Role of the JAK-STAT pathway in proliferation and differentiation of human hypertrophic scar fibroblasts induced by connective tissue growth factor

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Abstract. The aim of this study was to investigate whether the JAK-STAT pathway participates in the processes of proliferation and differentiation induced by connective tissue growth factor (CTGF) in human hypertrophic scar fibroblasts (hHSF). hHSF were grown as primary cultures, then treated with or without CTGF. Western blotting was used to detect JAK1, JAK2, JAK3, TYK2, STAT1, STAT2, STAT3, STAT4, STAT5 and STAT6 protein expression in the hHSF at various time points after stimulation with CTGF. Immunofluorescence and the electrophoretic mobility shift assay (EMSA) were used to identify the interacting signalling molecules and to investigate their activation, respectively. After the signalling molecules of interest were selected, a specific inhibitor (STAT1 ASODN) was used to block the JAK-STAT pathway. The MTT assay was used to detect the proliferation of hHSF, and differentiation was assessed by evaluating changes in α-SMA expression by RT-PCR. Based on the results of Western blotting, immunofluorescence and EMSA, proliferation and differentiation were much higher in hHSF treated with CTGF (p<0.05). After blocking the pathway with STAT1 ASODN, hHSF proliferation was markedly, though not entirely, inhibited, while α-SMA expression was not significantly altered. JAK1 and STAT1 are therefore likely to participate in the proliferation and differentiation of hHSF induced by CTGF. However, though the STAT1 signalling pathway plays a key role in the CTGF-induced proliferation of hHSF, it is not the only pathway controlling this process. JAK1 may be an upstream element of STAT1, and may also participate in the CTGF-induced proliferation of hHSF. These results elucidate the signal transduction mechanism of CTGF-induced hHSF proliferation, and may aid in the development of a novel method for the inhibition of scar fibrosis and contraction.

Introduction

Fibroblasts are the main cells affecting the growth of hypertrophic scars (HS), and their proliferation and differentiation are regulated by growth factors. Connective tissue growth factor (CTGF) was recently identified as a growth factor that intensively promotes fibrosis. In a previous study, our prophase results showed that CTGF stimulated the proliferation and differentiation of human hypertrophic scar fibroblasts (hHSF) (1); however, the signal transduction mechanisms behind this effect have yet to be determined. Previous studies have indicated that JAK-STATs participate in numerous diseases through the accommodation of multiple signal transduction pathways (2,3). However, there are few reports on the relationship between JAK-STATs and the CTGF-induced proliferation and differentiation of hHSF. Therefore, we investigated the activation of JAK-STATs in hHSF stimulated by CTGF in order to identify the functional signalling molecules in this pathway.

Materials and methods

Patients and samples. Samples were obtained from 6 patients that developed a hypertrophic scar 3-12 months post-surgery. The hypertrophic scars were red and hard to the touch, and displayed hyperemia and pruritus. Research was carried out following international and national regulations. The study was approved by the Local Ethics Committee of our institution, and all patients gave informed written consent.

Cell cultivation. hHSF were cultured in 10% calf serum (CS) (Hali Bioengineering Co., Chengdu, China), DMEM (Gibco, USA), 0.25% trypsin (Gibco) and phosphate buffered saline (PBS) (Zhongshan Jinqiao Biotechnology Co., Ltd., Beijing, China).
For the initial experiments, cells were divided into two groups: Group 1, hHSF without CTGF (control); and Group 2, hHSF with CTGF (experimental group).

Stimulation of hHSF by CTGF. DMEM was combined with 10% CS, then DMEM without CS was added. The cells were starved for 18-24 h, then the supernatant was poured off. DMEM was added to 10 ng/ml CTGF, and then the fibroblasts were cultured under standard conditions (37°C, 5% CO₂, humidity 95-100%) for 0, 5, 10, 20, 30, 45, 60 and 90 min, followed by protein extraction.

Western blotting. To detect the activation of the JAK STAT proteins in the two groups, Western blotting was conducted using bovine serum albumin (BSA), rabbit antibodies (all from Cell Signaling Technology, Beverly, MA, USA), with the exception of phospho-JAK3 and phospho-STAT4, which were from Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA, 0.45 µm polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA), a Bio-Rad electrophoresis apparatus, electrotransfection apparatus, and gel imaging system (all from Bio-Rad Laboratories, Hercules, CA, USA), and developer and fixer (analytical grade; Pierce, Rockford, IL, USA). Western blotting was conducted according to routine methods using monoclonal and polyclonal antibody concentrations of 1:1000 and 1:4000, respectively (1).

The results of Western blotting were analysed using single-factor analysis of variance (ANOVA) and the t-test with SPSS 10.0 statistical analysis software. Data are presented as the mean ± SD. A value of p<0.05 indicated a significant difference.

Immunofluorescence staining protocol. To determine the nuclear translocation of STAT1 in the two groups, immunofluorescence analysis was conducted using 4% paraformaldehyde fixative, 0.01 mol/l PBS (pH 7.4), BSA, Triton X-100, phospho-STAT1 rabbit monoclonal antibody (Cell Signaling Technology) and 5'-FITC-labelled goat anti-rabbit IgG. A routine indirect staining method was used (4). In a previous study (5), STAT1 and 5'-FITC-labelled goat anti-rabbit IgG. A routine indirect staining protocol was used. Immunofluorescence staining was performed using bovine serum albumin (BSA), rabbit antibodies (all from Cell Signaling Technology, Beverly, MA, USA). Western blotting was conducted according to routine methods using monoclonal and polyclonal antibody concentrations of 1:1000 and 1:4000, respectively (1).

The results of Western blotting were analysed using single-factor analysis of variance (ANOVA) and the t-test with SPSS 10.0 statistical analysis software. Data are presented as the mean ± SD. A value of p<0.05 indicated a significant difference.

Detection of differences in the CTGF-induced proliferation and differentiation of hHSF after STAT1 ASODN transfection. hHSF were further divided into groups as follows: Group 1, STAT1 ASODN + CTGF; Group 2, CTGF alone; Group 3, STAT1 ASODN alone; and Group 4, control group using GAPDH as the reference gene. The RT assay was used to detect the proliferation of hHSF in all four groups. To determine the extent of differentiation, STAT1 and α-SMA levels were evaluated by RT-PCR (Takara, Kyoto, Japan) using the primers STAT1: GgggggagaaGcacGacGcG (forward), and GCCCACCATGCACATGAT (reverse); α-SMA: CTGAAGAGCATCCGACAC (forward), and GAC TCCATCCTAATGAAAT (reverse); GAPDH: GCAGTGFCA AAGTGGAGATTG (forward), and GCAGAAGGGCGGGA GATG (reverse). The RT mixture was composed of 8 µl MgCl₂, 4 µl 10X RT buffer, 4 µl dNTP, 1 µl RNase suppressor, 2 µl oligo-dT and 2 µl AMV-RT, added to 10.5 µl RT buffer, 1 µg RNA and DEPC ddH₂O, for a total volume of 20 µl. Reaction conditions were 50°C for 30 min, 99°C for 5 min and 5°C for 5 min, followed by denaturing at 94°C for 2 min, then 30 cycles of denaturing at 94°C for 30 sec, annealing at 60°C for 30 sec, and elongation at 72°C for 30 sec, and a final elongation at 72°C for 10 min (STAT1 ASODN or α-SMA), or denaturing at 94°C for 2 min, then 30 cycles of denaturing at 94°C for 30 sec, annealing at 55°C for 30 sec and elongation at 72°C for 30 sec, and a final elongation at 72°C for 10 min (GAPDH). Tranfected hHSF were maintained at 4°C.
Electrophoresis was conducted at 100 V for 30 min with 10 µl PCR production and 2 µl 10X loading buffer. Images were captured on a Gel Doc 1000 system.

Results

Since an increase in proliferation and in the expression of α-SMA was observed in hHSF stimulated by CTGF, the JAK-STAT pathway was considered to be activated. Two forms of JAK-STAT protein (t-) were identified: non-phosphorylated and phosphorylated (p-). The phosphorylated proteins appeared early during the cultivation, increased gradually over time, peaked at 30 min, and decreased after 30 min. Among the proteins tested, only the ratio of p-JAK1/t-JAK1 to p-STAT1/t-STAT1 was increased following CTGF stimulation (Fig. 1). Time points analysed were 5, 30 and 90 min.

The results of immunofluorescence revealed that phosphorylated STAT1 peaked at 30 min. The control group (+++) (Fig. 2a) exhibited bright fluorescent green staining, and equal distribution of staining between the nucleus and cytoplasm. The CTGF group (+++ to ++++) (Fig. 2b) exhibited marked bright green staining, and the nuclear fluorescence was much brighter than the cytoplasmic fluorescence.

EMSA was used to detect the DNA binding ability and persistence of STAT1. The ability of STAT1 to bind DNA was correlated to the concentration of CTGF. Binding increased gradually at CTGF concentrations of 0 ng/ml, 5 ng/ml, 7.5 ng/ml, 10 ng/ml and 15 ng/ml, peaked at 10 ng/ml CTGF, then decreased (Fig. 3a).

hHSF were stimulated with 10 ng/ml CTGF and binding was evaluated at various time points (0, 10, 20, 30, 45, 60, 90 and 120 min). SIF was found to be activated at 10 min, peaked at 45-60 min and decreased gradually after 60 min. The binding ability of the CTGF group was much higher than that of the control group (Fig. 3b).

Discussion

Hypertrophic scars are caused by localised concrescence after damage to the dermis (1). Their pathological characteristics are over-proliferation, differentiation of hHSF and over-deposition of extracellular matrix proteins, in particular collagen (1). Elucidating the mechanisms behind the development of hypertrophic scars would aid in their prevention in the field of trauma medicine and in reconstructive burn care. The main strategy for the inhibition of fibrosis is the suppression of the
proliferation and differentiation of hHSF (1). CTGF is a growth factor that specifically and electively interferes in the formation of connective tissue by promoting fibrosis and stimulating hHSF to differentiate into myofibroblasts (1). The inhibition of CTGF is a specific and effective method for preventing organ fibrosis. However, there are few reports discussing its signal transduction mechanism in hHSF.

The JAK-STAT pathway is a key pathway that participates in cytokine signal transduction, many physiological functions, including haematogenesis, and numerous pathological diseases, including tumours, arthritis deformans, brain injuries and bronchial asthma (2,3,6,7). The JAK-STAT pathway also participates in the proliferation, differentiation and aging of nerve cells, and has an intimate correlation with the pathological processes of the nervous system (7). Numerous cytokines, including interferon-α, -β and -γ, and interleukin-2, -4 and -6, and growth factors, including epidermal and platelet-derived growth factors, growth hormones, and leptin, regulate the proliferation and differentiation of cells through the JAK-STAT pathway. JAK-STATs are affected by the expression of CTGF induced by advanced glycation end-product (AGE) in renal interstitial fibroblasts in patients with diabetic nephropathy (8). Studies have shown that IL-6, INF-γ, PDGF and EGF mediate the signal transduction of cell surface receptors to the nucleus and regulate gene transcription through the JAK-STAT pathway. The process involves the phosphorylation of JAKs, cytoplasmic STATs, the formation of SIF compounds, nuclear translocation and binding with SIE (sis-inducing element) in the promoter region.

The present study aimed to examine the role of the JAK-STAT pathway in the proliferation and differentiation of hHSF induced by CTGF. The results indicate that JAK1 and STAT1 activity was increased following cTGF stimulation, and was to some extent correlated with cTGF-induced proliferation and differentiation of hHSF. The fluorescence intensity of phosphorylated STAT1 and the DNA binding ability of STAT1 were markedly increased following the activation of CTGF. The proliferation of hHSF was markedly, but not entirely, inhibited after STAT1 ASODN transfection. α-SMA expression before and after blocking STAT1 ASODN, indicating differentiation, was not significantly altered. In conclusion, STAT1 plays a key role in the CTGF-induced proliferation of hHSF, but is not the only pathway controlling this process. JAK1 may be an upstream element of STAT1, and may participate in the induction of hHSF proliferation by CTGF. These results elucidate the signal transduction mechanism of CTGF-induced hHSF proliferation, and may aid in the development of a novel method for the inhibition of scar fibrosis and contraction.
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