Abstract. The aim of the present study was to investigate the effect of electroacupuncture (EA) on cognitive deficits, and the underlying mechanism following cerebral ischemia-reperfusion (I/R) via the calmodulin (CaM)-calmodulin-dependent protein kinase type IV (CaMKIV)-cyclic adenosine monophosphate response elements binding protein (CREB) intracellular signaling pathway in the hippocampus. In total, 45 adult female Sprague-Dawley rats were randomly divided into three groups, namely the sham group, the middle cerebral artery occlusion (MCAO) group and the MCAO + EA group. Rats in the MCAO and MCAO + EA groups were modeled for post-stroke cognitive impairment. EA was performed at the Baihui and Shenting acupuncture points for 30 min/day for one week in the MCAO + EA group. Behavioral testing was analyzed using a step-down apparatus, while 2,3,5-triphenyltetrazolium chloride was used to detect the infarct volume and lesion size. In addition, CaM activity was assessed by cyclic nucleotide-dependent phosphodiesterase analysis, and the protein expression levels of CaM, CaMKIV, phosphorylated (p)-CaMKIV, CREB and p-CREB were analyzed by western blot analysis. The cerebral I/R injured rat model in the MCAO group was established successfully with regard to the infarct volume and neuronal lesion size, as compared with the sham group. EA was demonstrated to effectively improve the cognitive ability, as measured by the step-down apparatus test, and decrease the infarct volume when compared with the MCAO group (P<0.05). The step-down apparatus test for the EA-treated rats revealed improved learning and reduced memory impairment when compared with the MCAO group. Furthermore, CaM activity and CaM protein expression levels in the MCAO + EA group were lower compared with those in the MCAO group (P<0.05). By contrast, the protein expression levels of CaMKIV, p-CaMKIV, CREB and p-CREB were significantly reduced in the MCAO group when compared with the sham group (P<0.05), although the expression levels increased following EA treatment when compared with the MCAO group (P<0.05). Therefore, cognitive repair benefited from EA, and the main intracellular signaling pathway in the hippocampus was mediated by CaM-CaMKIV-CREB. EA effectively inhibited the expression and activity of CaM, while further enhancing the expression of CaMKIV and CREB, and their associated phosphorylated functions.

Introduction

Electroacupuncture (EA) is a therapy for the treatment of neurological dysfunction that functions by stimulating specific areas in the body. This therapeutic method originated in China >2,000 years ago, and over time EA has been accepted as conventional medicine in clinical practice worldwide (1-4). Statistical analyses of clinical results have indicated that the effects of EA on strokes are significant (5). Stroke is a major cause of several complications, including cognitive impairment, with ~25% of patients suffering from cognitive impairment three months after a stroke (6). In addition, up to 75% of stroke survivors may be considered to have selective cognitive impairment, which commonly involves memory, orientation, language and attention (7).
Cognitive rehabilitation and medication have been used to enhance cognition in patients who have had a stroke (8,9); however, there is no one reliable method or medication that has been demonstrated in clinical practice. An increasing number of clinical trials have revealed that acupuncture exhibits positive effects following stroke, not only as a complementary and alternative medicine for post-stroke rehabilitation, but also as a preventative strategy that may induce cerebral ischemic tolerance (10-12).

Although the detailed mechanism underlying cognitive impairment remains unclear, neuronal excitotoxicity, the over-release of toxic neural transmitters and neuronal apoptosis have been demonstrated to contribute to the pathological process (8,9). Neuronal excitotoxicity is triggered by intrinsic or extrinsic stimuli, which eventually result in the activation of caspases and nucleases, subsequently causing cell destruction (13).

Excitotoxicity via calcium-permeable glutamate receptors is considered to be a critical trigger in ischemia-induced brain damage. Experimental models have revealed that excitotoxicity may be inhibited using glutamate receptor antagonists, such as the non-competitive N-methyl-D-aspartate type glutamate receptor (NMDAR) antagonist, MK-801 (14). In contrast to an NMDA blockade, the inhibition of specific postsynaptic NMDAR signaling by preconditioning neuroprotection may induce neuroprotection against cerebral ischemia-reperfusion (I/R) injury. Preconditioning has been associated with increased phosphorylation of cyclic adenosine monophosphate (cAMP) response element-binding (CREB) protein (15) and CREB-dependent transcription (16). Phosphorylation of CREB at Ser133 is involved in the regulation of neuronal plasticity and memory formation, and is required for glutamate- and Ca2+-dependent neuronal survival during development (17). In neurons of the central nervous system, CREB phosphorylation is induced by the synaptic activation of NMDARs, which occurs downstream of Ca2+/CaM-dependent protein kinase (CaMK) activation (18). CaMKII and CaMKIV are able to regulate CREB activity; however, CaMKIV is specifically associated with the activation and transcription of trophic CREB (19). CaMKIV is a nuclear serine/threonine kinase that phosphorylates CREB at Ser133 and its transcription partner, CREB binding protein (20), thereby activating trophic gene transcription. Therefore, these observations indicate that CaM, CaMKIV and CREB activation are important for neuronal survival.

A previous study demonstrated that EA directly affected the biochemical materials associated with neuronal survival, including Ca2+, glutamate and NMDA; however, the detailed mechanisms underlying the intracellular signaling pathway in the hippocampus are yet to be fully elucidated (21). In the present study, EA was hypothesized to improve cognitive impairment in cerebral I/R-injured rats by adjusting the CaM-CaMKIV-CREB intracellular signaling function in the hippocampus.

**Materials and methods**

**Animals.** In total, 45 female Sprague Dawley rats (weight, 270±20 g) were obtained from Shanghai SLAC Laboratory Animal Co. Ltd. (Shanghai, China) and housed in the Animal Center of Fujian University of Traditional Chinese Medicine (TCM; Fuzhou, China). All the animals were housed under pathogen-free conditions with a 12-h light/dark cycle and free access to food and water. The experimental protocol was approved by the Institutional Animal Care and Use Committee of Fujian University of TCM.

The 45 rats were randomly divided into three groups, which included the sham group (rats underwent sham surgery), the middle cerebral artery occlusion (MCAO) group (rats underwent MCAO) and the MCAO + EA group (rats underwent MCAO and received EA intervention).

**Establishment of the cerebral I/R injure rat model.** Following overnight fasting, the rats were anesthetized with 10% chloral hydrate (3 ml/kg; Sigma-Aldrich, St. Louis, MO, USA) through intraperitoneal injection. Subsequently, 0.100 0.149 mm nylon surgical thread (Wego Holding Co., Ltd., Weihai, China) was inserted into the left internal carotid artery to block the middle cerebral artery when the blunted distal end met resistance. Following 2 h of occlusion, the thread was removed to allow complete reperfusion of the ischemic area. A sham procedure was carried out as aforementioned, without the occlusion of the middle cerebral artery (21,22).

**EA intervention.** At 2 h after the surgery, rats in the MCAO + EA group received EA treatment for 30 min. The complete treatment period was 7 days. Baihui and Shenting acupuncture points in the governing vessel were selected for this study. Needles (0.3 mm diameter) were inserted to a depth of 2-3 mm and connected with the EA device (Model G6805; SMIF, Shanghai, China) with a dispersive wave of 1 and 20 Hz.

**Step-down avoidance test.** In the step-down inhibitory avoidance task, a rat is placed on an elevated platform in a dark compartment (20x20x60 cm), adjacent to a wall of an arena. When the rat steps down and places four paws onto the arena-floor grid, the rat receives a mild foot shock (36 V) and learns to associate the exploration of the arena with the punishment. On subsequent exposure to the same environment, the animal may increase the latency prior to ‘stepping down’ onto the floor grid, or may avoid stepping down. Rats were habituated to the handling procedure on the day prior to the test for 3 min. Following any intervention, the rats were placed onto the platform again. The first time period spent prior to stepping down onto the grid (latency period) and the frequency (number of errors) of stepping down from the platform within 3 min were recorded. In cases where the rats did not step-down from the platform within 3 min, the number of errors was recorded as ‘0’, and the latency period was ‘180 sec’. Step-down latencies and errors were recorded as a measure of memory retention (23-25).

**Histopathological staining with 2,3,5-triphenyl tetrazolium chloride (TTC).** Rats from each group were decapitated following anesthetization with 10% chloral hydrate (3 ml/kg). The brains were immediately removed and placed in ice/water at −20°C for 15 min to ensure rigidity. Subsequently, the brains were cut into coronal sections of 2-mm thickness at the middle of the connecting line between the prefrontal cortex and the optic chiasma, after which the samples were immersed in
2% TTC (Sigma-Aldrich) (to avoid light) for 15 min (37°C) and treated with 4% parafomaldehyde (Sigma-Aldrich) for fixation for 24 h. Finally, images of the brains were captured by camera (SX20; Canon, Inc., Tokyo, Japan). Analysis of the ischemia cerebral damage was performed as described previously (22,26).

Phosphodiesterase (PDE) activity. A modified three-step PDE-1 assay was used to determine the CaM-dependent activation of PDE-1. In the PDE-1 assay, 3,5-cyclic-nucleotide PDE, 2-mM cAMP and 100 μM CaM (Sigma-Aldrich) in 0.5 ml Tris buffer solution (40 mM Tris-chloride, 0.1 mM MnCl₂, and 0.01 mM CaCl₂ in distilled water at pH 7.5) were incubated with increasing concentrations of native tehranolide (1x10⁻⁶-9x10⁻⁴ M) for 10 min at 30°C. The reaction was stopped by placing the test tubes in a boiling water bath for 2 min, and then cooling. The 5-AMP in the reaction product was cleaved into adenosine and inorganic phosphate by incubation with 5'-nucleotidase (100 µl; Sigma-Aldrich) for 10 min, and the reaction was terminated by adding 0.05 ml trichloroacetic acid (55% w/v) and centrifuging (10,800 x g) until clear. The clear supernatants were decanted into test tubes with Fiske-Subbarow reagent (Sigma-Aldrich). A blue color reaction was allowed to develop in the presence of inorganic phosphorus for 10 min, and the absorbance was measured at 660 nm using spectrophotometry (SmartSpec Plus; Bio-Rad Laboratories, Inc.) (30).

Statistical analysis. Data are presented as the mean ± standard error of the mean. Statistical comparisons were conducted by one way analysis of variance using the SPSS software package, version 18.0 (SPSS, Inc., Chicago, IL, USA), where P<0.05 was considered to indicate a statistically significant difference.

Results

Effect of EA on the step-down avoidance test and infarct volume. As shown in Fig. 1, MCAO markedly affected the memory of the rats, while EA treatment was shown to successfully repair this ability. The step-down latency of the MCAO group rats was significantly shorter when compared with the sham group (P<0.05), and was prolonged by EA (P<0.05). During the 3-min test, the number of errors in the MCAO group rats was significantly fewer when compared with the sham group and MCAO + EA group rats (Fig. 1; P<0.05).

With regard to the infarct volume analyses, EA was revealed to significantly reduce the infarct volume caused by cerebral I/R. The sham group rats exhibited no trauma in the brain, while the MCAO group rats exhibited a large infarct area (23.98±5.04%; P<0.05), which was significantly decreased following EA (15.71±3.16%; P<0.05; Fig. 2).

Effect of EA on the levels of CaM activity and CaM protein expression. Notably, MCAO was found to promote CaM activity with 5% non-fat milk for 2 h, and subsequently incubated with primary antibodies (1:1,000 dilution) against CaM (cat. no. sc-137079; Santa Cruz Biotechnology, Inc., Dallas, TX, USA), CaMKIV (cat. no. 4032; Cell Signaling Technology, Inc., Danvers, MA, USA), p-CaMKIV (cat. no. sc-28443-R; Santa Cruz Biotechnology, Inc.), CREB (cat. no. 9197; Cell Signaling Technology, Inc.), p-CREB (cat. no. 9196; Cell Signaling Technology, Inc.) and β-actin (cat. no. 4970; Cell Signaling Technology, Inc.) overnight at 4°C. Next, the blots were incubated with a horseradish peroxidase-conjugated anti-rabbit or anti-mouse secondary antibodies (1:5,000; cat. nos. 7074 and 7076; Cell Signaling Technology, Inc.) for 50 min. β-actin was used as a loading control. The blots were developed using a commercially available enhanced chemiluminescence kit (Bio-Rad Laboratories, Inc.), and examined using a Bio-Image Analysis System (ChemiDoc™ Imaging systems; Bio-Rad Laboratories, Inc., Hercules, CA, USA) (27-29).

Western blot analysis for the determination of CaM, CaMKIV, phosphorylated (p)-CaMKIV, CREB and p-CREB protein expression levels. Hippocampi from three groups were homogenized in non-denaturing lysis buffer (20 mM Tris (pH 7.5), 150 mM NaCl, 1% Triton X-100, 1% NP-40, 2 mM sodium pyrophosphate, 25 mM β-glycerophosphate, 1 mM EDTA, 1 mM Na₂VO₃, 0.5 μg/ml leupeptin) and centrifuged at 12000 x g for 15 min. The supernatants were collected and frozen at -80°C prior to immunoblotting. Protein concentration was determined using a Bio–Image Analysis System (ChemiDoc™ Imaging Systems; Bio-Rad Laboratories, Inc.). In total, a 50-μg protein sample obtained from the CA1 region of the hippocampus was loaded onto a 12% SDS-PAGE gel. Following electrophoresis, the proteins were electroblotted onto polyvinylidene difluoride membranes (Sigma-Aldrich). The blots were blocked...
According to the PDE analyses, CaM activity was promoted by MCAO and inhibited by EA (P<0.05), and the same result was observed for CaM protein expression (P<0.05; Fig. 3).

Figure 2. Histopathological examinations in each group. (A) Pathological features of cerebral ischemia, where red tissues indicate normal tissue and white sections indicate infarction. (B) Quantitative analysis of the infarct volume comparison between the MCAO and MCAO + EA groups. *P<0.05, vs. MCAO group. MCAO, middle cerebral artery occlusion; EA, electroacupuncture.

Figure 3. (A) Activity levels and (B) protein expression levels of CaM in each group. (C) Quantitative analyses of the protein expression levels of CaM, which were in accordance with the western blot analysis results. *P<0.05, vs. sham group; *P<0.05, vs. MCAO group. CaM, calmodulin; MCAO, middle cerebral artery occlusion; EA, electroacupuncture.

Figure 4. (A) Protein expression levels of CaMKIV, p-CaMKIV, CREB and p-CREB in each group. (B) Quantitative analyses of the protein expression levels of CaMKIV, p-CaMKIV, CREB and p-CREB, which were in accordance with the western blot analysis results. *P<0.05, vs. sham group; *P<0.05, vs. MCAO group. CaM, calmodulin; MCAO, middle cerebral artery occlusion; EA, electroacupuncture; CaMKIV, calmodulin-dependent protein kinase type IV; CREB, cyclic adenosine monophosphate response elements binding protein; p-CREB, phosphorylated CREB; p-CaMKIV, phosphorylated CaMKIV.

Effect of EA on the protein expression levels of CaMKIV, p-CaMKIV, CREB and p-CREB. Protein expression levels of CaMKIV, p-CaMKIV, CREB and p-CREB were shown to decrease following MCAO and increase with EA treatment.
MCAO severely inhibited the protein expression of CaMKIV and pCaMKIV in the hippocampus (P<0.05), while EA repaired the expression of these proteins and promoted CaMKIV and CREB phosphorylation (P<0.05). Furthermore, CREB and p-CREB presented a similar variation trend where MCAO reduced CREB and p-CREB expression (P<0.05), while EA promoted their expression (P<0.05; Fig. 4).

Discussion

Therapeutics in clinical stroke treatment has led researchers to question the feasibility of neuroprotection. Novel insights into the cellular events responsible for neuronal death and an improved understanding of the toxic and trophic roles of excitatory neurotransmission are creating new avenues for therapeutic research. The presence of protective signaling cascades downstream of NMDAR activation, such as enhanced antioxidant defenses, results in the suppression of proapoptotic signaling and the maintenance of trophic signal events (31).

In the present study, according to the pathological features of cerebral ischemia, where red tissues indicate normal tissue and white sections indicate infarction, EA was demonstrated to reduce the infarct volume. As the infarct volume decreased, the behavior study was estimated by the step-down avoidance test. Using this test in previous studies (23,24,32), EA was shown to improve the memories of rats following stroke. Previous clinical studies and meta-analyses have demonstrated that EA exerts a positive effect on cognitive function when compared with no acupuncture, medicine or rehabilitation (11,12,33). Furthermore, a series of clinical trials have shown that acupuncture regulates the release of neurochemicals, hemorhology, cerebral microcirculation, metabolism, neuronal activity, and the function of specific brain regions (10,34,35). Animal studies have revealed that the effects of acupuncture therapy on stroke may possibly be mediated through the inhibition of post-ischemic inflammatory reactions, the stimulation of neurogenesis and angiogenesis, and the influence on neural plasticity (36,37).

Recently, improvement in cognitive dysfunction by EA following stroke has attracted increasing interest. Physical rehabilitation is not the only treatment aim, but also cognitive improvement is closely associated with the quality of daily life (38).

Transport of Ca²⁺/CaM from the surface membrane to the nucleus activates CaMK kinase (CaM KK) and the substrate, CaMKIV, the CREB kinase. This classical cellular signaling pathway is considered to be closely associated with cognitive function in the hippocampus. Ca²⁺ binding to CaM, and the consequent activation of Ca²⁺/CaM-dependent protein kinases combined with the CaM kinase family, contribute strongly to synaptic potentiation, learning and memory. Additional CaM kinases and CaMKIV form a CaMK cascade within the nucleus. Neuronal activity and Ca²⁺/CaM drive CaMKK to phosphorylate and activate nuclear CaMKIV, which phosphorylates CREB and CREB-binding protein (39). In the present study, a notable finding was that the expression and activity of CaM in the MCAO group was significantly increased, in contrast to the sham and EA groups. Thus, the pathological process is yet to be fully elucidated. However, the expression levels of the additional proteins varied as predicted. EA promoted CaMKIV, p-CaMKIV, CREB and p-CREB protein expression. According to a previous study, the inhibitory effect of EA on NF-κB activation led to the inhibition of cerebral cell apoptosis and an improved cognitive ability (21). Therefore, the CaM-CaMKIV-CREB pathway may be an additional important cellular signaling pathway involved in cognitive improvement.

The present study is preliminary investigation that used rat models; thus, the effect of EA on post-stroke cognitive impairment requires detailed evaluation in clinical practice. The aim of the present study was to explain the detailed mechanism underlying the effects of EA on cognitive impairment from a novel perspective; however, the mechanisms of EA on stroke are complex, comprehensive and wide. Therefore, this cellular signaling pathway may not be the only neuronal signaling pathway involved in cognitive impairment.

In conclusion, to the best of our knowledge, the present study is the first to demonstrate that EA exhibits excellent cognitive repair properties, with the underlying mechanism closely associated with the CaM-CaMKIV-CREB signaling pathway.

Acknowledgements

This study was supported by a grant from the National Natural Science Foundation of China (no. 81273835).

References


