hepatocellular carcinoma	(137,138)
Gnoni <i>et al</i> , 2009		Quercetin	⊥ACC	Rat hepatocytes	(144)
Vahlensieck <i>et al</i> , 1994; Weatherly <i>et al</i> , 2004		Soraphen A	⊥CC	<i>In vitro</i>	(145,146)
Potze <i>et al</i> , 2016	SCD	Betulinic acid	⊥SCD	Colon cancer, HeLa cells	(147)
Lee and Sung, 2016		Platyphylloside	⊥SCD	Preadipocytes	(141)

Wang and Tian, 2001	HIF-1	EGCG	⊥HIF-1 inducible pathways	Breast cancer	(137)
Wei <i>et al</i> , 2015		Oroxylin A		Breast cancer	(148)
Jung <i>et al</i> , 2013		Resveratrol		Breast cancer	(149)
Chen <i>et al</i> , 2015		Baicalein		Gastric cancer	(150)
Liu <i>et al</i> , 2009		Methylalpinumisoflavone		Breast cancer	(151)
Jiang <i>et al</i> , 2017		Eupafolin		Hepatocellular carcinoma	(152)
Ansó <i>et al</i> , 2010	HIF-1 $\alpha$	Quercetin	⊥Protein expression	<i>In vitro</i>	(153)
Sun <i>et al</i> , 2015		Resveratrol			(154)
Wang <i>et al</i> , 2011		EGCG			(155)

#### I, Amino acid metabolism

First author/s, year	Target	Flavonoids	Mode of action	Cancer model	(Refs.)
Nenkov <i>et al</i> , 2021	ASCT-2	Resveratrol	⊥ Regulation of ASCT-2	Hepatoma	(156)
Fan <i>et al</i> , 2022	GLS	Curcumin	⊥GLS	Renal cell carcinoma	(157)
Wang <i>et al</i> , 2020	GLS	Betulinic acid	⊥GLS	<i>In vitro</i> study	(158)
Pournourmohammadi <i>et al</i> , 2017	GDH	EGCG	⊥GDH	<i>In vitro</i> study	(159)
Hay, 2016	PYCR1	Shikonin	⊥PYCR1	T-cell leukemia/lymphoma	(160)

6PGD, 6-phosphogluconate dehydrogenase; ACC, acetyl CoA carboxylase; ACLY, ATP citrate lyase; ASCT2, alanine, serine, cysteine transporter 2; CHO, Chinese hamster ovary cells; EGCG, epigallocatechin-3-gallate; FAS, fatty acid synthase; G6PD, glucose-6-phosphate dehydrogenase; GDH, glutamine dehydrogenase; GLS, glutaminase; GLUT, glucose transporter; HIF, hypoxia-inducible factor; HK, hexokinase; IDH, isocitrate dehydrogenase; LDH, lactate dehydrogenase; LDHA, lactate dehydrogenase A; OXPHOS, oxidative phosphorylation; PD-L1, programmed death-ligand 1; PDH, pyruvate dehydrogenase; PFK, phosphofructokinase; PKM2, pyruvate kinase M2; PYCR1, pyrroline-5-carboxylate reductase 1; SCD, stearoyl CoA desaturase; TK, transketolase.

## References

- (1) Liu Y. Inhibitors of basal glucose transport and their anticancer activities and mechanism. Ohio University; 2012.
- (2) Liu Y, Cao Y, Zhang W, Bergmeier S, Qian Y, Akbar H, et al. A small-molecule inhibitor of glucose transporter 1 downregulates glycolysis, induces cell-cycle arrest, and inhibits cancer cell growth in vitro and in vivo. *Molecular cancer therapeutics* 2012;11:1672-82. doi: <https://doi.org/10.1158/1535-7163>
- (3) Gunnink LK, Alabi OD, Kuiper BD, Gunnink SM, Schuiteman SJ, Strohhenn LE, et al. Curcumin directly inhibits the transport activity of glut1. *Biochimie* 2016;125:179-85. doi: <https://doi.org/10.1016/j.biochi.2016.03.014>
- (4) Wood TE, Dalili S, Simpson CD, Hurren R, Mao X, Saiz FS, et al. A novel inhibitor of glucose uptake sensitizes cells to fas-induced cell death. *Molecular cancer therapeutics* 2008;7:3546-55. doi: <https://doi.org/10.1158/1535-7163.MCT-08-0569>
- (5) Chan DA, Sutphin PD, Nguyen P, Turcotte S, Lai EW, Banh A, et al. Targeting glut1 and the warburg effect in renal cell carcinoma by chemical synthetic lethality. *Science translational medicine* 2011;3:94ra70-94ra70. doi: DOI: 10.1126/scitranslmed.3002394
- (6) Wu K-H, Ho C-T, Chen Z-F, Chen L-C, Whang-Peng J, Lin T-N, et al. The apple polyphenol phloretin inhibits breast cancer cell migration and proliferation via inhibition of signals by type 2 glucose transporter. *Journal of food and drug analysis* 2018;26:221-31. doi: <https://doi.org/10.1016/j.jfda.2017.03.009>
- (7) Lin S-T, Tu S-H, Yang P-S, Hsu S-P, Lee W-H, Ho C-T, et al. Apple polyphenol phloretin inhibits colorectal cancer cell growth via inhibition of the type 2 glucose transporter and activation of p53-mediated signaling. *Journal of agricultural and food chemistry* 2016;64:6826-37. doi: <https://doi.org/10.1021/acs.jafc.6b02861>
- (8) Flaig TW, Gustafson DL, Su L-J, Zirrolli JA, Crighton F, Harrison GS, et al. A phase i and pharmacokinetic study of silybin-phytosome in prostate cancer patients. *Investigational new drugs* 2007;25:139-46. doi: <https://doi.org/10.1007/s10637-006-9019-2>
- (9) Dalva-Aydemir S, Bajpai R, Martinez M, Adekola KU, Kandela I, Wei C, et al. Targeting the metabolic plasticity of multiple myeloma with fda-approved ritonavir and metformin. *Clinical Cancer Research* 2015;21:1161-71. doi: <https://doi.org/10.1158/1078-0432.CCR-14-1088>
- (10) Yang J, Dong C, Wu J, Liu D, Luo Q, Jin X. Fructose utilization enhanced by glut5 promotes lung cancer cell migration via activating glycolysis/akt pathway. *Clinical and Translational Oncology* 2023;25:1080-90. doi: <https://doi.org/10.1007/s12094-022-03015-2>
- (11) De Marinis F, Rinaldi M, Ardizzoni A, Bruzzi P, Pennucci MC, Portalone L, et al. The role of vindesine and lonidamine in the treatment of elderly patients with advanced non-small cell lung cancer: A phase iii randomized fonicap trial. *Tumori Journal* 1999;85:177-82. doi: <https://doi.org/10.1177/030089169908500306>
- (12) Berruti A, Bitossi R, Gorzegno G, Bottini A, Alquati P, De Matteis A, et al. Time to progression in metastatic breast cancer patients treated with epirubicin is not improved by the addition of either cisplatin or lonidamine: Final results of a phase iii study with a factorial design. *Journal of clinical oncology* 2002;20:4150-9. doi: <https://doi.org/10.1200/JCO.2002.08.012>
- (13) Dai W, Wang F, Lu J, Xia Y, He L, Chen K, et al. By reducing hexokinase 2, resveratrol induces apoptosis in hcc cells addicted to aerobic glycolysis and inhibits tumor growth in mice. *Oncotarget* 2015;6:13703. doi: doi: 10.18632/oncotarget.3800
- (14) Boocock DJ, Faust GE, Patel KR, Schinas AM, Brown VA, Ducharme MP, et al. Phase i dose escalation pharmacokinetic study in healthy volunteers of resveratrol, a potential cancer chemopreventive agent. *Cancer Epidemiology Biomarkers & Prevention* 2007;16:1246-52. doi: <https://doi.org/10.1158/1055-9965.EPI-07-0022>
- (15) Mohanti BK, Rath GK, Anantha N, Kannan V, Das BS, Chandramouli BA, et al. Improving cancer radiotherapy with 2-deoxy-d-glucose: Phase i/ii clinical trials on human cerebral gliomas. *International Journal of Radiation Oncology\* Biology\* Physics* 1996;35:103-11. doi: [https://doi.org/10.1016/S0360-3016\(96\)85017-6](https://doi.org/10.1016/S0360-3016(96)85017-6)
- (16) Stein M, Lin H, Jeyamohan C, Dvorzhinski D, Gounder M, Bray K, et al. Targeting tumor metabolism with 2-deoxyglucose in patients with castrate-resistant prostate cancer and advanced malignancies. *The Prostate* 2010;70:1388-94. doi: <https://doi.org/10.1002/pros.21172>
- (17) Ko Y, Verhoeven H, Lee M, Corbin D, Vogl T, Pedersen P. A translational study “case report” on the small molecule “energy blocker” 3-bromopyruvate (3bp) as a potent anticancer agent: From bench side to bedside. *Journal of bioenergetics and biomembranes* 2012;44:163-70. doi: <https://doi.org/10.1007/s10863-012-9417-4>

- (18) El Sayed SM, Mohamed WG, Seddik M-AH, Ahmed A-SA, Mahmoud AG, Amer WH, et al. Safety and outcome of treatment of metastatic melanoma using 3-bromopyruvate: A concise literature review and case study. *Chinese journal of cancer* 2014;33:356. doi: 10.5732/cjc.013.10111
- (19) Xu D, Jin J, Yu H, Zhao Z, Ma D, Zhang C, et al. Chrysin inhibited tumor glycolysis and induced apoptosis in hepatocellular carcinoma by targeting hexokinase-2. *Journal of Experimental & Clinical Cancer Research* 2017;36:1-11. doi: <https://doi.org/10.1186/s13046-017-0514-4>
- (20) Li W, Zhang S, Zhao Y, Huang S, Zhao J. Molecular docking and molecular dynamics simulation analyses of urea with ammoniated and ammoxidized lignin. *Journal of Molecular Graphics and Modelling* 2017;71:58-69. doi: <https://doi.org/10.1021/acs.jafc.7b02120>
- (21) Li W, Hao J, Zhang L, Cheng Z, Deng X, Shu G. Astragaloside reduces hexokinase 2 through increasing mir-125b to inhibit the proliferation of hepatocellular carcinoma cells in vitro and in vivo. *Journal of Agricultural and Food Chemistry* 2017;65:5961-72. doi: <https://doi.org/10.1021/acs.jafc.7b02120>
- (22) Tao L, Wei L, Liu Y, Ding Y, Liu X, Zhang X, et al. Gen-27, a newly synthesized flavonoid, inhibits glycolysis and induces cell apoptosis via suppression of hexokinase ii in human breast cancer cells. *Biochemical pharmacology* 2017;125:12-25. doi: <https://doi.org/10.1016/j.bcp.2016.11.001>
- (23) Huang H-Q, Tang J, Zhou S-T, Yi T, Peng H-L, Shen G-B, et al. Orlistat, a novel potent antitumor agent for ovarian cancer: Proteomic analysis of ovarian cancer cells treated with orlistat. *International journal of oncology* 2012;41:523-32. doi: <https://doi.org/10.1002/mnfr.200800592>
- (24) Zhang Q, Liu Q, Zheng S, Liu T, Yang L, Han X, et al. Shikonin inhibits tumor growth of escc by suppressing pkm2 mediated aerobic glycolysis and stat3 phosphorylation. *Journal of Cancer* 2021;12:4830. doi: 10.7150/jca.58494
- (25) Chesney J, Clark J, Lanceta L, Trent JO, Telang S. Targeting the sugar metabolism of tumors with a first-in-class 6-phosphofructo-2-kinase (pfkfb4) inhibitor. *Oncotarget* 2015;6:18001. doi: 10.18632/oncotarget.4534
- (26) De Oliveira T, Goldhardt T, Edelmann M, Rogge T, Rauch K, Kyuchukov ND, et al. Effects of the novel pfkfb3 inhibitor kan0438757 on colorectal cancer cells and its systemic toxicity evaluation in vivo. *Cancers* 2021;13:1011. doi: <https://doi.org/10.3390/cancers13051011>
- (27) Wang Y, Qu C, Liu T, Wang C. Pfkfb3 inhibitors as potential anticancer agents: Mechanisms of action, current developments, and structure-activity relationships. *European Journal of Medicinal Chemistry* 2020;203:112612. doi: <https://doi.org/10.1016/j.ejmech.2020.112612>
- (28) Lea MA, Guzman Y, desBordes C. Inhibition of cancer cell growth by combined treatment with lactate dehydrogenase (ldha) inhibitors and either phenformin or inhibitors of 6-phosphofructo-2-kinase/fructose-2, 6-biphosphatase 3 (pfkfb3). *Cancer Research* 2016;76:32-. doi: <https://doi.org/10.1158/1538-7445.AM2016-32>
- (29) Jiang Y-x, Siu MK, Wang J-j, Leung TH, Chan DW, Cheung AN, et al. Pfkfb3 regulates chemoresistance, metastasis and stemness via iap proteins and the nf- $\kappa$ b signaling pathway in ovarian cancer. *Frontiers in oncology* 2022;12:748403. doi: <https://doi.org/10.3389/fonc.2022.748403>
- (30) Mateo J, De Bono JS, Ramanathan RK, Lustberg MB, Zivi A, Basset D, et al. A first-in-human phase i trial of ar-12, a pdk-1 inhibitor, in patients with advanced solid tumors. *American Society of Clinical Oncology*; 2013. <https://doi.org/10.1200/jco.2013.31.15>
- (31) Powell SF, Mazurczak M, Dib EG, Bleeker JS, Geeraerts LH, Tinguely M, et al. Phase ii study of dichloroacetate, an inhibitor of pyruvate dehydrogenase, in combination with chemoradiotherapy for unresected, locally advanced head and neck squamous cell carcinoma. *Investigational New Drugs* 2022;40:622-33. doi: <https://doi.org/10.1007/s10637-022-01235-5>
- (32) Dhar S, Lippard SJ. Mitaplatin, a potent fusion of cisplatin and the orphan drug dichloroacetate. *Proceedings of the National Academy of Sciences* 2009;106:22199-204. doi: <https://doi.org/10.1073/pnas.0912276106>
- (33) Liberti MV, Dai Z, Wardell SE, Baccile JA, Liu X, Gao X, et al. A predictive model for selective targeting of the warburg effect through gapdh inhibition with a natural product. *Cell metabolism* 2017;26:648-59. e8. doi: <http://dx.doi.org/10.1016/j.cmet.2017.08.017>
- (34) Liberti MV, Dai Z, Wardell SE, Baccile JA, Liu X, Gao X, et al. A predictive model for selective targeting of the warburg effect through gapdh inhibition with a natural product. *Cancer Research* 2018;78:5496-. doi: <https://doi.org/10.1158/1538-7445.AM2018-5496>
- (35) Ganapathy-Kanniappan S. Evolution of gapdh as a druggable target of tumor glycolysis? *Expert opinion on therapeutic targets* 2018;22:295-8. doi: <https://doi.org/10.1080/14728222.2018.1449834>

- (36) Ganapathy-Kanniappan S, Vali M, Kunjithapatham R, Buijs M, Syed L, Rao P, et al. 3-bromopyruvate: A new targeted antiglycolytic agent and a promise for cancer therapy. *Current pharmaceutical biotechnology* 2010;11:510-7. doi: <https://doi.org/10.1080/14728222.2018.1449834>
- (37) Megías-Vericat JE, Ballesta-López O, Barragán E, Montesinos P. Idh1-mutated relapsed or refractory aml: Current challenges and future prospects. *Blood and lymphatic cancer: targets and therapy* 2019;19-32. doi: DOI: 10.2147/BLCTT.S177913
- (38) Norsworthy KJ, Luo L, Hsu V, Gudi R, Dorff SE, Przepiorka D, et al. Fda approval summary: Ivosidenib for relapsed or refractory acute myeloid leukemia with an isocitrate dehydrogenase-1 mutation. *Clinical Cancer Research* 2019;25:3205-9. doi: <https://doi.org/10.1158/1078-0432.CCR-18-3749>
- (39) Heuser M, Palmisiano N, Mantzaris I, Mims A, DiNardo C, Silverman LR, et al. Safety and efficacy of bay1436032 in idh1-mutant aml: Phase i study results. *Leukemia* 2020;34:2903-13. doi: <https://doi.org/10.1038/s41375-020-0996-5>
- (40) Konteatis Z, Artin E, Nicolay B, Straley K, Padyana AK, Jin L, et al. Vorasidenib (ag-881): A first-in-class, brain-penetrant dual inhibitor of mutant idh1 and 2 for treatment of glioma. *ACS medicinal chemistry letters* 2020;11:101-7. doi: <https://doi.org/10.1021/acsmchemlett.9b00509>
- (41) Philip PA, Buyse ME, Alistar AT, Lima CM, Luther S, Pardee TS, et al. A phase iii open-label trial to evaluate efficacy and safety of cpi-613 plus modified folfirinox (mffx) versus folfirinox (ffx) in patients with metastatic adenocarcinoma of the pancreas. *Future Oncology* 2019;15:3189-96. doi: <https://doi.org/10.2217/fon-2019-0209>
- (42) Nyce JW. Autoinflammatory reaction in dogs treated for cancer via g6pd inhibition. *Case Reports in Veterinary Medicine* 2017;2017. doi: <https://doi.org/10.1155/2017/4275305>
- (43) Fang Z, Jiang C, Feng Y, Chen R, Lin X, Zhang Z, et al. Effects of g6pd activity inhibition on the viability, ros generation and mechanical properties of cervical cancer cells. *Biochimica et Biophysica Acta (BBA)-Molecular Cell Research* 2016;1863:2245-54. doi: <https://doi.org/10.1016/j.bbamcr.2016.05.016>
- (44) Arrieta O, Barrón F, Padilla M-ÁS, Avilés-Salas A, Ramírez-Tirado LA, Jiménez MJA, et al. Effect of metformin plus tyrosine kinase inhibitors compared with tyrosine kinase inhibitors alone in patients with epidermal growth factor receptor-mutated lung adenocarcinoma: A phase 2 randomized clinical trial. *JAMA oncology* 2019;5:e192553-e. doi: doi:10.1001/jamaoncol.2019.2553
- (45) Dalton KM, Lochmann TL, Floros KV, Calbert ML, Kurupi R, Stein GT, et al. Catastrophic atp loss underlies a metabolic combination therapy tailored for mycn-amplified neuroblastoma. *Proceedings of the National Academy of Sciences* 2021;118:e2009620118. doi: <https://doi.org/10.1073/pnas.2009620118>
- (46) Guo L, Shestov AA, Worth AJ, Nath K, Nelson DS, Leeper DB, et al. Inhibition of mitochondrial complex ii by the anticancer agent lonidamine. *Journal of Biological Chemistry* 2016;291:42-57. doi: <https://doi.org/10.1007/s11010-013-1723-6>
- (47) Fiorillo M, Lamb R, Tanowitz HB, Mutti L, Krstic-Demonacos M, Cappello AR, et al. Repurposing atovaquone: Targeting mitochondrial complex iii and oxphos to eradicate cancer stem cells. *Oncotarget* 2016;7:34084. doi: doi: 10.18632/oncotarget.9122
- (48) Sun RC, Board PG, Blackburn AC. Targeting metabolism with arsenic trioxide and dichloroacetate in breast cancer cells. *Molecular cancer* 2011;10:1-15. doi: <https://doi.org/10.1186/1476-4598-10-142>
- (49) Moncada S. Nitric oxide and oxygen: Actions and interactions in health and disease. *Redox Biology* 2015;5:421. doi: <https://doi.org/10.1016/j.redox.2015.09.034>
- (50) Oronsky B, Scicinski J, Reid T, Oronsky A, Carter C, Oronsky N, et al. Rrx-001, a novel clinical-stage chemosensitizer, radiosensitizer, and immunosensitizer, inhibits glucose 6-phosphate dehydrogenase in human tumor cells. *Discovery medicine* 2016;21:251-65. doi: doi.2016.04.rrx-001
- (51) Daneshmandi S, Cassel T, Higashi RM, Fan TW, Seth P. 6-phosphogluconate dehydrogenase (6pgd), a key checkpoint in reprogramming of regulatory t cells metabolism and function. *Elife* 2021;10:e67476. doi: <https://doi.org/10.7554/eLife.67476>
- (52) Le A, Cooper CR, Gouw AM, Dinavahi R, Maitra A, Deck LM, et al. Inhibition of lactate dehydrogenase a induces oxidative stress and inhibits tumor progression. *Proceedings of the National Academy of Sciences* 2010;107:2037-42. doi: <https://doi.org/10.1073/pnas.0914433107>
- (53) Zhou M, Zhao Y, Ding Y, Liu H, Liu Z, Fodstad O, et al. Warburg effect in chemosensitivity: Targeting lactate dehydrogenase-a re-sensitizes taxol-resistant cancer cells to taxol. *Molecular cancer* 2010;9:1-12. doi: <https://doi.org/10.1186/1476-4598-9-33>

- (54) Colen CB, Shen Y, Ghoddoussi F, Yu P, Francis TB, Koch BJ, et al. Metabolic targeting of lactate efflux by malignant glioma inhibits invasiveness and induces necrosis: An in vivo study. *Neoplasia* 2011;13:620-32. doi: <https://doi.org/10.1593/neo.11134>
- (55) Guan X, Morris ME. In vitro and in vivo efficacy of azd3965 and alpha-cyano-4-hydroxycinnamic acid in the murine 4t1 breast tumor model. *The AAPS journal* 2020;22:1-11. doi: <https://doi.org/10.1208/s12248-020-00466-9>
- (56) Guan X, Rodriguez-Cruz V, Morris ME. Cellular uptake of mct1 inhibitors ar-c155858 and azd3965 and their effects on mct-mediated transport of l-lactate in murine 4t1 breast tumor cancer cells. *The AAPS journal* 2019;21:1-10. doi: <https://doi.org/10.1208/s12248-018-0279-5>
- (57) Noble RA, Bell N, Blair H, Sikka A, Thomas H, Phillips N, et al. Inhibition of monocarboxylate transporter 1 by azd3965 as a novel therapeutic approach for diffuse large b-cell lymphoma and burkitt lymphoma. *Haematologica* 2017;102:1247. doi: 10.3324/haematol.2016.163030
- (58) Alwarawrah Y, Hughes P, Loiselle D, Carlson DA, Darr DB, Jordan JL, et al. Fasnall, a selective fasn inhibitor, shows potent anti-tumor activity in the mmtv-neu model of her2+ breast cancer. *Cell chemical biology* 2016;23:678-88. doi: <https://doi.org/10.1016/j.chembiol.2016.04.011>
- (59) Sadowski MC, Pouwer RH, Gunter JH, Lubik AA, Quinn RJ, Nelson CC. The fatty acid synthase inhibitor triclosan: Repurposing an anti-microbial agent for targeting prostate cancer. *Oncotarget* 2014;5:9362. doi: 10.18632/oncotarget.2433
- (60) Zhou W, Simpson PJ, McFadden JM, Townsend CA, Medghalchi SM, Vadlamudi A, et al. Fatty acid synthase inhibition triggers apoptosis during s phase in human cancer cells. *Cancer research* 2003;63:7330-7. doi: 10.1158/0008-5472.CAN-03-3645
- (61) Flavin R, Peluso S, Nguyen PL, Loda M. Fatty acid synthase as a potential therapeutic target in cancer. *Future oncology* 2010;6:551-62. doi: <https://doi.org/10.2217/fon.10.11>
- (62) Hardwicke MA, Rendina AR, Williams SP, Moore ML, Wang L, Krueger JA, et al. A human fatty acid synthase inhibitor binds  $\beta$ -ketoacyl reductase in the keto-substrate site. *Nature chemical biology* 2014;10:774-9. doi: 10.1038/NCHEMBIO.1603
- (63) Vázquez MJ, Leavens W, Liu R, Rodríguez B, Read M, Richards S, et al. Discovery of gsk837149a, an inhibitor of human fatty acid synthase targeting the  $\beta$ -ketoacyl reductase reaction. *The FEBS journal* 2008;275:1556-67. doi: <https://doi.org/10.1111/j.1742-4658.2008.06314.x>
- (64) Kridel SJ, Axelrod F, Rozenkrantz N, Smith JW. Orlistat is a novel inhibitor of fatty acid synthase with antitumor activity. *Cancer research* 2004;64:2070-5. doi: <https://doi.org/10.1158/0008-5472.CAN-03-3645>
- (65) Shiragami R, Murata S, Kosugi C, Tezuka T, Yamazaki M, Hirano A, et al. Enhanced antitumor activity of cerulenin combined with oxaliplatin in human colon cancer cells. *International journal of oncology* 2013;43:431-8. doi: <https://doi.org/10.3892/ijo.2013.1978>
- (66) Loomba R, Mohseni R, Lucas KJ, Gutierrez JA, Perry RG, Trotter JF, et al. Tvb-2640 (fasn inhibitor) for the treatment of nonalcoholic steatohepatitis: Fascinate-1, a randomized, placebo-controlled phase 2a trial. *Gastroenterology* 2021;161:1475-86. doi: <https://doi.org/10.1053/j.gastro.2021.07.025>
- (67) Lally JS, Ghoshal S, DePeralta DK, Moaven O, Wei L, Masia R, et al. Inhibition of acetyl-coa carboxylase by phosphorylation or the inhibitor nd-654 suppresses lipogenesis and hepatocellular carcinoma. *Cell metabolism* 2019;29:174-82. e5. doi: <https://doi.org/10.1016/j.cmet.2018.08.020>
- (68) Li E-Q, Zhao W, Zhang C, Qin L-Z, Liu S-J, Feng Z-Q, et al. Synthesis and anti-cancer activity of nd-646 and its derivatives as acetyl-coa carboxylase 1 inhibitors. *European Journal of Pharmaceutical Sciences* 2019;137:105010. doi: <https://doi.org/10.1016/j.ejps.2019.105010>
- (69) Beckers A, Organe S, Timmermans L, Scheys K, Peeters A, Brusselmans K, et al. Chemical inhibition of acetyl-coa carboxylase induces growth arrest and cytotoxicity selectively in cancer cells. *Cancer research* 2007;67:8180-7. doi: <https://doi.org/10.1158/0008-5472.CAN-07-0389>
- (70) Wang C, Xu C, Sun M, Luo D, Liao D-f, Cao D. Acetyl-coa carboxylase- $\alpha$  inhibitor tofa induces human cancer cell apoptosis. *Biochemical and biophysical research communications* 2009;385:302-6. doi: <https://doi.org/10.1016/j.bbrc.2009.05.045>
- (71) Bauer DE, Hatzivassiliou G, Zhao F, Andreadis C, Thompson CB. Atp citrate lyase is an important component of cell growth and transformation. *Oncogene* 2005;24:6314-22. doi: 10.1038/sj.onc.1208773



- (72) Hatzivassiliou G, Zhao F, Bauer DE, Andreadis C, Shaw AN, Dhanak D, et al. Atp citrate lyase inhibition can suppress tumor cell growth. *Cancer cell* 2005;8:311-21. doi: DOI 10.1016/j.ccr.2005.09.008
- (73) Skrypek K, Balog S, Eriguchi Y, Asahina K. Inhibition of stearyl-coa desaturase induces the unfolded protein response in pancreatic tumors and suppresses their growth. *Pancreas* 2021;50:219. doi: DOI: 10.1097/MPA.0000000000001737
- (74) Fritz V, Benfodda Z, Rodier G, Henriquet C, Iborra F, Avancès C, et al. Abrogation of de novo lipogenesis by stearyl-coa desaturase 1 inhibition interferes with oncogenic signaling and blocks prostate cancer progression in mice. *Molecular cancer therapeutics* 2010;9:1740-54. doi: <https://doi.org/10.1158/1535-7163.MCT-09-1064>
- (75) Li X, Chen Y-T, Hu P, Huang W-C. Fatostatin displays high antitumor activity in prostate cancer by blocking srebp-regulated metabolic pathways and androgen receptor signaling. *Molecular cancer therapeutics* 2014;13:855-66. doi: <https://doi.org/10.1158/1535-7163.MCT-13-0797>
- (76) Król SK, Kielbus M, Rivero-Müller A, Stepulak A. Comprehensive review on betulin as a potent anticancer agent. *BioMed research international* 2015;2015. doi: <https://doi.org/10.1155/2015/584189>
- (77) Ju H-Q, Lu Y-X, Chen D-L, Zuo Z-X, Liu Z-X, Wu Q-N, et al. Modulation of redox homeostasis by inhibition of mthfd2 in colorectal cancer: Mechanisms and therapeutic implications. *JNCI: Journal of the National Cancer Institute* 2019;111:584-96. doi: <https://doi.org/10.1093/jnci/djy160>
- (78) Fu C, Sikandar A, Donner J, Zaburanyi N, Herrmann J, Reck M, et al. The natural product carolacton inhibits folate-dependent c1 metabolism by targeting fold/mthfd. *Nature communications* 2017;8:1529. doi: <https://doi.org/10.1038/s41467-017-01671-5>
- (79) Okimoto T, Kotani H, Iida Y, Koyanagi A, Tanino R, Tsubata Y, et al. Pemetrexed sensitizes human lung cancer cells to cytotoxic immune cells. *Cancer Science* 2020;111:1910-20. doi: <https://doi.org/10.1111/cas.14401>
- (80) Rana RM, Rampogu S, Abid NB, Zeb A, Parate S, Lee G, et al. In silico study identified methotrexate analog as potential inhibitor of drug resistant human dihydrofolate reductase for cancer therapeutics. *Molecules* 2020;25:3510. doi: <https://doi.org/10.3390/molecules25153510>
- (81) Neradil J, Pavlasova G, Sramek M, Kyr M, Veselska R, Sterba J. Dhfr-mediated effects of methotrexate in medulloblastoma and osteosarcoma cells: The same outcome of treatment with different doses in sensitive cell lines. *Oncology Reports* 2015;33:2169-75. doi: <https://doi.org/10.3892/or.2015.3819>
- (82) Sharma R, Hoskins JM, Rivory LP, Zucknick M, London R, Liddle C, et al. Thymidylate synthase and methylenetetrahydrofolate reductase gene polymorphisms and toxicity to capecitabine in advanced colorectal cancer patients. *Clinical cancer research* 2008;14:817-25. doi: <https://doi.org/10.1158/1078-0432.CCR-07-0425>
- (83) Siddiqui NS, Godara A, Byrne MM, Saif MW. Capecitabine for the treatment of pancreatic cancer. *Expert opinion on pharmacotherapy* 2019;20:399-409. doi: <https://doi.org/10.1080/14656566.2018.1560422>
- (84) Kim ST, Kim SY, Lee J, Yun SH, Kim HC, Lee WY, et al. Oxaliplatin (3 months v 6 months) with 6 months of fluoropyrimidine as adjuvant therapy in patients with stage ii/iii colon cancer: Kcsg co09-07. *Journal of Clinical Oncology* 2022;40:3868-77. doi: doi: 10.1200/JCO.21.02962
- (85) Atkinson M, Murray A. Inhibition of purine phosphoribosyltransferases of ehrlich ascites-tumour cells by 6-mercaptopurine. *Biochemical Journal* 1965;94:64. doi: doi: 10.1042/bj0940064
- (86) Kodama M, Nakayama KI. A second warburg-like effect in cancer metabolism: The metabolic shift of glutamine-derived nitrogen: A shift in glutamine-derived nitrogen metabolism from glutaminolysis to de novo nucleotide biosynthesis contributes to malignant evolution of cancer. *BioEssays* 2020;42:2000169. doi: <https://doi.org/10.1002/bies.202000169>
- (87) Munshi PN, Lubin M, Bertino JR. 6-thioguanine: A drug with unrealized potential for cancer therapy. *The oncologist* 2014;19:760-5. doi: <https://doi.org/10.1634/theoncologist.2014-0178>
- (88) Elgogary A, Xu Q, Poore B, Alt J, Zimmermann SC, Zhao L, et al. Combination therapy with bptes nanoparticles and metformin targets the metabolic heterogeneity of pancreatic cancer. *Proceedings of the National Academy of Sciences* 2016;113:E5328-E36. doi: <https://doi.org/10.1073/pnas.1611406113>
- (89) Tannir NM, Motzer RJ, Agarwal N, Liu P-Y, Whiting SH, O'Keeffe B, et al. Cantata: A randomized phase 2 study of cb-839 in combination with cabozantinib vs. Placebo with cabozantinib in patients with advanced/metastatic renal cell carcinoma. *American Society of Clinical Oncology*; 2018. <https://doi.org/10.1200/JCO.2018.36.15>

- (90) Yamashita AS, da Costa Rosa M, Stumpo V, Rais R, Slusher BS, Riggins GJ. The glutamine antagonist prodrug jhu-083 slows malignant glioma growth and disrupts mtor signaling. *Neuro-Oncology Advances* 2021;3:vdad149. doi: doi:10.1093/naojnl/vdad149
- (91) Wang J-B, Erickson JW, Fuji R, Ramachandran S, Gao P, Dinavahi R, et al. Targeting mitochondrial glutaminase activity inhibits oncogenic transformation. *Cancer cell* 2010;18:207-19. doi: DOI 10.1016/j.ccr.2010.08.009
- (92) Melstrom LG, Salabat MR, Ding X-Z, Milam BM, Strouch M, Pelling JC, et al. Apigenin inhibits the glut-1 glucose transporter and the phosphoinositide 3-kinase/akt pathway in human pancreatic cancer cells. *Pancreas* 2008;37:426-31. doi: DOI: 10.1097/MPA.0b013e3181735ccb
- (93) Gonzalez-Menendez P, Hevia D, Rodriguez-Garcia A, Mayo JC, Sainz RM. Regulation of glut transporters by flavonoids in androgen-sensitive and-insensitive prostate cancer cells. *Endocrinology* 2014;155:3238-50. doi: <https://doi.org/10.1210/en.2014-1260>
- (94) Farhan M. Green tea catechins: Nature's way of preventing and treating cancer. *International journal of molecular sciences* 2022;23:10713.
- (95) Li S, Li J, Dai W, Zhang Q, Feng J, Wu L, et al. Genistein suppresses aerobic glycolysis and induces hepatocellular carcinoma cell death. *British journal of cancer* 2017;117:1518-28.
- (96) Yang Y, Wolfram J, Boom K, Fang X, Shen H, Ferrari M. Hesperetin impairs glucose uptake and inhibits proliferation of breast cancer cells. *Cell biochemistry and function* 2013;31:374-9. doi: <https://doi.org/10.1002/cbf.2905>
- (97) Azevedo C, Correia-Branco A, Araújo JR, Guimaraes JT, Keating E, Martel F. The chemopreventive effect of the dietary compound kaempferol on the mcf-7 human breast cancer cell line is dependent on inhibition of glucose cellular uptake. *Nutrition and cancer* 2015;67:504-13. doi: <https://doi.org/10.1080/01635581.2015.1002625>
- (98) Moreira L, Araújo I, Costa T, Correia-Branco A, Faria A, Martel F, et al. Quercetin and epigallocatechin gallate inhibit glucose uptake and metabolism by breast cancer cells by an estrogen receptor-independent mechanism. *Experimental Cell Research* 2013;319:1784-95. doi: <https://doi.org/10.1016/j.yexcr.2013.05.001>
- (99) Wei R, Mao L, Xu P, Zheng X, Hackman RM, Mackenzie GG, et al. Suppressing glucose metabolism with epigallocatechin-3-gallate (egcg) reduces breast cancer cell growth in preclinical models. *Food & function* 2018;9:5682-96. doi: doi:10.1039/c8fo01397g
- (100) Zhan T, Digel M, Küch EM, Stremmel W, Füllekrug J. Silybin and dehydrosilybin decrease glucose uptake by inhibiting glut proteins. *Journal of cellular biochemistry* 2011;112:849-59. doi: <https://doi.org/10.1002/jcb.22984>
- (101) Liu W, Li W, Liu H, Yu X. Xanthohumol inhibits colorectal cancer cells via downregulation of hexokinases ii-mediated glycolysis. *International Journal of Biological Sciences* 2019;15:2497. doi: <https://doi.org/10.1038/nruol.2010.47>
- (102) Wu H, Pan L, Gao C, Xu H, Li Y, Zhang L, et al. Quercetin inhibits the proliferation of glycolysis-addicted hcc cells by reducing hexokinase 2 and akt-mtor pathway. *Molecules* 2019;24:1993. doi: <https://doi.org/10.3390/molecules24101993>
- (103) Deng X, Liu R, Li J, Li Z, Liu J, Xiong R, et al. Design, synthesis, and preliminary biological evaluation of 3', 4', 5'-trimethoxy flavonoid salicylate derivatives as potential anti-tumor agents. *New Journal of Chemistry* 2019;43:1874-84. doi: DOI: 10.1039/c8nj04533j
- (104) Guo Y, Wei L, Zhou Y, Lu N, Tang X, Li Z, et al. Flavonoid gl-v9 induces apoptosis and inhibits glycolysis of breast cancer via disrupting gsk-3 $\beta$ -modulated mitochondrial binding of hkii. *Free Radical Biology and Medicine* 2020;146:119-29. doi: <https://doi.org/10.1007/s11010-013-1723-6>
- (105) Zhou Y, Lu N, Qiao C, Ni T, Li Z, Yu B, et al. Fv-429 induces apoptosis and inhibits glycolysis by inhibiting akt-mediated phosphorylation of hexokinase ii in mda-mb-231 cells. *Molecular carcinogenesis* 2016;55:1317-28. doi: <https://doi.org/10.1002/mc.22374>
- (106) Shan S, Shi J, Yang P, Jia B, Wu H, Zhang X, et al. Apigenin restrains colon cancer cell proliferation via targeted blocking of pyruvate kinase m2-dependent glycolysis. *Journal of agricultural and food chemistry* 2017;65:8136-44. doi: <https://doi.org/10.1021/acs.jafc.7b02757>
- (107) Feng J, Wu L, Ji J, Chen K, Yu Q, Zhang J, et al. Pkm2 is the target of proanthocyanidin b2 during the inhibition of hepatocellular carcinoma. *Journal of Experimental & Clinical Cancer Research* 2019;38:1-15. doi: <https://doi.org/10.1186/s13046-019-1194-z>
- (108) Aslan E, Guler C, Adem S. In vitro effects of some flavonoids and phenolic acids on human pyruvate kinase isoenzyme m2. *Journal of enzyme inhibition and medicinal chemistry* 2016;31:314-7. doi: <https://doi.org/10.3109/14756366.2015.1022173>

- (109) Jia L, Huang S, Yin X, Zan Y, Guo Y, Han L. Quercetin suppresses the mobility of breast cancer by suppressing glycolysis through akt-mtor pathway mediated autophagy induction. *Life sciences* 2018;208:123-30. doi: <https://doi.org/10.1016/j.lfs.2018.07.027>
- (110) Chen J, Xie J, Jiang Z, Wang B, Wang Y, Hu X. Shikonin and its analogs inhibit cancer cell glycolysis by targeting tumor pyruvate kinase-m2. *Oncogene* 2011;30:4297-306. doi: doi:10.1038/onc.2011.137
- (111) Mazlaghaninia M, Atri MS, Seyedalipour B. Scopoletin and morin inhibit lactate dehydrogenase enzyme activity, which is critical for cancer metabolism. *Hormozgan Medical Journal* 2019;23:e88269-e. doi: DOI:10.5812/HMJ.88269
- (112) Wu H, Cui M, Li C, Li H, Dai Y, Cui K, et al. Kaempferol reverses aerobic glycolysis via mir-339-5p-mediated pkm alternative splicing in colon cancer cells. *Journal of Agricultural and Food Chemistry* 2021;69:3060-8.
- (113) Bader A, Tuccinardi T, Granchi C, Martinelli A, Macchia M, Minutolo F, et al. Phenylpropanoids and flavonoids from *phlomis kurdica* as inhibitors of human lactate dehydrogenase. *Phytochemistry* 2015;116:262-8. doi: <https://doi.org/10.1016/j.phytochem.2015.03.007>
- (114) Li S, Wu L, Feng J, Li J, Liu T, Zhang R, et al. In vitro and in vivo study of epigallocatechin-3-gallate-induced apoptosis in aerobic glycolytic hepatocellular carcinoma cells involving inhibition of phosphofructokinase activity. *Scientific reports* 2016;6:28479. doi: <https://doi.org/10.1038/srep28479>
- (115) Gomez LS, Zancan P, Marcondes MC, Ramos-Santos L, Meyer-Fernandes JR, Sola-Penna M, et al. Resveratrol decreases breast cancer cell viability and glucose metabolism by inhibiting 6-phosphofructo-1-kinase. *Biochimie* 2013;95:1336-43. doi: <https://doi.org/10.1016/j.biochi.2013.02.013>
- (116) Dihal AA, van der Woude H, Hendriksen PJ, Charif H, Dekker LJ, IJsselstijn L, et al. Transcriptome and proteome profiling of colon mucosa from quercetin fed f344 rats point to tumor preventive mechanisms, increased mitochondrial fatty acid degradation and decreased glycolysis. *Proteomics* 2008;8:45-61. doi: <https://doi.org/10.1002/pmic.200700364>
- (117) Sellam LS, Zappasodi R, Chettibi F, Djennaoui D, Mesbah NY-A, Amir-Tidadi Z-C, et al. Silibinin down-regulates pd-11 expression in nasopharyngeal carcinoma by interfering with tumor cell glycolytic metabolism. *Archives of biochemistry and biophysics* 2020;690:108479.
- (118) Coricovac D, Dehelean CA, Pinzaru I, Mioc A, Aburel O-M, Macasoi I, et al. Assessment of betulinic acid cytotoxicity and mitochondrial metabolism impairment in a human melanoma cell line. *International Journal of Molecular Sciences* 2021;22:4870. doi: <https://doi.org/10.3390/ijms22094870>
- (119) Peeters TH, Lenting K, Breukels V, van Lith SA, van den Heuvel CN, Molenaar R, et al. Isocitrate dehydrogenase 1-mutated cancers are sensitive to the green tea polyphenol epigallocatechin-3-gallate. *Cancer & Metabolism* 2019;7:1-13. doi: <https://doi.org/10.1186/s40170-019-0198-7>
- (120) Icard P, Loi M, Wu Z, Ginguay A, Lincet H, Robin E, et al. Metabolic strategies for inhibiting cancer development. *Advances in Nutrition* 2021;12:1461-80.
- (121) Wei Z, Ye S, Feng H, Zeng C, Dong X, Zeng X, et al. Silybin suppresses ovarian cancer cell proliferation by inhibiting isocitrate dehydrogenase 1 activity. *Cancer Science* 2022;113:3032-43.
- (122) Frattaruolo L, Brindisi M, Curcio R, Marra F, Dolce V, Cappello AR. Targeting the mitochondrial metabolic network: A promising strategy in cancer treatment. *International journal of molecular sciences* 2020;21:6014.
- (123) Shuvalov O, Kirdeeva Y, Daks A, Fedorova O, Parfenyev S, Simon H-U, et al. Phytochemicals target multiple metabolic pathways in cancer. *Antioxidants* 2023;12:2012.
- (124) Bianchi G, Ravera S, Traverso C, Amaro A, Piaggio F, Emionite L, et al. Curcumin induces a fatal energetic impairment in tumor cells in vitro and in vivo by inhibiting atp-synthase activity. *Carcinogenesis* 2018;39:1141-50. doi: <https://doi.org/10.1093/carcin/bgy076>
- (125) Zheng X, Pan Y, Yang G, Liu Y, Zou J, Zhao H, et al. Kaempferol impairs aerobic glycolysis against melanoma metastasis via inhibiting the mitochondrial binding of hk2 and vdac1. *European journal of pharmacology* 2022;931:175226.
- (126) Brecht K, Riebel V, Couttet P, Paech F, Wolf A, Chibout S-D, et al. Mechanistic insights into selective killing of oxphos-dependent cancer cells by arctigenin. *Toxicology in vitro* 2017;40:55-65. doi: <https://doi.org/10.1016/j.tiv.2016.12.001>

- (127) Abotaleb M, Samuel SM, Varghese E, Varghese S, Kubatka P, Liskova A, et al. Flavonoids in cancer and apoptosis. *Cancers* 2018;11:28. doi: [doi.org/10.3390/cancers11010028](https://doi.org/10.3390/cancers11010028)
- (128) Patel K, Singh GK, Patel DK. A review on pharmacological and analytical aspects of naringenin. *Chinese journal of integrative medicine* 2018;24:551-60. doi: <https://doi.org/10.1007/s11655-014-1960-x>
- (129) Kicinska A, Jarmuszkiewicz W. Flavonoids and mitochondria: Activation of cytoprotective pathways? *Molecules* 2020;25:3060. doi: <https://doi.org/10.3390/molecules25133060>
- (130) Adem S, Comakli V, Kuzu M, Demirdag R. Investigation of the effects of some phenolic compounds on the activities of glucose-6-phosphate dehydrogenase and 6-phosphogluconate dehydrogenase from human erythrocytes. *Journal of biochemical and molecular toxicology* 2014;28:510-4. doi: DOI.10.1002/jbt.21592
- (131) Patrício JFS. Effects of pentose pathway inhibition on glucose metabolism in cell culture models of cancer using stable isotope tracers and nmr. Universidade de Coimbra (Portugal); 2022.
- (132) Khan A, Siddiqui S, Husain SA, Mazurek S, Iqbal MA. Phytochemicals targeting metabolic reprogramming in cancer: An assessment of role, mechanisms, pathways, and therapeutic relevance. *Journal of Agricultural and Food Chemistry* 2021;69:6897-928.
- (133) Lin R, Elf S, Shan C, Kang H-B, Ji Q, Zhou L, et al. 6-phosphogluconate dehydrogenase links oxidative ppp, lipogenesis and tumour growth by inhibiting lkb1-ampk signalling. *Nature cell biology* 2015;17:1484-96. doi: <https://doi.org/10.1038/ncb3255>
- (134) Li P, Tian W, Ma X. Alpha-mangostin inhibits intracellular fatty acid synthase and induces apoptosis in breast cancer cells. *Molecular cancer* 2014;13:1-11. doi: <https://doi.org/10.1186/1476-4598-13-138>
- (135) Funabashi H, Kawaguchi A, Tomoda H, Omura S, Okuda S, Iwasaki S. Binding site of cerulenin in fatty acid synthetase. *The Journal of Biochemistry* 1989;105:751-5. doi: doi.1922.105.5.105\_5\_751
- (136) Lee KH, Lee MS, Cha EY, Sul JY, Lee JS, Kim JS, et al. Inhibitory effect of emodin on fatty acid synthase, colon cancer proliferation and apoptosis. *Molecular medicine reports* 2017;15:2163-73. doi: <https://doi.org/10.3892/mmr.2017.6254>
- (137) Wang X, Tian W. Green tea epigallocatechin gallate: A natural inhibitor of fatty-acid synthase. *Biochemical and biophysical research communications* 2001;288:1200-6. doi: <https://doi.org/10.1006/bbrc.2001.5923>
- (138) Huang CH, Tsai SJ, Wang YJ, Pan MH, Kao JY, Way TD. Egcg inhibits protein synthesis, lipogenesis, and cell cycle progression through activation of ampk in p53 positive and negative human hepatoma cells. *Molecular nutrition & food research* 2009;53:1156-65. doi: <https://doi.org/10.1002/mnfr.200800592>
- (139) Li BH, Tian WX. Inhibitory effects of flavonoids on animal fatty acid synthase. *Journal of biochemistry* 2004;135:85-91. doi: <https://doi.org/10.1093/jb/mvh010>
- (140) Brusselmans K, Vrolix R, Verhoeven G, Swinnen JV. Induction of cancer cell apoptosis by flavonoids is associated with their ability to inhibit fatty acid synthase activity. *Journal of Biological Chemistry* 2005;280:5636-45. doi: <https://doi.org/10.1074/jbc.M408177200>
- (141) Lee M, Sung SH. Platyphylloside isolated from betula platyphylla inhibit adipocyte differentiation and induce lipolysis via regulating adipokines including pparγ in 3t3-l1 cells. *Pharmacognosy Magazine* 2016;12:276. doi: doi: 10.4103/0973-1296.192208
- (142) Khan A, Aljarbou AN, Aldebasi YH, Faisal SM, Khan MA. Resveratrol suppresses the proliferation of breast cancer cells by inhibiting fatty acid synthase signaling pathway. *Cancer epidemiology* 2014;38:765-72. doi: <https://doi.org/10.1016/j.canep.2014.09.006>
- (143) Gao Y, Islam MS, Tian J, Lui VWY, Xiao D. Inactivation of atp citrate lyase by cucurbitacin b: A bioactive compound from cucumber, inhibits prostate cancer growth. *Cancer letters* 2014;349:15-25. doi: <https://doi.org/10.1016/j.canlet.2014.03.015>
- (144) Gnoni GV, Paglialonga G, Siculella L. Quercetin inhibits fatty acid and triacylglycerol synthesis in rat-liver cells. *European Journal of Clinical Investigation* 2009;39:761-8. doi: <https://doi.org/10.1111/j.1365-2362.2009.02167.x>
- (145) Vahlensieck H, Pridzun L, Reichenbach H, Hinnen A. Identification of the yeast acc1 gene product (acetyl-coa carboxylase) as the target of the polyketide fungicide soraphen a. *Current genetics* 1994;25:95-100. doi: <https://doi.org/10.1007/BF00309532>

- (146) Weatherly SC, Volrath SL, Elich TD. Expression and characterization of recombinant fungal acetyl-coa carboxylase and isolation of a soraphen-binding domain. *Biochemical Journal* 2004;380:105-10. doi: <https://doi.org/10.1042/bj20031960>
- (147) Potze L, Di Franco S, Grandela C, Pras-Raves M, Picavet D, Van Veen H, et al. Betulinic acid induces a novel cell death pathway that depends on cardiolipin modification. *Oncogene* 2016;35:427-37. doi: <https://doi.org/10.1038/onc.2015.102>
- (148) Wei L, Zhou Y, Qiao C, Ni T, Li Z, You Q, et al. Oroxylin a inhibits glycolysis-dependent proliferation of human breast cancer via promoting sirt3-mediated sod2 transcription and hif1 $\alpha$  destabilization. *Cell death & disease* 2015;6:e1714-e. doi: <https://doi.org/10.1038/cddis.2015.86>
- (149) Jung K-H, Lee JH, Quach CHT, Paik J-Y, Oh H, Park JW, et al. Resveratrol suppresses cancer cell glucose uptake by targeting reactive oxygen species-mediated hypoxia-inducible factor-1 $\alpha$  activation. *Journal of Nuclear Medicine* 2013;54:2161-7. doi: <https://doi.org/10.2967/jnumed.112.115436>
- (150) Chen F, Zhuang M, Zhong C, Peng J, Wang X, Li J, et al. Baicalein reverses hypoxia-induced 5-fu resistance in gastric cancer ags cells through suppression of glycolysis and the pten/akt/hif-1 $\alpha$  signaling pathway. *Oncology reports* 2015;33:457-63. doi: <https://doi.org/10.3892/or.2014.3550>
- (151) Liu Y, Veena CK, Morgan JB, Mohammed KA, Jekabsons MB, Nagle DG, et al. Methylalpinumisoflavone inhibits hypoxia-inducible factor-1 (hif-1) activation by simultaneously targeting multiple pathways. *Journal of Biological Chemistry* 2009;284:5859-68. doi: DOI:<https://doi.org/10.1074/jbc.M806744200>
- (152) Jiang H, Wu D, Xu D, Yu H, Zhao Z, Ma D, et al. Eupafolin exhibits potent anti-angiogenic and antitumor activity in hepatocellular carcinoma. *International Journal of Biological Sciences* 2017;13:701. doi: doi: 10.7150/ijbs.17534
- (153) Ansó E, Zuazo A, Irigoyen M, Urdaci MC, Rouzaut A, Martínez-Irujo JJ. Flavonoids inhibit hypoxia-induced vascular endothelial growth factor expression by a hif-1 independent mechanism. *Biochemical pharmacology* 2010;79:1600-9.
- (154) Sun Y, Wang H, Liu M, Lin F, Hua J. Resveratrol abrogates the effects of hypoxia on cell proliferation, invasion and emt in osteosarcoma cells through downregulation of the hif-1 $\alpha$  protein. *Molecular medicine reports* 2015;11:1975-81.
- (155) Wang R-b, Zhou S-s, Li S-s. Cancer therapeutic agents targeting hypoxia-inducible factor-1. *Current medicinal chemistry* 2011;18:3168-89.
- (156) Nenkov M, Ma Y, Gaßler N, Chen Y. Metabolic reprogramming of colorectal cancer cells and the microenvironment: Implication for therapy. *International Journal of Molecular Sciences* 2021;22:6262. doi: <https://doi.org/10.3390/ijms22126262>
- (157) Fan W-h, Wang F-c, Jin Z, Zhu L, Zhang J-x. Curcumin synergizes with cisplatin to inhibit colon cancer through targeting the microrna-137-glutaminase axis. *Current Medical Science* 2022;42:108-17. doi: <https://doi.org/10.1007/s11596-021-2469-0>
- (158) Wang G, Wang Y-Z, Yu Y, Yin P-H, Xu K. The antitumor activity of betulinic acid-loaded nanoliposomes against colorectal cancer in vitro and in vivo via glycolytic and glutaminolytic pathways. *Journal of Biomedical Nanotechnology* 2020;16:235-51. doi: <https://doi.org/10.1166/jbn.2020.2888>
- (159) Pournourmohammadi S, Grimaldi M, Stridh MH, Lavallard V, Waagepetersen HS, Wollheim CB, et al. Epigallocatechin-3-gallate (egcg) activates ampk through the inhibition of glutamate dehydrogenase in muscle and pancreatic  $\beta$ -cells: A potential beneficial effect in the pre-diabetic state? *The international journal of biochemistry & cell biology* 2017;88:220-5. doi: <https://doi.org/10.1016/j.biocel.2017.01.012>
- (160) Hay N. Reprogramming glucose metabolism in cancer: Can it be exploited for cancer therapy? *Nature Reviews Cancer* 2016;16:635-49. doi: <https://doi.org/10.1038/nrc.2016.77>