Different tumoricidal effects of interferon subclasses and p53 status on hepatocellular carcinoma development and neovascularization

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Abstract. Interferon (IFN) is known as a multifunctional cytokine. The aim of this study was to examine the different effects of IFN subclass; namely, IFN-α and IFN-ß, on hepatocellular carcinoma (HCC) growth especially in conjunction with angiogenesis that is known to play a pivotal role in the tumor growth. Furthermore, we also examined whether the p53 status in the tumor would alter the anti-tumoral effect of IFN against HCC growth since the p53 status reportedly affects the therapeutic effect of anti-angiogenic agents against cancer. When compared with IFN-α, IFN-ß exerted a more potent inhibitory effect on HCC growth, even after the tumor was established, along with suppression of neovascularization in the tumor. A single treatment with clinically comparable low doses of IFN-ß significantly inhibited HCC vascularization in the tumor. A single treatment with clinically comparable low doses of IFN-ß significantly inhibited HCC growth whereas the same dose of IFN-α did not. IFN-ß also significantly suppressed the tumor growth both in the p53-wild and p53-mutant HCC cells. Our in vivo study revealed that IFN-ß showed a more potent inhibitory effect on the endothelial cell proliferation than IFN-α as in the in vitro study. Collectively, IFN may be an alternative anti-angiogenic agent against HCC since it exerted a significant tumoricidal effect regardless of the host p53 status even at a low dose. A cautious approach may be also required in the clinical practice since even in a same IFN subclass (class-I), IFN-α and IFN-ß exert tumoricidal effects of different magnitudes on HCC.

Introduction

Hepatocellular carcinoma (HCC) is now the fifth most common cancer worldwide, and its incidence will further increase accounting for 500,000 new cases annually (1). Despite the available therapeutic options, the incidence is still nearly equal to the mortality rate. At present, liver transplantation is considered the only curative option for HCC. However, it is not feasible to apply this option for all patients with HCC since the number of donors is absolutely insufficient. Several other modalities, such as surgery, percutaneous ethanol injection (PEI), transcatheter arterial embolization (TAE), and radiofrequency ablation (RFA), are reportedly useful to improve the prognosis in patients with small HCC (1). For advanced HCC, chemotherapy is the only remaining option although its efficacy is very poor so far (2). Recently, the progress in implantable drug delivery system has allowed repeated arterial infusion of chemotherapeutic agents, such as cisplatin (CDDP) and 5-fluouracil (5-FU), for patients with advanced HCC (3). An alternative agent; namely, interferon (IFN), has also been reported to improve the survival rate in combination with 5-FU (4,5).

IFNs are a family of natural glycoproteins initially discovered on their basis of antiviral activity. These cytokines have various biological properties, including immunomodulation and anti-proliferative actions (6). Type-I IFN, IFN-α and IFN-ß have been widely used for the treatment of patients with chronic hepatitis C in the clinical practice (7,8). There is a growing body of evidence that IFN reduces the incidence of HCC in patients with chronic hepatitis C and the intrahepatic recurrence of HCC (9). Furthermore, several reports demonstrated a drastic HCC regression after IFN therapy (10). However, to date, few studies have shown tumoricidal differences between IFN-α and IFN-ß.

Any solid tumor that has not acquired its own blood supply can not grow to more than only a few millimeters in size, including HCC (11). One of the characteristic features of HCC in the clinical practice is hypervascularity. Several studies have shown that neovascularization and angiogenic factors, such as the vascular endothelial cell growth factor (VEGF), are significantly up-regulated in the human HCC samples (12,13). We previously reported that angiogenesis plays a pivotal role in the murine HCC development, and that suppression of the VEGF-signaling pathway markedly attenuated the tumor growth (14). IFN also has an anti-angiogenic activity both in vitro and in vivo (15). Although IFN-α and IFN-ß reportedly exert different anti-angiogenic
activities in vitro (16,17), the in vivo anti-angiogenic differences in HCC have not been reported yet.

HCC is molecularly complex, as nearly every carcinogenic mechanism is altered to some degree, and HCC cells harbor numerous genetic defects such as p53 (18). Mutations of p53 gene are detectable in ~50% of HCC especially in the late stage of HCC (19). Loss of the p53 gene function is associated with poorly differentiated HCC and a shorter survival time (18). It has been reported that the host p53 status affects the tumor response to anti-angiogenic therapy, and that transcription of p53 gene is induced by IFN (20). It is very important to examine whether the anti-angiogenic therapeutic effect of IFN is affected by the host tumoral p53 status or not for future clinical application.

In the present study, to evaluate the feasibility of future clinical application, we examined the different effects of IFN-α and IFN-β at clinically comparable low doses on the experimental HCC tumor development and angiogenesis. We also elucidated whether the p53 status in the tumor alters the anti-tumoral effect of IFN against HCC growth, and investigated the possible mechanisms involved.

Materials and methods

Compounds and cell lines. IFN-α and IFN-β were generously supplied by Hayashibara Biochemical Laboratories, Inc. (Okayama, Japan) and Toray Industries, Inc. (Tokyo, Japan), respectively. The p53-wild- and -mutant HCC cell lines, BNL.1 ME A.7R.1 (BNL) and PLC/PRF/5 (PLC) were obtained from the Japanese Cancer Research Resources Bank (Okayama, Japan). The primary human umbilical endothelial cells (EC) were purchased from Kurabo (Osaka, Japan). These cells were cultured in the respective recommended medium as described previously (21).

Animal treatment. A total of 60 male 6-week-old BALB/c (n=30) and BALB/c-nu/nu (n=30) mice were purchased from Japan SLC Inc. (Hamamatsu, Shizuoka, Japan). They were housed under controlled temperature conditions and relative humidity, with 10-15 air changes per hour (h) and light illumination for 12 h a day. To compare the tumoricidal difference between IFN-α and IFN-β, we transplanted 5x10^6 PLC cells into the flank of BALB/c-nu/nu mice. The mice were randomly divided into 4 groups (n=10 in each group).

After the mean tumor volume reached 200 mm^3, the mice of the phosphate buffer saline (PBS)-treated group served as a control (Cont). The mice in IFN-α group and IFN-β group received 1x10^6 IU of murine IFN-α and -β, respectively, twice a week with subcutaneous injection on the contralateral side of the tumor. The doses of these agents are reported almost comparable to those used in the clinical practice (22).

The next experiment was conducted to examine the effect of IFN-β on BNL tumor growth. In this experiment, administration of IFN-β started on day 0, and IFN treatment was similar to that of the PLC experiment. The tumor was measured twice a week as described previously (22), and the animals were allowed free access to food and water throughout the acclimation and experiment protocols. The mice were sacrificed 56 and 32 days after the tumor cell implantation in the PLC and BNL experiments, respectively. All animal procedures were performed according to approved protocols and in accordance with the recommendations for the proper care and use of laboratory animals.

Neovascularization in the tumor. To evaluate the expression of CD31 mRNA, which is widely used as neovascularization marker, we performed a semi-quantitative RT-PCR analysis. Tumors were immediately snap-frozen for RNA extraction (n=5 in each experimental group), and mRNA was extracted from the pool of tumors. The primer for the mouse CD31 was follows: sense, 5'-CGGTGGATGATTTGATGT-3'; anti-sense, 5'-ACCGTTCTTGTGGCTCTGT-3'. PCR was performed at 94˚C for 1 min, at 52˚C for 1 min, and at 72˚C for 1 min for 30 cycles. To prevent genomic DNA contamination, all RNA samples were subjected to DNase I digestion and checked by 30 cycles of PCR to confirm the absence of any amplified DNA. The PCR products (620 bp) were analyzed by electrophoresis on 1.5% agarose gel, and the products were visualized by staining with ethidium bromide. Densitometric analysis was performed by measuring the absorbance of each band with Fuji BAS 2000 image analyzer (Fuji Co., Tokyo, Japan). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as an internal control.

Apoptosis and VEGF expression in the tumor. Apoptosis was detected with DNA fragmentation products that were stained by in situ 3'-end labeling [terminal deoxynucleotidyl transferase-mediated dUTP nick-labeling (TUNEL)] with paraffin-embedded sections. In each tumor, the positive cells in 10 high-power fields at a magnification x400 were examined as described previously (23). For measurement of the VEGF protein level in the tumor, five tumors having the same size were chosen from each group, because a different size of tumor may cause different hypoxic conditions, which strongly induce VEGF (24). The tumor samples were prepared as described previously (25). After the protein concentration was equalized, the VEGF level was measured with an ELISA kit (R&D Systems, Minneapolis, MN, USA) in accordance with the supplier's instructions.

In vitro proliferation and assay. The in vitro proliferation was determined by MTT assay as described elsewhere (25). The cell proliferation was quantified via conversion of tetrazolium, 3-(4,5-diethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) by cells cultured in 12-well plate. MTT was added to each well at a final concentration IFN-α and -β (10, 10^2, 10^3 IU/ml). After 4-h incubation at 37˚C with MTT, the untreated MTT and medium were removed, and 2 ml of dimethyl sulfoxide were added to solubilize the MTT formazan. After gentle agitation for 10 min, the optical density of each well, which is directly proportional to the number of living cells, was measured with a 540-nm filter. The absorbance was read with an ELISA plate reader (n=6 per group).

Statistical analysis. To assess the statistical significance of inter-group differences in the quantitative data, Bonferroni's multiple comparison test was performed after One-way ANOVA. This was followed by Barlett's test to determine the homology of variance.
Results

Effects of IFN-α and IFN-β on HCC growth and neovascularization. We first examined the effects of clinically comparable doses of IFN-α and -β on the HCC growth. As shown in Fig. 1, IFN-β treatment, even after the tumor was established, showed a marked inhibitory effect on HCC development as compared with the control group (p<0.05), whereas IFN-α treatment did not. The tumor volume was determined by calipers at the indicated time-points. Each point represents the mean ± SD (n=10). *Statistically significant difference between the indicated experimental groups (p<0.05). Cont, PBS-treated control group. INF-α, IFN-β, IFN-α and IFN-β treated groups at dose of 1x10^4 IU/twice a week starting from day 14 (the black arrow), respectively.

Effects of IFN-α and IFN-β on EC proliferation. To elucidate whether the differential tumoricidal effect between IFN-α and -β via anti-angiogenesis was due to the cytotoxic sensitivity to the EC or not, we next examined the effects of IFN-α and -β on EC proliferation in vitro. Both IFN-α and -β inhibited the EC proliferation in a dose-dependent manner. However, the sensitivity was significantly different. IFN-β had a marked inhibitory effect on EC proliferation even at a low dose (10 IU/ml), whereas IFN-α did not exhibit such an inhibitory effect even at a ten-times higher dose (100 IU/ml). IFN-α first exerted a significant inhibitory effect at 1,000 IU/ml although its magnitude was very low as compared with IFN-β (Fig. 3). Neither IFN-α nor IFN-β suppressed the HCC proliferation in vitro even at a high dose (1000 IU/ml) (data not shown).

Effect of the host p53 status on the tumoricidal effect of IFN. We first confirmed the host p53 status of BNL and PLC by ELISA (Roche, Germany; and Oncogene, Cambridge, MA, USA), and found that BNL and PLC exerted p53-wild and mutant status, respectively (data not shown) as described previously (26). Since only IFN-β exerted an inhibitory effect on the HCC growth at the current dose, we employed IFN-β in the following study. We then examined whether the tumoricidal effect of IFN-β was altered by the host p53 status or not. As shown in Fig. 4, IFN-β exerted a marked inhibitory effect on
**Angiogenesis, apoptosis, and VEGF in the tumor.** To elucidate the possible mechanism of the inhibitory effect of IFN-β, we elucidated the neovascularization, apoptosis, and VEGF expression level in the BNL and PLC tumors. Similar to the tumoricidal effect, IFN-β significantly suppressed the intra-tumoral neovascularization both in the BNL (p53-wild) and PLC (p53-mutant) tumors almost along with inhibition of the tumor growth (Fig. 5). On the other hand, the TUNEL-positive apoptotic cells markedly increased in both the BNL and PLC tumors by treatment with IFN-β. The incidence of apoptosis in the tumor almost corresponded to the effect of tumor development inhibition (Fig. 6). We also examined the VEGF protein level in the tumor, and found that the VEGF expression was significantly attenuated by IFN-β in both the BNL and PLC tumor indicating that the anti-angiogenic activity by IFN was likely, at least partly, to mediate the VEGF inhibition (Fig. 7).
Discussion

IFN was originally identified because of its function in antiviral host defense, and type-I IFN, IFN-α, and IFN-β are widely used for eradication of hepatitis viruses in patients with chronic hepatitis (8). Recent studies have revealed that IFN also exerted a tumoricidal effect in several types of tumors, including HCC (10,27). Most of these studies showed that the tumoricidal effect of IFN was due to direct cytotoxicity against the tumor cells at relatively high doses (28). However, the long-term administration of high doses of IFN is unacceptable for the patients with chronic liver diseases. In the clinical practice, continuous administration of high doses of IFN resulted in several symptoms including fatigue, anorexia, weight loss, dizziness, and severe hematological disorders (28).

Since therapies aiming at destruction of the tumor vasculature can achieve rapid regression of the experimental tumors, the anti-angiogenic therapy has been under investigation around the world, including the use of gene therapy, anti-angiogenic recombinant proteins, monoclonal antibodies, and various drugs (29,30). The first anti-angiogenic agent approved in the clinical practice was Bevacizumab (Avastin), a humanized version of the VEGF monoclonal antibody, in
sensitivities of IFN-α signaling cascades of IFN-α. Although both IFN-α significantly suppressed the HCC growth via anti-angiogenesis. New drugs become widely available. In the current study, we observed that the significant increase in apoptosis in HCC cell proliferation is known as one of the characteristic features of tumor cell type specific. Alternatively, the sensitivity of the newly developed EC in HCC angiogenic activity of IFN is tumor cell type specific. Furthermore, IFN attenuated the expression of VEGF in the tumor, which is known as a survival factor for the EC. Taken together, IFN may first induce EC apoptosis, with subsequent secondary apoptosis in the tumor. Alternatively, IFN acts on both HCC cells and EC by down-regulation of the intra-tumoral VEGF and inhibition of EC proliferation, respectively. These biphasic effects of IFN may also play an important role in the tumoricidal effect against HCC.

In conclusion, IFN exerted a marked inhibitory effect on the HCC growth, even after the tumor was established, via anti-angiogenesis along with suppression of VEGF and augmentation of apoptosis in the tumor. IFN-ß significantly revealed more potent tumoricidal effect than IFN-α, and a clinically comparable low dose of IFN-ß inhibited the HCC growth regardless of the host p53 status. A cautious approach may be also required in the clinical practice since even in the same IFN subclass (class-I), IFN-α and IFN-ß exert tumoricidal effects of different magnitudes on HCC.

References


