Increased cytochrome c correlates with poor survival in aggressive lymphoma

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Abstract. Mitochondria are central to a variety of cellular processes, from metabolism to cell death. In this study, we demonstrated that an increase in the critical mitochondrial protein, cytochrome c, correlated with drug resistance in a cell culture model of aggressive lymphoma. Increased cytochrome c expression was also correlated with decreased survival in the aggressive diffuse large B-cell and mantle cell lymphomas, but not in the indolent follicular lymphoma. This suggests that an increased reliance on the mitochondria for energy allows tumor cells to be metabolic opportunists and contribute to tumor development and drug resistance.

Introduction

Apoptosis resistance and altered metabolism are common cancer cell traits. In a murine lymphoma model, selection for oxidative stress resistance results in concomitant resistance to apoptosis and the acquisition of a more-aggressive tumor phenotype. WEHI7.2 thymic lymphoma cells transfected with catalase (CAT2, CAT38) or selected for resistance to hydrogen peroxide (200R) demonstrate resistance to glucocorticoid-induced apoptosis (1,2), one component of the CHOP lymphoma chemotherapy regimen. The oxidative stress-resistant cells are also resistant to the cyclophosphamide, doxorubicin and vincristine components of CHOP (Tome et al, unpublished data). Catalase-overexpressing cells exhibit increased tumor growth compared to WEHI7.2 parental cells in a mouse xenograft model (2). Additionally, the oxidative stress-resistant cells demonstrate an altered metabolic profile, including the ability to generate ATP from alternative carbon sources such as glutamine (3,4). When treated with glucocorticoids, the oxidative stress-resistant cells are better able to maintain ATP levels as compared to WEHI7.2 cells (3). Experiments with an uncoupler of mitochondrial respiration showed that the CAT38 and 200R cells produce more ATP from mitochondria than the WEHI7.2 cells (4).

Mitochondria are central to cellular metabolism and the decision to undergo apoptosis (5). Thus, changes in critical mitochondrial proteins may explain the more-aggressive tumor phenotype of the resistant lymphoma variants. In particular, cytochrome c plays an integral role as an electron carrier in the mitochondrial respiratory chain, and the release of this protein from the mitochondrial intermembrane space results in commitment to death via apoptosis (5). In a previous study, we showed that cytochrome c release is delayed in the resistant variants (1), and that mitochondria isolated from these cells demonstrate an intrinsic resistance to cytochrome c release (Wilkinson et al, unpublished data). The present study examined whether alterations in cytochrome c protein levels correlate with the previously characterized more-aggressive tumor phenotype of the resistant cells. To test the potential clinical relevance of these findings, gene expression data from lymphoma specimens were analyzed.

Materials and methods

Mitochondrial protein levels in cell lines. WEHI7.2 murine thymic lymphoma cells and variants were maintained as previously described (1,2). Mitochondria were isolated as previously described (2), with the exception that the mitochondrial isolation buffer was prepared in the same manner as Wang et al (6). Catalase-overexpressing cells exhibit increased tumor growth compared to WEHI7.2 parental cells in a mouse xenograft model (2). Additionally, the oxidative stress-resistant cells demonstrate an altered metabolic profile, including the ability to generate ATP from alternative carbon sources such as glutamine (3,4). When treated with glucocorticoids, the oxidative stress-resistant cells are better able to maintain ATP levels as compared to WEHI7.2 cells (3). Experiments with an uncoupler of mitochondrial respiration showed that the CAT38 and 200R cells produce more ATP from mitochondria than the WEHI7.2 cells (4).

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Gene expression profiling data. The gene expression profiling (GEp) data sets used in this analysis were generated by the Leukemia/Lymphoma Molecular Profiling Project (LLMPP) research group (7-9) and are publicly available from http://llmpn.nih.gov/. Diagnostic specimens were collected from 240 de novo diffuse large B-cell lymphoma (DLBCL).
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Patients subsequently received multi-agent chemotherapy and were monitored to assess treatment outcome. GEPI was performed using the lymphochip microarray for D:lBcl and M:cl and the Affymetrix U133A and U133B microarrays for Fl.

Genes of interest were subjected to expressed sequence tag validation and statistical analysis as previously described (10). For cytochrome c expression in diffuse large B-cell, mantle cell and follicular lymphoma. Kaplan-Meier plots were used to compare patient survival with the highest cytochrome c expression (solid line) with that of patients with the lowest cytochrome c expression (dashed line). The percentage of patients used for analysis is indicated on each graph. Cyt c, cytochrome c; NDH, NADH dehydrogenase; SDH, succinate dehydrogenase; CCRI, cytochrome c reductase I; COX, cytochrome c oxidase; F_0F_6, ATPase F_0F_6.

Results

Increased mitochondrial cytochrome c protein expression in a drug-resistant lymphoma model. Cytochrome c protein levels were increased in mitochondria from CAT2, CAT38 and 200R cells relative to WEHI7.2 (Fig. 1A). Increased protein levels were also noted for AK-2, another mitochondrial intermembrane space protein (Fig. 1A). This protein supports optimal ATP synthesis by equilibrating adenylate pools (11). The increased levels of cytochrome c and AK-2 found in mitochondria from the oxidative stress-resistant lymphoma cells are proportionate to the degree of apoptotic resistance measured previously in intact cells (2). No difference in expression of the mitochondrial matrix protein Hsp60 was found in the cell variants. Thus, the changes observed in cytochrome c and AK-2 content cannot be explained by increased mitochondrial mass. These data suggest that the resistant variants have an increased mitochondrial respiratory capacity.

Correlation of increased oxidative phosphorylation (OXPHOS) proteins with poor prognosis in diffuse large B-cell lymphoma. Based on the characteristics of the lymphoma variants observed in this and a previous study (3), an increased expression of cytochrome c and other genes involved in OXPHOS is expected to correlate with a poor response to chemotherapy in lymphoma patients. To test this, we analyzed a GEPI data set from D:lBcl patient samples. A previous study used this GEPI data set to develop an outcome predictor score (OPS) that accurately predicted patient survival following chemotherapy (7). Ten genes encoding for proteins involved in OXPHOS, including cytochrome c, were included on the LLMP microarray (Table I). We first identified cytochrome c as significantly correlated with OPS since an increased expression was associated with poor patient prognosis (p<0.05; Table I and Fig. 1B). Additionally, the increased expression of NADH dehydrogenase, succinate dehydrogenase, cytochrome c reductase I, cytochrome c...
Table I. Correlation of mitochondrial protein gene expression in diffuse large B-cell lymphoma patient tumor samples with prognosisa.

<table>
<thead>
<tr>
<th>Gene</th>
<th>10% best prognosisb Mean</th>
<th>10% worst prognosisb Mean</th>
<th>t-test; best 10% vs. worst 10% P-value</th>
<th>Regression all patients P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cytochrome c</td>
<td>0.1230</td>
<td>0.4475</td>
<td>0.0498c</td>
<td>0.0001c</td>
</tr>
<tr>
<td>NADH dehydrogenase</td>
<td>0.0315</td>
<td>0.2270</td>
<td>0.0486c</td>
<td>0.0010c</td>
</tr>
<tr>
<td>Succinate dehydrogenase</td>
<td>0.0899</td>
<td>0.1310</td>
<td>0.7941</td>
<td>0.0531c</td>
</tr>
<tr>
<td>Cytochrome c reductase I</td>
<td>-0.1857</td>
<td>0.2987</td>
<td>0.0016c</td>
<td>&lt;0.0001c</td>
</tr>
<tr>
<td>Cytochrome c reductase II</td>
<td>0.2167</td>
<td>0.2987</td>
<td>0.5866</td>
<td>0.7159</td>
</tr>
<tr>
<td>Cytochrome c reductase bp</td>
<td>0.0367</td>
<td>0.0739</td>
<td>0.7155</td>
<td>0.6744</td>
</tr>
<tr>
<td>Cytochrome c oxidase</td>
<td>-0.3338</td>
<td>0.6360</td>
<td>0.0003c</td>
<td>&lt;0.0001c</td>
</tr>
<tr>
<td>ATPase F6F0</td>
<td>0.0022</td>
<td>0.1405</td>
<td>0.2291</td>
<td>0.0132c</td>
</tr>
<tr>
<td>ATPase F1β</td>
<td>0.1261</td>
<td>0.1604</td>
<td>0.8237</td>
<td>0.0872</td>
</tr>
<tr>
<td>ATPase F0F1</td>
<td>0.1425</td>
<td>0.1512</td>
<td>0.9531</td>
<td>0.3684</td>
</tr>
</tbody>
</table>

aPrognosis was measured by the outcome predictor score, a value which accurately predicts overall patient survival. bValues represent the mean relative mRNA expression on a log2 scale for the 10% of patients with the best and worst prognosis from the LLMPP microarray. cDenotes p≤0.05.

Discussion

This study places mitochondria in the crossroads of successful adaptation to oxidative stress and carcinogenesis. The mitochondrial changes observed in our lymphoma model extend previous findings of metabolic flexibility in the oxidative stress-resistant cells (3,4). Our attempts to use continuous ethidium bromide treatment as a technique to deplete mitochondrial DNA (13) with the WEHI7.2 cells were unsuccessful (data not shown). The inability to create WEHI7.2 rho0 cells lacking functional OXPHOS suggests that reliance on mitochondrial respiration is critical for lymphoma cells. This complements the observed increase in cytochrome c and AK-2 levels, as well as an increased ability to maintain mitochondrial ATP generation (3), in the resistant variants. A novel conclusion from the present study, building on our previous biochemical analyses (3,4), is that oxidative stress may provide selective pressure for the concurrent development of apoptosis resistance and the metabolic adaptations seen in malignant cells. Since lymphoma can arise in the context of chronic infection and inflammation, our investigations may have implications for both current and novel therapies.

Increasing the mitochondrial respiratory proteins allows the oxidative stress resistant variants to be metabolic opportunists. Studies in HeLa cells show that growth in galactose/glutamine instead of glucose leads to increased dependence on OXPHOS (14). This biochemical change occurs via increased respiratory chain protein levels without increasing the mitochondrial mass. Thus, an increase in respiratory chain proteins, such as cytochrome c, may indicate enhanced OXPHOS capability and account for metabolic flexibility in the aggressive variants.

Our analyses show an increased expression of cytochrome c and other mitochondrial respiration proteins in aggressive lymphoma. Oxidative phosphorylation is critical for successful lymphoma chemotherapy, and our data supports the hypothesis that targeting this metabolic pathway could be a viable therapeutic strategy.
lymphomas from patients with the worst prognosis, suggesting that results from the lymphoma cell model are clinically relevant. The up-regulation of respiratory chain proteins in the DLBCL patients with the worst prognosis is consistent with an ability to use alternative energy substrates such as glutamine. Some studies suggest that tumor cells suppress OXPHOS as a protective mechanism to limit the production of endogenous reactive oxygen species (12). However, in our oxidative stress-resistant model and in tumors arising in the context of chronic inflammation, the ability to tolerate increased oxidative stress may allow them to take advantage of increased mitochondrial respiration.

We have shown that an increased cytochrome c expression is correlated with decreased survival in the aggressive DLBCL and MCL. The up-regulation of alternate pathways for ATP production would allow cancer cells to be metabolic opportunists in a tumor microenvironment where specific nutrients are limiting. This could provide both a growth advantage during tumor development, as well as protection from apoptosis-inducing chemotherapy. A correlation between cytochrome c expression and overall survival was not observed in the indolent FL, suggesting that for a less-aggressive cancer, increased OXPHOS capability would be less critical. Additionally, the majority of follicular lymphoma cases are characterized by overexpression of \textit{BCL2} (8), and it is possible that this prominent survival advantage overshadows metabolic changes.

Our results suggest that an increased ability to use mitochondria to produce ATP contributes to tumor development. The WEHI7.2 variants with the mitochondrial alterations are resistant to multiple lymphoma chemotherapeutics (Tome \textit{et al}, unpublished data), suggesting that these mitochondrial characteristics also contribute to chemoresistance in the clinic. Targeting such mitochondrial alterations with novel chemotherapeutics may improve clinical outcome for these aggressive lymphomas.

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\textbf{References}