Anti-allergic effect of the naturally-occurring conjugated linolenic acid isomer, jacaric acid, on the activated human mast cell line-1

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Abstract. The present study aimed to investigate the immunomodulatory effect of jacaric acid, a naturally-occurring conjugated linolenic acid isomer that can be found in jacaranda seed oil, on the activated human mast cell line-1 (HMC-1). Our previous studies have demonstrated that jacaric acid only exerted minimal, if any, cytotoxicity on normal murine cells. In the present study, jacaric acid at concentrations ≤100 µM did not exhibit direct cytotoxicity on human peripheral blood mononuclear cells after 72 h of incubation, as determined by the MTT reduction assay. By contrast, jacaric acid could alleviate the calcium ionophore A23187 and phorbol 12-myristate 13-acetate-triggered allergic response in the HMC-1 cells at concentrations that were non-cytotoxic to the HMC-1 cells. Following pretreatment with jacaric acid, the secretion of two inflammatory mediators, β-N-acetylglucosaminidase and tryptase, as well as the T helper 2 cytokines [interleukin (IL)-4 and IL‑13] was significantly reduced in HMC-1 cells. The alleviation of allergic response was accompanied by downregulation of the matrix metalloproteinase-2 and -9 proteins and upregulation of the tissue inhibitor of metalloproteinase-1 protein. Collectively, the results indicated that the naturally-occurring jacaric acid exhibits a suppressive effect on the allergic response in activated human mast cells in vitro, and this could not be attributed to the direct cytotoxicity of jacaric acid on the treated cells.

Introduction

Conjugated fatty acids (CFAs) refer to the positional and geometric isomers of polyunsaturated FAs with conjugated double bonds, in which 2 carbon-carbon double bonds are separated solely by one carbon-carbon single bond (1). Common CFA isomers, such as conjugated linoleic acids (CLAs) and conjugated linolenic acids (CLNAs), can be found naturally in meat or dairy products of ruminant animals (2) and different plant seed oils (3), respectively. CLAs have been studied most extensively due to their diverse metabolic and physiological effects (4,5). In recent years, CLNAs have received increasing attention as their relative abundance in certain plant seed oils (30-70% of total lipids) was much higher compared to that of CLAs (<1% of total lipids) (3,6). Previous in vitro and in vivo studies have demonstrated that CLNAs exhibit pleiotropic physiological and pharmacological activities, including anti-carcinogenic, anti-inflammatory, anti-obese, antioxidative and immunomodulatory properties (7,8). An earlier study by Ike et al (9) showed that α-eleostearic acid (9Z, 11E, 13E-CLNA isomer) isolated from bitter gourd could induce interferon-γ production in mice treated with heat-inactivated Propionibacterium acnes, suggesting that T helper 1 (Th1) cellular immunity can be activated by α-eleostearic acid, which is responsible for the defense against intracellular parasitic infection. By contrast, Yamasaki et al (10) reported that consumption of pomegranate seed oil that is rich in punicic acid (9Z, 11E, 13Z-CLNA isomer) could ameliorate the function of B cells, which have a key role in the humoral immune response. These studies started to explain the immunostimulatory activities of CLNAs, however, the anti-allergic activity of CLNAs and their modulatory effects on mast cells have not yet been investigated. In the present study, jacaric acid (8Z, 10E, 12Z-CLNA isomer; Fig. 1) alleviated the allergic response in calcium ionophore A23187 (Iono) and phorbol 12-myristate 13-acetate (PMA)-activated human mast cells by reducing the release of inflammatory mediators and Th2 cytokines, and by modulating the protein expression levels of matrix metalloproteinases (MMPs).

Materials and methods

Chemicals and reagents. Jacaric acid (8Z, 10E, 12Z-CLNA isomer) used in the study, with an estimated purity >97%, was purchased from Larodan Fine Chemicals AB (Limhamn, Sweden). The stock solution (0.2 M) was prepared by dissolving the powder in sterile, cell culture-tested ethanol.
A total of 50 µl of 0.09 M sodium citrate solution (pH 4.7) was transferred to the well of a flat-bottomed 96-well plate. Following incubation, the stimulated cells were centrifuged at 2 µM Iono and 40 nM PMA at 37˚C for 6 h in a 6-well plate. The cell-free supernatant was transferred to a flat-bottomed 96-well microtiter plate and the treated cells (3x10⁶) were pre-treated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks. Following incubation, cells were centrifuged at 400 x g for 5 min, washed with phosphate-buffered saline three times and 3x10⁶ cells were transferred to a flat-bottomed 6-well microtiter plate in 1 ml complete RPMI medium, and were activated with 2 µM Iono and 40 nM PMA at 37˚C for 6 h for the further assays.

Cytotoxicity assay. Human peripheral blood mononuclear cells (PBMCs) were prepared from the fresh human buffy coats supplied by the Hong Kong Red Cross Blood Transfusion Service (Kowloon, Hong Kong, SAR, China). The PBMCs used were a generous gift from Professor C.K. Wong of the Department of Chemical Pathology, The Chinese University of Hong Kong (Shatin, Hong Kong, SAR, China) (11). All other chemicals, unless otherwise stated, were purchased from Sigma-Aldrich.

Culture of cell line. Human mast cell line-1 (HMC-1) was a generous gift from Professor C.K. Wong of the Department of Chemical Pathology, The Chinese University of Hong Kong (Shatin, Hong Kong, SAR, China) (11). The cells were maintained in RPMI-1640 medium (Gibco, Thermo Fisher Scientific, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (Gibco, Thermo Fisher Scientific) and 1% antibiotics (100 U/ml penicillin G, 100 µg/ml streptomycin sulfate and 0.25 µg/ml amphotericin B in 0.85% saline from Gibco, Thermo Fisher Scientific) in a humidified incubator containing 5% CO₂ in air at 37˚C.

The procedures for the stimulation of HMC-1 cells were modified from a previously described method (12). In brief, HMC-1 cells (3x10⁶ cells/ml) were pre-treated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks. Following incubation, cells were centrifuged at 400 x g for 5 min, washed with phosphate-buffered saline three times and 3x10⁶ cells were transferred to a flat-bottomed 6-well microtiter plate in 1 ml complete RPMI medium, and were activated with 2 µM Iono and 40 nM PMA at 37˚C for 6 h for the further assays.

Measurement of tryptase release in HMC-1 cells. The assessment was performed according to the manufacturer’s instructions in the Mast Cell Degranulation Assay kit (Millipore Corp., Billerica, MA, USA). HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by Iono and PMA at 37˚C for 6 h in a 6-well plate. Following incubation, the stimulated cells were centrifuged at 400 x g for 5 min, and 180 µl cell-free supernatant was transferred to the well of a flat-bottomed 96-well plate. A total of 20 µl of tryptase substrate was added to each well and the samples were incubated at 37˚C for 2 h. The absorbance at 405 nm was measured by the Benchmark microplate reader and the concentrations of tryptase in the samples were quantified with reference to a series of tryptase positive control solutions with concentrations ranging from 0 to 10,000 ng/ml.

Assessment of cytokine secretion by ELISA. To determine the secretion of interleukin (IL)-4 and IL-13, the corresponding ELISA kit was used (ExCell Biology, Inc., Shanghai, China). Briefly, HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by 2 µM Iono and 40 nM PMA at 37˚C for 6 h in a 6-well plate. The cell-free supernatant was transferred to another 96-well plate provided in the ELISA kit. Subsequently, the absorbance at 405 nm was measured by the Benchmark microplate reader.

Western blotting. Protein expression levels were determined by the western blotting technique with the aid of a panel of specific antibodies. HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated HMC-1 cells were stained with 1 mg/ml 4-nitrophenyl-N-acetyl-β-D-glucosaminidase and color development. The enzymatic reaction was terminated by adding 200 µl of sodium carbonate solution [0.4 M (pH 11.6)] and the absorbance at 405 nm was measured by the Benchmark microplate reader. The concentrations of NAG in the samples were expressed in U/ml, in which 1 unit of NAG will hydrolyze 1 nanomole of 4-nitrophenyl-N-acetyl-β-D-glucosaminidase to p-nitrophenol and N-acetyl-β-D-glucosaminide per min at pH 4.7 and 37˚C. The percentage of NAG release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) x100.

Measurement of β-N-acetylglucosaminidase (NAG) release in HMC-1 cells. The measurement was performed according to the manufacturer’s instructions in the NAG assay kit (Sigma-Aldrich) with slight modifications. Briefly, HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by 2 µM Iono and 40 nM PMA at 37˚C for 6 h in a 6-well plate. Following incubation, the stimulated cells were centrifuged at 400 x g for 5 min, and 50 µl cell-free supernatant or cell lysate was transferred to the well of a flat-bottomed 96-well plate. A total of 50 µl of 0.09 M sodium citrate solution (pH 4.7) containing 1 mg/ml 4-nitrophenyl-N-acetyl-β-D-glucosaminidase was added to the wells and incubated at 37˚C for 30 min for color development. The enzymatic reaction was terminated by adding 200 µl of sodium carbonate solution [0.4 M (pH 11.6)] and the absorbance at 405 nm was measured by the Benchmark microplate reader. The concentrations of NAG in the samples were expressed in U/ml, in which 1 unit of NAG will hydrolyze 1 nanomole of N-acetyl-β-D-glucosaminide to p-nitrophenol and N-acetyl-β-D-glucosaminide per min at pH 4.7 and 37˚C. The percentage of NAG release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) x100.

Cytokine secretion by ELISA. To determine the secretion of interleukin (IL)-4 and IL-13, the corresponding ELISA kit was used (ExCell Biology, Inc., Shanghai, China). Briefly, HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated cells (3x10⁶) were further stimulated by 2 µM Iono and 40 nM PMA at 37˚C for 6 h in a 6-well plate. The cell-free supernatant was transferred to another 96-well plate provided in the ELISA kit. Subsequently, the absorbance at 405 nm was measured by the Benchmark microplate reader.

Western blotting. Protein expression levels were determined by the western blotting technique with the aid of a panel of specific antibodies. HMC-1 cells (3x10⁶ cells/ml) were incubated with different concentrations of jacaric acid at 37˚C for 72 h in 175-cm² tissue culture flasks and the treated HMC-1 cells were stained with 1 mg/ml 4-nitrophenyl-N-acetyl-β-D-glucosaminidase and color development. The enzymatic reaction was terminated by adding 200 µl of sodium carbonate solution [0.4 M (pH 11.6)] and the absorbance at 405 nm was measured by the Benchmark microplate reader. The concentrations of NAG in the samples were expressed in U/ml, in which 1 unit of NAG will hydrolyze 1 nanomole of 4-nitrophenyl-N-acetyl-β-D-glucosaminidase to p-nitrophenol and N-acetyl-β-D-glucosaminide per min at pH 4.7 and 37˚C. The percentage of NAG release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) x100.
transferred to a 6-well plate and sensitized in the presence of 2 µM Iono and 40 nM PMA at 37°C for 6 h. After incubation, the cell pellet was collected and total protein was extracted by the cell lysis buffer. Protein concentration was measured by the Bradford reagent and the protein sample was resolved on 12% polyacrylamide gels and transferred to polyvinylidene difluoride membranes. The membrane was first incubated with the following primary antibodies: Rabbit anti-MMP-2 (CST-4022S), anti-MMP-9 (CST-3852S), anti-tissue inhibitor of metalloproteinase-1 (TIMP-1) (CST-9846S) (Cell Signaling Technology, Inc., Danvers, MA, USA) and mouse anti-β-actin antibody (SC-A5331) (Santa Cruz Biotechnology, Inc., Dallas, TX, USA), followed by incubation with mouse IgG horseradish peroxidase-conjugated secondary antibody (GE-NA934) (GE Healthcare, Buckinghamshire, UK) and finally developed with the enhanced chemiluminescence reagent (Santa Cruz Biotechnology, Inc.).

Statistical analysis. Each experiment was repeated at least three times and only the results of the most representative experiments are shown. The data are expressed as the arithmetic mean ± standard error. One-way analysis of variance was used for statistical analysis, and P<0.05 was considered to indicate a statistically significant difference.

Results

Jacaric acid exhibits no direct cytotoxicity on HMC-1 cells and PBMCs. To investigate whether jacaric acid was cytotoxic to the HMC-1 cells or PBMCs, the colorimetric MTT reduction assay was employed. Fig. 2A shows that jacaric acid at concentrations ≤ 8 µM exhibited no significant cytotoxicity to the HMC-1 cells (>90% cell viability) after 72 h of incubation when compared with the solvent control. Additionally, the viability of jacaric acid-treated HMC-1 cells following Iono- and PMA-activation was examined by the trypan blue exclusion assay and by the annexin V-GFP/PI dual staining assay. As shown in Fig. 2B and C, the cell viability of HMC-1 cells and the percentage of the necrotic cells respectively did not differ significantly between the jacaric acid-treated group and the solvent-treated control. Notably, the viability of the PBMCs remained ≥90% when the cells were incubated with ≤100 µM jacaric acid for 6, 24, 48 and 72 h (Fig. 3), suggesting that jacaric acid was neither cytotoxic to the HMC-1 cells, nor to the PBMCs under the prescribed experimental conditions.

Jacaric acid suppresses the release of NAG and tryptase from HMC-1 cells. Mast cells have a well-known central role in allergic response by releasing pharmacologically active inflammatory mediators, which cause the symptoms of allergic inflammatory reaction (15). Common inflammatory mediators, including NAG and tryptase, are potent markers of allergic disease (16). In the present study, the release of NAG and tryptase in HMC-1 cells was increased by 2-fold upon activation with Iono and PMA (data not shown), suggesting that the combined use of Iono and PMA can elicit an allergic response. To investigate whether jacaric acid can exhibit an anti-allergic response, jacaric acid-treated HMC-1 cells were stimulated by 2 µM Iono and 40 nM PMA, and the release of NAG and tryptase from HMC-1 cells was determined. As shown in Table I, pre-treatment of HMC-1 cells with jacaric acid at 4 µM could significantly suppress the secretion of NAG and tryptase in HMC-1 cells.

Table I. Jacaric acid suppresses β-N-acetylgalactosaminidase and tryptase release in HMC-1 cells.

<table>
<thead>
<tr>
<th>Concentration of jacaric acid, µM</th>
<th>NAG, %</th>
<th>Tryptase, U/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>31.80±3.82</td>
<td>503.93±25.73</td>
</tr>
<tr>
<td>2</td>
<td>25.85±2.02</td>
<td>496.38±26.46</td>
</tr>
<tr>
<td>4</td>
<td>16.32±1.02</td>
<td>408.85±20.38</td>
</tr>
</tbody>
</table>

Percentage of β-N-acetylgalactosaminidase (NAG) release was calculated as follows: % NAG release = (concentration of NAG in supernatant)/(concentration of NAG in supernatant and whole cell lysate) x100. *P<0.001. HMC-1, human mast cell line-1.
from sensitized HMC-1 cells. To confirm the anti-allergic effect of jacaric acid, the release of tryptase in HMC-1 cells was assessed by the Mast Cell Degranulation Assay kit. Jacaric acid could also inhibit the release of tryptase from sensitized HMC-1 cells (Table I). These results suggest that jacaric acid could alleviate the allergic response in sensitized HMC-1 cells.

Discussion

Mast cells are the central effector cells in allergic response and they are also involved in the defense against pathogens, typically by cross-linking of surface immunoglobulin E (IgE) receptors, or through IgE-independent mechanisms.
in non-pathogenic conditions (15,18). The HMC-1 cell line was derived from a patient with mastocytosis by Butterfield et al. (19) in 1988, which shows numerous characteristics of immature mast cells and contains classical mast cell-associated markers, such as the c-kit receptor, heparin, histamine and tryptase, and secretes a panel of bioactive cytokines and a variety of MMP (18). In the present study, the modulatory effects and the underlying action mechanisms of jacaric acid on the human mast cells using the HMC-1 cells as the in vitro model were examined. Pre-treatment of HMC-1 cells with jacaric acid at lower concentrations (<5 µM) could reduce the secretion of inflammatory mediators, including NAG and tryptase, when the HMC-1 cells were activated with Iono and PMA. The presence of allergens or stimulants elicits an inflammatory response, which leads to the increased production and release of Th2 cytokines, such as IL-4, IL-5 and IL-13, in mast cells (17). The present results showed that the release of IL-4 and IL-13 was suppressed following the pre-treatment of activated HMC-1 cells with jacaric acid. To further elucidate the anti-allergic effect of jacaric acid on HMC-1 cells, the expression levels of MMP-2, MMP-9 and TIMP-1 proteins were examined. MMPs belong to a family of Zn-dependent endopeptidases that are involved in the decomposition of extracellular matrix and basement membrane (20). Furthermore, MMP-2, MMP-9 and MMP-13 have a key role in tissue remodeling and repair through degradation of type IV collagen, which is the major component of the basement membrane (21). By contrast, the activation of MMP is inhibited by TIMP, and previous studies have suggested that MMP-2 and MMP-9 are inactivated by TIMP-1 and TIMP-2 (22,23). By western blotting, jacaric acid-treated HMC-1 cells showed an increase in the TIMP-1 protein expression level, accompanied by a decrease in the expression levels of MMP-2 and MMP-9 proteins. To the best of our knowledge, this is the first study demonstrating the in vitro anti-allergic effect of CLNAs on HMC-1 cells, and this is in line with earlier findings showing that CFAs, such as CLAs, may possess anti-allergic activities. For example, Jaudszus et al. (24) reported that 9Z, 11E-CLA could inhibit allergic sensitization and airway inflammation through peroxisome proliferator-activated receptor-γ dependent mechanism in mice, whereas others demonstrated that human healthy volunteers supplemented with 9Z, 11E-CLA resulted in a decreased plasma IgE level and reduced secretion of IL-5 by PBMCs (25,26). Despite these earlier findings, the underlying action mechanisms for the anti-allergic effects of CFAs, such as CLAs and CLNAs, remain poorly understood and further studies are required to elucidate the mechanisms, in vitro and in vivo, for the anti-allergic activity of CFAs. In addition, whether CLNA supplementation could ameliorate other types of hypersensitivity reactions, such as the ex vivo delayed-type hypersensitivity response as shown by CLAs (25), is also a noteworthy aspect that is currently under investigation.

Collectively, the present results suggest that jacaric acid may exhibit modulatory effects on human mast cells, as it was shown to alleviate the allergic response in HMC-1 cells activated by Iono and PMA. As jacaric acid was found to exert minimal direct cytotoxicity on normal human PBMCs, and apparently was non-toxic to mice (27), further elucidation of the immunomodulatory effects of jacaric acid, in vitro and in vivo, may provide better insights for the development of jacaric acid as a potential candidate for the treatment of certain allergic disorders with minimal toxicity and fewer side effects.
References