Molecular basis for the involvement of thymidine phosphorylase in cancer invasion

TAKENARI GOTANDA¹,², MISAKO HARAGUCHI², TOKUSHI TACHIWADA¹,², REIKO SHINKURA³, CHIHAYA KORIYAMA³, SUMINORI AKIBA³, MOTOshi KAWAHARA¹, KENRYU NISHIYAMA¹, TOMOYUKI SUMIZAWA², TATSUHIKO FURUKAWA², HIROMITSU MIMATA⁴, YOSHIO NOMURA⁴, SHIN-ICHI AKIYAMA³ and MASAYUKI NAKAGAWA¹

Departments of ¹Urology, ²Molecular Oncology, Field of Oncology, Course of Advanced Therapeutics; ³Department of Epidemiology and Preventive Medicine, Field of Human and Environmental Sciences, Course of Health Research, Kagoshima University Graduate School of Medical and Dental Sciences, Kagoshima 890-8520; ⁴Department of Urology, School of Medicine, Oita University, Oita 879-5593, Japan

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Abstract. Thymidine phosphorylase (TP), also known as platelet-derived endothelial cell growth factor (PD-ECGF), has been implicated in bladder cancer angiogenesis and invasion. However, the molecular basis of its role in invasion remains unclear. We investigated the expression of TP and 10 invasion-related genes in bladder cancers from 72 randomly selected patients by real-time two-step RT-PCR assay. We found that the expression levels of TP, MMP-9, uPA, and MMP-2 were significantly higher in invasive tumors than those in superficial tumors. Also, the expression level of TP significantly correlated with that of uPA, MMP-1, MMP-9, PAI-1 and VEGF. KK47/TP cells, bladder cancer cells that overexpress TP, had a higher expression of MMP-7 and MMP-9 than KK/CV cells that express lower level of TP in hypoxic condition. PC/TP cells, prostate cancer cells that overexpress TP, also had a higher expression of MMP-1 and MMP-7 than PC/CV cells that express no detectable TP. Taken together these data indicate that TP enhances the invasion of tumor cells through the induction of invasion-related genes.

Introduction

Cancer invasion is a complicated process involving the activity of multiple genes. As cancer cells undergo invasion, they must first attach to the target tissue's basal matrix and then degrade the basement membrane. Degradation of the extracellular matrix, or components of the basement membrane, is carried out mainly by proteases belonging to the matrix metalloproteinase family (MMPs) (1,2). Elevated expression of the MMP-1 (3), MMP-2 (4), MMP-3 and MMP-9 (5) was correlated with the bladder cancer tumor stage, grade or prognosis. The membrane-type matrix metalloproteinase-1 (MT1-MMP), that induces specific activation of the precursor of the MMP-2 (6), is associated with bladder cancer progression. Urokinase-type plasminogen activator (uPA) (7) is an enzyme that degrades proteins in tissue basement membranes and the ECM. uPA and MT1-MMP have also been correlated with the stage and prognosis of patients with bladder cancer. High expression of MMP-2, -3, -7 and MT1-MMP are unfavorable prognostic features in breast cancer, gastric carcinoma and neuroblastoma (8-11). Vascular endothelial growth factor (VEGF) is an important mediator of tumor angiogenesis (12).

We have shown that TP, an enzyme that is involved in pyrimidine nucleoside metabolism, is identical to PD-ECGF (13). TP stimulates chemotaxis of endothelial cells in vitro, and displays angiogenic activity in vivo (14). Site-directed mutagenesis indicates that promotion of vessel growth by TP is dependent on the enzyme activity of TP (15). Among the degradation products generated from thymidine by TP activity, 2-deoxy-D-ribose (2DR), a dephosphorylated product derived from 2-deoxy-D-ribose-1-phosphate (16,17), also displays chemotactic activity in vitro (14), and may function as a downstream mediator of TP. TP expression in stroma cells was associated with non-endometrioid-type carcinomas, high tumor grade, deep myometrial invasion, and advanced stage (18). In esophageal squamous cell carcinomas, statistically significant differences in serum TP level were observed depending on tumor size (P<0.01) and tumor depth (P<0.01).
There was also a significant correlation between the proportion of cells expressing TP in differentiated gastric adenocarcinomas and the depth of tumor invasion. These findings suggested that TP is involved in the invasiveness of differentiated gastric adenocarcinomas. TP was also significantly elevated in invasive bladder cancer and, thus, may have a functional role in bladder cancer invasion. However, in spite of its potential importance for understanding cancer invasion the molecular basis for the involvement of TP in invasion remains unclear.

We examined whether TP modulates invasion by affecting the expression of invasion-related genes. We determined the expression of 10 invasion-related genes and 2 other genes in bladder cancers from 72 randomly selected patients with a real-time two-step RT-PCR assay. We found that TP expression strongly correlated with the expression of uPA, MMP-1, MMP-9, PAI-1 and VEGF. Furthermore, KK/TP bladder cancer cells that overexpress TP had a higher MMP-7 and MMP-9 expression than KK/CV cells in hypoxic condition that are commonly observed in tumor tissues. The data suggest that targeting TP expression might represent a novel anti-invasion approach.

### Materials and methods

**Patient characteristics.** Between March 1996 and April 2000 we carried out transurethral resection (TUR) or total cystectomy in 72 patients with bladder cancer. Each tissue specimen was immediately snap frozen and stored at -80°C. We used these tissue specimens after approval of the local ethics committee. Pathological examination was performed by several pathologists, and pathological staging was diagnosed according to the Tumor-Nod-Metastasis (TNM) classification of stage. Tumor cells above the muscularis mucosa (MM) were labeled superficial, whereas those neoplastic cells located below the MM were considered invasive. There were 34 invasive- and 38 superficial-bladder cancer tissue samples.

**RNA extraction.** Total RNA from tissues and cells was isolated by Trizol (Life Technologies) according to the manufacturer's protocol. Isolated RNA was further purified with the RNeasy kit (Qiagen), according to the RNA clean-up protocol, and eluted in 50 μl of RNase-free distilled water (Life Technologies). The quantity of RNA was measured spectrophotometrically.

**cDNA synthesis.** cDNA was synthesized from total RNA. Reverse-transcription was performed with a random primer using the ReverTra Ace·kit, according to the manufacturer's protocol (Toyobo). cDNA (1 μl) was employed in quantitative real-time PCR (an equivalent of 50 ng total RNA).

**Preparation of quantitation standards.** Real-time quantitative RT-PCR primers targeting the VEGF, MMP-1, MMP-2, MMP-7, MMP-9, uPA, TIMP-2, PAI-1, TS, DPD, and TP genes were designed by using Primer Express software (Applied BioSystems, Japan, Ltd.) and the sequences are listed in Table I.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Forward primer</th>
<th>Reverse primer</th>
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<tbody>
<tr>
<td>MMP-1</td>
<td>5'-GGACCTGGAGGAAATCTTGCT-3'</td>
<td>5'-GTCCACCTTTCTATCTCATCAAAAT-3'</td>
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<tr>
<td>MMP-2</td>
<td>5'-CCTGATGGGCACCACTTTACAC-3'</td>
<td>5'-GAGCTCCTGAAATGCGCCCTTG-3'</td>
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<tr>
<td>MMP-7</td>
<td>5'-AATGTTAACCTCCCGCTATAGA-3'</td>
<td>5'-TGGAATATGTGATTCTCGAACCATCT-3'</td>
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<tr>
<td>MMP-9</td>
<td>5'-CCCGAGTGTAGTTGAACCA-3'</td>
<td>5'-CAGGACGAGGAGCCCTTG-3'</td>
</tr>
<tr>
<td>MMP-14</td>
<td>5'-GATGGGACCGGAAATTTTG-3'</td>
<td>5'-GGGAGGTAGCAGGAGTTGAGG-3'</td>
</tr>
<tr>
<td>uPA</td>
<td>5'-TCCGGAGAGCCTGAAGATG-3'</td>
<td>5'-GGTGACTTCAGGAGGTGTG-3'</td>
</tr>
<tr>
<td>TIMP-2</td>
<td>5'-ATACCCCCTCTGTAGCTTCATCG-3'</td>
<td>5'-TGTTGTTCAGGCTCTTCTCAG-3'</td>
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<tr>
<td>PAI-1</td>
<td>5'-AGGCTGACTCTCAGGAGGATCTCG-3'</td>
<td>5'-CGGTTACCTGATCTTCACCTTC-3'</td>
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<tr>
<td>DPD</td>
<td>5'-GATTCTCGTTATCGCACTACGTT-3'</td>
<td>5'-GTACAGCCCTGAAATGTTCCCTTG-3'</td>
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<tr>
<td>TS</td>
<td>5'-CGTGATGTTGCAATCATCG-3'</td>
<td>5'-ACATCGCCGCTACGCCCTGCTC-3'</td>
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<tr>
<td>TP</td>
<td>5'-AATGACGGAGGCGCTTGAGT-3'</td>
<td>5'-TCTTACTGGAATGGGAGCTTGTA-3'</td>
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<tr>
<td>VEGF</td>
<td>5'-CCTGCGGAGGAAATCTCTAT-3'</td>
<td>5'-TGATCCAGATACTTTGAGTG-3'</td>
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</table>
and the copy number calculated from the molecular weight of the plasmid. Plasmids encoding MMP-2, MMP-9, MMP-14, and TIMP-2 were kindly provided by Dr Motoharu Seiki.

Quantitative real-time PCR. A novel quantitative PCR analysis was performed using an ABI PRISM-7900 HT sequence detector (Applied Biosystems Japan Ltd.). The probe was labeled with a reporter fluorescent dye, either FAM (6-carboxy-fluorescein) or VIC, at the 5' end and a quencher fluorescent dye TAMRA (6-carboxy-tetramethyl-rhodamine) at the 3' end. The ABI PRISM-7900 HT sequence detector measured fluorescent emission that was synchronized with the thermal cycler during each extension step. Relative quantification of GAPDH and other mRNA was calculated by the comparative cycle threshold method. The Taqman human endogenous control plate (Applied Biosystems Japan Ltd.) was put into operation and GAPDH was used as a standard for the investigation. TaqMan® Universal PCR master mix (10 μl), 250 nmol/l probe and 900 nmol/l primer were used for each PCR run. Each reverse transcribed sample (1 μl) was added to 19 μl of the PCR mixture. The thermal cycling conditions comprised an initial UNG incubation at 50˚C, followed by AmpliTaq Gold activation at 95˚C for 10 min and then 40 cycles of 95˚C for 15 sec and annealing and extension at 60˚C for 1 min. Experiments were performed with duplicates for each data point. Each PCR run consisted of a 5-point calibration curve, a no-template control, and the 72 bladder cancer specimen cDNAs. Each quantitation was performed with the standard curve method.

Cell lines and induction of hypoxia. The human prostate cancer cell line, PC-3, was purchased from the American Type Culture Collection. PC/TP cells, i.e. PC-3 cells transfected with TP/PD-ECGF cDNA, and PC/CV cells, a mock transfectant, were maintained in endotoxin-free RPMI-1640 medium supplemented with 10% fetal bovine serum (FBS). KK/TP cells, human bladder cancer KK47 cells transfected with TP/PD-ECGF cDNA, and KK/CV cells, a mock transfectant, were maintained in RPMI-1640 medium supplemented with 10% fetal bovine serum (FBS). KK/TP and KK/CV cells were seeded at 1x10⁶ cells/well in 12-well plates, cultured under normoxic condition for 12-24 h and then under hypoxic condition for 16 h. Hypoxia was induced with the Gas Pak pouch anaerobic system (Becton Dickinson and Company, Cockeysville, MD).

TP activities. Enzyme activity was assayed by the spectrophotometric method and radiometric assay. For spectrophotometric assay, cell lysates were incubated with a potassium phosphate buffer (pH 6.5) and 10 mM thymidine at 37˚C for 1 h. The thymine formed was quantitated by the absorbance at 300 nm. Enzyme-specific activity was expressed as the amount of thymine (nmol) formed/mg protein/h. For radiometric assay, enzymatic activities were determined by measuring the conversion of [14C]-thymidine to [14C]-thymine as described previously (25) with slight modification. KK/TP cells were seeded at 1x10⁶ cells/well in 12-well plates and cultured with normoxic condition for 24 h and then under hypoxic condition for 24 h. [14C]-thymine was then added in each culture medium and incubated at 37˚C for 1 h. Aliquots of media were spotted on PEI-cellulose chromatography plates. Aliquots of 10 mM thymidine and 1 mM thymine were spotted as standards. The plates were developed in a solvent system of chloroform-methanol-acetic acid 17:3:1. The nucleoside and base were identified under UV light, scraped out and analyzed for radioactivity. The ratios of radioactivity of [14C]-thymine to total radioactivity were determined.

Statistical analyses. Statistical analysis was performed using the StatView software package (Abacus, Berkeley, CA). Logistic regression model was used to determine independent predictors of bladder cancer invasion (0, superficial bladder cancer; 1, invasive bladder cancer). All variables achieving statistical significance at P<0.05 level in the univariate analysis were considered in the multivariate analyses. We also calculated the 95% confidence intervals of these odds ratios. Correlations between the log mRNA levels of TP gene expression and others were evaluated by the Spearman's rank test, with P<0.05 being considered significant. A two-tailed Student's test was used with each individual experiment to determine the significance of any changes.
Correlation of TP and other genes with bladder cancer. The importance of TP and other genes for invasive bladder cancer was examined by comparing the expression of invasion-related gene/GAPDH ratios in superficial and invasive bladder cancers (N=72). The mean TP/GAPDH mRNA ratios was 59998 (SE=23394) for invasive bladder cancer and was 11939 (SE=10824) for superficial cancer (Fig. 1a). The difference was highly significant in a two-tailed Student's test (P<0.001).

The mRNA ratios to GAPDH of MMP-2 (P=0.024), MMP-9 (P<0.001), and uPA, (P=0.0026) were significantly higher in invasive bladder cancers than those in superficial bladder cancers (Fig.1b-d). Univariate and multivariate logistic analyses were conducted to make comparisons between superficial and invasive bladder cancer with respect to the expression of variable genes including TP. Univariate analyses confirmed the findings described above: TP, MMP-9, MMP-2 and uPA mRNA ratios relative to GAPDH significantly increased in invasive bladder cancer (Table II). In the multivariate analysis, only the TP/GAPDH mRNA ratio was significantly elevated among the genes examined.

Table II. Univariate and multivariate analysis of invasion-related factors using a logistic expression model in bladder cancer.

<table>
<thead>
<tr>
<th>Experimental Variables</th>
<th>Univariate</th>
<th>Multivariate</th>
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<tr>
<td></td>
<td>β</td>
<td>SE</td>
</tr>
<tr>
<td>TP</td>
<td>0.881</td>
<td>0.227</td>
</tr>
<tr>
<td>MMP-9</td>
<td>0.717</td>
<td>0.214</td>
</tr>
<tr>
<td>uPA</td>
<td>0.677</td>
<td>0.242</td>
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<tr>
<td>MMP-2</td>
<td>0.532</td>
<td>0.188</td>
</tr>
<tr>
<td>VEGF</td>
<td>0.439</td>
<td>0.228</td>
</tr>
<tr>
<td>MMP-14</td>
<td>0.287</td>
<td>0.179</td>
</tr>
<tr>
<td>PAL-1</td>
<td>0.247</td>
<td>0.157</td>
</tr>
<tr>
<td>MMP-7</td>
<td>0.168</td>
<td>0.106</td>
</tr>
<tr>
<td>MMP-1</td>
<td>0.154</td>
<td>0.113</td>
</tr>
</tbody>
</table>

Regression coefficients (βs) and its standard errors (SEs) were obtained from logistic analysis using the presence of invasion as a dependent variable. Explanatory variables were included in logistic models after log transformation. Covariates of the multivariate regression model were TP, TIMP-2, MMP-9, uPA and MMP-2.

Figure 2. Generation of bladder cancer cells stably expressing TP. Immunoblot of TP protein in KK47 cells transfected with TP cDNA. Cytosolic fractions (100 µg of protein) were separated by SDS-9.4% PAGE. TP levels were determined using an anti-TP monoclonal antibody and anti-α-tublin monoclonal antibody (a). Enzyme activities of TP was assayed by the spectro-photometric method. TP activity is expressed as moles of thymidine catalyzed to thymine per min per mg of protein. Each column and bar represents the mean ± SD (b).
Correlation between the expression levels of TP and other genes in bladder cancer. To determine if any correlation existed between the expression level of TP and other genes in bladder cancer, we measured the mRNA levels of TP and other genes and evaluated them by the Spearman’s rank test. The expression of TP was significantly correlated with that of uPA, MMP-1, MMP-9, MMP-7, PAI-1, DPD and VEGF (Table III). In contrast, there was no correlation between TS expression and TP expression (data not shown).

Overexpression of TP enhances the expression of invasion-related genes in bladder cancer KK47 cells. A correlation between TP and invasive gene expression suggested the possibility that TP expression might modulate the expression of invasion-related genes. To test this possibility we generated the TP-overexpressing bladder cancer (KK/TP) cells. Parental KK47 is a superficial transitional cell carcinoma (TCC) cell line and had endogenous TP expression and enzymatic activity. KK/TP1 and KK/TP2 cells had higher TP expression and enzymatic activity than mock transfected KK/CV1 and KK/CV2 cells (Fig. 2a and b).

We measured the expression level of invasive genes in KK/TP cells and compare them to those in KK/CV cells in various conditions using real-time quantitative PCR. The KK/TP cells had a higher expression of MMP-7 (Fig. 3a) and MMP-9 (Fig. 3b) than the KK/CV cells in hypoxic condition. In normoxic condition, the expression of MMP-9 in KK/CV and KK/TP cells were not correlated with the level of TP expression (data not shown). In contrast to MMP-7 and MMP-9, the expression of MMP-2 in these cells were not correlated with TP expression either in normoxic or hypoxic condition (data not shown).

The enhanced expression of invasion-related genes caused by TP overexpression was not suppressed by treatment of TPI. We next examined whether the enzymatic activity of TP is required for its function. We determined if inhibition of the TP activity with TPI, a specific TP inhibitor (Ki=2x10⁻⁸ M), could abrogate its effect. We assayed the level of TP enzymatic activity of KK/TP cells using a radiometric assay. Pretreatment with 100 μM of TPI for 24 h in normoxic condition and 24 h in hypoxic condition greatly reduced the TP enzymatic activity in the KK/TP cells to a much lower level compared to the mock-transfected KK/CV cells. Pretreatment with 100 and 500 μM of TPI for 24 h in normoxic condition and 24 h in hypoxic condition did not significantly alter the expression of MMP-9 in KK/TP2 cells (Fig. 4). Although TPI seemed to partially suppress MMP-9 expression in KK/TP1 cells, the expression level was higher than that of KK/CV cells. TPI was not cyto-toxic to the cells as it did not have any effect on the number of apoptotic cells at this concentration (data not shown).

Overexpression of TP enhances the expression of invasion-related genes in prostate cancer PC-3 cells. Prostate cancer is adenocarcinoma with hormone dependency. Cellular properties of these cells appear to be different from bladder cancer.
cells. Thus, we examined if overexpression of TP enhances the expression of MMPs in prostate cancer PC-3 cells. Expression levels of MMP-1 and MMP-7 in PC/TP cells were higher than those in PC/CV cells in normoxic condition (Fig. 5a and b). This suggest a role for TP in the regulation of invasive gene expression in various cell types.

Discussion

Recent studies suggest that bladder cancer invasion is regulated by several factors. These factors induce more aggressive tumors and a more unfavorable prognosis. The ubiquitous expression of many invasion-related genes in bladder cancer (i.e., VEGF, MMP-2, MMP-9, uPA, and TP) suggests that bladder cancer invasion is a multi-step process. However, the contribution of individual genes to invasion appears to vary between different cancers. To our knowledge, this is the first time that co-expression of 10 invasion-related genes in 72 human bladder cancers was quantified. The findings presented in Table I suggest that TP, MMP-7, uPA and MMP-2 are involved in the invasiveness of bladder cancer. Surprisingly, among these genes, TP had the highest coefficient. This suggest that TP is a strong risk factor for invasiveness in bladder cancer. Furthermore, TP expression was significantly correlated with the expression of invasion-related genes such as uPA, MMP-1, MMP-9, or VEGF in bladder cancer.

Our data further suggested that one mechanism by which TP modulates invasiveness is by enhancement of expression of invasive genes such as MMP-7 and MMP-9 in bladder cancer cells in hypoxic condition. Since many tumors contain a hypoxic micro-environment, the expression of TP may play important roles in the progression and invasion of bladder cancer. The mechanism of TP induction of MMP expression is still unclear. Because the treatment with TP inhibitor TPI did not suppress the enhancement of expression of MMP-9 in KK/TP2 cells, TP has other biological roles independent of its enzymatic activity. TP-transfectants showed augmented activity of Akt compared to the respective mock-transfectants (data not shown). PI-3K and Akt are mediators of Activator Protein-1 (AP-1) induction (25). The AP-1 sites are upstream of the transcriptional start sites of MMP-7, MMP-9 and MMP-1 and has long been thought to play a dominant role in the transcriptional activation of the MMP promoters (26). The AP-1 sites are also upstream of transcriptional start sites of uPA and VEGF. On the other hand, the promoter of MMP-2 lacks the adjacent AP-1 elements (26). AP-1 induced by activated Akt might enhance the transcription of the invasion-related genes. Alternatively, Akt/PKB activates NF-xB, a family of transcription factors that increase transcription of a wide range of genes. MMP-9 has an NF-xB binding site in its promoter region and is therefore expressed in an NF-xB-dependent manner (27). Thus, TP might induce MMP expression via Akt activation. We observed enhanced expression of MMPs in PC/TP cells even in normoxic condition. Under hypoxic condition, ‘hypoxia-response’ signaling system is activated. Ultimately, this signaling system upregulates the expression of a network of gene products such as VEGF and HIF-1α that increase the propensity of the cell to survive even in this adverse environment (28). TP might need to act in harmony with such proteins to enhance expression of MMPs in KK/TP cells. Although, the exact role of TP in invasion remains unclear, TP seems to enhance the expression of MMPs in each tumor. The compounds that could suppress TP expression might become promising anti-tumor drugs.

In summary, we have provided quantitative data to support the role of TP in invasive activity of bladder cancer cells. We have demonstrated that the expression level of TP is correlated with those of several invasive genes, and TP induced the expression of MMP-7 and MMP-9 in human bladder carcinoma KK47 cells. These studies promote the understanding of molecular mechanisms underlying bladder cell invasion and may allow the future development of anti-invasive drugs based on inhibition of TP-signaling pathways.

References


