Overexpression of N-cadherin and β-catenin correlates with poor prognosis in patients with nasopharyngeal carcinoma

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Abstract. An increasing amount of evidence demonstrates that epithelial-mesenchymal transition (EMT) is important in tumor invasion and metastases. The cell-cell adhesion molecule N-cadherin and the Wnt/β-catenin cascade protein β-catenin are two biomarkers of EMT. The present study aimed to measure the expression levels of N-cadherin and β-catenin in samples from patients with nasopharyngeal carcinoma (NPC) and evaluate their prognostic significance. N-cadherin and β-catenin mRNA was evaluated using reverse transcription-quantitative polymerase chain reaction in 26 NPC tissue samples and 8 nasopharyngeal epithelium samples. Protein expression of N-cadherin and β-catenin was also detected using immunohistochemistry in 128 archival NPC paraffin-embedded specimens. Finally, associations between clinical pathological parameters and prognostic values in NPC were evaluated. The results demonstrated that both the mRNA and protein levels of N-cadherin and β-catenin were significantly increased in NPC tissues compared with the controls. Enhanced expression of N-cadherin and β-catenin protein was strongly correlated with the status of lymph node metastasis and clinical stages in patients with NPC. Notably, high expression of N-cadherin and β-catenin proteins was significantly correlated with lower overall survival (OS) rate in patients with NPC. Finally, multivariate analysis demonstrated that expression of N-cadherin protein and clinical stages were independent prognostic factors for patients with NPC. Therefore, the present study demonstrated that N-cadherin and β-catenin expression may be used as potential prognostic biomarkers for patients with NPC.

Introduction

Epithelial-mesenchymal transition (EMT) is important for early tumor invasion and metastases (1). During this process, high expression of N-cadherin (cell-cell adhesion molecule) and β-catenin (important Wnt/β-catenin cascade protein) are hallmarks of EMT (2). N-cadherin and β-catenin are expressed normally in mesenchymal cells, but are upregulated in a various forms of human cancer, including gastric cancer, breast tumors and nasopharyngeal carcinoma (NPC), providing these cancer cells with more invasive and motile phenotypes (3). N-cadherin and β-catenin inactivation inhibits cell migration and metastatic formation (4). Furthermore, these two proteins are often associated with unfavorable prognoses in a diverse range of human malignancies (5,6). Notably, accumulating evidence suggests that N-cadherin and β-catenin are molecular targets in current cancer gene therapy and have been evaluated in preclinical and clinical studies (7,8). Thus, N-cadherin and β-catenin may be important for tumor development and progression.

NPC is a distinctive malignant disease that is prevalent in southern China, and has an annual incidence rate of ~20 per 100,000 people (9). Unlike other types of head and neck cancer, NPC exhibits features of regional lymph node metastases, and patients with NPC often present at an advanced clinical stage for diagnosis and/or treatment (10). Thus, it is imperative to understand the molecular changes in NPC that drive tumor progression and metastasis (11,12). Positive N-cadherin and β-catenin have been reported in multiple forms of human carcinoma to date with the exception of NPC. Therefore, the present study aimed to investigate the expression of N-cadherin and β-catenin in patients with NPC and further study their prognostic value.

Materials and methods

Tumor sample collection. A total of 128 NPC paraffin-embedded specimens were collected between January 2007 and October 2009 from Xiangya Hospital and Hunan Provincial Tumor Hospital, Central South University (CSU; Changsha, China). Individuals were excluded if they had a previous history of cancer and underwent radiotherapy or chemotherapy at diagnosis. All patients were classified and staged according to...
the sixth edition of the Union for International Cancer Control/ American Joint Committee on Cancer Tumor-Node-Metastasis staging system (13). Follow-up information was collected every 6 months after diagnosis or until mortality. The complete cohort characteristics of the 91 patients with NPC are listed in Table I.

Additionally, 26 fresh biopsy samples from patients with NPC and 8 nasopharyngeal epithelial (NPE) tissue samples were collected from Xiangya Hospital, CSU between August 2013 and October 2013. These samples were validated and diagnosed by qualified pathologists, and were used to determine the mRNA expression of N-cadherin and β-catenin. All tissue samples were infiltrated in RNAlater® Stabilization Solution (Qiagen, Inc., Valencia, CA, USA) and snap-frozen in liquid nitrogen. Prior informed consent was obtained, and the study protocol was approved by the Ethics Committee of Xiangya Hospital, CSU.

RNA isolation. Total RNA was isolated using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to the manufacturer's protocol. RNA was extracted using the RNeasy® Mini kit (Qiagen, Inc.) and treated with DNase I according to the manufacturer's protocol. The integrity and quality of RNA was confirmed using agarose gel electrophoresis and absorbance at 260 nm. Then, 2 μg total RNA was reverse transcribed to cDNA using a PrimeScript RT reagent kit with a DNA Eraser (Takara Biotechnology Co., Ltd., Tokyo, Japan) via the SuperScript® First-Strand Synthesis system (Thermo Fisher Scientific, Inc.) with random hexamer primers (Promega Corporation, Madison, WI, USA).

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). RT-qPCR was performed using the aforementioned cDNA. Primers for mRNA were designed and synthesized for N-cadherin (forward, 5'-GCTGCTGTAGGCTTCTGG-3' and reverse, 5'-GCACCTTGCACTTTCTGTG-3') and β-catenin (forward, 5'-GGAGAGCTCCGAGGTAT-3' and reverse, 5'-GGCTGTACGTGTATCCCA-3'). qPCR was performed using the Bio-Rad iQ™5 Multicolor Real-Time PCR Detection system (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The final PCR reaction mixture (10 µl) included 1 µl cDNA product, 4 µl DEPC water and 5 µl SYBR® Premix Ex Taq™ II (Takara Biotechnology Co., Ltd.). Reactions were incubated at 95°C for 30 sec, followed by 40 cycles at 95°C for 5 sec and 60°C for 30 sec. All reactions were run in triplicate with GAPDH as an internal reference (forward primer, 5'-GACTAAGGCTGAGGT-3' and reverse primer, 5'-AGTAGGGCTGCTGGA-3'). Gene expression levels were quantified using the 2-ΔΔCq method (14). The data were representative of the means of three experiments.

Immunohistochemistry. Tissue samples were fixed in 4% formaldehyde for 48 h at room temperature and then dehydrated by ethanol from 75-100% for 1 h at room temperature. The tissues were subsequently infiltrated by xylene twice for 30 min at room temperature, embedded in paraffin and cut into 4 µm sections. Following this, sections were deparaffinized by washing with xylene twice and rehydrated with ethanol from 75-100%, following antigen retrieval by heating in a microwave for 4 min at 100% power followed by 16 min at 20% power. After blocking peroxidase activity with 3% H2O2 for 5 min at room temperature, sections were blocked by 5% goat serum (Santa Cruz Biotechnology, Inc., Dallas, TX, USA) in PBS for 30 min at room temperature. Negative control slides were probed with 5% normal goat serum in PBS under the same experimental conditions. Primary antibodies were added in blocking buffer (horse serum (Santa Cruz Biotechnology, Inc.) 0.2% Triton X-100 and 0.1% bovine serum albumin in PBS) and incubated with sections at 4°C overnight. The primary antibodies were as follows: Rabbit polyclonal anti-N-cadherin (dilution, 1:300; #18203; Abcam, Cambridge, MA, USA) and rabbit monoclonal anti-β-catenin (dilution, 1:500; #32572; Abcam). Following sufficient rinses by PBS three times (5 min each), sections were immunostained with Polink-1 HRP DAB Detection system (PV-6000; Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd., Beijing, China). A light microscope and a digital color camera were used for examination and image capture of the slides.

Immunohistochemistry results were diagnosed and scored independently by two qualified pathologists without knowledge of the information of all patients with NPC. Assessments of N-cadherin and β-catenin proteins referred to methods described by Hui et al (15). The intensity of N-cadherin and β-catenin staining was scored as 0, 1, 2 and 3. Scores of positive cell percentage were assigned as 0 (<5%), 1 (5-25%), 2 (26-50%), and 3 (≥51%). The scores of each view were multiplied to give a final score of 0-9, and the final score of one sample was the mean of 10 microscopic fields. For final statistical analysis, tumors were divided into two groups: High expression of N-cadherin and β-catenin protein (scored 6-9) and low expression of N-cadherin and β-catenin protein (scored 0-5).

Statistical analysis. Statistical procedures were analyzed using SPSS version 15.0 (SPSS, Inc., Chicago, IL, USA). The χ2 test was used to analyze the potential associations between the expression of N-cadherin and β-catenin, and diverse NPC clinicopathological variables. The Kaplan-Meier survival analysis was applied to determine the prognostic value of N-cadherin and β-catenin proteins. In addition, the Cox proportional hazards model was used to perform multivariate analysis and to determine the final independent prognostic factors. P<0.05 was considered to indicate a statistically significant difference.

Results

Expression of N-cadherin and β-catenin mRNA is increased in NPC samples. In order to clarify the abnormal expression of N-cadherin and β-catenin at the transcriptional level, mRNA expression of N-cadherin and β-catenin was initially measured in 26 NPC specimens and 8 normal NPE samples was initially measured via RT-qPCR. Fig. 1A and B depict the individual expression of N-cadherin and β-catenin in NPC samples. Expression of N-cadherin and β-catenin was initially measured via RT-qPCR. Fig. 1A and B depict the individual expression of N-cadherin and β-catenin in NPC samples.
The present study measured the expression of N-cadherin and β-catenin in NPC. Following staining of N-cadherin and β-catenin proteins, these cases were categorized into two groups containing high and low N-cadherin or β-catenin expression. Expression of N-cadherin and β-catenin protein was positively correlated with lymph node metastasis status and advanced clinical stages in patients with NPC (P=0.001) (Table I). However, no significant differences were identified between expression levels of these proteins and parameters, including age (P=0.339) and gender (P=0.651).

Survival analysis. Of the 91 patients with NPC, 85 (93.4%) had intact follow-up information that was able to be further used for survival analysis. According to the expression levels of N-cadherin and β-catenin proteins, these cases were categorized into two groups containing high and low N-cadherin or β-catenin expression. Kaplan-Meier survival analyses demonstrated that patients with NPC with high N-cadherin protein expression had a significantly lower overall survival (OS) rate than those with low N-cadherin expression (P=0.001; Fig. 3A). Similarly, high β-catenin protein expression was significantly associated with a lower OS rate compared with low expression of β-catenin protein (P=0.01; Fig. 3B). Finally, Cox multivariate analysis determined that N-cadherin expression (P=0.017) and clinical stage (P=0.024) were independent factors with prognostic value in patients with NPC (Table II).

Discussion

The present study measured the expression of N-cadherin and β-catenin in NPC, which indicated that the mRNA and protein expression of the two were significantly elevated in the NPC tissues. Moreover, the data demonstrated that increased protein expression of N-cadherin and β-catenin was positively correlated with NPC lymph node metastasis and predicted a poorer prognosis in patients with NPC.

The cadherin family is comprised of various proteins and includes E-cadherin (epithelial), N-cadherin (neural), P-cadherin (placental) and nonclassical cadherins, such as OB-Cadherin, VE-Cadherin, K-cadherin, LI-cadherin, BR-cadherin, M-cadherin, R-cadherin and T-cadherin (16). E-cadherin has been studied as an epithelial cell molecule in a various forms of human cancer, including NPC (17). A number of studies have demonstrated that downregulated E-cadherin is associated with cancer metastasis and poor prognosis (18-20). Studies performed in vitro and in vivo have established the hypothesis that E-cadherin is a vital molecule in the EMT process (21).

By contrast, the well-known mesenchymal molecule N-cadherin has scarcely been investigated in NPC. In the present study, mRNA and protein expression of N-cadherin were markedly increased in the NPC tissues when compared with the noncancerous NPE tissues. Moreover, N-cadherin upregulation was highly correlated with lymph node metastasis and poor survival in patients with NPC (22). The aforementioned results are consistent with previous reports examining other forms of malignancy, including breast cancer, lung cancer and oral squamous cell carcinoma (23-25). However, it is important to note that decreased expression of N-cadherin has also been reported in several human malignancies, such as osteosarcoma (26), ovarian carcinoma (27), glioblastoma (28) and renal carcinoma (29). The opposing expression profile in these forms of cancer suggests that N-cadherin has a distinct expression pattern based on the diverse background of cancer, and its exact role in different types of cancer requires further study.

β-catenin is a critical component in the cadherin-catenin complex, which is essential in connecting actin filaments of cells to the cell-cell interface at adherens junctions (30). The aberrant expression and dysfunction of the N-cadherin/β-catenin complex leads to increased malignant capacity in cancer cells,
including increased cell motility and invasion (31). Multiple signaling pathways, such as phosphoinositide 3-kinase/protein kinase B and nuclear factor-κB, have been implicated in the mechanisms behind these unfavorable alterations in the complex (32,33). However, these mechanisms require further study in different forms of human cancer.

Finally, the prognostic significance of N-cadherin and β-catenin proteins in patients with NPC was investigated. To the best of our knowledge, the present study is the first to demonstrate that high expression of N-cadherin or β-catenin is strongly correlated with adverse prognosis in patients with NPC. These results indicate that N-cadherin and β-catenin may be used as a valuable biomarker during surveillance of patients with NPC.

In conclusion, the present study demonstrated that N-cadherin and β-catenin mRNA and protein expression are positively correlated with lymph node metastasis and prognosis in patients with NPC. As the current study was a retrospective clinical association analysis based on archival tissue specimens, further in vitro and in vivo studies are required to enable a comprehensive understanding of the potential function and related mechanism of the N-cadherin/β-catenin complex in NPC.

Acknowledgements

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Table II. Cox regression analysis of the progression-free survival and overall survival rates.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Progression-free survival</th>
<th>Overall survival</th>
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<tbody>
<tr>
<td></td>
<td>HR 95% CI P-value</td>
<td>HR 95% CI P-value</td>
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<tr>
<td>Gender (female/male)</td>
<td>1.665 0.710-3.416 0.251</td>
<td>1.436 0.815-3.317 0.342</td>
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<tr>
<td>Age (≤48/&gt;48)</td>
<td>1.16 0.816-1.221 0.914</td>
<td>0.875 0.657-1.149 0.764</td>
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<tr>
<td>β-catenin (high/low)</td>
<td>1.417 0.857-2.124 0.612</td>
<td>1.214 0.716-2.151 0.273</td>
</tr>
<tr>
<td>N-cadherin (high/low)</td>
<td>2.453 1.325-3.455 0.013</td>
<td>2.887 1.561-5.176 0.017*</td>
</tr>
<tr>
<td>Clinical stage (I and II/III and IV)</td>
<td>2.612 1.724-3.294 0.021</td>
<td>2.429 1.816-3.169 0.024*</td>
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*P<0.05. HR, hazard ratio; CI, confidence interval.

Figure 2. Representative images of N-cadherin and β-catenin as detected in 91 paraffin-embedded NPC tissues by immunohistochemistry. Brown denotes a positive signal. Expression of N-cadherin in (A) columnar epithelia and (B) NPC. Magnification, x200. Expression of β-catenin in (C) columnar epithelia and (D) NPC. Magnification, x400. Nuclei were counterstained by hematoxylin. NPC, nasopharyngeal carcinoma.

Figure 3. Expression of N-cadherin and β-catenin were significantly correlated with survival of patients with NPC. Kaplan-Meier estimates of overall survival rate for 85 NPC patients are presented according to (A) N-cadherin or (B) β-catenin expression. P-values were obtained using the log-rank test. NPC, nasopharyngeal carcinoma.
References


