Colon cancer cells treated with 5-fluorouracil exhibit changes in polylactosamine-type N-glycans

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Abstract. 5-Fluorouracil (5-FU) is the major chemotherapeutic agent for the treatment of colorectal carcinoma, which were found to have N-glycans containing polylactosamine on the cancer cell surface. Alterations in the expression and structure of polylactosamine glycans are associated with cellular differentiation and oncogenesis. However, little is known with regard to the correlation between the levels of polylactosamine expressed in colon cancer cells and the anticancer effect of 5-FU. In the present study, SW620 cells were treated with the half maximal inhibitory concentration (IC50; determined by MTT-assay) of 5-FU. Hoechst 33258 staining and flow cytometric analysis indicated that 5-FU administration resulted in apoptosis in SW620 cells. An increased percentage of cells in S phase was also observed among the SW620 cells treated with 5-FU. Under the same experimental conditions, a decrease in the 5-FU-induced inhibition of polylactosamine glycans was recorded. However, an increase in the activity of alkaline phosphatase was also observed. Furthermore, pretreatment of the SW620 cells with 5-FU inhibited the expression of β1,3-N-acetylgalactosaminyltran sferase-8 (β3Gn-T8) and cluster of differentiation (CD)147 in a time-dependent manner. Overall, changes in glycosylation were associated with the anticancer effect of 5-FU in the colon cancer cells. In conclusion, polylactosamine may be a useful target for the identification of substances with anticancer activity.

Introduction

5-Fluorouracil (5-FU) remains a widely used anticancer drug. An improvement in the response and survival rates in breast, head and neck cancer has been observed following administration of 5-FU and other chemotherapeutical agents. However, the largest impact has been observed in colorectal cancer. As a pyrimidine analogue, 5-FU interferes with nucleoside metabolism and is incorporated into RNA and DNA, finally leading to cell cycle arrest and apoptosis. There are numerous identified and potential modulators that contribute to the 5-FU response in cancer cells. For example, celecoxib combined with 5-FU has been shown to enhance tumor cell apoptosis and anticancer efficacy in a subcutaneous implantation tumor model of colon cancer (1). Also, the inhibition of O-glycosylation by benzyl-α-N-acetylgalactosamine resulted in significant antiproliferative activity of 5-FU against pancreatic cancer cells (2). In addition, 5-FU has the potency to change cellular glycosylation in various cell types (3).

Colon cancer cells frequently contain glycans at different levels or with fundamentally different structures than those observed on normal cells. Recently, Yang et al confirmed that colon cancer was associated with antigenic and structural changes in mucin-type O-glycans (4). In addition, Barrow et al identified that in human colon cancer cells, the suppression of core 1 galactose (Gal)-transferase is linked to a decrease in the Thomsen-Friedenreich (TF) and a corresponding increase in O-[2-(acetylamino)-2-deoxy-α-D-galactopyranosyl]-L-serine (Tn), sialyl-Tn and Core 3 glycans (5). In another study, core fucosylated high mannose N-glycans were detected in colorectal cancer tissues using hydrophilic interaction liquid chromatography with matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (6). Furthermore, Hahne et al (7) analyzed the N-glycosylation profiles of two colon carcinoma cell lines, SW480 (primary tumor) and SW620 (metastatic tumor). There was a significant downregulation of high-mannose glycans in the metastatic cells. Therefore, it is necessary to identify changes in glycans and to investigate the consequences of blocking or modifying glycosylation in colon cancer cells following 5-FU treatment. However, little is known with regard to glycan changes following 5-FU treatment during colon cancer progression.

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The glycosylation of polylactosamine uses repeating Galβ1-4-glucosyl (Glc)-N-acetyl-(Ac)-β1-3 disaccharide units that are preferentially joined to β1-GlcNAc-linked antennae that are connected to the trimannosyl core of the complex-type N-linked oligosaccharides (8). It has been reported that highly metastatic colon cancer cells synthesize more N-glycans that contain polylactosamine than cells with low metastasis (9). In the present study, the human colon cancer cell line SW620 was pretreated with the half maximal inhibitory concentration (IC_{50}) of 5-FU for a short term. The apoptosis induction and cell cycle arrest that occurred within the first 48 h was examined. Under the same experimental conditions, the correlation between the levels of polylactosamine expressed in colon cancer cells and the anticancer effect of 5-FU were also investigated. Characterization of the glycan changes in response to short-term drug treatment in colorectal cancer cells will facilitate an improved understanding of the multiple mechanisms involved in drug response. Alterations in glycans result from disruptions in the expression levels of the glycosyltransferases. β3Gn-T8 is an enzyme involved in the synthesis of polylactosamine on β1,6-branched N-glycans in colon cancer (10). Western blot analysis revealed that the expression of β3Gn-T8 in the SW620 cells was markedly suppressed by 5-FU (Fig. 5B). CD147 is a transmembrane glycoprotein containing polylactosamine-type glycans (11).

Materials and methods

Cell culture. The human colon cancer cell line SW620 was purchased from the Institute of Biochemistry and Cell Biology, Chinese Academy of Science (Shanghai, China), and cultured in RPMI-1640 medium (Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum at 37°C under 5% CO₂.

Cell viability assays in vitro. SW620 cells were seeded onto a 96-well plate, with 5x10³ cells in 180 µl culture medium being added to each well. Next, the cells were exposed to different concentrations of 5-FU [0.01 x peak plasma concentration (PPC), 0.1 x PPC, 1 x PPC and 10 x PPC] for 48 h. The effect of 5-FU on cell viability was then determined for each time-point by adding 20 µl MTT (5 mg/ml; Sigma-Aldrich, St. Louis, MO, USA) to each well and incubating the cells for 4 h. The reaction was stopped by the addition of 150 µl dimethyl sulfoxide, and the absorbance of the samples was then measured at 570 nm by means of an ELISA plate reader (BioTek Instruments, Inc., Winooski, VT, USA). The experiments were repeated at least three times. The concentration of 5-FU causing 50% inhibition (IC_{50}) was determined by plotting the toxin concentrations versus percentages of inhibition (on a Probit scale), as calculated from the absorbance data.

Hoechst 33258 staining. The cells were seeded at a density of 5x10³ cells/ml in culture medium onto a 3.5-cm sterile plate and incubated with 40 µg/ml 5-FU for 48 h. Subsequently, the cells were fixed with 4% formaldehyde. The fixed cells were then incubated with 1 mg/ml nuclear fluorochrome Hoechst 33258 (Beyotime Biotechnology, Jiangsu, China) at room temperature for 10 min in the dark. The stained cells were observed under a fluorescence microscope.

Flow cytometry. Apoptosis was identified and quantified using flow cytometry with Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide (PI) double-staining. In brief, 2x10⁶ SW620 cells were treated with 40 µg/ml 5-FU for 48 h, washed twice with cold PBS and then gently re-suspended in 195 µl binding buffer. Thereafter, 5 µl Annexin V-FITC (20 µg/ml) and 5 µl PI (20 µg/ml; both Beyotime Biotechnology) were added. The cells were gently vortexed and incubated for 15 min while protected from light. Subsequent to adding 400 µl IX binding buffer to each tube, the cells were analyzed using flow cytometry (Becton-Dickinson, Mountain View, CA, USA) within 1 h.

Cell cycle analysis. The cells were treated with 40 µg/ml 5-FU for 48 h. The cultured cells were trypsinized and fixed with 70% ethanol at 4°C overnight prior to being stained with PI using freshly prepared staining solution. The distribution of cells in the different phases of the cell cycle was measured by flow cytometric analysis. The data were analyzed using CellQuest software (Becton-Dickinson).

Western blot analysis. Subsequent to 5-FU treatment (40 µg/ml) for 24 or 48 h, western blot analysis was conducted using standard methods. Briefly, equal amounts of protein (30 µg/lane) from total cell lysates were separated by 10% SDS-PAGE and transferred onto a polyvinylidene difluoride membrane. The membrane was blocked with 1% bovine serum albumin in Tris-buffered saline buffer [10 mM Tris and 150 mM NaCl (pH 7.9)] containing 0.05% Tween 20, and the proteins were analyzed using specific antibodies as follows: Horseradish peroxidase-conjugated secondary antibodies and an enhanced chemiluminescence (ECL) kit were used for detection (both Beyotime Biotechnology). Rabbit anti-human β1,3-N-acetylglucosaminyltransferase-8 (β3Gn-T8) polyclonal antibody was purified by our laboratory (12). Goat-anti-human cluster of differentiation 147 (CD147) antibody and mouse anti-human β-actin antibody were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA).

Flow cytometric analysis of cellular glycosylation. The cells were washed, collected from the plates and then centrifuged at 1,500 x g for 3 min; the precipitate was resuspended in 100 µl PBS. Next, the cells were incubated with 0.5 µg/ml biotin-conjugated Lycopersicon esculentum agglutinin lectin (LEL; Sigma-Aldrich) for 1 h at 37°C. Cells were then washed and bound lectin was detected with phycoerythrin-conjugated Streptavidin (Sigma) for 30 min at 37°C. Cell samples were subjected to flow cytometry, with unstained cells serving as controls. Fluorescence histograms and mean fluorescence data were created and analyzed using CellQuest software (Becton-Dickinson).

Lectin blot. The cells were harvested and lysed, proteins extracted from the cells were electrophoresed by 10% SDS-PAGE and polyvinylidene difluoride membranes were prepared as mentioned in the western blot analysis method. Following blocking with Carbo-Free Blocking Solution (Vector labs, Burlingame, CA, USA), the membranes were incubated with 2 µg/ml of various biotinylated *Lycopersicon esculentum* agglutinin (tomato) lectins (LEL, TL; Vector labs) for 30 min. The reactive bands were
detected with a diluted horseradish peroxidase-conjugated streptavidin (Vector labs) and then visualized using an ECL system (GE Healthcare, Little Chalfont, UK).

**Measurement of alkaline phosphatase (ALP) activity.** For assessment of ALP, the ALP detection kit (Nanjing Jiancheng Bioengineering Institute, Nanjing, China) was used according to the manufacturer’s instructions. The cells were seeded at a density of 1x10^4 cells/ml and treated with 40 µg/ml 5-FU prior to being assayed for ALP activity. Next, the homogenates were centrifuged at 12,000 x g for 30 min at 4˚C. The supernatants were subjected to quantification of protein with a Bradford assay and to an ALP activity assay using a detection kit. The enzymatic activities were expressed as U/g (protein).

**Statistical analysis.** Values are expressed as the mean ± standard deviation. P<0.05 was used to indicate a statistically significant difference. Statistical analyses were performed using SPSS 11.5 (SPSS, Inc., Chicago, IL, USA) and each experiment was repeated three times.

**Results**

**Determination of the IC_{50}-value of 5-FU.** In the present study, the effect of the conventional cytotoxic drug, 5-FU, was studied on the viability of SW620 cells. The PPC of 5-FU was 10 µg/ml (13), therefore, the cells were treated with 5-FU at concentrations of 0.1, 1, 10 and 100 µg/ml for 48 h. As shown in Fig. 1, 5-FU caused an evident reduction of cell viability, and the inhibitory effect was dose-dependent. The IC_{50}-value of 5-FU on SW620 cells was calculated to be 13 µg/ml. All the successive experiments were therefore performed on SW620 cells treated with 13 µg/ml 5-FU.

**Induction of apoptosis in SW620 cells by 5-FU treatment.** To check whether 5-FU has a role in the regulation of apoptosis, the morphological changes of SW620 cells were examined using Hoechst 33258 staining. When the cells were stained with Hoechst 33258, live cells with uniformly light blue nuclei were observed under a fluorescence microscope, while apoptotic cells exhibited bright blue nuclei due to karyopyknosis and chromatin condensation, and necrotic cells can not be stained. As shown in Fig. 2A, the number of Hoechst 33258-positive apoptotic cells following 5-FU treatment was increased compared with the control, demonstrating that the cells were apoptotic. Furthermore, the apoptosis triggered by 5-FU was confirmed by annexin V-FITC and PI double-staining and quantification by flow cytometry. As shown in Fig. 2B, incubation of SW620 cells

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with 13 µg/ml 5-FU for 48 h increased the percentage of apoptotic cells to 9.29%, compared with the control, which contained only 1.05% apoptotic cells (P<0.05).

**5-FU increases cell cycle arrest.** 5-FU is an antimetabolite that acts by specifically blocking cells in the S-phase. As shown in Fig. 3, 5-FU caused a significant increase in the percentage of SW620 cells in S phase, and a decrease in cells in G₀/G₁ and G₂/M phases following incubation with 5-FU for 48 h. Compared with the untreated cells, the percentage of cells in S-phase was increased from 22.36 to 54.48% (P<0.05). However, in the presence of 5-FU, the percentage of SW620 cells in G₀/G₁ phase was decreased from 58.95 to 28.62%, while the percentage of cells in G₂/M phase was decreased from 18.68 to 16.89% (P<0.05).

**5-FU decreases polylactosamine levels in colon cancer cells.** The plant lectins LEL preferentially recognize glycopeptides containing longer polylactosamine repeats. As shown in Fig. 4A, the treatment of SW620 cells with 5-FU markedly reduced LEL staining, indicating the effectiveness of 5-FU treatment in reducing lactosamine addition to N-glycans (P<0.05). The intensity of LEL staining is presented as the X-mean value. As shown in Fig. 4A, treatment of SW620 cells with 5-FU markedly reduced LEL staining, indicating the effectiveness of 5-FU treatment in reducing polylactosamine. In the untreated cancer cells (control group) and the cells treated with 13 µg/ml 5-FU for 24 h (5-FU 24 h group) or 48 h (5-FU 48 h group), the X-mean was 5.53, 5.12 and 4.69, respectively. (B) The X-mean values of the colorectal cancer cells by flow cytometric analysis at different time points. (C) Analysis of polylactosamine by lectin blot at different time points. FU, fluorouracil; LEL, *lycopersicon esculentum* agglutinin lectin.
membranes. The data revealed that polylactosamine levels were generally decreased in the glycoproteins in the SW620 cells following 5-FU treatment (Fig. 4C). The lectin blot assay revealed similar results to those of the flow cytometric assay. These data indicated that 5-FU generally altered the polylactosamine structures of the N-glycans among the glycoproteins of SW620 cells. To the best of our knowledge, the present study is one of the first to focus on the changes of polylactosamines in human colon cancer cells affected by 5-FU.

5-FU increases cell differentiation. The structures of polylactosamines are often characteristic for different cell types and stages of differentiation. ALP has been used to monitor the differentiation effect of certain anticancer compounds. Therefore, in the present study, the ALP activity was observed. Untreated SW620 cells exhibited relatively low ALP activity (Fig. 5A), while in cells treated with 13 µg/ml 5-FU, the activity of ALP reached high levels in a time-dependent manner. There was a significant difference between the control and treated cells (P<0.05).

Changes in the expression of β3Gn-T8 and CD147 following 5-FU treatment. Western blot analysis revealed that the expression of β3Gn-T8 in the SW620 cells was markedly suppressed by 5-FU (Fig. 5B). Treatment of SW620 cells with 5-FU also resulted in a marked decrease in CD147 at the protein level in a time-dependent manner (Fig. 5B). These results were consistent with those of the lectin blot analysis in vitro.

Discussion

Multiple pathways are known to be targeted by 5-FU, and as a consequence, its effect may be modulated by a number of genes and gene products within those pathways. Not all the major targets and pathways affected by 5-FU are known, which limits the possibilities of predicting the effectiveness of the drug in individual patients. Therefore, understanding the mechanism that contributes to the response of cancer cells to 5-FU is important for determining how to control tumor growth and therapy. The present study has specifically focused on colon cancer, a major malignancy with insufficient treatment options and one of the leading causes of cancer mortality throughout the world. In the present study, the polylactosamine structures of N-glycans in glycoproteins were demonstrated to be generally altered following 5-FU treatment in colon cancer cells, although the data are not conclusive.

As a major anticancer drug for the treatment of colorectal carcinoma, 5-FU has modest clinical activity at standard doses, and in general, dosing is limited by its safety profile. As a result, decisions with regard to the therapeutic dose of 5-FU are challenging for clinicians. In the present study, the IC_{50}-value of 5-FU on colon cancer cells over a 48-h treatment period was determined in vitro using the MTT assay. The obtained IC_{50}-value in regard to a 48-h exposure of SW620 to 13 µg/ml 5-FU was comparable with that previously observed (14). Since the PPC of 5-FU in colon cancer patients is ~10 µg/ml, the data of the present study are clinically relevant. However, it was confirmed that treatment with 5-FU was able to induce variable degrees of apoptosis in cultured cells. Apoptosis is a form of cell death defined by a characteristic set of morphological and biochemical changes. The data in the present study, obtained from Hoechst 33258 staining and flow cytometric analysis, indicated that 5-FU administration also resulted in apoptosis in SW620 cells. In addition, pretreatment with 5-FU induced an accumulation of SW620 cells in the S phase of the cell cycle. Observations from the present study have shown that 5-FU has a significant role in the therapy of human colon cancer.

Altered glycosylation is a universal feature of cancer cells, and certain glycan structures are well-known markers for tumor progression (15). It is well known that the expression of N-glycans as constituents of cell surface glycoproteins is essential for the regulation of various processes in tumor cell biology. The amount of tetra-antennary and triantennary N-glycans is often associated with the increased amount of polylactosamine (16). This is in particular due to the β1,6 branch of the N-glycans being one of the favored sites for polylactosamine substitutions (17). There are two major roles with regard to the function of polylactosamines. Alterations in polylactosamines of tumor cells are correlated with carcinogenesis, invasion and metastasis (18). Thus, the inhibition of glycan synthesis reduces tumorigenicity. Furthermore, the structures of polylactosamines are often characteristic of different cell types and stages of differentiation (19). Saitoh et al showed that the polylactosamine content was decreased after colon cancer CaCo-2 cells were differentiated and lost their tumorigenicity (9). The results of the present study further revealed that 5-FU specifically affected the expression of polylactosamine, as indicated by lectin blot and flow cytometric assays. In addition, ALP activity was examined as a marker for cellular differentiation in human colon cancer cells (20). Similar to the change of glycans, treatment with 5-FU increased ALP activity in cancer cells in the present study. Thus, it may be hypothesized that microenvironmental changes in the synthesis of glycans greatly affect their synthetic efficiency and also their structures. However, it has been challenging to analyze polylactosamine in a comprehensive and
quantitative manner, since a very low amount of glycans is available in biological samples.

The polylactosamine of N- and O-glycans is coordinately synthesized by the alternate action of β1-4-galactosyltransferases and β3Gn-Ts (21). In different types of glycoconjugates and cells at various stages of differentiation, the presence of β3Gn-T is also completely different. In comparison to normal tissue, the majority of colorectal cancer tissues that have been examined have been found to have significantly higher levels of the β3Gn-T8 transcript (10). The result of the present study revealed that the expression of β3Gn-T8 in SW620 cells was evidently suppressed by 5-FU. Notably, the potential anticancer activity of 3-azido-3-deoxythymidine, which is able to inhibit the synthesis of polylactosamine, is able to be modulated by combining it with 5-FU (22,23). In this manner, the differential expression of β3Gn-T8 in SW620 cells that is responsible for polylactosamine chain elongation may be affected by 5-FU and further studies are required to confirm this.

Furthermore, polylactosamine glycosylation is performed preferentially to select proteins. CD147 is a major carrier of β1,6-branched polylactosamine sugars on tumor cells (24). In the present study, the treatment of SW620 cells with 5-FU resulted in a marked decrease of CD147 at the protein level. Traditionally, the biosynthesis of the glycans in glycoproteins is regulated by a number of factors, including i) Expression of associated glycosyltransferases and/or glycosidases, ii) proper locations and iii) the functional machinery of sugar nucleotides (25). The present observations of downregulated CD147 expression lead to the hypothesis that one of the mechanisms underlying the anticancer effect of 5-FU on colon cancer is the suppression of the biosynthesis of polylactosamines. However, it remains to be elucidated whether other genes exist that encode for proteins with this activity.

In conclusion, 5-FU has been shown to exhibit anticancer effects by the induction of apoptosis and cell cycle arrest. 5-FU also decreased polylactosamine in colon cancer cells. It may thus be a novel drug target candidate. In conclusion, the present study indicates that polylactosamine may be a potential target that contributes to the 5-FU response in colon cancer cells. It may thus be a novel drug target candidate for the treatment of colon cancer.

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