Identification of candidate biomarkers for epithelial ovarian cancer metastasis using microarray data

SU LI¹, HUA LI², YING XU³ and XIAOMEI LV¹

¹Department of Obstetrics and Gynecology, Jinan Central Hospital Affiliated to Shandong University; ²Department of Obstetrics and Gynecology, The People's Hospital of Zhangqiu, Jinan, Shandong 250014; ³Department of Obstetrics and Gynecology, Shandong Coal Taishan Sanatorium, Taian, Shandong 271000, P.R. China

Received May 26, 2016; Accepted May 8, 2017

DOI: 10.3892/ol.2017.6707

Abstract. Epithelial ovarian cancer (EOC) is a common cancer in women worldwide. The present study assessed effective biomarkers for the prognosis of EOC metastasis. The GSE30587 dataset, containing 9 EOC primary tumor samples and 9 matched omental metastasis samples, was analyzed. Following normalization, the differentially expressed genes (DEGs) between these samples were identified using the limma package for R. Subsequently, pathway enrichment analysis was performed using ClueGO, and a protein-protein interaction (PPI) network was constructed using the Search Tool for the Retrieval of Interacting Genes database. The microRNA (mRNA/miR)-target network was established using the multiMiR package. A set of 272 DEGs was identified in metastatic EOC samples, including 189 upregulated and 83 downregulated genes. Collagen type I α 1 chain (COL1A1), COL1A2, collagen type XI α 1 chain (COL11A1) and thrombospondin 3-kinase/protein kinase B (PI3K/Akt), focal adhesion and extracellular matrix (ECM)-receptor interaction signaling pathways. THBS1 and tissue inhibitor of metalloproteinase (TIMP)3 were two dominant nodes in the PPI network and were key in the miRNA-target network, being targeted by hsa-miR-1. Multiple DEGs and miRNAs were identified as potential biomarkers for the prognosis of EOC metastasis in the present study, which likely affected metastasis by regulating the PI3K/Akt, ECM-receptor interaction and cell adhesion signaling pathways. In addition, THBS1 and TIMP3 were identified as potential targets of hsa-miR-1.

Introduction

Epithelial ovarian cancer (EOC) is the sixth largest cause of cancer-associated mortality in women globally (1). In 2012, ~22,280 and 69,565 cases of EOC were estimated for the USA and Europe, respectively (2). EOC accounts for 90% of cases of ovarian cancer and is characterized by metastasis (3). Typically, primary EOC tumors disseminate within the peritoneal cavity, primarily into the omentum (4). Only once the tumor cells have spread into the peritoneal cavity may EOC be diagnosed, which often results in a poor prognosis (5).

Numerous studies have assessed the mechanisms involved in EOC metastasis. Scotton et al (6) demonstrated that C-X-C motif chemokine receptor 4 was the only chemokine receptor expressed in ovarian cancer cells. This restricted expression is proposed to be a major step in ovarian cancer metastasis. Disrupting cell adhesion promotes tumor progression. The downregulation of the adhesion molecules cluster of differentiation (CD)82 and CD9 has been reported to be associated with the progression of ovarian cancer, particularly metastasis (7). Another study reported that the tumorigenicity-associated protein mucin 1 serves a function in EOC metastasis (8).

MicroRNAs (miRNAs/miRs) are small non-coding RNAs that serve key functions in the development of numerous types of cancer, including EOC, by regulating gene expression (9). A previous study examined the alteration of miRNAs during the development of EOC and, as expected, identified numerous differentially expressed miRNAs, including the overexpression of miR-200a, 200b, 200c and 141 (1). However, there are few reports of miRNAs associated with EOC metastasis.

A recent study identified differentially expressed genes (DEGs) between EOC primary tumors and metastases by microarray profiling (4). However, this previous study primarily concerned copy number variations (CNVs), which refers to variations caused by gene rearrangement, and the upregulation of the transforming growth factor β signaling pathway. The results of this previous study suggested that although the clone (the altered genes corresponding to the CNVs) in metastasis and primary tumors was different, the tumor cells were adapting to the omental environment. Despite these results, the function of numerous other DEGs and their interactions in EOC remain unclear. Therefore, the present study re-analyzed the GSE30587 microarray dataset (4) to...
identify DEGs between primary tumor and omental metastatic tumor EOC cells. Furthermore, the present study performed term and pathway enrichment analyses, and protein-protein interaction (PPI) network construction. The present study also combined the DEG data with information on miRNAs in multiple databases to predict miRNA-target interactions. Through these comprehensive bioinformatical methods, the present study assessed effective biomarkers for the prognosis of EOC metastasis.

Materials and methods

Data resources. The GSE30587 microarray dataset (4) was downloaded from the Gene Expression Omnibus (GEO) database (www.ncbi.nlm.nih.gov/geo). Of the dataset, 9 primary tissue samples (control samples) and 9 matched omental metastatic tumor samples (metastatic samples) from patients with serous EOC were used in the present study. The platform used for the detection of this microarray data in the study by Brodsky et al (4) was the GeneChip™ Human Gene 1.0 ST Array (Affymetrix; Thermo Fisher Scientific, Inc., Waltham, MA, USA).

Pretreatment and differential analysis. Expression profiles from probe level and annotation profiles from the dataset were downloaded from the GEO database. Raw data in the expression profiles were preprocessed via robust multi-array average (RMA) normalization (10), allowing the expression values from probe level to correspond with those of the gene level, in accordance with the annotation profile. The average probe expression value was considered to be the gene expression value. The DEGs between control and metastatic samples were identified using the limma package (version 3.22.7) of R software (11). The cut-off values for DEG selection were a fold-change in expression of ≥1.5 and P<0.05.

Term and pathway enrichment analyses. The Cytoscape plugin ClueGO (11), which facilitates pathway enrichment analysis and classification of enriched terms, was used to perform the enrichment analysis. Information in the Kyoto Encyclopedia of Genes and Genomes (http://www.genome.jp/kegg/pathway.html) database was combined. Based on the results of ClueGO, a κ coefficient that reflected the association between two pathways or two functional terms was calculated, with a threshold of 0.4. Similar functional terms were given the same color. The Pathview package (version 1.4.2) of R software (12), which reveals the location of DEGs in a pathway, was used to present the enriched pathway. P<0.05 was considered to indicate a statistically significant pathway selection.

PPI network analysis of the DEGs. The Search Tool for the Retrieval of Interacting Genes (STRING) database (13) is a comprehensive database containing coexpression, co-occurrence, text-mining, fusion and protein interaction information. STRING uses a combined score (0-1) to assess reliability; the higher the score, the more reliable the interaction. In the present study, a combined score of 0.4 was used to establish the PPI network, which was visualized using Cytoscape. Each protein in the network served as a node, and the degree of a

<table>
<thead>
<tr>
<th>KEGG pathway no.</th>
<th>KEGG pathway</th>
<th>DEGs involved</th>
<th>No. of DEGs</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>KEGG:00500</td>
<td>Starch and sucrose metabolism</td>
<td>AMY1A, MY1B, AMY1C, AMY2A, MY2B</td>
<td>5</td>
<td>2.04×10⁻⁴³</td>
</tr>
<tr>
<td>KEGG:04145</td>
<td>Phagosome</td>
<td>C1R, THBS2, THBS1, COLEC12, COMP, and others</td>
<td>11</td>
<td>3.85×10⁻¹⁷</td>
</tr>
<tr>
<td>KEGG:04151</td>
<td>PI3K/Akt</td>
<td>COL11A1, COL1A1, COL1A2, THBS1, THBS2, and others</td>
<td>21</td>
<td>6.36×10⁻¹⁰⁻⁵</td>
</tr>
<tr>
<td>KEGG:04510</td>
<td>Focal adhesion</td>
<td>COL11A1, COL1A1, COL1A2, THBS1, THBS2, and others</td>
<td>19</td>
<td>6.2×10⁻¹⁰⁻⁵</td>
</tr>
<tr>
<td>KEGG:04512</td>
<td>ECM-receptor interaction</td>
<td>AMY1A, AMY1B, MY1A, MY2A, MY2B</td>
<td>18</td>
<td>7.51×10⁻¹⁰⁻⁵</td>
</tr>
<tr>
<td>KEGG:04514</td>
<td>Cell adhesion molecules</td>
<td>C1R, THBS2, THBS1, COLEC12, COMP, and others</td>
<td>18</td>
<td>7.51×10⁻¹⁰⁻⁵</td>
</tr>
<tr>
<td>KEGG:04940</td>
<td>Type I diabetes mellitus</td>
<td>HLA-A, HLA-B, HLA-E, HLA-F, HLA-G</td>
<td>5</td>
<td>7.51×10⁻⁴³</td>
</tr>
<tr>
<td>KEGG:04973</td>
<td>Glycogen metabolism</td>
<td>AMY1A, AMY1B, AMY1C, AMY2A, AMY2B</td>
<td>5</td>
<td>7.51×10⁻⁴³</td>
</tr>
<tr>
<td>KEGG:04974</td>
<td>Protein digestion and absorption</td>
<td>COL11A1, COL12A1, COL1A1, COL1A2, COL3A1, and others</td>
<td>10</td>
<td>1.64×10⁻⁴⁰⁻⁶</td>
</tr>
<tr>
<td>KEGG:05146</td>
<td>Amoebiasis</td>
<td>COL11A1, COL1A1, COL1A2, COL3A1, COL4A1, and others</td>
<td>9</td>
<td>5.81×10⁻⁵⁻⁵</td>
</tr>
<tr>
<td>KEGG:05150</td>
<td>Staphylococcus aureus infection</td>
<td>C1QA, C1R, C1S, FCGR2C, PPR3</td>
<td>5</td>
<td>2.21×10⁻³⁰⁻³</td>
</tr>
<tr>
<td>KEGG:05320</td>
<td>Autoimmune thyroid disease</td>
<td>HLA-A, HLA-B, HLA-E, HLA-F, HLA-G, HLA-A, HLA-B, HLA-E, HLA-F, HLA-G</td>
<td>5</td>
<td>1.73×10⁻³⁰⁻³</td>
</tr>
<tr>
<td>KEGG:05330</td>
<td>Allograft rejection</td>
<td>HLA-A, HLA-B, HLA-E, HLA-F, HLA-G, HLA-A, HLA-B, HLA-E, HLA-F, HLA-G</td>
<td>5</td>
<td>3.82×10⁻³⁰⁻³</td>
</tr>
<tr>
<td>KEGG:05332</td>
<td>Graft-versus-host disease</td>
<td>HLA-A, HLA-B, HLA-E, HLA-F, HLA-G, HLA-A, HLA-B, HLA-E, HLA-F, HLA-G</td>
<td>5</td>
<td>6.07×10⁻⁴⁰⁻⁴</td>
</tr>
<tr>
<td>KEGG:05416</td>
<td>Viral myocarditis</td>
<td>HLA-A, HLA-B, HLA-E, HLA-F, HLA-G, HLA-A, HLA-B, HLA-E, HLA-F, HLA-G</td>
<td>5</td>
<td>7.27×10⁻⁴⁰⁻⁴</td>
</tr>
</tbody>
</table>

KEGG, Kyoto Encyclopedia of Genes and Genomes; DEGs, differentially expressed genes; PI3K/Akt, phosphoinositide 3-kinase/protein kinase B; ECM, extracellular matrix.
node was defined as the number of interactions with other nodes. Hub genes were nodes with ≥20 degrees.

Construction of the miRNA-target regulatory network. The multiMiR package (version 3.0.2) (14) of R contains the miRNA-target interaction information from 14 databases, including three validated databases (miRecords version 4, miRTarBase version 4.5 and TarBase version 6), eight predicted databases (DIANA-microT-CDS version 5, E1MMo version 5, MicroCosm version 5, miRanda, miRDB version 4, PicTar version 2, PITA version 6 and TargetScan version 6.4) and three miRNA-disease/drug association databases [miR2Disease (version January, 2010), Pharmaco-miR (version 5.2) and PhenomiR (version 2.0)]. The present study extracted the miRNA-target interaction that appeared in at least two validated databases to establish the miRNA-target regulatory network. The network was subsequently visualized using Cytoscape.

Results

Identification of DEGs in metastatic EOC. The present study identified a total of 272 DEGs between the control and metastatic EOC samples, including 189 upregulated genes and 83 downregulated genes (Fig. 1).

Enriched signaling pathways of DEGs in metastatic EOS. With a predefined threshold of P<0.05, the present study demonstrated that the DEGs identified in metastatic EOS were significantly enriched in signaling pathways associated with cellular signaling transduction and cell adhesion (Fig. 2; Table I), including the phosphoinositide 3-kinase/protein kinase B (PI3K/Akt) signaling pathway. This pathway included collagen type I α 1 chain (COL1A1), COL1A2, collagen type XI α 1 chain (COL11A1) and thrombospondin (THBS)1. The DEGs were also associated with the focal adhesion signaling
pathway, including \textit{COL1A1}, \textit{COL1A2}, \textit{COL11A1} and \textit{THBS1}, the extracellular matrix (ECM)-receptor interaction signaling pathway, including \textit{COL1A1}, \textit{COL1A2}, \textit{COL11A1} and \textit{THBS1}, and the cell adhesion signaling pathway, including activated leukocyte cell adhesion molecule and \textit{CD2}. DEGs enriched in the ECM-receptor interaction signaling pathway were all upregulated, including certain collagen genes and \textit{THBS} (Fig. 3).

PPI network of DEGs in metastatic EOS. Using the STRING database, a PPI network consisting of 493 interactions of 146 DEGs was constructed (Fig. 4). The majority of the DEGs were upregulated, with the exception of 18 downregulated DEGs. A total of 14 hub genes were identified, including \textit{COL1A1} (degree=37), matrix metallopeptidase (\textit{MMP})2 (degree=36), decorin (degree=35), \textit{COL3A1} (degree=29), \textit{COL1A2} (degree=29), \textit{MMP14} (degree=26), \textit{COL5A1} (degree=26), secreted protein acidic and cysteine rich (degree=25), \textit{COL4A1} (degree=25), \textit{THBS1} (degree=24), fibronectin 1 (degree=24), \textit{THBS2} (degree=22), tissue inhibitor of metalloproteinase (\textit{TIMP})3 (degree=21) and fibrillin 1 (degree=20).

Integrated miRNA-target gene regulatory network. The present study focused on the 14 hub genes, and assessed their miRNA-target associations further. The miRNA-target regulatory network was based on interactions in the aforementioned validated databases. \textit{THBS1} and \textit{TIMP3} were the dominant targets identified and interacted with multiple miRNAs (Fig. 5). \textit{THBS1} was predicted to be the target of the following eight miRNAs: hsa-miR-98-5p, hsa-let-7d-5p, hsa-miR-155-5p, hsa-let-7b-5p, hsa-miR-132-3p, hsa-miR-30a-3p, hsa-miR-30a-5p and hsa-miR-1. \textit{TIMP3} was predicted to be the target of seven miRNAs as follows: hsa-miR-124-3p, hsa-miR-21-5p, hsa-miR-181b-5p, hsa-miR-221-3p, hsa-miR-222-3p, hsa-miR-335-5p and hsa-miR-1.

Discussion

The present study identified several DEGs in metastatic EOS. Of these DEGs, certain collagen (\textit{COL11A1}, \textit{COL1A1} and \textit{COL1A2}) and \textit{THBS} (\textit{THBS1} and \textit{THBS2}) genes were associated with the PI3K/Akt, ECM-receptor interaction and cell adhesion signaling pathways. These DEGs were also hub genes in the PPI network constructed. \textit{THBS1} and \textit{TIMP3} dominated the miRNA-target network and were targeted by hsa-miR-1.

Hepatocyte growth factor (HGF) aids in the regulation of cell growth and motility. A previous study reported that HGF serves a crucial function in tumor metastasis by enhancing cell motility and increasing proteolytic activity in metalloproteinases (15). The PI3K/Akt signaling pathway is a crucial kinase cascade involving HGF-induced metastasis and invasion (16).

Figure 2. Enriched signaling pathways of the differentially expressed genes in metastatic EOC compared with primary EOC. EOC, epithelial ovarian cancer.
In uveal melanoma cells, activating the PI3K/Akt signaling pathway decreases cell adhesion, and thus promotes motility and migration (17). In glioma cells, the PI3K/Akt signaling pathway may regulate tumor cell proliferation and migration (18). Expression of collagen genes is often regulated via the PI3K/Akt signaling pathway. In hepatic stellate cells, collagen genes may be regulated by fascin, a component of actin bundles, through the focal adhesion kinase/PI3K/Akt signaling pathway (19). In normal human dermal fibroblasts, the transcription of collagen genes may be stimulated by interleukin-13 via the PI3K/Akt signaling pathway (20). In the present study, certain collagen genes, including COL11A1, COL1A1 and COL1A2, were identified as DEGs in metastatic EOC and were enriched in the PI3K/Akt signaling pathway, suggesting that these collagen genes may also serve functions in EOC through this signaling pathway, particularly in metastasis.

ECM proteolysis allows cancer cells to invade and is thus associated with migration in multiple types of cancer (21). A previous study demonstrated that THBS1 is an adhesive glycoprotein that regulates cell-cell and cell-ECM interactions. A previous study demonstrated that THBS1 expression is associated with, and may function as a biomarker for the prognosis of, ovarian cancer (25). Another study demonstrated that downregulating THBS1 in ovarian cancer promotes tumor migration (26). According to comparative proteomic analysis, THBS1 is associated with cell adhesion, and differentially expressed between low malignant potential and highly proliferative EOC cell lines (27). THBS2 serves a function in cell-ECM adhesion (28). Furthermore,
THBS2 is one of ten signature genes associated with cell adhesion, and is associated with metastasis and poor overall survival time in patients with serous ovarian cancer (29). Downregulated by the inhibition of the Hedgehog signaling pathway, THBS1 is associated with ECM-ovarian cancer cell receptor interaction (30). The enrichment analysis performed in the present study demonstrated that THBS1 and THBS2 are associated with the cell adhesion and ECM-receptor interaction signaling pathways, suggesting they may serve key functions in EOC metastasis via regulating these two pathways.

TIMP3 inhibits MMPs, which are associated with ECM degradation. In osteosarcoma, lack of TIMP3 expression increases tumor cell proliferation and promotes migration (31). Arpino et al (32) demonstrated that TIMP3 serves a key function in the regulation of uterine ECM degradation during embryo implantation. Furthermore, TIMP3 was a key DEG identified in metastatic EOC in the present study. Although TIMP3 was not enriched in ECM-associated signaling pathways in the present study, TIMP3 was associated with THBS1 in the PPI network, suggesting that TIMP3 may serve a function in the ECM-receptor interaction signaling pathway during EOC metastasis.

Figure 4. Protein-protein interaction network of the differentially expressed genes. Red nodes, upregulated genes; green nodes, downregulated genes.
Since hsa-miR-1 may decrease tumor cell proliferation in numerous types of cancer and is therefore considered a tumor suppressor. However, a previous study demonstrated that the upregulation of hsa-miR-1 was associated with increased tumor cell growth in relapsed ovarian tumors compared with ovarian primary tumors (33). In cardiac tissues, hsa-miR-1 may target TIMP3 (34) and is predicted to target THBS1 in heart failure (35). However, targeting of TIMP3 and THBS1 by hsa-miR-1 has not yet been reported in EOC. In the present study, THBS1 and TIMP3 were predicted as targets of hsa-miR-1, suggesting that hsa-miR-1 may target the two genes during EOC metastasis.

In conclusion, multiple DEGs and miRNAs were identified as potential biomarkers for the prognosis of EOC metastasis. These DEGs were associated with the PI3K/Akt, ECM-receptor interaction and cell adhesion signaling pathways. In addition, THBS1 and TIMP3 were predicted to be targets of hsa-miR-1. However, these predictive results require validation by further study.

Acknowledgements

The present study was supported by the Surface Project of Shandong Provincial Foundation (grant no. ZR2013HM097).

References

et al: Heart structure-specific transcriptomic atlas


