Survivin promotes the invasion of human colon carcinoma cells by regulating the expression of MMP-7

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Abstract. Increased expression levels of survivin are crucial for invasion activity in several types of human cancer, including colon carcinoma. However, the molecular mechanisms whereby survivin regulates cancer invasion have not been completely elucidated. To the best of our knowledge, this study is the first to investigate the role of matrix metalloprotease-7 (MMP-7) in cell invasion that is induced by survivin by using in vitro assays, including western blot, immunofluorescence and qPCR analyses. The results demonstrated that the ectopic expression of survivin significantly promoted the invasive activity of colon carcinoma cells (SW620 and HCT-116) and resulted in increased levels of MMP-7 activation. By contrast, the small interfering RNA (siRNA)-based knockdown of survivin markedly reduced expression levels. Compared with the controls, knockdown of MMP-7 by siRNA in colon carcinoma cells led to reduced invasibility, whereas no obvious changes were observed when MMP-7 expression was silenced in survivin-overexpressing colon carcinoma cells. These findings demonstrate that MMP-7 is crucial for survivin-mediated invasiveness, suggesting that the survivin-mediated MMP-7 signaling pathway is a potential therapeutic target for the treatment of colon carcinoma.

Introduction

Survivin, a member of the family of inhibitor of apoptosis (IAP) proteins, contains 142 amino acid residues. Survivin functions as a key regulator of mitosis and programmed cell death by inhibiting apoptosis and promoting cell proliferation (1). Studies have identified that survivin is able to promote the migratory/invasive properties of several types of human cancer, including melanoma, prostate cancer and breast cancer (2-4). Survivin has been demonstrated to regulate the invasion ability of colorectal cancer (CRC) (5,6). However, little is known regarding the functional mechanism of survivin in CRC invasion. A better understanding of the mechanisms by which survivin regulates aggressive cellular behaviors may lead to the production of novel therapeutic targets for colon cancer.

The extracellular matrix (ECM) provides a structural framework to support cells and maintains cell functions by mediating cell-cell or cell-ECM interactions. Degradation of ECM components is mainly controlled by matrix metalloproteinases (MMPs), a large group of enzymes (7). The MMPs comprise a family of zinc-dependent endopeptidases that consist of >21 human proteases and are important in carcinogenesis. One of the smallest known members of the MMP family, MMP-7, was first identified as an enzyme of the involuting rat uterus and was subsequently determined to be an important marker in human cancer progression (8). For example, MMP-7 has been reported to be overexpressed in numerous types of human malignancy, including ovarian, prostate, gastric and breast cancer (9-12), and is important in cancer progression as it promotes tumor invasion. Furthermore, the ability of CRC cells to migrate and invade has been demonstrated to correlate with an increased expression of MMP-7 (13-15). In addition, the overexpression of MMP-7 has been reported to contribute to clinically aggressive behaviors and is a significant predictor of patient survival in CRC (16,17). Certain studies have indicated that upregulation of survivin and MMP-7 expression levels correlates with metastatic CRC and is significantly associated with liver metastasis of colon cancer (18,19). However, the functional correlation between survivin and MMP-7 remains largely unknown.

The present study identified an alternative role for survivin in tumor progression. It demonstrates that the increased invasion of colon carcinoma cells that overexpress survivin is mediated by the activation of MMP-7. Furthermore,
overexpression of survivin enhances MMP-7 expression, whereas silencing endogenous survivin in colon cancer cell lines results in a decreased MMP-7 expression. These findings suggest the molecular mechanism by which survivin enhances cancer invasion. In addition, these data indicate that MMP-7 is a potential mediator of this process.

Materials and methods

Cell culture. 293FT cells were cultured and maintained in Dulbecco's modified Eagle's medium with 10% fetal bovine serum, and used as packaging cells for lentivirus production. RPMI-1640 supplemented with 10% fetal bovine serum was used to separately culture SW620 and HCT-116 cells (American Type Culture Collection, Manassas, VA, USA).

Expression vector construction and viral production. Full-length survivin cDNA was generated by PCR and subcloned into the pReceiver-Lv08 expression vector (Guangzhou Fulengen Co., Ltd., Guangzhou, China). Survivin expression in colon carcinoma cells was ablated using short hairpin RNA (shRNA). Sense (GGACCACCGCAUCUCAGA) and antisense (UGUAGAGCAGCGGGGC) oligonucleotides for the double-stranded cassette were designed and the scrambled sense (UAGAGCAGCGGGGC) and antisense (CACCCUAACAACCGGUAG) sequences were used. These oligonucleotides were synthesized and generated by PCR and subcloned into the psi-U6 vector (Guangzhou Fulengen Co., Ltd.). The 293FT cells were transfected with pReceiver-Lv105/survivin, psi-U6/survivin shRNA or the control plasmid pReceiver-Lv105 and psi-U6 using the ViraPower™ Lentiviral Expression system, the BLOCK-iT™ Lentiviral RNAi Expression system and Lipofectamine® 2000 (Invitrogen Life Technologies, Carlsbad, CA, USA) according to the manufacturer's instructions. The colon carcinoma cells (SW620 and HCT-116) were infected by serial dilution with the virus-containing medium and the stably-transfected cells were selected using puromycin for one week.

siRNA transfection. A small interfering RNA (siRNA) pool against MMP-7 was purchased from Guangzhou Fulengen Co., Ltd. The siRNA was dissolved in siRNA buffer (Guangzhou Fulengen Co., Ltd.) at a concentration of 100 μM and transfected into cells using Lipofectamine 2000. The gene-silencing effect was measured by western blotting 48 h after transfection.

qPCR. To measure the mRNA levels of cell-cycle regulators, total RNA was reverse transcribed using a PrimeScript™ RT Reagent kit (Takara Bio, Inc., Shiga, Japan). qPCR was performed using SYBR® Premix Ex Taq™ II (Takara Bio, Inc.) on a StrataGene Mx3005P system. The primer sequences used were as follows: Forward: TTCTCAAGGACCACCGCATC and reverse: GCCAAGTCTGGCTCTGATC for survivin; and forward: GAGTGAGCTAAGTGGAAC and reverse: CTATGACGCGGGAGTTTACAT for MMP-7. GAPDH was used as an endogenous control. The time points for colon carcinoma cells being transfected with MMP-7-specific siRNA were 12, 24, 48, 72 and 96 h. All samples were normalized to internal controls and fold-changes were calculated using relative quantification ($2^{-\Delta\Delta C_{t}}$).

Western blot analysis. Protein was extracted by solubilizing cells in RIPA buffer containing a protease and phosphatase inhibitor cocktail (Keygen Biotech. Co. Ltd., Nanjing, China). Protein concentration was quantitated with Bradford protein assay reagent (Keygen Biotech. Co. Ltd.), and 40 μg of total protein was loaded onto 4-12% polyacrylamide gels, separated and then transferred onto a polyvinylidene difluoride membrane. The membrane was probed with mouse anti-Survivin, MMP-7 (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA) and proteins were detected with enhanced chemiluminescence detection reagents (Keygen Biotech. Co. Ltd.). Protein loading was normalized by probing the membrane with anti-GAPDH antibodies.

Immunofluorescence analysis. Cell lines were plated on culture slides (Corning Incorporated, Tewksbury, MA, USA) and, after 24 h, were rinsed with phosphate-buffered saline (PBS) and fixed in methanol-acetone for 30 min at room temperature. The cells were permeabilized with 0.5% Triton X-100 for 30 min and blocked for 45 min in 10% bovine serum albumin (Sigma-Aldrich, St. Louis, MO, USA) in PBS, and then incubated with primary monoclonal antibodies in PBS for 2 h at room temperature. After three washes in PBS, the slides were incubated for 1 h in the dark with secondary goat anti-mouse antibodies (Invitrogen Life Technologies). After another three washes, the slides were stained with 4',6-diamidino-2-phenylindole (Sigma-Aldrich) for 5 min to visualize the nuclei and were examined using an Eclipse TE300 microscope (Nikon, Tokyo, Japan).

Boyden chamber assay. Biocoat™ Growth Factor-Reduced Matrigel™ invasion chambers (8.0 μm PET membrane; 24-well cell culture inserts; BD Biosciences, Franklin Lakes, NJ, USA) were preincubated in RPMI-1640 supplemented with 1% fetal bovine serum for 1 h. Harvested cells (2x10⁶ cells) in 100 μl RPMI-1640 supplemented with 1% fetal bovine serum were replated onto the upper chamber and placed in the lower chamber with 500 μl RPMI-1640 supplemented with 10% fetal bovine serum. After 20 h, the cells were fixed with methanol. Non-migrated cells on the upper side of the filter were removed with a cotton swab and cells on the underside of the filter were stained with hematoxylin. Images were captured using an Eclipse TE300 microscope. Relative cell migration was determined by the mean number of migrated cells in eight random fields on the underside of the filter. For each experiment, three independent filters were analyzed.

Statistical analysis. Statistical analysis was performed using SPSS software, version 13.0 (SPSS, Inc., Chicago, IL, USA) and the data are expressed as the mean ± standard deviation. Comparisons were conducted using Student’s t-test. P<0.05 was considered to indicate a statistically significant difference.

Results

Overexpression of survivin enhances colon carcinoma cell invasion. To investigate the potential role of survivin in cell
invasion, cell invasion was examined in human colon carcinoma SW620 and HCT-116 cells overexpressing survivin (pRe-Sur) were collected and survivin protein levels were analyzed by western blotting. GAPDH expression was the internal control for loading of protein. (B) Colon carcinoma cells were transfected with survivin control vector and survivin (pRe-Sur), and the motility of the cells was evaluated by using Boyden chamber assay. Original magnification, x400. Bars represent the mean ± SD. *P<0.01. pRe-Sur, pReceiver-survivin.

Knockdown of survivin impairs colon carcinoma cell invasion. To confirm the role of survivin in the invasion of colon carcinoma, RNAi-mediated knockdown of survivin was performed to suppress survivin in SW620 and HCT-116 cells.

Survivin regulates MMP-7 expression in colon carcinoma cells. To examine the effect of survivin on MMP-7 expression, ectopic expression and siRNA-based knockdown of survivin were performed in colon carcinoma SW620 and HCT-116 cells. Western blot analysis and immunofluorescence demonstrated
that the overexpression of survivin induced upregulation of MMP-7 (Fig. 3A and E). By contrast, MMP-7 expression levels were markedly suppressed in survivin-silenced colon carcinoma cells (Fig 3B and E). Furthermore, decreased expression levels of survivin led to a dose-dependent reduction in MMP-7 expression levels (Fig. 3C and D; P<0.01).

**Role of MMP-7 in survivin-mediated invasion.** To confirm the importance of MMP-7 in survivin-induced cell invasion, RNAi-mediated silencing of MMP-7 was performed in SW620 and HCT-116 cells and resulted in effective reduction of the endogenous MMP-7, as determined by qPCR (Fig. 4A). Notably the optimum reduction required 48 h of RNAi-mediated silencing. Subsequently, the protein expression of survivin and MMP-7 was assessed following cell culture for 48 h. As shown in Fig. 4B, MMP-7 expression levels were reduced successfully using RNAi, as revealed by western blot analysis. Notably, the knockdown of MMP-7 did not alter the expression levels of survivin in colon carcinoma cells in comparison with the controls. Compared with the controls, reduced MMP-7 expression levels in the colon carcinoma cells significantly decreased the cell capacity for invasion.

Figure 3. MMP-7 expression in survivin- or survivin shRNA-transfected SW620 and HCT-116 cells. (A) Cells were collected and MMP-7 protein levels were analyzed by western blotting. Survivin increased MMP-7 activation in survivin-transfected cell lines. (B) Knockdown of survivin expression levels decreased MMP-7 expression levels in Sur-shRNA-transfected cell lines, as identified by western blotting. GAPDH expression was the internal control for loading of protein. Expression levels of MMP-7 after survivin siRNA infection at different doses (nM) in SW620 cells by (C) qPCR and (D) western blot analysis. (E) Immunofluorescence staining reveals increased expression levels of survivin and MMP-7 in survivin-overexpressing cells (pRe-Sur) and reduced expression levels in survivin-silenced cells (Sur-shRNA), in comparison with the vector controls. Original magnification, x400. MMP-7, metalloprotease-7; pRe-Sur, pReceiver-survivin; shRNA, short hairpin RNA; siRNA, small interfering RNA.
invasion, as evaluated by Boyden chamber assay (P<0.01). However, no significant difference was observed in the cells also overexpressing survivin (P≥0.05) (Fig. 4C).

Discussion

CRC is the third most common type of cancer worldwide. The majority of individuals who are diagnosed with this disease are already at an advanced stage (20). Despite significant advances in early diagnosis and treatment, the survival rate of patients has not yet markedly improved, which is mainly due to the high rate of localized recurrence and distant metastasis following surgical resection (21). The exact mechanisms underlying the aggressive progression of CRC remain to be clarified. Therefore, elucidating the underlying mechanisms that drive CRC cell invasion and metastasis may aid in the exploitation of strategies for effective CRC treatment.

Survivin is the smallest member of the IAP family of anti-apoptotic proteins. It is highly expressed in tumors and is thought to promote tumor progression through apoptotic inhibition and mitotic chromosomal alignment (1,22). Studies have demonstrated that overexpression of survivin is correlated with increased invasion and metastasis of human cancer, such as melanoma, prostate cancer, breast cancer and endometrial carcinoma (2-4,23). Similar to these observations, the present study, to the best of our knowledge, has demonstrated for the first time that the overexpression of survivin is capable of enhancing cell invasion in the SW620 and HCT-116 colon carcinoma cell lines. Studies performed with the use of chemically synthesized siRNAs or plasmid/viral vectors encoding shRNAs have suggested that RNAi-mediated survivin knockdown is able to suppress cancer cell proliferation and promote caspase-dependent apoptosis in a number of human tumor cell models (24). Studies from our laboratory and findings of other studies have demonstrated that inhibition of survivin using shRNA and siRNA influences the biological features of colorectal carcinoma cells and inhibits invasion and metastasis of the human colon carcinoma cell line SW480 in vitro (5,25). Similar to these observations, the present study found that RNAi-mediated knockdown of survivin is capable of significantly impairing cell invasion in SW620 and HCT-116 colon carcinoma cells. Therefore, through two methods of upregulating and inhibiting the expression of survivin in CRC cells, the results of the present study strongly indicate that survivin is required for tumor invasion in CRC. Notably, it has been demonstrated that inhibition of survivin is an effective therapeutic strategy in preclinical and clinical studies (26). The aforementioned studies suggest that survivin may be used as a potential target for cancer treatment in CRC patients.

Figure 4. Role of MMP-7 in the survivin promotion of colon carcinoma cell motility. (A) Colon carcinoma cells were transfected with scrambled siRNA (the control) or MMP-7-specific siRNA, then cells were collected after 12, 24, 48, 72 and 96 h and MMP-7 mRNA levels were analyzed by qPCR. (B) Survivin and MMP-7 protein levels were analyzed in the colon carcinoma cells and in the cells overexpressing survivin by using western blotting 48 h after transfection with MMP-7 siRNA. (C) Silencing MMP-7 expression in colon carcinoma cells reduced their motility ability, as evaluated by Boyden chamber assay. However, decreased MMP-7 expression levels did not correlate with significant changes in the motility ability of cells overexpressing survivin. *P<0.01 compared with the control. MMP-7, metalloproteinase-7; siRNA, small interfering RNA.
Studies have revealed that survivin enhances cell proliferation and suppresses apoptosis through a number of pathways (27,28). The exact mechanisms by which survivin exerts this effect on the invasion ability of CRC remain largely unknown. In the present study, overexpression of survivin enhanced cell invasion and resulted in increased MMP-7 activation in SW620 and HCT-116 colon carcinoma cells. Conversely, knockdown of survivin using RNAi decreased MMP-7 expression levels. Notably, decreased expression levels of survivin led to a dose-dependent decrease in MMP-7 expression levels. However, knockdown of MMP-7 expression did not affect the expression levels of survivin in colon carcinoma cells compared with that in the controls. Similarly, Chu et al demonstrated that survivin knockdown downregulates the expression of MMPs in other types of colon carcinoma cell (6). Furthermore, the present study includes the primary investigation of the role of MMP-7 in cell invasion induced by survivin. Silencing of MMP-7 expression in the colon carcinoma cells reduced the cell invasion ability. By contrast, MMP-7 knockdown in transfected-survivin colon carcinoma cells did not contribute to significant changes in cell invasion. Overall, these results demonstrate that survivin may promote the invasion of colon carcinoma cells through the regulation of MMP-7 expression.

In conclusion, these data strongly suggest that upregulation of MMP-7 expression levels by survivin contributes to cell invasion, and that MMP-7 is a putative therapeutic target in the invasive progression of colon carcinoma. Additional investigation is necessary to elucidate the regulatory mechanism between survivin and MMP-7.

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References