Neuroprotective effect of matrine on MPTP-induced Parkinson's disease and on Nrf2 expression

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Abstract. The incidence rate of Parkinson's disease (PD) is ≤2% in Chinese individuals >65 years old, accounting for 40% of the global total of PD patients. The pathogenesis of PD is not yet clear, and oxidative stress-induced mitochondrial dysfunction is considered to be the main reason for the onset of PD. Studies have shown that matrine exhibits good antioxidant activity. Thus, the present study aimed to observe the protective effect and mechanism of matrine on 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced dopaminergic neuron damage. A total of 25 C57BL male mice were randomly divided into 5 groups, consisting of the control group (group A), the MPTP group (group B) and three matrine (4, 8 and 16 mg/kg) plus MPTP treatment groups (groups C, D and E, respectively). Results from a pole-climbing test and locomotor activity experiments were recorded. The mice were sacrificed 4 days later and brain dissection was performed. The levels of superoxide dismutase (SOD) and glutathione (GSH) were assessed. The expression level of tyrosine hydroxylase (TH) in the ventral midbrain was studied by western blot analysis. The expression level of nuclear factor erythroid 2-related factor 2 (Nrf2) in the ventral midbrain was studied by immunofluorescence analysis. The expression level of TH and levels of TH-positive cells. Western blotting results showed that the expression of Nrf2 in the ventral midbrain decreased significantly in the PD mice, and that the mice administered matrine showed higher expression of TH and levels of TH-positive cells. Western blotting results showed that the expression of Nrf2 in the ventral midbrain decreased significantly in the PD mice, and that matrine was able to reverse this phenomenon. In conclusion, by promoting antioxidant-related Nrf2 signaling pathways in the ventral midbrain, matrine can inhibit the oxidative damage of dopamine neurons in PD.

Introduction

Oxidative stress is involved in the development and progression of various neurodegenerative diseases, including Parkinson's disease (PD), Alzheimer's disease and amyotrophic lateral sclerosis (ALS). PD is a progressive neurodegenerative disease with an unknown pathogenesis, and the loss of substantia nigra dopaminergic neurons is characteristic of its lesions (2,3). Oxidative stress-induced mitochondrial dysfunction is hypothesized to be the main reason for the pathogenesis of PD (6). After neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) induction, PD can be modeled \textit{in vitro}. MPTP can cross the blood-brain barrier, and is converted into its active metabolite methyl-4-phenylpyridine (MPP+), which due to affinity with the dopamine transporter, is thus selectively transported to the mitochondria of the dopaminergic neurons. As a neurotoxic metabolite, MPP+ can block cell respiration and promote active oxygen \[, reactive oxygen species (ROS) formation\], thus inducing the death of dopaminergic neurons (7).

While traditional levodopa replacement therapy and certain non-steroidal anti-inflammatory drugs can reduce the clinical symptoms, they do not prevent progression of the disease and long-term medication can cause serious adverse reactions (8,9). Matrine can be obtained from plants of the \textit{Sophora} genus and has always been used in traditional Chinese medicine to treat inflammation (10). Matrine has been shown to produce a wide range of pharmacological effects...
and has been used to treat a variety of diseases, including viral hepatitis, neuropathic pain and isoproterenol-induced heart disease (11-13). In addition, significant antitumor effects have been found in gastric cancer, rhabdomyosarcoma, acute myeloid leukemia and breast cancer (14,15), and studies have shown that matrine exhibits antioxidant effects in a number of diseases. PD is mainly caused by damage to dopamine neurons, and oxidative stress is one of its important pathogenetic factors. There is little literature on the interaction between matrine and the MPTP-induced damage to mouse dopaminergic neurons in PD. Accordingly, the present study investigated whether matrine has a protective effect on dopaminergic neurons, and the viral mechanisms involved were studied.

Materials and methods

Materials. C57BL, 7 to 8-month-old, male mice (weighing 20-25 g) were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. (Beijing, China). The mice were housed in a thermostatically controlled environment with set lighting conditions (lighting time, 7:30 a.m. to 7:30 p.m.). A total of 25 mice were randomly divided into five groups, namely the control group (group A), the MPTP group (group B) and three matrine (4, 8 and 16 mg/kg) plus MPTP treatment groups (groups C, D and E, respectively). The control group received saline by intraperitoneal injection (30 mg/kg/day for 4 days), and the MPTP group was continuously administered an intraperitoneal injection of 30 mg/kg MPTP for 4 days (once a day) to create the PD mouse model. The matrine + MPTP groups were treated with different doses of matrine (4, 8 and 16 mg/kg) in advance, 8 h prior to intraperitoneal injection with MPTP.

The study was approved by the Ethics Committee of the College of Basic Medical Sciences, Jilin University (Changchun, Jilin, China).

Equipment, drugs and reagents. An ultra-pure water system (Milli-Q Synthesis) was purchased from Millipore (Darmstadt, Germany) and an automatic embedding machine (model no. EG-1140C) was purchased from Leica Microsystems, Inc. (Buffalo Grove, IL, USA). A slicing machine (model no. X-202A) was purchased from Guangdong Yi Mai Technology Co., Ltd. (Guangdon, China), an inverted phase contrast microscope was obtained from Olympus Corporation (model no. BX51), and constant current regulator electrophoresis (model no. DYC-40C) and semi-dry transfer equipment (model no. DYY-8B) instruments were purchased from Beijing Liuyi Biotechnology Co., Ltd. (Beijing, China). Matrine (catalog no. CDS016735), MPTP (catalog no. M0896) and rabbit anti-mouse tyrosine hydroxylase (TH) antibody (catalog no. T8700) were purchased from Sigma-Aldrich (Millipore). Rabbit anti-mouse nuclear factor E2-related factor 2 (Nrf2; catalog no. 12721P) and rabbit anti-mouse β-actin (catalog no. 12620; dilution, 1:10,000) antibodies were purchased from Cell Signaling Technology, Inc. (Danvers, MA, USA). The concentrated DAB kit was purchased from Zhongshan Golden Bridge Biotechnology Co., Ltd. (Beijing, China; catalog no. ZLI-9017), the superoxide dismutase (SOD) test kit (catalog no. A001-3) and the glutathione (GSH) test kit (catalog no. A006) were purchased from Nanjing Jiancheng Bioengineering Institute (Nanjing, Jiangsu, China), and the CytoBuster protein extraction reagent was purchased from Novagen Inc. (Madison, WI, USA; catalog no. 7109-3).

Establishment and execution of a mouse model of PD. Under a constant temperature and lighting conditions (lighting time, 7:30 a.m. to 7:30 p.m.), the mice were continuously treated for 4 days, and suspension and climbing experiments were conducted every day. On the fourth day, all mice were decapitated, then the mouse brain striatum and substantia nigra were isolated; one portion was used for protein extraction, and the other portion was stored at room temperature after being embedded in paraffin.

Suspension experiment. The C57BL mice were placed on a horizontal wire of ~1.5 mm in diameter, suspended 30 cm from the ground, and the hang time was recorded to detect mouse limb coordination. Scoring criteria: 0-5 sec, 0 points; 6-10 sec 2 points; 11-15 sec, 3 points; 16-20 sec, 4 points; and >20 sec, 5 points.

Pole-climbing test. A tube of ~30 cm in length and ~1 cm in diameter was gauze wrapped and a wooden ball was attached to the top. Mice were placed on top of the wooden ball and the time required for the mouse to traverse from the top to the bottom of the tube was recorded. Time were compared prior to and after PD modeling.

SOD and GSH detection. The levels of SOD and GSH were detected according to the manufacturer's instructions provided in the SOD (catalog no. A001-3) and GSH (catalog no. A006) test kits (Nanjing Jiancheng Bioengineering Institute).

TH antigen test. The paraffin sections were made into frozen sections and dried at room temperature for 15 min, the stained tissue was marked by Pap Pen, soaked for 10 min in PBS to remove the OCT and then film-fixed at room temperature for 30 min with 4% paraformaldehyde. The sections were treated for 10 min with 2.2% Triton X-100, and then washed with PBS twice and blocked with 10% normal goat serum in PBS at room temperature for 1 h. TH antibody was added (1:3,000 dilution) and the section was incubated at 4˚C, followed by being washed with PBS twice. The Alexa Fluor 488-labeled goat anti-rabbit immunoglobulin G secondary antibody was added and the section was incubated at 37˚C for 1 h prior to being washed again with PBS twice. Fluorescence microscopy was performed on five randomly selected fields, and the number of positive cells was counted and analyzed statistically.

Nigrostriatal protein extracts. The nigrostriatal tissue was homogenized, then washed twice with ice-cold PBS and centrifuged at 400 x g for 5 min. The supernatant was discarded, and the 100X protease inhibitors and CytoBuster protein extraction reagent were added to the precipitate, pipetted and mixed at room temperature for 15 min, prior to centrifugation at 12,000 x g for 15 min at 4˚C. The resulting supernatant contained the desired protein.
Western blotting. The extracted protein was isolated using 10% SDS-PAGE gel separation and a 5% stacking gel, and then semi-transferred to nitrocellulose membranes using incubation in TBST containing 5% BSA at room temperature for 2 h. Nrf2 rabbit anti-mouse antibodies (1:1,000 dilution) were added and the membranes were incubated at 4˚C overnight. The next day, the membrane was washed with 0.1% TBST 3 times, for 5 min each, and the horseradish peroxidase (HRP)-labeled goat anti-rabbit secondary antibody was added for incubation at room temperature for 1 h. 0.1% TBST was used to wash the membrane, using Supersignal West Femto/ Pico HRP-sensitive chemiluminescent substrate for coloration. Rabbit anti-mouse β-actin (1:10,000 dilution) was used as an internal control. All experiments were repeated at least 3 times.

Statistical. Using SPSS 15.0 statistical software (SPSS, Inc., Chicago, IL, USA), multiple sets of data were compared using a one-way analysis of variance, and differences between two groups of data were compared using the Student-Newman-Keuls analysis method. All data are presented as the mean ± standard deviation. P<0.05 was considered to indicate a statistically significant difference.

Results

Effect of matrine on the suspension ability of mice with PD. The control mice exhibited a mean suspension ability score of 5. Compared with the control group, the mean score in the MPTP mice was significantly lower (P<0.001), while matrine administration significantly alleviated this phenomenon (P=0.004). The suspension ability score increased with increasing matrine concentration (P<0.001) (Fig. 1).

Effect of matrine on pole-climbing ability. Compared with the control group, MPTP mice exhibited a significantly reduced capacity for pole-climbing, with a significantly longer climb time (P<0.001). The administration of matrine was able to significantly alleviate this phenomenon (P=0.008), and the pole-climbing ability recovered more significantly with increasing matrine concentration (P<0.001) (Fig. 2).

Mouse brain tissue SOD and GSH assay. Compared with the control group, the MPTP mice exhibited significantly lower SOD and GSH activity in the brain tissues (P<0.001), while the administration of matrine significantly alleviated this phenomenon (P<0.001). The brain activity recovery increased more significantly with increasing matrine concentration (P<0.001) (Fig. 3).

Immunohistochemistry. In the comparison of dopamine TH expression in the substantia nigra and the striatum, relative to the control group, the number of TH-positive cells of the MPTP group significantly decreased (P<0.001). In the matrine-treated mice, a higher expression level of TH and a greater number of TH-positive cells was evident (Fig. 4).
Western blot analysis detecting the expression of Nrf2. The relative intensity of antioxidant-related Nrf2 molecules decreased significantly in the MPTP group, while in the matrine-treated mice, stronger Nrf2 expression was exhibited in the substantia nigra and striatum, which increased in a concentration-dependent manner (Fig. 5).

Discussion

PD is a progressive neurodegenerative disease whose main symptoms include bradykinesia, resting tremors and rigidity (16-18). In the present study, using an MPTP-induced mouse PD model, reduced motion bradykinesia, which is typical of PD, was apparent along with other symptoms of the disease. Oxidative stress is the cause of a number of neurodegenerative diseases, including PD (6). Due to a high metabolic rate, brain tissue is prone to hypoxia, and this may lead to an increase in reactive oxygen species (ROS) and oxidative stress. SOD is one of the key enzymes in the oxidative stress defense and is key to good health (19). GSH, formed from a combination of glutamic acid, cysteine and glycine, is a tripeptide containing a thiol group, and has antioxidant and detoxification integration (20). The activity of SOD and GSH may reflect the free radical scavenging ability of the body.

When PD occurs, brain tissue SOD and GSH activity is significantly reduced. In the present study, SOD and GSH activity was significantly reduced in the cerebral tissue of the MPTP mice. In addition, immunofluorescence showed a significant reduction in the TH expression of MPTP mice at the substantia nigra and the striatum. The described animal model meets the requirements established for PD in this study.

Matrine obtained from plants of the Sophora genus has always been used in traditional Chinese medicine to treat inflammation (10). Studies have shown that matrine can prevent the steatohepatitis caused by a high carbohydrate diet (21) through its antioxidant effects. PD is mainly caused by a loss of dopamine neurons, and oxidative stress is important in the pathogenesis. Little literature is available on whether matrine can protect dopamine neurons in mice with MPTP-induced PD and the specific mechanisms involved. The results of the present study showed that following the administration of matrine treatment, a number of the typical symptoms of PD significantly improved, SOD and GSH activity appeared significantly increased in the brain tissue, and a higher level of TH expression and more TH-positive cells were evident. This suggests that matrine may have a significant therapeutic effect on PD.
Nrf2 can regulate the expression of 200 genes, including a number of antioxidant genes (22). Nrf2 protects against the islet β cell damage caused by acute oxidative stress (23). The present study further examined the matrine treatment of PD by regulating the expression of Nrf2 to investigate how matrine protect dopamine neurons. The results showed that Nrf2 expression in the substantia nigra and the striatum, in a concentration-dependent manner. In summary, the results of the present study confirmed that matrine exhibited a significant therapeutic effect in mice with PD and demonstrated that the mechanism of matrine treatment in PD may be the inhibition of oxidative damage of dopamine neurons by the promotion of antioxidant-related Nrf2 signaling pathways.

References