**DPD is a molecular determinant of capecitabine efficacy in colorectal cancer**

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Received August 23, 2006; Accepted November 1, 2006

**Abstract.** Capecitabine is a fluoropyrimidine-based drug that offers physicians a more convenient treatment for advanced colorectal cancer (CRC), with manageable toxicity and anti-tumor activity comparable to that of continuous-infusion therapies with 5-fluorouracil (5-FU). However, there are no validated and established predictive factors for clinical outcome of capecitabine efficacy in CRC. The gene expressions of the pyrimidine metabolism enzymes dihydropyrimidine dehydrogenase (DPD), thymidine phosphorylase (TP) and thymidylate synthase (TS) have previously been shown to be response determinants of fluoropyrimidine-based drugs in various tumors. Therefore, we investigated whether intratumoral mRNA expression levels of these genes are also associated with the clinical outcome of patients with metastatic CRC treated with first-line capecitabine. Thirty-seven patients with metastatic CRC were enrolled in this study and treated with single agent capecitabine. The intratumoral mRNA levels of DPD, TP and TS were assessed from paraffin-embedded tissue samples using laser-capture-microdissection methods and quantitative real-time PCR. There were 20 women and 17 men with a median age of 61 years (range 49-74). The median progression-free survival was 6.7 months (95% CI, 4.8-11.6 months), with a median follow-up of 14.4 months (range 1.3-18.7 months). Complete response was observed in 1 (3%), partial response in 6 (20%), stable disease in 14 (47%) and progressive disease in 9 (30%) patients (response was inevaluable in 7 patients). Higher gene expression levels of DPD were associated with resistance to capecitabine (P=0.032; Kruskal-Wallis test). Patients with a lower mRNA amount of DPD (≤0.46) had a longer progression-free survival compared with patients that had a higher mRNA amount (8.0 vs. 3.3 months; adjusted P=0.048; log-rank test).

This pilot study suggests that intratumoral gene expression levels of DPD may be useful in predicting the clinical outcome of patients with metastatic CRC with first-line single agent capecitabine treatment. Our data should be validated in larger and prospective clinical trials.

**Introduction**

In the 1950s 5-fluorouracil (5-FU) was first synthesized by Heidelberger et al (1). Fifty years later, 5-FU still remains an important component of many standard treatments in the multimodal therapy of colorectal cancer (CRC) (2). It is well established that continuous-infusion 5-FU administration, in combination with leucovorin, is more effective and associated with a better toxicity profile than intravenous bolus application (3). However, intravenous 5-FU administration is costly, inconvenient for patients and potentially associated with morbidity, including hematological toxicity and gastrointestinal toxicity (2). To overcome these disadvantages but still provide the benefits of continuous-infusion dosing, oral fluoropyrimidines were developed.

Capecitabine (Xeloda®) is a prodrug that undergoes a three-step enzymatic conversion to 5-FU (4). This oral fluoropyrimidine was already shown to offer a more convenient treatment for advanced CRC, with manageable toxicity and antitumor activity comparable to that of continuous-infusion therapies with 5-FU. In fact, two randomized clinical studies comparing capecitabine to the parenteral treatment with 5-FU and leucovorin demonstrated that the rate of objective response in patients treated with capecitabine was moderately improved (5,6). In addition, capecitabine demonstrated clinically meaningful safety advantages and the convenience of an oral agent.
Although capecitabine has clinical activity in patients with colorectal cancer, primary and acquired treatment resistance is common. Therefore, the potential to identify those individuals most likely to benefit from a given treatment assumes increasing importance. In previous studies, the expression levels of dihydropyrimidine dehydrogenase (DPD), thymidine phosphorylase (TP) and thymidylate synthase (TS) have been implicated as molecular response determinants for 5-FU based chemotherapies not only in metastatic CRC but other cancers as well (7-13). However, the role of these factors as response determinants for capecitabine has not been fully studied. TS, a rate limiting enzyme of DNA synthesis, is the major target for the anti-tumor activity of fluoropyrimidines and thus historically has been considered as the main candidate respose determinant of tumors to 5-FU-based therapy. Recently, some of the present authors (J.L., J.N.N. and A.J.) performed a study showing a significant correlation between the predominant immunohistochemical reaction pattern of TS and response (but not between immunohistochemical score and response) to capecitabine in the same CRC patients as in this study (14). TP is thought to be part of an activation pathway that converts the pyrimidine base 5-FU to the nucleotide formFdUMP, which is the actual TS inhibitor. Preclinical studies have shown that overexpression of TP, catalyzing the final step in conversion of capecitabine to 5-FU, is associated with increased sensitivity to capecitabine or oxifluridine, due to greater conversion to 5-FU (15-17). Meropol et al (J Clin Oncol 22: abs. 3520, 2004) have reported that higher TP levels are associated with response to a capecitabine-based regimen although the protocol included irinotecan as well. DPD is a catabolic enzyme that degrades 5-FU and thus its activity should theoretically bear an inverse relationship to 5-FU efficacy. This prediction has been confirmed repeatedly in the case of 5-FU-based chemotherapies, but at this time, we know of no reports concerning the relationship between DPD expression and tumor response to capecitabine (8,9,13).

The availability of FFPE tissue specimens of tumors from a group of CRC patients who were treated with capecitabine as a first-line single agent provided the opportunity to obtain an unambiguous evaluation of DPD, TP and TS gene expressions as response determinants to this drug without the potentially obscuring effects of other co-administered agents. In this study, we used laser capture microdissection to isolate tumor areas in FFPE tissue samples and then measured relative mRNA levels of DPD, TP and TS by quantitative RT-PCR to determine if the expressions of these genes are associated with the clinical outcome of patients with metastatic CRC treated with first-line capecitabine.

Materials and methods

Patients. The study included 37 patients with confirmed metastatic CRC with at least one measurable lesion. Biopsies from the primary tumor were obtained at operation. The study was performed according to the Helsinki II Declaration and approved by the regional ethics committee.

All 37 patients received first-line capecitabine based chemotherapy. They received capecitabine 1250 mg/m² twice daily as part of a phase II trial. The treatment was given for a period of 14 days followed by a 7-day rest period. The clinical evaluation and response criteria of all patients in the study are listed below.

Clinical evaluation and response criteria. CT imaging for response was performed every 6 weeks. In general, responders to therapy were classified as those patients whose tumor burden had decreased by 50% or more for at least 6 weeks. Patients with evaluable but non-measurable disease, whose tumor and all evidence of disease had disappeared, were classified as showing complete response (CR). Responders with anything less than complete response were simply categorized as demonstrating partial response (PR). Non-responders were, likewise, divided into two separate classification groups. The first of these, progressive disease (PD), was defined as a 25% or more increase in tumor burden (compared to the smallest measurement) or the appearance of new lesions. Further, non-responsive patients, who did not progress within the first 12 weeks following the start of capecitabine, were classified as having stable disease (SD).

Microdissection. Paraffin-embedded tumor blocks were reviewed for quality and tumor content by a pathologist. Tens micrometer-thick sections were obtained from the identified areas with the highest tumor concentration. Sections were mounted on uncoated glass slides. For histology diagnosis, three representative sections, consisting of the beginning, the middle and the end of sections of the tissue were stained with H&E by the standard method. Before microdissection, sections were deparaffinized in xylene for 10 min and hydrated with 100%, 95% and finally 70% ethanol. The sections were washed in H₂O for 30 sec and then stained with nuclear fast red (NFR, American MasterTech Scientific, Inc., Lodi, CA) for 20 sec and rinsed in H₂O for 30 sec. Samples were then dehydrated with 70%, 95% and 100% ethanol for 30 sec each, followed by xylene for 10 min. The slides were completely air-dried. If the histology of the sample was homogeneous and contained >90% tissue of interest, the specimen was dissected from the slides using a scalpel. All other sections of interest were selectively isolated by laser capture microdissection (P.A.L.M. Microsystem, Leica, Wetzlar, Germany) according to the standard procedure (18). The dissected particles of tissue were transferred to a reaction tube containing 400 μl of RNA lysis buffer.

RNA isolation and cDNA synthesis. The tissue samples to be extracted were placed in a 0.5 ml, thin walled tube containing 400 μl of 4 M diithiothreitol (DTT)-GITC/sarcosine (4 M guanidinium isothiocyanate, 50 mM Tris-HCl, pH 7.5, 25 mM EDTA) (Invitrogen; #15577-018). The samples were heated at 92°C for 30 min and then transferred to a 2 ml centrifuge tube. To the tissue suspensions were added 50 μl of 2 M sodium acetate, pH 4.0, followed by 600 μl of freshly prepared phenol/chloroform/isooamyl alcohol (250:50:1). The tubes were vortexed for 15 sec, placed on ice for 15 min and then centrifuged at 13,000 rpm for 8 min in a chilled (8°C) centrifuge. The upper aqueous phase (250-350 μl) was carefully removed and placed in a 1.5 ml centrifuge tube. Glycogen (10 μl) and 300-400 μl of isopropanol were added and the
Applied Biosystems). Cycling conditions were 50˚C for 2 min, reference dye, to a final volume of 20 μl (all reagents from PE System (TaqMan ®) Perkin-Elmer (PE) Applied Biosystems, detection method [ABI PRISM 7900 Sequence detection gene (ß-actin) was done using a fluorescence based real-time Quantitation of DPD, TP and TS and an internal reference Real-time PCR quantification of mRNA expression. described (19).

Response Genetics, Inc., Los Angeles, CA; United States in 50 μl of 5 mM Tris (this is a proprietary procedure of samples air-dried for 15 min. The pellet was re-suspended remaining ethanol was removed with a 20 μl pipette and the samples were quick-spun for 15 sec at 13,000 rpm. The centrifuge. The supernatant was poured off and 500 μl of 75% ethanol was added. The tubes were centrifuged at 13,000 rpm for 7 min in a chilled (8˚C) centrifuge. The supernatant was poured off and 500 μl of 75% ethanol was added. The tubes were centrifuged at 13,000 rpm for 6 min in a chilled (8˚C) centrifuge. The supernatant was carefully poured off so as not to disturb the RNA pellet and the samples were quick-spun for 15 sec at 13,000 rpm. The remaining ethanol was removed with a 20 μl pipette and the samples air-dried for 15 min. The pellet was re-suspended in 50 μl of 5 mM Tris (this is a proprietary procedure of Response Genetics, Inc., Los Angeles, CA; United States patent number 6,248,535). cDNA was prepared as previously described (19).

Real-time PCR quantification of mRNA expression. Quantitation of DPD, TP and TS and an internal reference gene (ß-actin) was done using a fluorescence based real-time detection method [ABI PRISM 7900 Sequence detection System (TaqMan®) Perkin-Elmer (PE) Applied Biosystem, Foster City, CA, USA]. The PCR reaction mixture consisted 1200 nM of each primer, 200 nM probe, 0.4 U of AmpliTaq Gold Polymerase, 200 nM each dATP, dCTP, dGTP, dTTP, 3.5 mM MgCl₂ and 1X Taqman buffer A containing a reference dye, to a final volume of 20 μl (all reagents from PE Applied Biosystems). Cycling conditions were 50˚C for 2 min, 95˚C for 10 min, followed by 46 cycles at 95˚C for 15 sec and 60˚C for 1 min. The primers and probes used are listed in Table I.

Gene expression values (relative mRNA levels) are expressed as ratios (differences between the Ct values) between the gene of interest and an internal reference gene (ß-actin) that provides a normalization factor for the amount of RNA isolated from a specimen. In validating our assays, we found that gene expression values were reproducible and reliable if Ct values were below a certain maximum. This number varied somewhat for each gene but in general, we considered Ct values below 37 to be sufficiently reliable for gene expression measurements. Those gene expressions with Ct's above 37 were not reported.

Statistical analysis. Tumor response to capecitabine and progression-free survival were our primary endpoints in this study. The progression-free survival time was calculated as the period from the first day of capecitabine until the first observation of disease progression or death from any cause. If a patient had not progressed or died, progression-free survival was censored at the time of the last follow-up.

Gene expression values are expressed as ratios between two absolute measurements, that of the gene of interest and that of the internal reference gene, ß-actin. The associations between gene expression levels and response to capecitabine (complete response, partial response, stable disease, and progressive disease) were evaluated by the Kruskal-Wallis test. To evaluate the associations between the expression level of each gene and progression-free survival, the expression level was categorized into a low and a high value at optimal cutpoints. The maximal χ² method of Miller and Siegmund (20) and Halpern (21) was used to determine which gene expression (optimal cutpoint) best segregated patients into poor- and good-prognosis subgroups (in terms of likelihood of pro-gression-free survival).

All reported P-values were two-sided. The analyses were performed using the SAS statistical package version 9.0 (SAS Institute Inc., Cary, NC).

Results
There were 20 female and 17 male patients with a median age of 61 (range 49-74) in our subject population. All patients were assessable for associating gene expression levels of DPD, TP and TS with response and progression-free survival. The median progression-free survival was 6.7

### Table I. Primers and probes.

<table>
<thead>
<tr>
<th>Gene</th>
<th>GenBank Accession</th>
<th>Forward primer (5'-3')</th>
<th>Reverse primer (5'-3')</th>
<th>Taqman probe (5'-3')</th>
</tr>
</thead>
<tbody>
<tr>
<td>ß-actin</td>
<td>NM_001101</td>
<td>GAGCGGCCGTACAGCTTT</td>
<td>TCCCTTAATGTACCCAGAGATT</td>
<td>ACCACACGGCGAGCGG</td>
</tr>
<tr>
<td>DPD</td>
<td>NM_000110</td>
<td>AGGACGCAAAGGGTTTTTGG</td>
<td>GTCGCGGAGTTCTTACTGA</td>
<td>CAGTCCCTACAGTCTCCAGT</td>
</tr>
<tr>
<td>TP</td>
<td>NM_001953</td>
<td>CCGTACGGACGAACTCTTCT</td>
<td>GCTGTTGTTGAGTGGCAGGT</td>
<td>CAGCCAGATGGTGAGACACCAG</td>
</tr>
<tr>
<td>TS</td>
<td>NM_001071</td>
<td>GCTCCGTTGTCGTCTTTCA</td>
<td>CCGTGAATGTCGCCCAAT</td>
<td>TCGCCAGCTACGCCTTCA</td>
</tr>
</tbody>
</table>

DPD, dihydropyrimidine dehydrogenase; TP, thymidine phosphorylase; TS, thymidylate synthase.

### Table II. Demographic and clinical parameters of patients with advanced CRC treated with first-line capecitabine.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Frequency</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Median age, years (range)</td>
<td>61 (49-74)</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>20</td>
<td>54</td>
</tr>
<tr>
<td>Male</td>
<td>17</td>
<td>46</td>
</tr>
<tr>
<td>Response</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Complete response</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>Partial response</td>
<td>6</td>
<td>20</td>
</tr>
<tr>
<td>Stable disease</td>
<td>14</td>
<td>47</td>
</tr>
<tr>
<td>Progressive disease</td>
<td>9</td>
<td>30</td>
</tr>
<tr>
<td>Inevaluable/early off study</td>
<td>7</td>
<td></td>
</tr>
</tbody>
</table>

samples vortexed for 10-15 sec. The tubes were placed at -20°C for 30-45 min to precipitate the RNA. The samples were then centrifuged at 13,000 rpm for 7 min in a chilled (8˚C) centrifuge. The supernatant was poured off and 500 μl of 75% ethanol was added. The tubes were centrifuged at 13,000 rpm for 6 min in a chilled (8˚C) centrifuge. The supernatant was carefully poured off so as not to disturb the RNA pellet and the samples were quick-spun for 15 sec at 13,000 rpm. The remaining ethanol was removed with a 20 μl pipette and the samples air-dried for 15 min. The pellet was re-suspended in 50 μl of 5 mM Tris (this is a proprietary procedure of Response Genetics, Inc., Los Angeles, CA; United States patent number 6,248,535). cDNA was prepared as previously described (19).
months (95% CI, 4.8-11.6 months), with a median follow-up of 14.4 months (range 1.3-18.7 months). Complete response was observed in 1 (3%), partial response in 6 (20%), stable disease in 14 (47%) and progressive disease in 9 (30%) patients (response was evaluable in 7 patients) (Table II).

Gene expression levels of DPD, TP and TS. Gene expression of DPD, TP and TS was measurable in all 37 samples (100%). The median gene expression levels, relative to the internal reference gene (ß-actin), of the analyzed genes are listed in Table III.

Gene expression levels and response of patients receiving first-line capecitabine-based chemotherapy. The only factor that showed a significant correlation between response and gene expression levels was DPD (Table IV and Fig. 1). The patient with a complete response had a median gene expression level of 0.01x10 -3, patients with a partial response 0.06x10 -3, patients with stable disease 0.29x10 -3 and patients with progressive disease 0.20x10 -3 (P=0.032, Kruskal-Wallis test).

Gene expression levels and progression-free survival in patients receiving first-line capecitabine-based chemotherapy. Gene expression cut-off values that best segregated patients into poor- and good prognosis subgroups (in terms of likelihood of progression-free survival) were defined using the maximal $\chi^2$ method of Miller and Siegmund (20) and Halpern (21). The log-rank test was used to evaluate the association between gene expression levels and survival for each single gene. The best cut-off value for DPD gene expression was found to be 0.47x10 -3, with 31 DPD expression levels below this value ('low' expression) and 6 above the cut-off value ('high' expression). The median survival of patients with low DPD mRNA levels was 8.0 months (95% CI, 5.3-12.9 months) and 3.3 months (95% CI, 1.9-4.8 months) in patients with high DPD mRNA levels (P=0.048; log-rank test) (Table V and Fig. 2A). The association between expression levels of the other genes and progression-free survival did not show significant results or relevant trends, as shown in Table V and Fig. 2B and C.

Discussion

In this report, we present data on the relationship between clinical outcomes in 37 patients with metastatic CRC receiving first-line capecitabine chemotherapy and mRNA expression of TS, TP and DPD, which have often been previously implicated in the anti-tumor activity of 5-FU based therapies (7-13). Among these 3 molecular factors, we found that only intratumoral mRNA expression levels of DPD were significantly associated with response to capecitabine treatment. Also, we showed that patients with a lower mRNA

Table III. Gene expression levels relative to the internal reference gene ß-actin of the analyzed genes.

<table>
<thead>
<tr>
<th>Gene</th>
<th>No. of Patients</th>
<th>mRNA expression levels relative to ß-actin x 10^{-3}</th>
</tr>
</thead>
<tbody>
<tr>
<td>DPD</td>
<td>37</td>
<td>0.22 (0.01-0.96)</td>
</tr>
<tr>
<td>TP</td>
<td>37</td>
<td>1.58 (0.55-7.22)</td>
</tr>
<tr>
<td>TS</td>
<td>37</td>
<td>1.36 (0.31-6.09)</td>
</tr>
</tbody>
</table>

DPD, dihydropyrimidine dehydrogenase; TP, thymidine phosphorylase; TS, thymidylate synthase.

Table IV. mRNA expression level of genes and tumor response to capecitabine therapy in patients with advanced colorectal cancer.

<table>
<thead>
<tr>
<th>Gene</th>
<th>CR</th>
<th>Median (Min-Max)</th>
<th>Partial response</th>
<th>Stable disease</th>
<th>Progressive disease</th>
<th>P value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>DPD</td>
<td>1</td>
<td>0.01 (0.01-0.37)</td>
<td>6 0.06 (0.01-0.37)</td>
<td>14 0.29 (0.05-0.96)</td>
<td>9 0.20 (0.09-0.67)</td>
<td>0.032</td>
</tr>
<tr>
<td>TP</td>
<td>1</td>
<td>3.04 (0.64-2.00)</td>
<td>6 1.48 (0.64-2.00)</td>
<td>14 1.58 (0.55-7.22)</td>
<td>9 1.58 (0.92-2.60)</td>
<td>0.47</td>
</tr>
<tr>
<td>TS</td>
<td>1</td>
<td>0.97 (0.52-2.04)</td>
<td>6 0.86 (0.52-2.04)</td>
<td>14 1.66 (0.63-6.09)</td>
<td>9 1.36 (0.31-4.53)</td>
<td>0.49</td>
</tr>
</tbody>
</table>

*The Kruskal-Wallis test. DPD, dihydropyrimidine dehydrogenase; TP, thymidine phosphorylase; TS, thymidylate synthase.
amount of DPD had a longer progression-free survival compared with patients that had a higher mRNA amount. DPD represents a key enzyme of 5-FU metabolism. This rate-limiting enzyme of 5-FU catabolism inactivates >80% of the drug in the liver (22). It has been suggested that low levels of DPD may increase bioavailability of the drug, thereby improving response. Clinical studies have already shown in patients with metastatic CRC receiving intravenous 5-FU based chemotherapy that low intratumoral DPD expression is significantly associated with a better response to 5-FU (13,23). Thus, since capecitabine is a pro-drug of 5-FU, it is reasonable to expect that low DPD expression would similarly be associated with response and prognosis of metastatic CRC patients treated with this drug. Recent studies by Tsuji et al (24) and Ichikawa et al (9) found a correlation between intratumoral DPD expression in patients with colorectal cancer and response to another 5-FU pro-drug, the oral fluoropyrimidine uracil/tegafur (UFT). Both of their studies, which included patients with stage II/III as well as metastatic colorectal cancer, demonstrated that high gene expression levels of DPD were associated with resistance to the treatment with the oral fluoropyrimidines. These data and our findings indicate that intratumoral DPD expression is an efficient predictive factor not only in intravenous but also in oral 5-FU-based chemotherapy.

Although the range and median value of intratumoral TS gene expression were lower in the 7 patients with showing response to capecitabine (Table IV) the levels of difference did not reach statistical significance. However, because the trend in the survival curves between high and low TS expressors was in the expected direction, it is probably premature to exclude TS as a response determinant for capecitabine until a study is done with a larger number of responding patients. These results do suggest though that the relationship between TS expression and response to capecitabine may not be as strong as that for the parent drug 5-FU.

### Table V. mRNA expression level of genes and time to progression in patients with advanced colorectal cancer receiving capecitabine therapy.

<table>
<thead>
<tr>
<th>Gene</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>P-value a</th>
</tr>
</thead>
<tbody>
<tr>
<td>DPD</td>
<td>≤0.46</td>
<td>31</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>&gt;0.46</td>
<td>6</td>
<td>3.64</td>
</tr>
<tr>
<td>TP</td>
<td>≤1.20</td>
<td>7</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>&gt;1.20</td>
<td>30</td>
<td>2.84</td>
</tr>
<tr>
<td>TS</td>
<td>≤2.35</td>
<td>32</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>&gt;2.35</td>
<td>5</td>
<td>3.06</td>
</tr>
</tbody>
</table>

aAdjusted. DPD, dihydropyrimidine dehydrogenase; TP, thymidine phosphorylase; TS, thymidylate synthase.

TP putatively plays a crucial double role in the anti-tumor activity of capecitabine, first cleaving the molecule to generate free 5-FU, then in the reverse reaction, converting the resulting 5-FU to FdUMP (25). The prediction derived
from this particular biological role of TP would be that high levels of TP should increase response to 5-FU-based therapy, a prediction that was supported by in vitro studies (15-17). In the present study, we did not find that high intratumoral TP expression was associated with better response to capecitabine treatment in the current set of patients; indeed, the trend we saw was in the opposite direction. However, this trend is consistent with two earlier studies regarding the relationship between TP expression and response to fluoropyrimidines. We previously had unexpectedly found that, contrary to the above model, the highest TP expressors in a group of metastatic CRC treated with 5-FU-based therapy actually had the worst response (13). To account for this unanticipated result, we hypothesized that the role of TP as an angiogenic agent (when it is known as platelet-derived endothelial cell growth factor) causes it to be an unfavorable prognostic factor that supersedes its function as an activating agent of 5-FU, while moreover, a high TP expression level does not necessarily guarantee a greater enzymatic activity of TP due to low levels of the co-substrate deoxyribose-1-phosphate in tissues. In agreement, Jakob et al (26) also found that high TP expression predicted a poor response in rectal cancer treated with 5-FU-based radiochemotherapy, a result that these workers also hypothesized as probably due to the second role of TP as an angiogenic factor.

In conclusion, by using laser-capture microdissection and real-time RT-PCR, we were able to show in patients with metastatic CRC receiving first-line capecitabine that high intratumoral mRNA expression levels of DPD were significantly associated with resistance to chemotherapy, while those of TS or TP did not reach significant association. Furthermore, we could show that patients with a lower mRNA amount of DPD had a longer progression-free survival compared with patients that had a higher mRNA amount. These data indicate that DPD may be a useful predictive and prognostic factor in patients with metastatic colorectal cancer: indeed, these data indicate that DPD may be a useful predictive and prognostic factor in patients with metastatic colorectal cancer.

References