miR-219-5p suppresses the proliferation and invasion of colorectal cancer cells by targeting calcyphosin

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Abstract. MicroRNAs (miRNAs) are small non-coding RNAs involved in an array of biological processes, and their dysregulation is associated with tumor development and progression. One such miRNA, miR-219-5p, is abnormally expressed in patients with colorectal cancer (CRC). In the present study, reverse transcription-quantitative polymerase chain reaction was performed to measure miR-219-5p expression in cells from both CRC tumors, and surrounding healthy tissue. MTT and invasion assays were used to determine the role of miR-219-5p in regulating CRC cell proliferation and invasion, respectively. A luciferase assay was then performed to assess the binding of miR-219-5p to the CAPS gene that encodes calcyphosin protein. The present study confirmed that miR-219-5p expression is significantly downregulated in CRC tissue. miR-219-5p knockdown promoted the growth of HCT-8 cells and increased the expression of calcyphosin protein (CAPS). On the other hand, overexpressing miR-219-5p inhibited HCT-8 cell growth and invasion, and downregulated CAPS expression. In addition, CAPS was identified as the functional downstream target of miR-219-5p by directly targeting its 3'-untranslated region. Therefore, miR-219-5p may function as a tumor suppressor by decreasing CAPS expression, and subsequently inhibit tumor proliferation and invasion. These results indicate that novel therapeutic strategies that increase miR-219-5p expression may be developed to treat CRC.

Introduction

Colorectal cancer (CRC) is one of the most common malignancies worldwide, and has multiple risk factors, associated with lifestyle, genetics and the environment (1). Currently, the main therapeutic strategy used to treat CRC is surgical resection, however its effectiveness is limited in patients with locally invasive or metastatic disease. Therefore, understanding the mechanisms of CRC disease progression is essential in identifying potential biomarkers and improving CRC patient prognosis and treatment.

MicroRNAs (miRNAs) are small non-protein coding RNAs 20-24 nucleotides long, that negatively regulate gene expression by binding to the 3’ untranslated region (3’UTR) of corresponding target messenger RNAs (mRNAs) (2). Abnormal miRNA expression is thought to contribute to tumorigenesis and carcinoma progression of various human cancers (3). Previous studies have demonstrated that analysis of the genes targeted by miRNA may identify altered miRNA regulatory networks involved in tumor pathogenesis; and miRNA itself may be used to identify cancer-specific signatures for CRC diagnosis (4). A number of studies have demonstrated that downregulating the expression of tumor suppressor miRNAs, such as miR-219-5p, may increase CRC development and progression (5). miR-219-5p inhibits papillary thyroid carcinoma cell growth and invasion by targeting estrogen receptor 1 (6), and targets glypican-3, inhibiting hepatocellular carcinoma cell proliferation (7). Furthermore, miR-219-5p regulates the receptor tyrosine kinase pathway in glioblastoma by targeting the epidermal growth factor receptor (8), and may regulate Sall4 in CRC cells (5).

Calcyphosine, a calcium binding protein of 24 kDa, was originally isolated from the canine thyroid cDNA library as a major phosphorylated substrate for protein kinase A following stimulation of thyroid cells by thyrotropin (8). As it is phosphorylated in a cAMP-dependent manner, it is considered to be implicated in the cross-signaling between these cascades to coordinate cellular proliferation and differentiation (9). Calcyphosine is also expressed in the brain, salivary glands and lungs of dogs (10). Moreover, calcyphosine functions as a candidate for cross-talk between cAMP-mediated and inositol trisphosphate/Ca2+ mediated signal transduction pathways (11). Calcyphosine was thus identified as a tumor-specific protein that may serve as a tumor marker for a novel subgroup of ependymomas and as a potential drug target for therapy in pediatric brain tumors (12). The present study aims to identify a novel target gene of miR-219-5p and demonstrate that calcyphosin protein (CAPS) is targeted by miR-219-5p in human CRC, to elucidate the mechanism by which miR-219-5p...
suppresses CRC progression. This would allow the clinical value of miR-219-5p in treating CRC patients to be evaluated.

**Materials and methods**

**Tissue specimens and cell culture.** Fresh CRC tissue specimens, and specimens from adjacent non-tumor tissues, were collected from 34 patients with CRC, who received surgical treatment at the Affiliated Hospital of Nantong University between May 2013 and May 2014. None of the patients had received prior neoadjuvant treatment. Samples were immediately frozen in liquid nitrogen, and stored at -80°C. All human tissue was collected using protocols approved by the Ethics Committee of Affiliated Hospital of Nantong University.

**Cell culture and transfection.** Human ileocecal colorectal adenocarcinoma HCT-8 cells obtained from the Type Culture Collection of Chinese Academy of Sciences (Shanghai, China) were maintained in RPMI-1640 Medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (FBS, Invitrogen; Thermo Fisher Scientific, Inc.) in a humidified incubator with a mixture of 5% CO₂ at 37°C. HCT-8 cells were then transfected with a scrambled miR-219-5p inhibitor (Thermo Fisher Scientific, Inc.), miR-219-5p mimics, or miRNA, which acted as a negative control (NC). For CAPS functional analysis, the coding sequence of CAPS was amplified and sub cloned into the pcDNA3.1 (+) vector (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer instructions, in order to induce overexpression of endogenous CAPS. Following this, HCT-8 cells were transfected either with an empty vector or a CAPS-expressing plasmid, using lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.).

**Reverse transcription-quantitative polymerase chain reaction (RT-qPCR).** Total RNA was extracted using a Trizol® Plus RNA purification kit (Invitrogen; Thermo Fisher Scientific, Inc.) and cDNA was synthesized from total RNA treated with DNase I using an Omniscript reverse transcription kit (Qiagen Inc., Valencia, CA, USA) according to the manufacturer's instructions, in order to induce overexpression of endogenous CAPS. Following this, HCT-8 cells were transfected either with an empty vector or a CAPS-expressing plasmid, using lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.).

**Western blot analysis.** Protein from tumor tissue was extracted by lysis buffer containing protease inhibitors. Wells were loaded with 10 µg/well of protein from each sample and then transferred to a polyvinylidene fluoride membrane. Blocking was subsequently carried out for 2 h using 5% non-fat milk in a mixture of Tris-buffered saline and Tween 20 (TBST). Following incubation with the primary monoclonal rabbit anti-human CAPS antibody (cat. no. ab186741, 1:1,000, Abcam, Cambridge, MA, USA) overnight at 4°C, the membranes were washed with TBST three times for 10 min, and then incubated with horseradish peroxidase-conjugated goat anti-rabbit secondary antibody (cat. no. ab97051, 1:5,000, Abcam) for 12 h at 4°C. The ECL western blotting detection reagent (GE healthcare life sciences, Chalfont, UK) was used to detect signals prior to film development. β-actin was used as the loading control.

**MTT assay.** The MTT assay was used to determine the effect of miR-219-5p on cellular proliferation. A total of 1x10⁴ cells containing 200 µl RPMI-1640 supplemented with 10% FBS were plated in each well of a 96-well plate. Following 1, 2, 3 days of incubation, 20 µl MTT (5 mg/ml; Sigma-Aldrich, St. Louis, MO, USA) was added, followed by 4 h incubation at 37°C in a 5% CO₂ incubator. The supernatant was subsequently removed and 150 µl dimethyl sulfoxide was added. To dissolve MTT crystals, all plates were shaken for 10 min at room temperature. The absorbance values of each sample were read at 490 nm using a micro plate reader (Model 550, Bio-Rad Laboratories, Shanghai, China). Each experiment was repeated ≤3x.

**Colony formation assay.** Following transfection of miR-219-5p mimics (50 nmol/l), miR-219-5p inhibitor (50 nmol/l) or scramble (NC; 50 nmol/l), HCT-8 cells were plated in a 24-well plate for 24 h. The cells were then collected and seeded (1,000 cells/well) in a fresh 6-well plate for 14 days. Surviving colonies (>50 cells per colony) were counted following fixation with 1:1 methanol and aceticone, and staining with 5% Gentian Violet (ICM Pharma Pte. Ltd., Singapore, Singapore). The experiment was carried out in triplicate.

**Transwell assay.** A transwell assay (BD Biosciences, Franklin Lakes, NJ, USA) was used to perform cell invasion according to the manufacturer's instructions. Mimics and inhibitors of miR-219-5p transfected cells were harvested 24 h following
transfection. 2.0x10^5 transfected cells or untreated cells in serum-free medium were then added to each upper insert pre-coated with Matrigel matrix, and 600 µl 10% FBS medium was added to the matched lower chamber. Following incubation, non-invasive cells were removed from the upper surface of the Transwell membrane by a cotton swab. Invasive cells on the lower membrane surface were fixed with methanol, stained with 0.1% crystal violet, photographed, and counted.

**Dual-luciferase reporter assay.** HCT-8 cells were seeded in 24-well plates (1x10^5 cells/well) and incubated for 24 h prior to transfection. For the reporter gene assay, the cells were co-transfected with either 0.5 µg pGL3-CAPS-3'UTR wild-type or pGL3-CAPS-3'UTR mutant plasmid, 0.05 ng pRL-TK control vector (Promega Corporation, Madison, WI, USA), and 50 nmol/l either miR-219-5p mimic or scramble, using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). The firefly and renilla luciferase activities were measured consecutively through a dual luciferase assay (Promega Corporation) 24 h after transfection. Luciferase intensity was measured using the Dual Luciferase Reporter assay system (Promega Corporation,) according to the manufacturer instructions.

**Statistical analysis.** All statistical analyses were performed using SPSS software ver. 17.0 (SPSS, Inc., Chicago, IL, USA). Results were compared using either Student’s t-test or χ² test. One-way analysis of variance (ANOVA) test was used to analyze the significance of differences between groups of various differentiations. All data were presented as mean ± standard deviation (SD). P<0.05 was considered to indicate a statistically significant difference.

**Results**

Over-expression of miR-219-5p inhibits proliferation of CRC cells in vitro. RT-qPCR detected significantly decreased miR-219-5p expression in CRC tissue compared with the adjacent non-tumor tissue (P=0.003). U6 snRNA was used as internal control. The proliferation of human ileocecal colorectal adenocarcinoma HCT-8 cells transfected with miR-219-5p mimics, inhibitor, or scramble (control), was measured by MTT assay. The number of colonies were evaluated by a colony formation assay of HCT-8 cells after treatment with miR-219-5p mimics, inhibitor, or scramble (control). *P<0.05 miR-219-5p mimic vs. scramble; #P<0.05 miR-219-5p inhibitor vs. scramble.

CAPS-3'UTR is a potential functional target of miR-219-5p. Previous studies have demonstrated that miR-219-5p suppresses the expression of several genes, such as Sal14, by targeting their 3'-UTR in CRC (5). To identify new potential target genes of miR-219-5p, the websites TargetScan (http://www.targetscan.org/) and miRanda (http://www.microrna.org/) were used. Bioinformatic analysis demonstrated that miR-219-5p directly targets the CAPS gene.
To validate this, the wild-type and mutant sequences of the CAPS 3’UTR were cloned into luciferase reporter vectors (sequences shown in Fig. 2A). The luciferase assay revealed that miR-219-5p significantly suppressed the luciferase activity of CAPS containing a wild-type 3’UTR, however it did not suppress the activity of CAPS containing a mutant 3’UTR (P<0.05, Fig. 2B). This confirms that miR-219-5p targets the 3’UTR of CAPS.

miR-219-5p mimics significantly decreased CAPS protein and mRNA levels compared to controls, whereas miR-219-5p inhibitors increased levels of CAPS mRNA and protein in HCT-8 cells (Figs. 2C and E). This was confirmed by western blot analysis (Fig. 2D).

Furthermore, the mRNA level of CAPS in the 34 pairs of tumor and adjacent non-tumor tissue were analyzed by RT-qPCR. CAPS mRNA levels were significantly higher in CRC tissue than in healthy tissue (P<0.001, Fig. 3A). Furthermore, a significant negative correlation was observed between miR-219-5p and CAPS expression in CRC samples (r²=-0.47, P<0.001, Fig. 3B). Taken all together, the results of the current study suggest that miR-219-5p decreases the expression of CAPS in CRC by directly targeting CAPS 3’UTR.

Restoration of miR-219-5p inhibits CAPS-mediated CRC cell invasion. Western blot analysis indicated that HCT-8 cells transfected with the pcDNA3.1 (+)-CAPS may enhance CAPS protein expression compared with cells transfected with an empty vector control (Fig. 4A). Furthermore, the Transwell assay demonstrated that HCT-8 cells transfected with miR-219-5p mimic and pcDNA3.1 (+)-CAPS overexpression of miR-219-5p (312±12.46) exhibited significantly decreased migratory capacity compared with CAPS-induced CRC cells transfected with CAPS and a scramble (403±18.53; Fig. 4B, P<0.05).

Discussion

Recently, the role of miRNAs in tumorigenesis and their regulatory function in a number of biological processes associated with cancer has been investigated. In human cancer, miRNAs are usually located in genomic breakpoint regions and function as tumor suppressor genes or oncopgenes during tumor development and progression (14). miR-219-5p may act as a tumor manipulator in various types of cancers by targeting downstream oncogenic molecules, thus attenuating
the malignant features of hepatocellular carcinoma (7), papillary thyroid carcinoma (6), glioblastoma (15) and colorectal cancer (5).

In the present study, analysis by RT-qPCR demonstrated that miR-219-5p is downregulated in CRC tissue compared with adjacent non-tumor tissues. Evidence gathered from numerous studies highlights the significance of miRNAs as being vital in the regulation of numerous physiological processes such as cell proliferation, differentiation and apoptosis (16-18). Both the MTT and colony formation assays demonstrated that miR-219-5p significantly suppressed the growth and proliferation of CRC cells in vitro. By contrast, downregulating miR-219-5p expression promoted CRC cell proliferation.

Numerous studies have indicated that miRNAs are deregulated in CRC, and the importance of this deregulation is highlighted by the importance of the target genes they regulate (19-21). The current study sought to identify other potential targets of miR-219-5p to further understand how miR-219-5p suppresses tumors in CRC. Target prediction algorithms identified the binding sites for miR‑219‑5p in the 3'-UTR of CAPS.

CAPS belongs to the calmodulin superfamily of calcium-binding proteins and is regulated by cyclic AMP through protein phosphorylation (22). It is phosphorylated in a cAMP-dependent manner and thus may be implicated in the cross-signaling between these cascades to coordinate cell proliferation and differentiation (9), suggesting its involvement in carcinogenesis. It has been proven that CAPS is expressed in many tumors; overexpression of CAPS occurs in ependymoma (23), lung cancer (24), ovarian cancer (25) and endometrial cancer (26).

In conclusion, the present study identified CAPS as a direct target of miR-219-5p in CRC cells. CAPS was upregulated...
in CRC tissues, and miR-219-5p expression in CRC tissues was inversely correlated with CAPS expression: Increasing miR-219-5p expression downregulated the expression of CAPS protein, while knockdown of miR-219-5p increased CAPS protein levels. Furthermore, CAPS-induced cell invasion was reversed by miR-219-5p expression. Therefore, miR-219-5p may function as a tumor suppressor and inhibit tumor proliferation and invasion by regulating CAPS expression.

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References