Synergistic effects of combined treatment with simvastatin and exemestane on MCF-7 human breast cancer cells

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Received May 12, 2014; Accepted January 27, 2015

DOI: 10.3892/mmr.2015.3406

Abstract. Breast cancer is associated with high levels of incidence, morbidity and mortality; therefore, the identification of effective chemopreventive strategies is crucial. It is important for clinicians to be able to identify the populations at risk who would benefit from chemoprevention, and the interventions that are effective and safe. The aim of the present study was to investigate the combined effects of simvastatin and exemestane on MCF-7 human breast cancer cells. The anti-proliferative effects of simvastatin and exemestane, alone and in combination, on the growth of MCF-7 human breast cancer cells were assessed by MTT assay. The synergism between the two drugs was determined in vitro using the combination index (CI) analysis. Cell cycle distribution and apoptosis were analyzed by flow cytometry, and alterations to the signaling pathway in MCF-7 cells were examined by immunoblotting following treatment with various regimens. The results of the MTT assay indicated that the combined treatment of simvastatin and exemestane significantly decreased the viability of MCF-7 estrogen receptor-positive (ER+) human breast cancer cells, as compared with those that were treated with the individual drugs (CI<1). In addition, coadministration of exemestane and simvastatin was shown to result in marked inhibition of tumor cell proliferation, significant cell cycle arrest at G₀/G₁ phase and induction of apoptosis, as compared with that of the control and individual drug-treated cells. Furthermore, the results of the present study indicated that these synergistic effects may be associated with the B-cell lymphoma 2 (Bcl-2)/Bcl-2-associated X protein apoptotic pathway and the mitogen-activated protein kinase/mammalian target of rapamycin/p70S6 kinase growth pathway. The combination of exemestane and simvastatin generated synergistic effects on MCF-7 ER+ breast cancer cells, indicating that the combination of these drugs may be a potential therapeutic strategy for the treatment of hormone-dependent breast cancer. The combination of the two inhibitors markedly increased the efficacy, as compared with the single-agent treatment, suggesting that combination treatment could become a highly effective approach for breast cancer. The results of the present study suggested that this combination of drugs has therapeutic potential, and requires further mechanistic and biomarker investigations in clinical trials.

Introduction

Breast cancer is the most prevalent type of cancer diagnosed in the worldwide female population (1). In patients with breast cancer, ~60% of pre-menopausal and ~75% of post-menopausal females have hormone-dependent (estrogen receptor positive [ER+]) carcinomas (2), and are therefore suitable for endocrine therapy, which is a therapeutic strategy that aims to suppress the mitogenic effects of estrogen on breast cancer cells (3). There are various types of hormonal therapies that may be used to treat ER+ breast cancers. Recently, aromatase inhibitors (AIs) have been considered the primary choice for hormonal treatment of ERα+ breast cancer in postmenopausal females (4). Third-generation AIs include the non-steroidal triazole derivatives, anastrozole and letrozole, which act as competitive inhibitors. Furthermore, a steroidal derivative of androstenedione, exemestane, has been shown to be an effective alternative to tamoxifen. Previous studies have demonstrated that exemestane is superior to tamoxifen, with regards to its effects on disease progression, incidences of locoregional and distant relapses, and contralateral breast cancers (2,5). However, despite advances in breast cancer treatment, ~25–40% of patients will eventually develop metastatic disease, which is largely incurable (6). Systemic chemotherapy is currently considered the standard treatment for patients with metastatic breast cancer (7). In addition, despite the success of the most recent generation of AIs, the eventual occurrence of adverse effects, including bone loss, fractures (8) and acquired resistance (9), reinforce the importance of searching for novel potent and specific agents with lower side effects, which may reverse the acquired resistance and extend the benefits of AIs.

Statins are one of the most frequently prescribed medications, which are used to decrease the risk of cardiovascular events and overall mortality (10). Statins are known to

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Key words: synergistic effect, combination treatment, simvastatin, exemestane, MCF-7
decrease high blood cholesterol levels through suppression of hepatic cholesterol biosynthesis (11). Statins have a similar structure to that of 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) and competitively inhibit HMG-CoA reductase, an enzyme that catalyzes the rate-limiting step in cholesterol biosynthesis, the conversion of HMG-CoA to mevalonate (12). As well as producing cholesterol, the mevalonate pathway produces numerous non-sterol products, including ubiquitine, dolichol, isopentenyladenine and prenyl groups, which are essential for the isoprenylation of intracellular second messenger mitogenic signaling proteins, such as Ras and other small G proteins (12). These non-sterol isoprenoid byproducts are important regulators of numerous oncogenic properties, including angiogenesis, proliferation and migration (13,14). Since mevalonate is synthesized from HMG-CoA, HMG-CoA reductase inhibitors, also known as statins, reduce the entry of mevalonate into the pathway. Previous studies have demonstrated the anti-neoplastic effects of statins in vitro (15-18). Nielsen et al (19) reported that statin use in Danish patients with cancer was associated with reduced cancer-associated mortality in a large observational study that included >295,000 patients with cancer.

Simvastatin is the most commonly used lipid-lowering statin drug, which is derived from lovastatin. Simvastatin has been shown to exhibit anti-proliferative and apoptotic activity against numerous types of cancer cell lines, including colon, prostate and breast (16-18). The anti-tumor mechanisms of simvastatin have been investigated and numerous potential underlying mechanisms have been identified, including suppression of downstream signaling of epidermal growth factor receptors, and attenuation of extracellular signal-regulated kinases 1/2, nuclear factor-κB, c-Jun N-terminal kinases, phosphoinositide 3-kinase/Akt (17), generation of reactive oxygen species (20), activation of inducible nitric oxide species resulting in increased levels of nitric oxide (21), as well as down regulation of B-cell lymphoma 2 (Bcl-2) and activation of Bcl-2-associated X protein (Bax) (15). Furthermore, a large Danish nationwide prospective cohort study demonstrated that use of simvastatin, a highly lipophilic statin, reduced the recurrence risk by 10 fewer cases per 100 females over 10 years among Danish females with Stage I-III breast cancer (22).

Therapeutic strategies using a combination of drugs in order to enhance the efficacy of cancer treatment have recently garnered attention. The drugs may act together synergistically to inhibit tumor progression through the regulation of various signaling pathways, or reverse the cancer-specific upregulated cell proliferation or evasion of apoptosis (23). The present study chose to evaluate combinations of statin drugs, based upon previously reported cytotoxic experience in breast cancer cell lines (24,25). The present study tested the hypothesis that the combination of simvastatin and exemestane may suppress the growth of ER+ breast cancer, and investigated the effects of combined exemestane and simvastatin treatment on breast cancer cell function, including cell survival, cell cycle, cell apoptosis and alterations in signaling pathways.

Materials and methods

Cell culture. The MCF-7 human breast cancer cell line was kindly provided by the Laboratory of Molecular Biology of Anhui Medical University (Anhui, China). The cells were purchased from American Type Culture Collection (Manassas, VA, USA) and were cultured in Dulbecco's modified Eagle's medium (DMEM; HyClone Laboratories, Inc., Logan, UT, USA) supplemented with 10% heat-inactivated fetal bovine serum, 100 U/ml penicillin, 100 µg/ml streptomycin and 2 mM L-glutamine, which were all purchased from Sijiqing Biological Engineering Materials (Hangzhou, China), and cells were maintained at 37°C in a humidified incubator containing 5% CO₂ and 95% air. The cells were harvested with trypsin-EDTA once they had reached the exponential growth phase.

Reagents and antibodies. Exemestane was provided by Pfizer Inc. (New York, NY, USA) and was dissolved in dimethyl sulfoxide (DMSO; Sigma-Aldrich, St. Louis, MO, USA) at 100 mM, in order to produce a stock solution. Simvastatin was purchased from Sigma-Aldrich and was dissolved in DMSO to a stock concentration of 100 mM. The drugs were stored at -20°C and diluted with culture medium prior to use. Final concentrations of exemestane were 3.125, 6.25, 12.5, 25, 50 and 100 µM, and simvastatin were 1.5625, 3.125, 6.25, 12.5, 25 and 50 µM. The final concentration of DMSO in the DMEM was kept at <0.1%, and equal amounts of the solvent were added to the control cells. The following primary antibodies were used: Phosphorylated (p-)mitogen-activated protein kinase (MAPK), p-mammalian target of rapamycin (mTOR), mTOR, P70S6 kinase (K), p-P70S6K (Cell Signaling Technology, Inc., Danvers, MA, USA); Bcl-2, Bax and β-actin (Santa Cruz Biotechnology, Inc., Dallas, TX, USA).

Growth inhibition assay. The anti-proliferative effects of exemestane and simvastatin on the cells were evaluated using an MTT (Sigma-Aldrich) assay. Exponentially growing cells were seeded in 96-well plates (1x10⁴ cells/well). The cells were incubated overnight for cell attachment and recovery. Following treatment with various concentrations of exemestane or simvastatin for 72 h, 20 µl MTT solution (5 mg/ml) was added to each well, and the plates were incubated for a further 4 h at 37°C. The colored formazan product was dissolved in 150 µl DMSO. The 96-well plates were then agitated for 10 min at room temperature in order to thoroughly dissolve the MTT product. The optical density (OD) of each well was measured at a wavelength of 490 nm on an ELISA plate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The percentage of inhibited cell growth resulting from each concentration that resulted in 50% cell growth inhibition, as calculated as follows: [(OD490(control cells)−OD490(treated cells))/OD490(control cells)]x100%. The half maximal inhibitory concentration (IC₅₀) of the drugs was determined as the drug concentration that resulted in 50% cell growth inhibition, as compared with the growth of the control cells, following 72 h exposure to the drugs. Six replicate wells were used for each drug concentration. Experiments were repeated at least three times and performed in triplicate.

Measurement of synergy. The anti-proliferative effects of the interaction between exemestane and simvastatin were assessed by measuring the combination-index (CI), a quantitative representation of the pharmacological interaction between two drugs. The combined effect of exemestane and
simvastatin was assessed using the median effect analysis method, as previously described by Chou and Talalay (26). The two drugs were combined in a fixed ratio of doses, which typically corresponded to 0.125, 0.25, 0.5, 1, 2, and 4 times that of the individual IC_{50} values. The CI values of interactions between exemestane and simvastatin were assessed using CompuSyn 1.01 software (CompuSyn, Inc., Paramus, NJ, USA); CI<1, CI=1, and CI>1 were considered to indicate synergistic, additive and antagonistic effects, respectively (26).

Cell cycle analysis by flow cytometry. Equal numbers of MCF-7 cells (1x10^6/well) were seeded in six-well dishes and were incubated for 24 h prior to treatment with exemestane, simvastatin or a combination of the two drugs for 72 h. The adhered cells were harvested by trypsinization (Sijiqing Biological Engineering Materials), washed twice with phosphate-buffered saline (PBS; Sigma-Aldrich) and fixed overnight in 70% ethanol (Sijiqing Biological Engineering Materials) at 4˚C. The ethanol was removed and the cells were washed a further two times with PBS, prior to resuspension in 1 ml propidium iodide/Triton X-100 staining solution [PBS containing 0.1% Triton X-100 (Sigma-Aldrich), 200 µg/ml RNase A (Sigma-Aldrich) and 50 µg/ml propidium iodide (Sigma-Aldrich)] in the dark for 30 min. The cell cycle distribution was measured by flow cytometry using a FACScan system equipped with Cell Quest software (BD Biosciences, San Jose, CA, USA). The percentage of cells in the G_0/G_1, S, and G_2/M phases was calculated using ModFit LT™ 4.0 software (Verity Software House, Topsham, ME, USA) in order to determine the cell cycle distribution.

Annexin V assay for the assessment of apoptosis. The cells in the exponential growth phase were plated (1x10^6/well) in six-well plates, allowed to attach overnight and treated with IC_{50} values of exemestane and simvastatin, either alone or in combination, for 72 h. Following 72 h of treatment, the adherent and floating cells were collected, washed twice with precooled (4˚C) PBS and resuspended in 400 µl binding buffer. The cells were then incubated with 5 µl Annexin V-fluorescein isothiocyanate (BestBio, Shanghai, China) at room temperature in the dark for 15 min, followed by an incubation with 10 µl propidium iodide (40 µg/ml; Sigma-Aldrich) at room temperature in the dark for 5 min. Following incubation, the stained cells were analyzed using a FACScan system equipped with Cell Quest software. Untreated cells were used as controls.

Western blot analysis. The MCF-7 cells treated with or without the drugs were washed with ice-cold PBS and scraped into lysis buffer (HyClone Laboratories, Inc.). The lysates were centrifuged at 16,853 x g for 30 min at 4˚C, and the supernatants were collected. Briefly, the protein concentration of each sample was determined using a Bicinchoninic Acid Protein Assay kit (Pierce, Rockford, IL, USA). Equal amounts of protein (5 µl; 0.62 mg/ml) from each sample were separated by 10% SDS-PAGE (HyClone Laboratories, Inc.) and were transblotted to polyvinylidene difluoro (PVDF) membranes (Millipore, Billerica, MA, USA). The membranes were blocked with a solution of PBS containing 5% milk and 0.1% Tween 20 (HyClone Laboratories, Inc.) for 2 h. The PVDF membranes were then probed with the following specific primary antibodies: p-MAPK, MAPK, p-mTOR (9208P; rabbit monoclonal), mTOR (2983P; rabbit monoclonal), p70S6K (2708P; rabbit monoclonal) and p-p70S6K (9234P; rabbit monoclonal), which were used at a dilution of 1:1,000 and were purchased from Cell Signaling Technologies, Inc. (Danvers, MA, USA), as well as Bel-2 (ab32124; rabbit monoclonal), Bax (ab32503; rabbit monoclonal) and β-actin (ab133626; mouse monoclonal) were used at a dilution of 1:500 in Tris-buffered saline with 0.1% Tween 20 (TBST) and were purchased from Abcam, (Cambridge, MA, USA) at 4˚C overnight. Following rinsing with TBST three times, the PVDF membranes were incubated with a horseradish peroxidase-conjugated secondary antibody (1:5,000) at room temperature for 1 h. Positive bands were detected using Enhanced Chemiluminescence reagents (EMD Millipore, Billerica, MA, USA). β-actin was used as a loading control.

Statistical analysis. Values are expressed as the mean ± standard deviation, obtained from at least three independent experiments. Student’s t-test and one-way analysis of variance were used to determine the significant differences between the control and treatment groups. Data processing was performed using the SPSS version 16.0 software package (SPSS Inc., Chicago, IL, USA). P<0.05 was considered to indicate a statistically significant difference.

Results

Exemestane or simvastatin alone inhibit the growth of MCF-7 cells. The anti-proliferative effects of exemestane and simvastatin as single agents on MCF-7 cells were determined using an MTT assay. The MCF-7 cells were treated with various concentrations of exemestane (3.125-100 µM) or simvastatin (1.5625-50 µM) for 72 h. A dose-dependent decrease in cell viability was observed following treatment with either exemestane or simvastatin. The IC_{50} values were 28.02±2.806 µM and 10.93±1.615 µM for exemestane and simvastatin respectively, following 72 h exposure (Fig. 1). Therefore, 28 µM exemestane and 11 µM simvastatin were used for all of the subsequent experiments.

Synergistic interaction between exemestane and simvastatin in MCF-7 cells. To investigate the effects of exemestane combined with simvastatin on MCF-7 cells, the cells were exposed to various concentrations of exemestane and simvastatin for 72 h. Combined treatment with the two agents induced increased levels of cell death, as compared with treatment with either exemestane or simvastatin alone. In the cells treated with exemestane and simvastatin concurrently, the CI values were all <1, with mean CI values of 0.25. These results indicated a synergistic interaction between exemestane and simvastatin on the growth inhibition of MCF-7 cells (Fig. 2). In support of this result, photomicrographs demonstrated that treatment with exemestane or simvastatin alone had only a minor effect on the number of MCF-7 cells and their morphology, whereas combined treatment resulted in a marked reduction of cell proliferation after 72 h of treatment.

Detection of cell cycle distribution using flow cytometry. To elucidate the mechanisms by which exemestane and
simvastatin inhibit the proliferation of MCF-7 cells, the cell cycle distribution was analyzed by flow cytometry. Treatment with either exemestane or simvastatin increased the population of cells in G₀/G₁ phase, with a concomitant decrease of cells in S phase (P<0.05) (Fig. 3). In addition, combined treatment with exemestane and simvastatin further increased the percentage of MCF-7 cells in G₀/G₁ phase, as compared with the cells treated with either exemestane or simvastatin alone (P<0.01). These results indicated that these two drugs may exert synergistic growth-inhibitory effects, resulting in a cell cycle arrest in G₀/G₁ phase.

Effects of exemestane or simvastatin, either alone or in combination, on cell apoptosis. To examine whether the observed suppression of growth was due to an enhanced rate of apoptosis, the apoptotic rates of the cells treated with exemestane and simvastatin, either alone or in combination, were determined using Annexin V-propidium iodide staining. Annexin V staining is markedly more sensitive for detecting apoptosis, as compared with the methods based on hypodiploid DNA content. Treatment with exemestane combined with simvastatin significantly enhanced apoptosis of the cells, as compared with the treatment with either drug alone. Individual treatment with exemestane and simvastatin resulted in 13.37 and 18.05% apoptotic cells, respectively, whereas 37.08% Annexin V-positive cells were observed following combined treatment with the two drugs (Fig. 4). These data indicated that concurrent exposure to exemestane and simvastatin resulted in synergistic interaction in MCF-7 cells.

Exemestane and simvastatin alone or in combination modify the expression levels of mitogen-activated protein kinase (MAPK) and mTOR/p70S6K signaling-associated proteins in MCF-7 cells. The main downstream effect of MAPK activation is inhibition of the mTOR signaling pathways, which have been causally associated with breast cancer cell proliferation, motility and invasiveness. Therefore, the present study analyzed the expression levels of p-MAPK and reduced expression levels of p-mTOR and p-p70S6K, whereas the total MAPK, mTOR and p70S6K expression levels were unchanged in response to treatment with simvastatin, exemestane or a combination of the two (Fig. 5).
Effects of simvastatin and exemestane treatment on the expression levels of Bcl-2 and Bax. To further evaluate the potential synergistic mechanisms of exemestane and simvastatin, the protein expression levels of Bcl-2 and Bax in MCF-7 cells were detected by western blotting. Combined treatment of exemestane with simvastatin resulted in a marked reduction in the protein expression levels of Bcl-2, and an increase in the expression levels of Bax, as compared with those in the control cells and those treated with simvastatin or exemestane alone (Fig. 6).

Discussion

Breast cancer is a hormone-dependent disease that relies on the mitogenic effects of estrogen to drive carcinogenesis. AIs are currently used as the standard first-line treatment to significantly reduce the risk of recurrence for postmenopausal females with ER+ metastatic breast cancer, as they have been proven to be more effective than tamoxifen (5). An open-label, randomized, phase III study conducted by the European Organisation for the Research and Treatment of Cancer reported a significant improvement in median progression-free survival and overall response rate for exemestane treatment, as compared with tamoxifen (27). However, despite the proven clinical efficacy of AIs in the treatment of breast cancer, de novo and acquired drug resistance often occurs and presents a major obstacle to successful therapy. In addition, patients with breast cancer treated with AIs have a higher incidence of AI-associated musculoskeletal symptoms. An International Standard Randomised Controlled Trial reported that females receiving exemestane experienced significantly higher rates of arthralgia (28). A previous study demonstrated that Armosain® (exemestane) combined with simvastatin was able to significantly increase bone mineral density, thus suggesting that simvastatin may have potential therapeutic application in the treatment of osteoporosis, to counterbalance the adverse effects of exemestane (29). Furthermore, numerous preclinical studies have shown that statins possess anti-proliferative, anti-angiogenic, anti-metastatic and pro-apoptotic properties in various types of cancer cell (17,18). In addition, some epidemiological studies have also demonstrated that statins are associated with a lower incidence of invasive breast cancer, these findings suggest that statins may contribute to the primary prevention of breast cancer (19,22). However, to the best of our knowledge, the effects of statins on endocrine therapies for patients with ER+ breast cancer have remained to be elucidated. Therefore, the present study investigated the
efficacy of simvastatin, either alone or in combination with exemestane, on the MCF-7 ER+ breast cancer cell line. The results of the present study demonstrated that simvastatin enhanced the inhibitory effects of exemestane on the growth of ER+ breast cancer cells. When simvastatin was combined with exemestane, the concentration of exemestane required to inhibit cancer cell growth was significantly reduced. These results indicated the potential importance of combined treatment approaches for increasing the efficacy of exemestane, and lessening its associated side-effects.

Simvastatin is a widely used cholesterol-adjusting drug, which selectively inhibits HMG-CoA reductase, leading to decreased cholesterol biosynthesis. Population-based studies have demonstrated that treatment with simvastatin is associated with a substantial reduction in the risk of breast cancer (19,22). Furthermore, the ex vivo tumor cell inhibition of simvastatin and its additive effects upon combination with cisplatin or docetaxel, provide the basis for epidemiological and clinical studies on statins, potentially directed toward co-medication in future treatment regimens (30). Cholesterol is a steroid hormone precursor, and the majority of cases of breast cancer are considered hormone responsive (31). Previous studies in genetic- or diet-induced hypercholesterolemic murine models have demonstrated an obvious association between high lipid levels and breast cancer progression (32). Furthermore, a primary metabolite of cholesterol, oxysterol 27-hydroxycholesterol, has been shown to promote the growth of ER+ breast cancer in in vivo models (33). Therefore, the reversal of these processes by the oral lipid-lowering drug simvastatin is an attractive anti-cancer strategy.

The results of the present study indicated that simvastatin and exemestane inhibited the proliferation of MCF-7 cells in a concentration-dependent manner. Combined treatment of simvastatin with exemestane for 72 h resulted in a marked increase in the inhibition of cell growth, as compared with treatment with exemestane or simvastatin alone. The present study also demonstrated an enhanced effect on cell cycle progression and apoptosis. MAPK is a key kinase that has an essential role in energy homeostasis and regulates processes associated with the development of cancer, including cell proliferation and survival (34,35), cell cycle arrest (34) and protein synthesis (35). Despite uncontrolled cellular proliferation in breast cancer, which is theoretically expected to create a large demand for cellular energy, there is histological evidence that phosphorylation of MAPK at Thr-172 is down-regulated, particularly in tumors of high histological grade that are associated with axillary node metastasis (36). The MAPK signaling pathway not only promotes cell proliferation, but also induces cell apoptosis and is known to be upregulated in cancer cells (37). The main downstream effect of MAPK activation is the inhibition of the mTOR signaling pathways. Misirkic et al (38) reported that in simvastatin-treated glioma cells, inhibition of mTOR, and its substrate p70S6K, resulted in the activation of the mTOR negative regulator MAPK. This result is concordant with the previously reported ability of statins to activate MAPK in hepatic and colorectal cancer cells in vitro (39), as well as to inhibit the Akt/mTOR signaling pathway in renal cancer cells (40). In the present study, combined treatment of exemestane with simvastatin caused a large decrease in the relative protein expression levels of p-MAPK and decreased the expression levels of p-mTOR, as compared with those in the control or single-agent treatment groups. p70S6K is one of the main downstream effectors of the mTOR signaling pathway, and its activated form p-p70S6K was also shown to be inhibited in the present study. To the best of our knowledge, the present study was the first to examine the individual and combined effects of simvastatin and exemestane on MCF-7 ER+ breast cancer cells, and to investigate the underlying apoptotic and growth pathways involved.

The results of the present study also demonstrated that simvastatin and exemestane increased the expression levels of Bax and decreased the expression levels of Bcl-2. The progression of cancer depends on the balance between pro-apoptotic proteins, such as Bax and anti-apoptotic proteins, such as Bcl-2 (15,41). Bcl-2 and Bax are key apoptosis regulators in numerous types of cells, which in response to treatment may lead to the activation of apoptosis (15). The results of the present study indicated that regulation of Bax and Bcl-2 protein expression may be involved in combination treatment-induced cell death.

In conclusion, the results of the present study indicated that simvastatin combined with exemestane may have synergistic effects on cell proliferation and induce cell cycle arrest and apoptosis of MCF-7 human breast cancer cells in vitro. The present study further confirmed that the synergistic effects of these two agents may involve the Bax/Bcl-2 apoptotic pathway and the MAPK/mTOR/p70S6K growth pathway. The anti-tumor effects of simvastatin are complex and remain to be fully elucidated; however, these findings provided direct evidence of its efficacy on ER+ breast cancer cells when used in combination with exemestane. Furthermore, the results of the present study suggested that the combination of simvastatin and exemestane may be a potential therapeutic strategy used to treat breast cancer; however, the synergistic effects of these two drugs require a large-scale clinical trial for further validation.

**Acknowledgements**

The present study was supported by grants from the Anhui Provincial Science and Technology Agency Foundation of China (grant nos. 1301042214, 12070403072 and KJ2012A157). The authors would also like to thank Mrs. YuanYuan, The Central Laboratory of Binhu Hospital, The Third Affiliated Hospital of Anhui Medical University (Anhui, China), for experimental instruction. The authors declare that they have no conflict of interest.

**References**

Simvastatin


