Preparation of a composite fibrous membrane loaded with mesalazone and metronidazole by interlaced electrospinning

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Abstract. Novel composite fibrous membranes loaded with mesalazine and metronidazole were fabricated via interlaced electrospinning. The fibers were characterized by scanning electron microscopy and FTIR transmission spectra techniques. These characterizations were performed in the aim of optimizing the experimental conditions which allowed us to obtain good morphology of fibrous membrane loaded drugs. The in vitro release experiments revealed that mesalazine and metronidazole were released continuously from the loaded drug fibrous membrane. The fibrous membrane-loaded drugs also showed excellent stability. Compared to those of other drug delivery systems, the main advantage of these two fibrous membrane-loaded drugs is that they can be directly implanted as a lesion after surgery to inhibit recurrence in Crohn’s disease.

Introduction

Crohn’s disease (CD) is an inflammatory bowel disease and its cause is obscure (1,2). Since the symptoms of CD are similar to those of other intestinal disorders, such as irritable bowel syndrome and ulcerative colitis, it can be difficult to diagnose. Ulcerative colitis causes inflammation and ulcers in the top layer of the lining of the large intestine. In CD, all layers of the intestine may be involved, and normal healthy bowel can be found between sections of diseased bowel.

CD is an ongoing disorder that causes inflammation of the digestive tract, also referred to as the gastrointestinal (GI) tract. CD may affect any area of the GI tract, from the mouth to the anus, but most commonly affects the lower part of the small intestine, called the ileum. The swelling extends deep into the lining of the affected organ. The swelling causes pain and makes the intestines empty frequently, resulting in diarrhea.

Treatment may include drugs, nutritional supplements, surgery or a combination of these options. The goals of treatment are to control inflammation, correct nutritional deficiencies, and relieve symptoms such as abdominal pain, diarrhea and rectal bleeding. At present, treatment helps control the disease by lowering the number of times a person experiences a recurrence, but there is no cure. Treatment for CD depends on the location and severity of the disease, complications and the response of an individual to previous medical treatments when treated for recurring symptoms (3).

Chemically, mesalazine (MEZ) is a 5-aminosalicylic acid (4). It is an anti-inflammatory drug structurally related to salicylates, which is active in inflammatory bowel disease (5). Fig. 1A shows its molecular structure. MEZ diminishes inflammation by inhibiting cyclooxygenase and lipoxygenase, thereby decreasing the production of prostaglandins, leukotrienes (CTB4) and hydroxyeicosatetraenoic acids (HETEs), respectively. It appears to be topical rather than systemic. Common oral administration facilitates absorption of MEZ in the small intestine, reduces the time of drug arrival in the colon and causes side effects, such as diarrhea, nausea, cramping, flatulence and headache (6). The recurrence of ulcerative colitis is related to re-breeding infection of clostridium bacteria.

Metronidazole (7) is an amebicide antiprotozoal and antibiotic effective against anaerobic bacteria and certain parasites. Its molecular structure is shown in Fig. 1B. It is the drug of choice for first episodes of mild-to-moderate Clostridium difficile infection (8). Metronidazole kills Bacteroides fragilis and Clostridium perfringens more rapidly than clindamycin (9,10). It also involves the immune system, hampering adhesion between leukocytes and endothelial cells, and inhibiting immigration of inflammatory cells (11). The combination of MEZ and metronidazole alleviates the suffering of the patient during active episodes of ulcerative colitis.

Prevention of post-operative recurrences in CD is a complex problem. In the past, the strategy was to administer no treatment until clinical recurrence. An alternative approach is to administer no treatment, and then to treat the patient according to the severity of the endoscopic recurrence.

Electrospinning is a useful and relatively simple method for producing fibers with a submicrometer diameter range. Using this method, it is possible to obtain ultra-fibers from various types of polymers, such as polyolefins (12), polyamides (13), polyelectrolytes (14), polyurethanes (15), polypeptides and DNA (16,17) as well as polymers with special properties (18). In recent years, probably more than 100 different polymers, copolymers and mixtures of polymers have been successfully used to obtain fibers with diameters of less than 1 μm. Since
MEZ and metronidazole have difficulty dissolving in the same solvent, we developed a new fibrous membrane by interlaced electrospinning, with concurrent loading of the above two drugs. The amount of loaded drugs may be adjusted to a suitable range. These two fibrous membrane-loaded drugs can then be directly implanted as a lesion after surgery, improving the therapeutic index and reducing drug dosage, toxicity and side effects.

Materials and methods

Materials. PLGA (Mn 70,000; LA:GA 50:50) was purchased from the Daigang Biomaterials Co., Ltd. (Jinan, China). MEZ and metronidazole were purchased from the Laifu Chemical Drug Co., Ltd. (Zhousan, China) and the Yancheng Pharmacy (Yancheng, China), respectively. Chloroform, dimethylformamide and ethanol were of analytical grade and used without further purification. Methanol was of chromatographic grade. Tetrabutylammonium bromide was obtained commercially.

Preparation of the fibrous membrane-loaded drugs. Preparation of the different solutions was as follows. PLGA was dissolved in chloroform to prepare a 10% wt solution. Incorporation of the metronidazole (~2% by weight of the polymer) was carried out by dissolving the drug directly in the as-prepared polymer solution. A suitable amount of MEZ was dissolved in DMF, and then chloroform was added (chloroform/DMF, 3/2 weight ratio). PLGA was dissolved in the mixed solvent system to prepare a 10% wt solution.

An interlaced electrospinning setup schematic diagram is shown in Fig. 1. Electrospinning was carried out using a variable high-voltage power supply, which produces voltages ranging from 0 to 10 kV. It consists of two positive-power supplies attached to a blunt steel needle and one negative-power supply attached to the counter electrode. The syringes containing the different solution were attached to the blunt needle and were put into a syringe pump ejecting 50 µl of the solutions per min through the blunt needle. A stainless steel drum (d=15 cm) which may slowly rotate served as the counter electrode. It was covered with aluminum foil for the collection of the different fibers.

Characterization of the fibrous membrane-loaded drugs. Morphological appearances of ultra-fine mats were observed on a scanning electron microscope (SEM; Hitachi S-2350) after gold coating. Fourier transform infrared (FT-IR) spectra of celluloses were obtained with a Nicolet Magna-IR 560 spectrophotometer.

In vitro drug release. Preparation of phosphate-buffered saline (PBS) was as follows. The phosphate buffer solution was prepared by mixing 0.2 M monobasic potassium phosphate (KH₂PO₄) aqueous solution with 0.2 M sodium hydroxide (NaOH) aqueous solution to attain a pH of 6.8.

A piece of the fibrous membrane loaded with the drugs (~2x2 cm²) was placed in 40 ml of 0.05 M, pH 6.8 PBS. The test was performed in a 37°C incubator-shaker at 50 rpm. At appropriate intervals, 2 ml of the supernatant was removed and replenished with an identical volume of fresh buffer. The drug concentrations were determined by a high-performance liquid chromatograph (LabAlliance model 500; USA). Each sample was assayed in triplicate. Then, the accumulative amount of the released drug was calculated as a function of incubation time. The chromatographic conditions (19) were as follows: chromatographic column was a Hypersil-ODS C18 column. The mobile phase was a 80/20 mixture (v/v) of pH 6.5 PBS and methanol, including 0.01 M tetrabutylammonium bromide. The flow rate was 1.0 ml/min, and the detection wavelength was 240 nm.

Stability test. The stability of the fibrous membrane-loaded drugs was tested. Several samples were put in a desiccator at 60°C and a humidity dryer at 25°C as well as 90% relative humidity, respectively, and were kept for 12 days. Moreover, lightfastness experiment was also carried out in a light tester (Huanghai Ls3000; Shanghai, China). Illumination was 4,500±500 Lx. The samples were taken out on the 6th and 12th day, and then their appearance, the amount of loaded drug and rate of release were investigated.

Results and Discussion

The choice of pharmacological treatment for the prevention of post-operative recurrence in CD relies on an informed estimate of the risk of recurrence. Due to different pharmacological action, the combination of MEZ and metronidazole has been applied to decrease the risk of recurrence. The two composite fibrous membrane-loaded drugs were produced by interlaced electrospinning (Fig. 2).

Fig. 3 shows the morphological appearance of the fibers loaded with MEZ or metronidazole. The average diameter of the two fiber samples was ~800 nm. The fibrous surface was smooth, and no drug crystals were detected by electronic microscopy, either on the surface of the fibers or outside the fibers, as shown in Fig. 3. This was attributed to the solubility...
of metronidazole in PLGA/chloroform solution, while MEZ was soluble in PLGA/DMF/chloroform solution. When the solution jet was rapidly drafted and the solvents evaporated quickly, it was difficult to achieve phase-separation, and the drug was prone to remain inside the fiber where there was enough solvent left. When the fiber became dry, the drug was encapsulated inside.

FTIR spectra of the different samples are presented in Fig. 4. Fig. 4c shows characteristic bands of PLGA ~1,756 cm\(^{-1}\) of C=O, 2,900-3,000 cm\(^{-1}\) of C-H, 1,189 and 1,131 cm\(^{-1}\) of C-O, which were also observed in the case of two PLGA fibers loaded with the drugs (Fig. 4b and d). In addition, many new peaks appeared in the IR spectra of the loaded drug PLGA fibers, and the characteristic bands of PLGA after incorporating the drugs strengthened, indicating that the drug was mixed with PLGA, and there existed an interaction between PLGA and the drugs.

To determine the effect of the loaded amount on drug release, the in vitro release experiment was carried out by incubating fibrous membranes containing 1 or 2% wt of MEZ in 0.05 M, and pH 6.8 PBS solutions at 37°C. The results are shown in Fig. 5. No burst release of the drugs was observed, indicating the perfect inclusion of the drugs inside the fibers. With an increase in time, the released amount of MEZ from the fibrous membranes increased steadily. The higher the loaded drug amount, the more rapid the MEZ release rate from the fibrous membrane was. For example, the percentages of drug release of the fibrous membrane loaded with 1 and 2% wt MEZ were 45 and 57% at 15 days, respectively. It showed that the drug release behavior was able to be controlled by adjusting the loaded drug amount. Xu et al (20) also discovered a similar phenomenon when they investigated the in vitro release of BCNU-loaded PEG-PLLA ultra-fine fibers.

Fig. 6a and b illustrates the drug release behaviors of the composite fibrous membranes loaded with 2% wt mesalazine and 2% wt metronidazole. Both drugs from the fibrous membrane were composed of a first rapid release phase and a gradual release phase. The MEZ release rate was slightly more rapid than that of metronidazole, although the molecular weight of MEZ (594) was higher than that of metronidazole (177). Mesalazine may create intramolecular or intermolecular hydrogen bonds in solution, which depress the interaction between polymers and MEZ. Due to degradation of the polymer, corrosion also promoted drug release with an increase in time. Moreover, the dosage of MEZ and metronidazole in the fibrous membranes may be controlled easily through adjustment of the volume ratio of the two spin solutions.
Table I shows the effect of different factors on the stability of the fibrous membrane-loaded drugs. The appearance and color of the samples were not changed. Compared to the initial sample, the loaded drug amount only slightly varied, while the drug release percentage increased slightly on the 12th day. This may be due to the degradation of PLGA after testing.

In conclusion, Crohn's disease is one of the most difficult diseases to therapeutically manage in the field of gastroenterology. We firstly prepared a composite fibrous membrane loaded with MEZ and metronidazole by interlaced electrospinning. The two drugs were entrapped in PLGA fibers, respectively, and there was some interaction between the drugs and the polymer. The in vitro release experiment revealed that the interlaced fibrous membrane was able to be used to control release of the two drugs. The loaded drug amount and the drug release rate may be adjusted by changing the volume ratio of the two spin solutions and drug concentrations, respectively. Fibrous membrane-loaded drugs exhibit excellent stability.

References