Novel herbal medicine LA16001 ameliorates cisplatin-induced anorexia

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Abstract. Chemotherapy frequently causes anorexia in cancer patients, which has been associated with poor disease prognosis. Several therapeutic strategies for the treatment of chemotherapy-induced anorexia are available; however, their adverse effects limit their clinical use. Herbal medicines have a long history of use for the treatment of various diseases, including cancer, and recent research has demonstrated their safety and efficacy. In the present study, combinations of herbal medicines were designed based on traditional Korean medicine, and their effects were investigated on chemotherapy-induced anorexia. Herbal mixtures were extracted, composed of Atractylodes japonica, Angelica gigas, Astragalus membranaceus, Lonicera japonica Thunb., Taraxacum platycarpum H. Dahlstedt and Prunella vulgaris var. asiatica (Nakai) Hara. The mixtures were termed LCBP-Anocure-16001-3 (LA16001, LA16002, LA16003). A cisplatin-induced anorexic mouse model was used to evaluate the putative effects of the extracts on chemotherapy-induced anorexia. Treatment with LA16001 was revealed to prevent body weight loss, and all three extracts were demonstrated to improve food intake. When the molecular mechanisms underlying the orexigenic effects of LA16001 were investigated, altered expression levels of ghrelin, leptin and interleukin-6 were revealed. Furthermore, LA16001 was reported to induce phosphorylation of Janus kinase 1 and signal transducer and activator of transcription 3. In addition, LA16001 administration increased the number of white blood cells and neutrophils. These results suggested that the herbal formula LA16001 may be able to prevent chemotherapy-induced anorexia and may have potential as a novel therapeutic strategy for the adjuvant treatment of patients with cancer.

Introduction

Chemotherapeutic agents used for the treatment of patients with cancer have been associated with serious adverse effects, including tiredness, nephrotoxicity, anemia, leukopenia, nausea, vomiting and loss of appetite (1-3). Vomiting and appetite loss in patients with cancer have been linked to the development of anorexia (4-6). Numerous patients with cancer who suffer from chemotherapy-induced anorexia have been reported to struggle with the continuation of treatment, thus resulting in poor disease prognosis and impaired quality of life (6,7). Megestrol acetate (MGA) is a steroidal drug that is currently used as an appetite stimulant (8-11), as it can increase appetite and promote weight gain in patients with chemotherapy-induced anorexia (12,13). However, the use of MGA has been associated with numerous adverse effects, including excessive weight gain, fluctuations in appetite and gastrointestinal complications (14,15). Our previous study demonstrated that the traditional Korean medicine sip-jeon-dae-bo-tang (SJDBT) was more effective in improving food intake in cisplatin-induced and tumor-induced anorexic mouse models compared with MGA (16,17). Furthermore, the traditional Kampō medicine rikkunshito has been demonstrated to ameliorate cisplatin-induced anorexia in rats and humans (18,19). These findings suggested that herbal formulas used in traditional Eastern medicine may have potential as alternative therapeutic strategies for the prevention and treatment of chemotherapy-associated anorexia in patients with cancer.

A herbal formula termed LCBP-Anocure-16001 (LA16001) has been designed based on traditional Korean medicine and SJDBT, and its effects have been investigated on chemotherapy-induced anorexia (20). The formula consists of a combination of herbal extracts from Atractylodes japonica (AJ), Angelica gigas (AG), Astragalus membranaceus (AM),...
which are included in SJDBT, with extracts from Lonicera japonica Thunb. (LJ), Taraxacum platycarpum H. Dahlstedt (TP), and Prunella vulgaris var. asiatica (Nakai) Hara (PV), at a 1:1:1:1:1:1 ratio. AJ, AG and AM have been used in traditional Korean medicine for the treatment of anorexia, fatigue, general weakness, and for immunopotentiation (21-26). In Korean tradition, LJ and TP have been reported to alleviate drug-induced toxicity and fever, and exert anti-inflammatory and diuretic effects (27,28), whereas PV has traditionally been used for its antiviral and antibacterial properties, and for the treatment of indigestion (29,30).

In the present study, a cisplatin-induced anorexic mouse model was employed to investigate the effects of LA16001 on appetite. LA16001 was demonstrated to increase food intake and body weight, and its effects were more potent compared with SJDBT and MGA. The molecular mechanisms underlying its effects appeared to involve altering the levels of ghrelin, leptin and interleukin (IL)-6, potentially via regulation of the Janus kinase (JAK) 1/signal transducer and activator of transcription (STAT) 3 pathway. In addition, LA16001 was demonstrated to prevent chemotherapy-associated leukopenia and neutropenia. These results suggested that LA16001 may have potential as an alternative herbal therapeutic strategy for the treatment of patients with chemotherapy-induced anorexia.

Materials and methods

Herbal extracts and reagents. The herbal formulas were composed of extracts from AJ, AG, AM, LJ, TP and PV, at w/w ratios of 1:1:1:1:1:1, 1:2:1:2:2:2 or 2:2:1:1:1. The herbal formulas were termed LCBB-Anocure-16001, 16002 and 16003 (LA16001, LA16002, LA16003), respectively. Herbal medicines were obtained from Kyung Hee Herb Pharm (Wonju,) and Omniherb (Seoul, Korea). LA16001 was composed of 40 g each of AJ, AG, AM, LJ, TP and PV; LA16002 was composed of 25 g each of AJ, AG and AM and 50 g each of LJ, TP and PV; LA16003 was composed of 50 g each of AJ, AG and AM and 25 g each of LJ, TP and PV. The ingredients were placed in 15-fold volume of distilled water and boiled at ~100°C for 3 h. The extracts were filtered using 75 µm microfilters. The filtered residues were placed in a 15-fold volume of distilled water, boiled at ~100°C for 3 h and refiltered through 75-µm microfilters. The extracted liquid was concentrated under low pressure at 60°C. Finally, 81.8 g (LA16001), 84 g (LA16002) and 93.3 g (LA16003) dried extract was obtained; the yields were 34.08, 37.33 and 41.47%, respectively. The extracts were prepared according to the Good Manufacturing Practice guidelines (31,32). SJDBT was obtained from Hanpoong Korea. Mice (n=4/group) were randomly assigned to the following groups: Normal, control, cisplatin, LA16001, LA16002, LA16003, SJDBT, MGA (Sigma-Aldrich; Merck KGaA), AJ, AG, AP, LJ, TP and PV, and were maintained at 23±3°C (humidity, 55±15%), under a 12-h light/dark cycle. Access to food and water was ad libitum, apart from during the experimental period. All groups (except the normal group) were fasted for 24 h prior to the start of the experiment (the normal group was used as a control for starvation). The control group did not receive any drug treatment, only starvation; this group was used as a control for the cisplatin group. All groups (except the normal and control groups) were induced to become anorexic through administration of cisplatin. Therefore, the cisplatin group was used as a control for the drug treatments. The drug was administered at the concentration of 1,000 mg/kg on days 1-3. After 24 h, 8 mg/kg cisplatin was intraperitoneal injected into the mice. During fasting, water was continuously provided and food was provided following the oral administration of drugs. Food intake and body weight were measured daily throughout the duration of the study (Fig. 1A). An equal amount of food was given at the same time each day and mice were weighed the following day. All mice were sacrificed at the time labeled “Sac” in the animal study design presented in Fig. 1A. Blood and adipose tissue, stomach, and hypothalamus were obtained from the sacrificed mice.

The present study was approved by the Animal Ethics Committee of Kyung Hee University [approval no. KHUASP (SE)-016-100].

ELISA and blood analysis. Samples from stomach, fat and hypothalamic tissue were isolated from mice sacrificed 4 h subsequent to the administration of cisplatin (n=4 mice/group), and lysed with radioimmunoprecipitation assay lysis buffer (Biosesang, Inc., Seongnam, Korea). Lysates were centrifuged at 196 x g for 20 min at room temperature, and supernatants were collected and used for the assessment of target proteins using ELISA. The levels of ghrelin (cat. no. EZRGRA-90K; EMD Millipore, Billerica, MA, USA), leptin (cat. no. DY498; R&D Systems, Inc., Minneapolis, MN, USA) and IL-6 (cat. no. 1555240; BD Biosciences, San Jose, CA, USA) were measured using ELISA kits according to the manufacturer’s protocol. The blood of the mice sacrificed 3 days subsequent to the administration of cisplatin was used. Control group and cisplatin group, LA16001 group mice were used, 4 mice/group. Exsanguination was performed, and whole blood was analyzed using the HEMAVET®950 hematology system (Drew Scientific Inc., Miami Lakes, FL, USA); SigmaPlot, version 2001 (Systat Software, Inc., San Jose, CA, USA) was used for analysis of the blood data. To anesthetize, 400 µl/mouse 1.2% avertin solution was intraperitoneally injected [1.2% avertin solution, 0.5 g 2,2,2-tribromoethanol powder (Sigma-Aldrich; Merck KGaA)] dissolved in 1 ml 2-methyl-2-butanol (Sigma-Aldrich; Merck KGaA) at 55°C, and 39 ml PBS], the avertin solution was filtered through a Nalgene 0.22-µm filter (Thermo Fisher Scientific, Inc., Waltham, MA, USA). Western blot analysis. Adipose tissue was isolated from mice. To obtain protein, the adipose tissue was lysed using radioimmunoprecipitation assay buffer (Biosesang, Inc.) and was centrifuged at 24,562 x g at 4°C for 20 min. Protein concentration was measured using a Bradford assay. Equal amounts of extracted protein samples (20 µg) were separated by 8% SDS-PAGE and transferred onto polyvinylidene difluoride membranes (GE Healthcare Life
Sciences, Chalfont, UK). Membranes were blocked in 1% bovine serum albumin (Sigma-Aldrich; Merck KGaA) and 2% skimmed milk for 1 h at room temperature. Membranes were then incubated at 4°C overnight with the following primary antibodies: Anti-phosphorylated (p)-JAK1 (1:1,000; cat no. 3331; Cell Signaling Technology, Inc., Danvers, MA, USA), anti-p-STAT3 (1:1,000; cat no. 9131; Cell Signaling Technology, Inc.) and anti-α-tubulin (1:10,000; cat no. T-5168; Sigma-Aldrich; Merck KGaA). Subsequently, membranes were incubated with horseradish peroxidase-conjugated secondary antibody (1:2,000 in 1x PBS with 0.05% Tween-20; cat. no. 5450-0010; KPL, Inc., Gaithersburg, MD, USA) for 1 h at room temperature. Protein bands were visualized using Enhanced Chemiluminescence kit solution (DoGen, Seoul, Korea) and quantified using ImageJ software version 1.49v (National Institutes of Health, Bethesda, MD, USA) and normalized to α-tubulin.

Statistical analysis. The data are expressed as the mean ± standard deviation and experiments were performed on 4 mice/group. The statistical significance of the differences between groups was assessed using Student's t-test. Statistical analysis was performed using Microsoft Excel 2013 (Microsoft Corporation, Redmond, WA, USA). P<0.05 was considered to indicate a statistically significant difference. One-way analysis of variance was additionally performed with post hoc test (Tukey), using SPSS software (version 23; IBM Corp., Armonk, NY, USA).

Results
LA16001-3 increases food intake in cisplatin-induced anorexic mice. Mice were fasted for 24 h and intraperitoneally injected with 8 mg/kg cisplatin. Experimental drugs were orally administrated 24 h before cisplatin injection. To investigate the effects of the experimental drugs on food intake and body weight, mice were maintained for 3 days following the injection of cisplatin (Fig. 1A). The results of the present study demonstrated that following cisplatin administration, food intake and body weight decreased (Figs. 1B and C, respectively). Notably, LA16001, LA16002, LA16003, SJDBT and MGA alleviated cisplatin-induced loss of appetite (Fig. 2A), and LA16001 and MGA significantly reversed decreases in body weight (Fig. 2B). LA16001 appeared to have more potent orexigenic effects on food intake, compared with all other treatment groups (Fig. 2A). In addition, all treatment groups demonstrated significantly reversed cisplatin-induced loss of appetite, with LA16001 exerting the most pronounced effect (Fig. 3A). Furthermore, LA16001, AP, AG and TP treatment significantly reversed loss of body weight in experimental mice, again with LA16001 being the most effective (Fig. 3B). These results...
Figure 2. Treatment with LA16001-3 increases food intake and body weight in cisplatin-induced anorexic mice. Mice were fasted for 24 h and orally administered with LA16001, LA16002 or LA16003 (1,000 mg/kg), SJDBT (678.4 mg/kg) or MGA (100 mg/kg). After 24 h, mice received an intraperitoneal cisplatin (8 mg/kg) injection. (A) Food intake and (B) body weight gain were monitored daily throughout the duration of the study. Data are presented as the mean ± standard deviation. *P<0.05. LA16001-3, LCBP-Anocure 16001-3; SJDBT, sip-jeon-daebotang; MGA, megestrol acetate.

Figure 3. Effects of LA16001 on food intake and body weight in cisplatin-induced anorexic mice. Mice were fasted for 24 h and orally administered with LA16001, AJ, AG, AP, LJ, TP and PV (1,000 mg/kg). After 24 h, mice received an intraperitoneal cisplatin (8 mg/kg) injection. (A) Food intake and (B) body weight gain were monitored daily throughout the duration of the study. Data are presented as the mean ± standard deviation. *P<0.05. LA16001, LCBP-Anocure 16001; AJ, Atractylodes japonica; AG, Angelica gigas; AM, Astragalus membranaceus; LJ, Lonicera japonica Thunb.; TP, Taraxacum platycarpum H. Dahlstedt; PV, Prunella vulgaris var. asiatica (Nakai) Hara.
suggested that LA16001 may be able to counteract cisplatin-induced anorexia in vivo.

LA16001 increases the levels of appetite-associated hormones in cisplatin-treated mice. A total of 4 h post-cisplatin injection, the hormone levels of ghrelin, leptin and IL-6 were assessed in mouse tissue and serum samples. Mice in the cisplatin and LA16001 groups exhibited increased ghrelin levels in stomach tissue samples compared with mice in the fasting control group (Fig. 4A). Cisplatin was demonstrated to reduce leptin levels in fat and hypothalamic tissue, whereas LA16001 appeared to counteract this effect (Figs. 4B and C). IL-6 levels were assessed in serum and fat tissue samples. IL-6 levels in fat tissue were not altered following cisplatin administration; however, they were significantly decreased in serum samples. Notably, LA16001 administration was
demonstrated to significantly increase fat tissue and serum IL-6 levels (Figs. 4D and E). These results indicated that LA16001 administration increased the levels of ghrelin, leptin and IL-6 in cisplatin-treated mice.

LA16001 activates the JAK1/STAT3 signaling pathway in fat tissue. A total of 4 h post-cisplatin injection, the phosphorylation status of JAK1 and STAT3 was investigated in fat tissue samples. LA16001 administration resulted in significantly
increased protein expression levels of p-JAK1 and p-STAT3 compared with the cisplatin group (Figs. 5A and B). These results suggested that the molecular mechanisms underlying the orexigenic effects of LA16001 in cisplatin-induced anorexic mice may involve the activation of the JAK1/STAT3 intracellular signaling pathway.

LA16001 prevents immunosuppression in cisplatin-induced anorexic mice. White blood cell and neutrophil counts were performed 3 days after cisplatin administration to evaluate the effects of LA16001 on chemotherapy-induced immunosuppression. Treatment with cisplatin was demonstrated to reduce the numbers of white blood cells (Fig. 6A) and neutrophils (Fig. 6B) in mouse whole blood samples. Notably, LA16001 administration appeared to counteract the effects of cisplatin on immune cell numbers (Fig. 6). These results suggested that LA16001 may mitigate the immunosuppressive effects associated with cisplatin treatment.

Discussion

Chemotherapy induces various side effects, including anorexia. Anticancer drugs are toxic to healthy cells as they share characteristics with rapidly dividing cancer cells. The healthy cells that are primarily affected are hair follicle cells, and cells of the mouth, stomach, large intestine and the mucous membrane of the anus, which exhibit a rapid growth rate compared with other cells (33,34). Anorexia is particularly induced by gastrointestinal toxicity (35). Anorexia is a frequent adverse event following cancer chemotherapy, and has been associated with poor disease prognosis and treatment discontinuation (1).

Previous studies have suggested that herbal formulas used in traditional Eastern medicine may have potential as alternative strategies for the treatment of chemotherapy-induced anorexia (16,36,37). In the present study, the herbal formula LA16001 was developed, and was demonstrated to effectively counteract the development of chemotherapy-induced anorexia in mice. LA16001 prevented the loss of appetite and body weight following cisplatin administration. Ghrelin is a hormone critical for appetite regulation, and its levels have been reported to decrease following food intake (38). Leptin is a hormone functioning as an appetite suppressant, and its levels increase following feeding (39). IL-6 is a proinflammatory cytokine that has been revealed to function together with leptin to increase appetite (40). Patients with cancer with anorexia due to anticancer drugs exhibit increased ghrelin and decreased leptin (41). Leptin and IL-6 have been demonstrated to act through the regulation of the JAK1/STAT3 intracellular signaling pathway. Furthermore, LA16001 counteracted the immunosuppressive effects of cisplatin, as demonstrated by an increase in immune cell numbers. The present results suggested that LA16001 may have potential as an alternative therapeutic strategy for the prevention and treatment of chemotherapy-associated anorexia in patients with cancer. Further studies are required to validate the orexigenic effects of LA16001, and to investigate the cellular and molecular mechanisms underlying its actions.

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