R-spondin 2 promotes proliferation and migration via the Wnt/β-catenin pathway in human hepatocellular carcinoma

XINGUANG YIN1,2, HUIXING YI3, LINLIN WANG4, WANXIN WU5, XIAOJUN WU5 and LINGHUA YU1

1Centre for Gastroenterology and Hepatology, The First Affiliated Hospital of Jiaxing College; 2Centre for Gastroenterology and Hepatology, The Maternity and Child Health Care Hospital Affiliated to Jiaxing College, Jiaxing, Zhejiang 314001; 3Intensive Care Unit, The Second Affiliated Hospital of Zhejiang University, Hangzhou, Zhejiang 310009; 4Department of Basic Medicine Sciences, School of Medicine, Zhejiang University, Hangzhou, Zhejiang 310058; 5Department of Pathology, The First Affiliated Hospital of Jiaxing College, Jiaxing, Zhejiang 314001, P.R. China

Received March 6, 2016; Accepted February 13, 2017

Abstract. Hepatocellular carcinoma (HCC) is a leading cause of malignant disease-associated mortality, particularly in China. The R-spondin 2 (RSPO2) gene is evolutionarily conserved in vertebrates and is involved in developmental and physiological processes. Importantly, RSPO2 has been reported to be associated with colon cancer and potentiate the Wnt/β-catenin signaling pathway. In the present study, enhanced expression of RSPO2 in HCC was observed using tissue microarray. Similarly, the expression level of RSPO2 was higher in HepG2, Huh7 and Hep3B cells but lower in Bel7404 and QGY7703 cells compared with human normal QSG7701 liver cells. Subsequently, gain-of-function studies indicated that RSPO2 promotes the proliferation and migration of QGY7703 cells based on lentivirus-based gene delivery. Furthermore, it was revealed that p21 and leptin, rather than vascular endothelial growth factor-A, are involved in the function of RSPO2 in QGY7703 cells. Particularly, the signal transducer and activator of transcription 3 (STAT3) and Wnt/β-catenin signaling pathways are involved in this process. Overexpression of RSPO2 resulted in the elevated expression of phosphorylated STAT3, β-catenin and c-Myc. Therefore, the present study is beneficial to the understanding of RSPO2-involved liver cancer transformation and drug discovery.

Introduction

Hepatocellular carcinoma (HCC) is one of the most common cancer types and a major cause of cancer-associated mortality worldwide (1). The majority of HCC cases are caused by chronic hepatitis B or hepatitis C infections (2). HCC, characterized by rapid recurrence and poor survival, remains a challenging disease to treat (3). As HCC is not sensitive to radiotherapy or chemotherapy, surgery is the only effective treatment (4). However, the rate of recurrence is high and metastasis is common following surgery, which leads to the poor prognosis for HCC (3). Therefore, it is necessary to understand the molecular mechanisms underlying the growth and metastasis of HCC, which may help to identify effective diagnosis and therapeutic targets to improve the survival. However, the associated molecular mechanisms of HCC progression are not well understood.

The R-spondin (RSPO) protein family consists of four homologous members, which are evolutionarily conserved in vertebrates and are involved in a broad range of developmental and physiological processes (5,6). RSPO1 is important for sex determination (7); RSPO2 is required for limb, laryngeal-tracheal and lung development (8); RSPO3 is critical for placentation formation (9); and the mutation of RSPO4 results in inherited anonychia (10). The association between RSPO and cancer has not been extensively studied. It has been reported that RSPO2 and RSPO3 insertional activation is observed in the mouse mammary tumor virus model system (11,12). Administration of RSPO1 protein to mice induces rapid crypt cell proliferation, which causes a marked increase in the size of the small intestine (13). Seshagiri et al (14) identified that RSPO2 and RSPO3 transcript fusion occurs in 10% of colon tumors. In addition, it was found that RSPO fusions occur exclusively in tumors without adenomatous polyposis coli mutations, indicating that RSPO genes have a role in the activation of Wnt signaling and tumorigenesis (14). Subsequently, RSPO gene fusions were also observed in a subset of colon tumors in the Japanese population, and forced expression
of the RSPO gene was revealed to increase the growth of colorectal cells (15).

A number of previous studies have indicated that RSPOs may potentiate Wnt signaling via stimulation of the leucine-rich repeat-containing G-protein coupled receptors (LGR) 4, LGR5 and LGR6 (16-18). Wnt proteins, comprising a large family of extracellular, lipid-modified glycoproteins, are crucial for embryonic development and cell proliferation, regulation, differentiation, survival and tissue homeostasis in adults (19,20). It was demonstrated that RSPOs cooperate with Wnts during development, particularly by promoting the transcriptional activity of β-catenin, which is an important mechanism of the Wnt signaling pathway (5,16). It has been revealed that canonical Wnt/β-catenin signaling serves an important role in numerous cancer types, including lung, breast, brain, colorectal and liver tumors (21-23). Interaction between RSPOs and LGR4, as well as the intracellular signaling proteins, promotes phosphorylation of LRPs/6, stabilizes β-catenin expression and promotes its transcriptional activity (18). Activation of Wnt/β-catenin results in the increased expression level of its target genes, including cyclin D1 and c-Myc, which are important for driving tumorigenesis in numerous types of cancer (24).

In the present study, high expression of RSPO2 was observed in certain HCC cell lines. Tissue microarray also revealed that the expression of RSPO2 is increased in primary tumors compared with the adjacent normal tissues. Functional study revealed that overexpression of RSPO2 enhances the cell proliferation and anchorage-independent growth of the human liver QGY7703 cancer cell line. In addition, RSPO2 overexpression may promote cell motility, which was demonstrated using Transwell and wound healing assays. Similar to a previous study (25), the present study also revealed that overexpression of RSPO2 is involved in Wnt/β-catenin activation via increasing the expression of β-catenin and the downstream gene c-Myc. The present study revealed the functional role of RSPO2 in HCC and indicated that RSPO2 may be a potential drug target for patients with liver tumors.

Materials and methods

Patients and liver tissue samples. A total of 72 human liver tissues were obtained from 24 patients with HCC who had undergone surgical resection between January 2013 and December 2014 at the First Affiliated Hospital of Jiaxing College (Jiaxing, China) with written informed consent and ethical approval from the Local Ethics Committee of Jiaxing College. Patients did not receive radiotherapy or chemotherapy prior to surgery.

Tissue microarray. Tissue samples (24 tumor tissues, 24 paired non-tumor adjacent tissues and 24 normal tissues from 24 HCC patients) were formalin-fixed and paraffin-embedded. The samples were used to construct tissue microarrays (TMAs) using a Beecher Instrument (Sun Prairie, WI, USA) as described previously (26). A total of 3 tissue cylinders of 0.6 mm in diameter were punched from each sample. Subsequent to sectioning (4 mm for each section), tissue slides were baked at 60°C for 2 h, and kept at 4°C for subsequent analysis. TMAs were stained with hematoxylin and eosin (H&E). A trained pathologist reevaluated H&E-stained samples to determine the tumor stage and grade according to the WHO criteria (27). The slides with tissue sections were subjected to immunohistochemical detection of RSPO2, using the primary antibody against RSPO2 (dilution, 1:50; cat. no. ab73761; Abcam, Cambridge, MA, USA). Following rinsing with 1X PBS, slides were incubated with biotinylated anti-rabbit IgG for 30 min (dilution, 1:1,000) using Vectastain Elite ABC kit (cat. no. PK-6100; Vector Laboratories, Inc., Burlingame, CA, USA). The detection was achieved using the avidin-biotin peroxidase method with a diaminobenzidine chromogen kit (cat. no. SK-4100; Vector Laboratories, Inc.). The TMAs were evaluated for RSPO2 expression by a trained pathologist and were scored as strong (+++) with >80% positive cells, moderate (++ with 30-80% positive cells, weak (+) with observed <30% stained cells or absent (-). The subcellular localization of the staining was noted for each score using CI microscopy by NIS-Elements F under x100 magnification.

Cell culture. The QSG-7701 cell line derived from human normal liver tissue and the human hepatoma cell lines Bel7404, QGY7703, HepG2, Huh7, 293T and Hep3B were purchased from the Cell Bank of the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China). QSG-7701 and QGY7703 cells were cultured in RPMI-1640 (cat. no. SH30809.01; HyClone; GE Healthcare Life Sciences, Logan, UT, USA) medium containing 10% fetal bovine serum (FBS) (cat. no. 10099141; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA). Bel7404, HepG2, Huh7 and Hep3B cells were cultured in Dulbecco's modified Eagle's medium (DMEM; cat. no. SH30243.01B; HyClone; GE Healthcare Life Sciences) with high glucose, supplemented with 10% FBS. All cells were cultured in an atmosphere of 95% air and 5% CO2 under humidified conditions.

Western blot analysis. All cultured cells were lysed using radioimmunoprecipitation assay lysis and extraction buffer (cat. no. 89900; Thermo Fisher Scientific, Inc.). Total proteins were subjected to concentration determination using a commercial bicinchoninic acid quantification kit, according to the manufacturer's protocol (cat. no. 23227; Thermo Fisher Scientific, Inc.). Lysates from QGY7703 cells or HCC tissue (15 μg) were subjected to 12% SDS-PAGE for protein separation and then electrophoretically transferred to nitrocellulose membranes (Axogen Scientific, Union City, CA, USA). Subsequent to being blocked by PBS containing 5% fat-free milk, the nitrocellulose membranes were incubated with rabbit polyclonal antibody for RSPO2 (cat. no. ab73761; dilution, 1:1,000; Abcam), β-catenin (cat. no. 8480; dilution, 1:1,000; Cell Signaling Technology, Inc., Danvers, MA, USA), c-Myc (cat. no. 5605; dilution, 1:1,000; Cell Signaling Technology, Inc.), eif4e (cat. no. ab51745; dilution, 1:1,000; Abcam), phosphorylated signal transducer and activator of transcription 3 (p-STAT3) (cat. no. 9130; dilution, 1:1,000; Cell Signaling Technology, Inc.), leptin (cat. no. ab3583; dilution, 1:1,000; Abcam), p21 (cat. no. SC-397; dilution, 1:500; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) and rabbit polyclonal antibody for β-actin (cat. no. 10303001;
dilution, 1:5,000; Harmonious One Biotechnology, Shanghai, China) overnight at 4°C and then incubated with horseradish peroxidase-conjugated rabbit IgG (cat. no. 7074; dilution, 1:3,000; Cell Signaling Technology, Inc.) for 1.5 h at room temperature. The immunolabeled proteins were detected using a commercial enhanced chemiluminescent detection kit (cat. no. 108070002; Harmonious One Biotechnology, Shanghai, China). Results were quantified by a luminescent digital image analyzer Bio-Spectrum600 (UVP, Upland, CA, USA). Band intensity was assessed using a Gel-Pro analyzer (V6.3; Media Cybernetics, Inc., Rockville, MD, USA).

**Lentivirus vector construction.** The total RNA from 5x10^6 HepG2 cells was extracted using the phenol-chloroform method following TRIzol (cat. no. 15596-026; Invitrogen; Thermo Fisher Scientific, Inc.) lysis, according to the manufacturer’s protocol. The cDNA was prepared by reverse transcription using a random primer (D3801; Takara Biotechnology Co., Ltd., Dalian, China) at 37°C for 1 h, following the denaturation of total RNA by heating for 5 min at 37°C, followed by immediate chilling on ice. The full coding region of the RSPO2 gene was isolated following denaturation at 95°C for 5 min, 36 cycles of 95°C for 30 sec, 56°C for 30 sec and 72°C for 45 sec, and then subjected to post-elongation for 10 min at 72°C. The mixture for polymerase chain reaction (PCR) was composed of 5 µl of 10X PCR buffer, 0.2 mM dNTP, 0.2 µM RSPO2-EcoRI-F primer (5'-CCGCGATCCTATTGCTGCTGCTGAC-3'), 2 U PFU polymerase (cat. no. 101060002; Harmonious One Biotechnology Pvt. Ltd., Beijing, China) with the selective marker gene puromycin by a classic ligation and transfection of 293T cells. Briefly, 1.5x10^5 of pseudotyped recombinant lentivirus was performed

**Lentivirus production and cells transduction.** Lentivirus production and cells transduction. E. coli (Thermo Fisher Scientific, Inc.) and DH5alpha (Invogen Biotechnology Pvt. Ltd., Beijing, China) with the lentiviral vector pLV-mCherry (2A) puro-RSPO2 or pLV-mCherry (2A) puro were trypsinized and suspended in 200 µl serum-free medium and were seeded into the Transwell chamber by adding DMEM without FBS into the upper and bottom chamber overnight at 37°C. QGY7703 cells transfected with pLV-mCherry (2A) puro-RSPO2 or pLV-mCherry (2A) puro were trypsinized and suspended in 200 µl serum-free medium and were seeded in the upper chamber at a density of 1.5x10^5 cells per well. Following growth for 10 days, the cells were subjected to fixation by methanol for 10 min and then stained with 0.1% crystal violet at room temperature for 30 min. Following removal of the dye, colonies containing >50 cells were counted using a TiS microscopy by NIS-Elements Viewer (version 4.2; Nikon, Tokyo, Japan) under x100 magnification.

**Migration assay.** A 24-well Transwell chamber with 8.0 µm pore size (Costar; Corning Incorporated, Corning, NY, USA) was used for the migration assay. The pre-balance of the Transwell chamber was performed by adding DMEM without FBS into the upper and bottom chamber overnight at 37°C. QGY7703 cells transfected with pLV-mCherry (2A) puro-RSPO2 or pLV-mCherry (2A) puro were trypsinized and suspended in 200 µl serum-free medium and were seeded in the upper chamber at a density of 1.5x10^5 cells per well. The bottom chamber was filled with DMEM containing 10% FBS. Following incubation for 24 and 48 h at 37°C, the non-migrating cells were removed by soft scratch with small cotton swabs and rinsed with 1X PBS. Migrated cells were then dried, fixed with methanol and stained with 0.1% crystal violet at room temperature for 30 min. The transmembrane cells were counted using a TiS microscopy by NIS-Elements Viewer (version 4.2, Nikon) under x100 magnification.

**Viability analysis.** The human hepatoma QGY7703 cell line with stable overexpression of RSPO2 and the control QGY7703 cells transfected with empty vectors were seeded onto 96-well plates at the density of 2.0x10^4 per well. Cells were analyzed using an MTT assay at day 1, 2, 3, 4 and 5 subsequent to cell seeding. Briefly, 100 µg of MTT (cat. no. 0793-1G; Amresco, LLC, Solon, OH, USA) was added to each well. Following incubation for 4 h at 37°C, the purple formazan crystals generated from viable cells were dissolved by adding 100 µl of dimethyl sulfoxide to each well. The absorbance of each well was then read at 570 nm.

**Anchorage-independent growth analysis.** QGY7703 cells transfected with pLV-mCherry (2A) puro-RSPO2 or pLV-mCherry (2A) puro were trypsinized and suspen

**Wound healing assay.** RSPO2-overexpressed QGY7703 cells or control cells were plated onto a 24-well plate with 4x10^4 per well and incubated for 24 h at 37°C. Linear scratch wounds were then produced using a 10 µl pipette tip on the confluent cell monolayer. The medium was replaced with the serum-free medium. Images were captured at 0, 12, 24, 36 and 72 h and the wounding size was quantified and analyzed by NIS-Elements Viewer (version 4.2, Nikon) under x100 magnification.

**Statistical analysis.** Data are presented as the mean ± standard deviation. A two-tailed Student’s t-test was employed to evaluate the differences between groups. P<0.05 was considered to indicate a statistically significant. The differences of indexes between tumor tissues and paired non-tumor adjacent tissues
were analyzed using the Wilcoxon signed-rank test. Differences between groups were analyzed by the Mann-Whitney U test. Data were processed with R Studio (v1.0; Boston, MA, USA).

Results

Expression of RSPO2 in various HCC cells. The expression level of RSPO2 was detected in various HCC cells by western blot analysis. As presented in Fig. 1, increased expression levels of RSPO2 were observed in HepG2, Huh7 and Hep3B cells compared with the human normal liver QSG-7701 cell line. However, markedly decreased expression of RSPO2 was found in the human hepatoma Bel7404 and QGY7703 cell lines compared with QSG-7701 cells.

Overexpression of RSPO2 in liver cancer tissues. The expression of RSPO2 in clinical samples of HCC was analyzed. A total of 24 pairs of human hepatic carcinoma and matched adjacent non-tumor tissues or normal tissues were examined using immunohistochemical staining with an antibody against human RSPO2. Samples were considered RSPO2-positive if either the cell nucleus or cytoplasm stained positive. As presented in Fig. 2A and B, the staining of RSPO2 was primarily observed in the cytoplasm of cancer cells. Fig. 2A presents representative examples from the tissue microarray for each RSPO2 staining score, ranging from 0 to +++. Intense expression of RSPO2 in tumor tissue (score ++ or ++++) was identified in 16/24 patients (66.7%), whereas in other patients (33.3%) a weak immunoreactivity (score +) was detected. However, the expression of RSPO2 in non-tumor adjacent tissues was significantly lower compared with tumor tissues (Wilcoxon signed-rank test, P=0.007). Moderate expression of RSPO2 (score ++) in non-tumor adjacent tissue was found in 6 of 24 patients (25%), whereas the remaining 75% exhibited weak expression of RSPO2 (score +). Similar expression patterns were found in the paired normal tissues. The expression of RSPO2 in the paired normal tissues was significantly lower compared with tumor tissues (Wilcoxon signed-rank test, P=0.001). Representative images of RSPO2 expression in the paired normal, adjacent and tumor tissues are shown in Fig. 2A.

Statistical analyses were performed to examine the association between RSPO2 expression and the clinicopathological characteristics of hepatic carcinoma. As shown in Table I, no association was observed between the expression of RSPO2 and patient age or gender, tumor grading and tumor staging in patients with HCC. A small sample size (n=24) may be the reason that no statistically significant results were identified.

RSPO2 overexpression enhances the proliferation and anchorage‑independent growth of QGY7703 cells. To define the function of RSPO2 in HCC, RSPO2 overexpression in the QGY7703 cell line was achieved by lentivirus delivery (Fig. 3A). The cell growth of QGY7703 cells with stable overexpression of RSPO2 was firstly evaluated by MTT assay. RSPO2 overexpression significantly enhanced soft agar growth of QGY7703 cells, as evidenced by the decrease in colony number and size compared with the control QGY7703 cells that were infected with the pLV-mCherry (2A) puro empty vector (P<0.01; Fig. 3B).

Table I. Characteristics and RSPO2 expression of patients with hepatocellular carcinoma.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Patients, n (%)</th>
<th>Absent or weak, n</th>
<th>Moderate to high, n</th>
<th>P-value</th>
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</thead>
<tbody>
<tr>
<td>Age ≤50 years</td>
<td>13 (54)</td>
<td>6</td>
<td>7</td>
<td>0.155</td>
</tr>
<tr>
<td>&gt;50 years</td>
<td>11 (46)</td>
<td>2</td>
<td>9</td>
<td></td>
</tr>
<tr>
<td>Gender Male</td>
<td>18 (75)</td>
<td>7</td>
<td>11</td>
<td>0.766</td>
</tr>
<tr>
<td>Female</td>
<td>6 (25)</td>
<td>1</td>
<td>5</td>
<td></td>
</tr>
<tr>
<td>Tumor grading I</td>
<td>3 (3)</td>
<td>0</td>
<td>3</td>
<td>0.286</td>
</tr>
<tr>
<td>II</td>
<td>19 (79)</td>
<td>7</td>
<td>12</td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>2 (8)</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>Tumor staging II</td>
<td>4 (17)</td>
<td>1</td>
<td>3</td>
<td>0.996</td>
</tr>
<tr>
<td>III</td>
<td>20 (83)</td>
<td>7</td>
<td>13</td>
<td></td>
</tr>
</tbody>
</table>

P-value calculated using Mann-Whitney U test. RSPO2, R-spondin 2.
RSPO2 overexpression promotes the migration of QGY7703 cells. To study the effect of RSPO2 on the motility of QGY7703 cells, Transwell and wound healing assays were performed. In the Transwell assay, the percentage of cells that migrated through the membrane was significantly increased in cells with RSPO2 overexpression compared with the control cells transfected with empty vector (Fig. 4A). The wound healing assay results also demonstrated that RSPO2 overexpression significantly enhanced the migration of QGY7703 cells compared with the control cells (Fig. 4B).

RSPO2 overexpression potentiates the activation of Wnt/β-catenin. Previous studies have reported that the RSPO2 gene is involved in the activation of the Wnt/β-catenin pathway. To improve the understanding of the biological role of the RSPO2 gene in HCC and the underlying mechanism of the aforementioned findings, the expression level of nuclear β-catenin was analyzed in QGY7703 cells. As shown in Fig. 5, the expression level of nuclear β-catenin was significantly increased in QGY7703 cells with stable overexpression of RSPO2 gene compared with the negative control group. The expression of c-Myc, one of the target genes of Wnt/β-catenin signaling, was also analyzed. Consistently, c-Myc gene expression was significantly increased in QGY7703 cells with RSPO2 stable overexpression (Fig. 5). The present data indicated that overexpression of RSPO2 is involved in Wnt/β-catenin activation via increasing the expression of β-catenin and its downstream genes.

RSPO2 regulates proliferation-associated genes and signaling pathways. To further elucidate the molecular mechanism underlying RSPO2-induced cell proliferation, the...
proliferation-associated genes p21, leptin and VEGF-A, and the STAT3 signaling pathway were investigated. The p21 and leptin genes exhibited significantly reduced and increased expression, respectively (Fig. 5). However, the expression level of VEGF-A did not show a notable difference between control and RSPO2-overexpressed QGY7703 cells. The elevated expression of phosphorylated STAT3 indicated that the STAT3 signaling pathway may be involved in RSPO2-induced cell proliferation.

Discussion

The carcinogenesis of HCC is a multi-factorial, multi-step and complex process. Previous studies have documented that the bidirectional interactions between tumors and hepatic stellate cells (HSCs) compose an amplification loop to enhance metastatic growth in the liver (28,29). Previous studies revealed that the RSPO family may promote HSC activation by enhancing the canonical Wnt pathway (30,31). In the present study, the data provided evidence that RSPO2 expression may contribute to malignant biological behavior in HCC. Additional large-scale investigations are required to pinpoint the link between the expression level of RSPO2 and the clinical characteristics of human HCC. The present study demonstrated that the expression of RSPO2 is upregulated in various HCC cell lines. Paired HCC lesions and adjacent non-cancer tissues were found to express RSPO2 differently. The tumor tissues exhibited significantly increased expression of RSPO2 compared with adjacent non-tumor tissues. Furthermore, the present data demonstrated that RSPO2 overexpression enhances the cell proliferation and anchorage-independent growth of human liver QGY7703 cell lines. In addition, RSPO2 overexpression may also promote the motility of QGY7703 cells. Study of the underlying molecular mechanism indicated that overexpression of RSPO2 may be associated with Wnt/β-catenin activation via increasing the expression of β-catenin and its
downstream gene c-Myc. Taken together, the present results revealed the potential functional role of RSPO2 in HCC and indicated that RSPO2 may be a potential drug target for patients with liver tumors.

Previous studies have revealed that RSPOs are involved in the activation of Wnt signaling, which is important for tumorigenesis (14,32). However, the association between RSPO and cancer has not been extensively studied. Studies on the role of RSPO2 in cancer primarily focus on colon tumors (14,15). A recent study reported that high-copy amplifications of RSPO2 gene were observed in 231 HCC cases via whole exome sequencing (25). In the present
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study, significantly increased expression levels of RSPO2 were detected in HCC tissues compared with the adjacent non-tumor tissues or paired normal tissue. Similarly, an increased expression level of RSPO2 was observed in HepG2, Huh7 and Hep3B cells compared with human normal liver QSG-7701 cell lines. High-copy amplifications of RSPO2 may aid the explanation of the phenomenon of increased expression of RSPO2 in tumor tissues.

In the present study, altered p21 and leptin expression levels were observed, which may contribute to RSPO2-induced proliferation on QGY7703 cells. Further work requires an elucidation of the spatial-temporal association between RSPO2 and p21 or leptin. Furthermore, systematic investigation of pivotal molecules located in the STAT3 signaling pathway may aid understanding of the role of RSPO2 in liver cancer transformation. Other signaling pathways may be examined to systematically elucidate this molecular mechanism, potentially contributing to identification of novel HCC therapeutics.

In conclusion, the present study revealed that R-spondin 2 promotes proliferation and migration in various HCC cell lines via the Wnt/β-catenin pathway. However, additional studies are required to confirm the tumor-promoting effects of R-spondin2 in mouse models.

Acknowledgements

The present study was funded by grants from the Jiaxing Municipal Science and Technology Project (grant no. 2014AY21030-1), the Zhejiang Science and Technology Public Welfare Project (grant no. 2015C33279), the Zhejiang Provincial Natural Science Fund (grant nos. LY16H030016 and LY17H030012), the Anesthesiology Center in North of Zhejiang Province and the Jiaxing Key Laboratory of Neurology and Pain Medicine (2015-02-2).

References


