Effect of isoflurane + N₂O inhalation and propofol + fentanyl anesthesia on myocardial function as assessed by cardiac troponin, caspase-3, cyclooxygenase-2 and inducible nitric oxide synthase expression

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Abstract. The aim of this study was to evaluate the effect of isoflurane + N₂O inhalation and propofol + fentanyl anesthesia on myocardial function as assessed by cardiac troponin T (cTnT). A total of 60 patients were randomized into two groups: isoflurane + N₂O inhalation (n=30) and propofol + fentanyl anesthesia (n=30). The findings demonstrated that there was no significant difference between the two experimental groups in terms of cTnT levels, demographic properties or hemodynamic parameters. Isoflurane + N₂O inhalation and propofol + fentanyl anesthesia, respectively, were also investigated in a rat model of myocardial infarction. Myocardial cell damage, inflammation and oxidative stress levels, caspase‑3, cyclooxygenase‑2 and oxidative stress levels, caspase‑3, cyclooxygenase‑2 protein expression in the isoflurane + N₂O inhalation group were significantly higher than that of the propofol + fentanyl anesthesia group (P<0.01). In conclusion, isoflurane + N₂O inhalation was significantly higher than that of the propofol + fentanyl anesthesia group (P<0.01). In conclusion, isoflurane + N₂O inhalation and propofol + fentanyl anesthesia are not associated with risks for myocardial function.

Introduction

Myocardial infarction (MI), commonly referred to as a heart attack, occurs when blood flow stops to part of the heart causing damage to the heart muscle (1). With increasing morbidity and death rates, it has become a great threat to human health. The leading cause of MI at the acute phase is acute heart failure (2). With the development of medical techniques, thrombolysis, interventional stent and bypass surgery have greatly decreased mortality rates of the conversion from acute MI to acute heart failure (3). However, heart failure can become chronic. Priority is given to remodeling of cardiac fibrosis for chronic MI. Excessive cardiac fibrosis remodeling causes heart failure.

As an inhalation anesthetic, isoflurane is does not irritate the respiratory system (4). When used as an anesthesia for teenagers, adults and the elderly, it is stable in induction with high recovery quality (5). N₂O is not toxic, has strong analgesic effects and patients are quickly awoken (5). Consequently, it is widely employed in clinics.

With high lipid solubility and low water solubility, blood concentrations of N₂O peak after intravenous injection of 2.5 mg/kg of propofol after 2 min (6). Redistribution quickly occurs, resulting in a quick decrease in blood concentration (5). Characterized by quickly taking effect without drug accumulation and patients being easily awoken, propofol metabolizes in the liver and its elimination half life is 30-60 min (7). Fentanyl is a newly discovered narcotic opiate analgesic, which has similar pharmacological functions to other opiates, such as relieving pain and calming (8). The efficacy of fentanyl peaks 3-5 min after intravenous injection and metabolizes through the liver. Its terminal half time is 2 to 4 h (9). With specific pharmacokinetic features, it takes effect fast and peaks after 1.6 min of intravenous injection. It can be degraded via non-specific esterase in red blood cells and tissues (10). With a terminal half life of 0.1 to 0.6 h, its clearance rates are not influenced by hepatic and renal functions, which is a key advantage in addition to its high safety index, short waking time, reduced respiratory depression and stable hemodynamics (11).

In the present study, the effect of isoflurane + N₂O inhalation and propofol + fentanyl anesthesia on myocardial function was investigated, as assessed by cardiac troponin in patients or the established rat model.
Materials and methods

Ethics statement. This study was approved by the Ethics Committee (grant no. 20081009) of Zhongshan Hospital affiliated to Xiamen University (Xiamen, China). Written informed consent was obtained from all participants who were involved in the study. All procedures involving experimental animals were performed in accordance with the protocols that were approved by the Committee for Animal Research of Xiamen University and complied with the Guideline for the Care and Use of Laboratory Animals.

Study population and administration of anesthesia. All patients were aged 20-45 years and had been admitted to The First Affiliated Hospital of Xiamen University between July 2013 and December 2014. Patients were randomly split into two equal groups. The following exclusion criteria was applied to all patients in the present study: i) Patients with ongoing myocardial damage (LV dysfunction, electrocardiographic abnormalities or elevated troponin); and ii) patients exhibited new onset or persistent symptoms suggestive of myocarditis. The following exclusion criteria was followed: i) exclusion for history of systemic viral disease; and ii) exclusion for relevant coronary artery disease (CAD) on angiography. One group received isoflurane + N₂O inhalation (n=30) and the other group received propofol + fentanyl anesthesia (n=30). In the isoflurane + N₂O inhalation group, patients were given 1.5% isoflurane + NO/0₂ (50/50%) after induction with thiopental sodium (5 mg/kg) + fentanyl (1 µg/kg). In the propofol + fentanyl anesthesia group, patients were given propofol (0.2 mg/kg/min) and fentanyl (0.2 µg/kg/min), and ventilated with 50% O₂ and 50% air, after induction with propofol (3 mg/kg) and fentanyl (5 µg/kg).

Animals and experimental procedures. A total of 20 male Sprague-Dawley rats (aged 8 weeks) were housed at 22-24°C, 12-h light/dark cycle and 50-60% humidity with free access to food and water. All Sprague-Dawley rats were randomly assigned into two groups: One group received isoflurane + N₂O inhalation (n=10) and the other group received propofol + fentanyl anesthesia (n=10). In the isoflurane + N₂O inhalation group, rats were performed with 1.5% isoflurane + NO/0₂ (50/50%) in the buffer for 30 min after induction with thiopental sodium (5 mg/kg) + fentanyl (1 µg/kg). In the propofol + fentanyl anesthesia group, rats were administered propofol (0.2 mg/kg/min) and fentanyl (0.2 µg/kg/min), and ventilated with 50% O₂ and 50% air in the buffer for 30 min after induction with propofol (3 mg/kg) and fentanyl (5 µg/kg).

To establish MI in rats, all the rats were anesthetized with 35 mg/kg of pentobarbital and carefully catheterized with a stump needle. An incision was cut along the left side of sternum, the thorax was opened and the pericardium was exposed. Left coronary artery (LCA) was ligated via using a 6-0 prolene suture and the chest was closed.

Hematoxylin and eosin (H&E) staining. Hearts were harvested from the experiment rats and perfusion-fixed with 4% buffered formalin (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) for 24 h at room temperature. Tissue samples were horizontally sectioned and embedded in paraffin. Tissue sections were cut into 5-µm thick slices and subsequently stained with H&E. Tissue samples were stained with H&E (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany) and visualized using enhanced chemiluminescence reagent (Beyotime Institute of Biotechnology, Shanghai, China). The stained sections were analyzed using ImagePro Plus 6.0 software (Media Cybernetics, Inc., Rockville, MD, USA).

Detection index. Blood samples (500 µl) were obtained from rats after treatment with isoflurane + N₂O inhalation and propofol + fentanyl anesthesia, and transferred to sterile tubes without EDTA and heparin prior to centrifugation at 3,000 x g for 10 min at 4°C, and serum was collected and storage at -20°C. Nuclear factor (NF)-κB of p65 (ml003404), interleukin (IL)-6 (ml102828), superoxide dismutase (SOD; ml540172), glutathione (GSH; ml531010), glutathione peroxidase (GSH-PX; ml097316) and malondialdehyde (MDA; ml022446) activities were measured using ELISA kits (Shanghai Enzyme-linked Biotechnology Co, Shanghai, China). Caspase-3 and -9 activity was measured using Ac-DEVD-pNA/LEHD-pNA (C1115 and C1157; Beyotime Institute of Biotechnology; Nanjing, China) and incubated for 2 h at 37°C. Activities were determined using an ELISA plate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA).

Western blot analysis. Total protein from rat hippocampi was extracted using protein lysis buffer containing protease inhibitor (PMSF; Beyotime Institute of Biotechnology; Nanjing, China) at 4°C for 30 min and centrifuged for 10 min at 4°C at 12,000 x g. Total cellular proteins (50 µg) were separated by 12% SDS-PAGE and electrotransferred to polyvinylidene difluoride membranes (Bio-Rad Laboratories, Inc.). The membrane was blocked with 5% skimmed milk powder in Tris-buffered saline with Tween-20 (TBST) for 1 h at 37°C and incubated with the following primary antibodies: Anti-cyclooxygenase-2 (sc-7951; COX-2; 1:3,000), anti-inducible nitric oxide synthase (sc-649; iNOS; 1:3,000) and anti-β-actin (sc-7210; 1:5,000; all Santa Cruz Biotechnology, Inc., Dallas, TX, USA) at 4°C overnight. Subsequently, the membrane was washed with TBST three times for 15 min and incubated with an anti-mouse or anti-rabbit IgG secondary antibody conjugated to horseradish peroxidase (sc-2004, 1:5,000; Santa Cruz Biotechnology, Inc.) for 1 h at 37°C and visualized using enhanced chemiluminescence reagent (Beyotime Institute of Biotechnology, Jiangsu, China). Protein bands were analyzed using Bio-Rad Laboratories Quantity One software 3.0 (Bio-Rad Laboratories, Inc.).

Statistical analysis. Data are presented as the mean ± standard deviation and were analyzed using SPSS 13.0 for Windows (SPSS, Inc., Chicago, IL, USA). One-way analysis of variance tests were performed followed by the Student-Newman-Keuls test. P<0.05 was considered to indicate a statistically significant difference.

Results

Basic characteristics of patients. In the isoflurane + N₂O inhalation group, the mean age of patients was 37.81±6.71 years, their body weight was 65.59±2.12 kg and the female: male ratio was 20:0 (Table I). In the propofol + fentanyl anesthesia group, the mean age of patients was 38.33±6.21 years, their body
weight was 66.48±1.89 kg and the female:male ratio was 20:0 (Table I).

Hemodynamic parameters in patients. Prior to induction, as shown in Table II, the preanesthetic and anesthetic hemodynamic parameters of patients in the isoflurane + N\(_2\)O inhalation group were similar to those of patients in the propofol + fentanyl anesthesia group (P>0.05). However, the pulse rates of those in the isoflurane + N\(_2\)O inhalation group were lower than that of the propofol + fentanyl anesthesia group (P<0.05; Table II).

Cardiac troponin T (cTnT) levels in patients. Levels of cTnT and troponin from the two patient groups are presented in Tables III and IV. There was no significant inter-group difference between the isoflurane + N\(_2\)O inhalation group and propofol + fentanyl anesthesia group when assessing the level of cTnT and troponin (P>0.05; Tables III and IV).

H&E staining in rat models of myocardial infarction (MI). To examine the effect of isoflurane + N\(_2\)O inhalation and propofol + fentanyl anesthesia on myocardial function in rats, a model of MI was induced using male Sprague-Dawley rats. As shown in Fig. 1, there was no notable difference between these two experimental groups in terms of myocardial cell damage.

Inflammation levels in rat models of MI. To examine whether the effect of isoflurane + N\(_2\)O inhalation and propofol + fentanyl anesthesia on inflammation levels in rats, NF-κB of p65 and IL-6 activities were measured. As shown in Fig. 2, NF-κB of p65 and IL-6 activities of isoflurane + N\(_2\)O inhalation group were significantly lower than those of the propofol + fentanyl anesthesia group (P<0.05). These findings demonstrated that propofol + fentanyl anesthesia possesses anti-inflammation effects in rats.

Oxidative stress levels in rat models of MI. To further investigate the effect of isoflurane + N\(_2\)O inhalation and propofol + fentanyl
anesthesia on oxidative stress levels in rats, the activities of SOD, GSH, GSH-PX and MDA were measured using ELISA kits. As shown in Fig. 3, the activities of SOD, GSH and GSH-PX of isoflurane + N₂O inhalation group were lower and those of propofol + fentanyl anesthesia group; whereas the activity of MDA in the isoflurane + N₂O inhalation group was significantly lower than that of the propofol + fentanyl anesthesia group. These findings demonstrated that propofol + fentanyl anesthesia also possesses anti-oxidative effects in rats.

Caspase-3 and -9 activity. Using ELISA kits, the effects of isoflurane + N₂O inhalation and propofol + fentanyl anesthesia on caspase-3 and -9 activity were determined in the rat models of MI. As shown in Fig. 4, caspase-3 and -9 activities in the isoflurane + N₂O inhalation group were significantly suppressed, as compared with the propofol + fentanyl anesthesia group (P<0.05). The study indicated that propofol + fentanyl anesthesia inhibited caspase-3/9 activity to reduce heart cell apoptosis in in rats of MI.

COX-2 protein expression. Using western blot analysis, the effect of isoflurane + v inhalation and propofol + fentanyl anesthesia on COX-2 protein expression levels was determined. Western blot analysis demonstrated that COX-2 protein expression levels in the isoflurane + N₂O inhalation group were significantly inhibited, as compared with the propofol + fentanyl anesthesia group, which suggested that propofol + fentanyl anesthesia weakened COX-2 protein
expression and prevented inflammation in rats of MI (P<0.05; Fig. 5).

**iNOS protein expression.** To further confirm the effect of isoflurane + N\textsubscript{2}O inhalation and propofol + fentanyl anesthesia on iNOS protein expression, iNOS protein expression levels were detected using western blot analysis. As shown in Fig. 6, iNOS protein expression in the isoflurane + N\textsubscript{2}O inhalation group was significantly higher than that of the propofol + fentanyl anesthesia group (P<0.05). These findings demonstrated that propofol + fentanyl anesthesia also promoted iNOS protein expression in rats of MI.

**Discussion**

MI is an ischemic heart disease. When the coronary artery or branches have lesions, stenosis or blocking occurs, which causes myocardial ischemic-anoxic injury or necrosis (12). Neither traditional therapy nor newer intervention therapy has been able to achieve satisfactory effects for diffuse injuries of coronary arterioles (13). This is mainly as blood micro-circulation at the ischemic region cannot be improved.

In this study, the pulse rates of the isoflurane + N\textsubscript{2}O inhalation group were lower than that of the propofol + fentanyl anesthesia group. Clinically, the judgment of anesthesia depth for patients is based on heart rate, blood pressure and body movement (14). However, the dose of propofol was decreased in anesthetic depth monitoring, which suggests that blood pressure and heart rate are inadequate for judging anesthesia depth, which may lead to excessive dosages of anesthetic drugs (6). Our study showed that no significant inter-group difference existed between the isoflurane + N\textsubscript{2}O inhalation group and the propofol + fentanyl anesthesia group in terms of the levels of cTnT and troponin.

N\textsubscript{2}O, which is colorless, odorless and non-irritating, is an inorganic gas with a low blood/gas distribution coefficient (5). In addition to rapid induction, quick waking and non-irritation of the respiratory tract, N\textsubscript{2}O does not damage important organs, such as the heart, lung, liver and kidneys (15). It does not participate in bio-conversion or degradation in vivo, thus it is predominantly discharged when breathing (16). There is minimal evaporation through skin without accumulation; therefore, N\textsubscript{2}O is an ideal inhaled anesthetic (17). As anesthesia performance of N\textsubscript{2}O is low and weak, a high concentration of single use may cause anoxia; thus, its concentration should not be higher than 60% (18). We found that NF-κB of p65 and IL-6, caspase-3 and 9 activities in the isoflurane + N\textsubscript{2}O inhalation group were suppressed, compared with the propofol + fentanyl anesthesia group.

Fentanyl-propofol is a common intravenous anesthesia (19). However, respiratory depression of fentanyl during surgery is rather common and hyperpathia after surgery occurs frequently (19). Propofol is an alkylphenolsedative-hypnotic drug and its advantages include quick effects, short hold time and fast waking (20). It is eliminated through the liver without accumulation. It also has weak antiemetic and analgesic functions (21). The greatest disadvantage of propofol is dose-dependent respiratory inhibition and the fact that it is influenced by injection rates (22). Large dosages induce notable decreases in blood pressure, heart rate, hyoxemia, bradypnea or apnea. With short-term effect, fentanyl is a new opium analgesics (23). It can be degraded quickly through non-specific esterase in blood and tissues and its elimination rate is not affected by hepatorenal functions (24). Characterized by taking effect quickly, strong
abirritation, quick recovery times and a lack of accumulation, it rarely exhibits side effects, such as decreased blood pressure, declined heart rate and breath inhibition, which are related with dosage and injection rates (25). COX-2 and iNOS protein expression levels in the isoflurane + N₂O inhalation group were inhibited and activated, respectively, as compared with the propofol + fentanyl anesthesia group. In conclusion, isoflurane + N₂O inhalation and propofol + fentanyl anesthesia were demonstrated to not affect myocardial function.

References